Epithelium-generated neuropeptide Y induces smooth muscle contraction to promote airway hyperresponsiveness

Shanru Li,1,2,3 Cynthia Koziol-White,1,4 Joseph Jude,1,4 Meiqi Jiang,1,4 Hengjiang Zhao,1,4 Gaoyuan Cao,1,4 Edwin Yoo,1,4 William Jester,1,4 Michael P. Morley,1,2,3 Su Zhou,1,2,3 Yi Wang,1,2,3 Min Min Lu,1,2,3 Reynold A. Panettieri Jr.,1,4 and Edward E. Morrisey1,2,3,5,6

Asthma is one of the most common chronic diseases globally and can be divided into presenting with or without an immune response. Current therapies have little effect on nonimmune disease, and the mechanisms that drive this type of asthma are poorly understood. Here, we have shown that loss of the transcription factors forkhead box P1 (Foxp1) and Foxp4, which are critical for lung epithelial development, in the adult airway epithelium evokes a non-Th2 asthma phenotype that is characterized by airway hyperresponsiveness (AHR) without eosinophilic inflammation. Transcriptome analysis revealed that loss of Foxp1 and Foxp4 expression induces ectopic expression of neuropeptide Y (Npy), which has been reported to be present in the airways of asthma patients, but whose importance in disease pathogenesis remains unclear. Treatment of human lung airway explants with recombinant NPY increased airway contractility. Conversely, loss of Npy in Foxp1- and Foxp4-mutant airway epithelium rescued the AHR phenotype. We determined that NPY promotes AHR through the induction of Rho kinase activity and phosphorylation of myosin light chain, which induces airway smooth muscle contraction. Together, these studies highlight the importance of paracrine signals from the airway epithelium to the underlying smooth muscle to induce AHR and suggest that therapies targeting epithelial induction of this phenotype may prove useful in treatment of noneosinophilic asthma.

Introduction
The contribution of the immune system and airway smooth muscle to asthma phenotype, including airway hyperresponsiveness (AHR), has been well studied, and most therapies are directed toward inhibiting the associated inflammatory response in this disease. Asthma can be divided into either disease involving a Th2 inflammatory/eosinophilic immune response or disease lacking eosinophilic involvement (1, 2). Current asthma therapies are not curative and have little effect on asthma lacking immune involvement. The role of epithelium in mediating the asthma diathesis has been studied, but the precise molecular mechanisms by which airway epithelium mediates AHR remain unknown. Importantly, whether paracrine signals originating in the airway epithelium can promote and exacerbate the asthmatic phenotype remains unclear. In this report, we show that the critical airway epithelial transcription factors forkhead box P1 (FOXP1) and FOXP4 act coordinately to restrict expression of neuropeptide Y (NPY) specifically in airway epithelium. Importantly, loss of FOXP1 and FOXP4 leads to increased NPY expression that can, in turn, induce airway hyperresponsivity in a paracrine manner through activation of smooth muscle myosin light-chain phosphorylation. This phenotype can also be induced in mouse and human lung explants by treatment with recombinant NPY, revealing the importance of epithelial/smooth muscle paracrine signaling in promoting the asthma phenotype in both mouse and human lungs.

Results and Discussion
Loss of Foxp1 and Foxp4 in postnatal airway epithelium increases expression of goblet cell–associated proteins MUC5AC, MUC5B, CLCA3, and TFF2, whose expression is increased in asthmatic airways (3). To assess whether the disruption of airway epithelial homeostasis caused by loss of Foxp1 and Foxp4 evoked AHR, a defining phenotype of asthma, we performed lung resistance and airway compliance testing after exposure to methacholine on adult controls (Scgb1a1-Cre) and Scgb1a1-Cre Foxp47/7 Foxp46/6 (herein referred to as Foxp1/4DKO) mutants. The Scgb1a1 line results in a highly effective deletion of genes, including Foxp1 and Foxp4, in postnatal airway epithelium (3). While methacholine challenge increased airway resistance in a dose-dependent manner in control animals, the Foxp1/4DKO mutants responded with an increase in airway resistance (RL) and tissue damping (G), though there was no difference between control and mutants in Newtonian resistance (RN) (Figure 1, A and B, and Supplemental Figure 1, A–C; supplemental material available online with this article; doi:10.1172/JCI81389DS1). Additionally, at baseline, the Foxp1/4DKO mutants exhibited greater airway resistance, suggesting that an underlying increase in bronchomotor tone exists even

Authorship note: R.A. Panettieri Jr. and E.E. Morrisey contributed equally to this work.
Conflict of interest: The authors have declared that no conflict of interest exists.
Submitted: February 9, 2015; Accepted: March 1, 2016.
in the absence of contractile stimulation of the airways (Figure 1, A and B). To eliminate the effect of baseline differences in lung resistance testing, we compared the airway resistance response to methacholine using precision cut lung slices (PCLSs) from control and Foxp1/4DKO mutants (4–6). These experiments showed that Foxp1/4DKO mutants exhibited greater airway resistance both in vivo and in the PCLS system (Figure 1C). The AHR response in unchallenged Foxp1/4DKO mutants was similar to that of control mice challenged with Aspergillus fumigates (Supplemental Figure 1D). Moreover, the AHR response in Foxp1/4DKO mutants was further amplified after exposure to A. fumigatus challenge (Supplemental Figure 1E). Our findings are consistent with those seen in asthma patients, who often manifest a diminished forced expiratory volume in 1 second (FEV1) (more obstruction) and greater methacholine sensitivity after challenge (7).

Rodent models of AHR, including those that are sensitive to ovalbumin, house dust mite, and A. fumigatus challenge, have a significant inflammatory component (8, 9). We examined the lungs of Foxp1/4DKO mutants for expression of the eosinophilic inflammatory marker IL-13 and neutrophilic inflammatory markers IL-17, IL-6, and keratinocyte-derived chemokine (KC) to assess whether loss of Foxp1 and Foxp4 evoked an inflammatory response. Surprisingly, Foxp1/4DKO mutants did not exhibit increased levels of any of these inflammatory cytokines compared with control mice (Figure 1D). We did observe a modest increase in the numbers of neutrophils, but not macrophages, eosinophils, or lymphocytes in the bronchoalveolar lavage (BAL) fluid from Foxp1/4DKO mutants (Supplemental Figure 2A).

Transcriptome analysis revealed 109 genes significantly (P < 0.05) up- or downregulated upon genetic inactivation of Foxp1 and Foxp4. We focused our attention on those genes that were upregulated and that could act in a paracrine fashion upon airway smooth muscle cells to confer AHR. Among the 10 upregulated genes coding for secreted proteins, NPY expression was changed by the greatest extent (Figure 1E and Table 1). The increase in Npy expression was confirmed by ISH (Figure 1F) and quantitative real-time PCR (Q-PCR) (Supplemental Figure 2, B and C). NPY is a 36-aa polypeptide and is expressed in the central and peripheral nervous system and in other cell lineages, including endothelium, platelets, immune cells, and enteric neurons in the gut (10–15). Although NPY is one of the most abundant peptides in the peripheral nervous system, the role of NPY in asthma remains unclear (16, 17). Interestingly, increased Npy expression is associated with a mouse model of allergic airway inflammation, and polymorphisms in the NPY gene predict severity of asthma (18). Figure 1. Loss of Foxp1 and Foxp4 in airway epithelial cells promotes AHR and induces ectopic expression of NPY. (A and B) Lung resistance to cumulative doses of methacholine was measured. Deficiency in FOXP1 and FOXP4 in epithelial cells markedly enhances methacholine-induced lung resistance demonstrated in airway resistance (RL) (A) and dynamic elastance (B) (n = 6 for control and DKO, respectively) and in the mouse PCLS assay (C) (control n = 3, 30 slices; DKO n = 8, 19 slices). (D) Loss of Foxp1 and Foxp4 induced AHR without evoking airway eosinophilic inflammation, as demonstrated by no significant increase in IL-13, IL-17 (n = 11 for control, 8 for DKO), IL-6, and KC (n = 3 for WT and 4 for DKO, respectively). Heat map from the microarray analyses showing that Npy expression was increased by in the secreted proteins category in Foxp1/4DKO mutants. (F) ISH showing increased Npy expression in the epithelium of Foxp1/4DKO mutants (arrowheads). Ai, airway. **P < 0.01. Scale bar: 200 μm.
increased levels of Npy observed in the Foxp1/4^DKO mutant airway epithelium generated a paracrine signal to promote airway smooth muscle contraction rather than increased mass.

Two FOXP1 and FOXP4 DNA–binding sites are located in an evolutionarily conserved region approximately –0.8 kb to the transcriptional start of the mouse Npy gene (Supplemental Figure 2I). ChIP assays showed that FOXP1 and FOXP4 bind to these sites in chromatin from mouse lungs (Supplemental Figure 2J). Luciferase reporter constructs containing this ~0.8 kb region were repressed by FOXP1, demonstrating that Npy is a transcriptional target of FOXP1 and FOXP4 (Supplemental Figure 2K).

To determine whether NPY potentiates airway smooth muscle contraction, we incubated mouse and human PCLSs (4–6) with recombinant NPY and characterized methacholine-mediated bronchoconstriction. These experiments demonstrated that NPY amplifies methacholine-induced bronchoconstriction (Figure 2, A and B, and Supplemental Figure 3).

Phosphorylation of myosin light chain (pMLC) is a critical signaling event required for airway smooth muscle contraction (20, 21). We next examined the level of pMLC in primary human airway smooth muscle (HASM) cells. While methacholine increased pMLC as expected, Npy treatment in addition to methacholine significantly increased pMLC levels (Figure 2C). Activation of Rho kinase mediated sensitization of bronchoconstriction to contractile agonists in HASM. Brady-

Table 1. Cell surface genes upregulated in Foxp1/4^DKO mutant lungs

<table>
<thead>
<tr>
<th>Gene</th>
<th>Fold change</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sgpi5b</td>
<td>13.3</td>
</tr>
<tr>
<td>Cpn1</td>
<td>4.8</td>
</tr>
<tr>
<td>Npy</td>
<td>30.2</td>
</tr>
<tr>
<td>Celo2a</td>
<td>8.3</td>
</tr>
<tr>
<td>Egf1</td>
<td>3.5</td>
</tr>
<tr>
<td>Lgr7</td>
<td>1.8</td>
</tr>
<tr>
<td>Gch1</td>
<td>1.7</td>
</tr>
<tr>
<td>Retnla</td>
<td>2.2</td>
</tr>
<tr>
<td>Fide7</td>
<td>4.2</td>
</tr>
<tr>
<td>Gpx3</td>
<td>2</td>
</tr>
<tr>
<td>Sgfa1A1</td>
<td>–2.8</td>
</tr>
<tr>
<td>Itih2</td>
<td>–1.7</td>
</tr>
<tr>
<td>Ccl20</td>
<td>–9.5</td>
</tr>
<tr>
<td>Csn2</td>
<td>–3.3</td>
</tr>
</tbody>
</table>

Npy gene are associated with an increased risk for asthma (18, 19). Assessment of changes in the amount of airway smooth muscle in Foxp1/4^DKO mutants did not reveal any changes compared with controls (Supplemental Figure 2, D–H). These data indicate that

Figure 2. NPY amplifies methacholine-induced bronchoconstriction in human airways. (A) Human PCLSs were treated with 10 nM recombinant NPY for 18 hours, and cumulative dose curves to methacholine measuring luminal diameter narrowing were determined. (B) Pretreatment with NPY markedly enhances maximal bronchoconstriction induced by methacholine. (C) NPY treatment alone has little effect on pMLC; however, pretreatment with NPY markedly enhances methacholine-induced pMLC. Graphical data represent experiments performed in triplicate and are expressed as arbitrary densitometry units. Mch, methacholine. (D) NPY had little effect on peak calcium levels induced by contractile agonists BK or histamine, indicating that NPY does not act to induce calcium mobilization through these 2 pathways. (E) NPY promotes Rho kinase activity by itself and in cooperation with BK as measured by Western blot. *P < 0.01; **P < 0.005.
kinin (BK) and histamine are known to increase peak calcium levels and bronchoconstriction in HASM cells (22). Treatment of HASM cells with NPY had little effect on peak cellular calcium levels over that induced by BK or histamine alone (Figure 2D). NPY alone, however, increased Rho kinase activity, and augmented BK induced Rho kinase activity above that induced by BK alone (Figure 2E).

To determine whether the phenotype observed in Foxp1/4DKO mutants is due to increased Npy expression, we genetically deleted Npy in Foxp1/4DKO mutants to determine whether this would attenuate the AHR phenotype. Foxp1/4DKO Npy–/– mutants (herein referred to as Foxp1/4 NpyTKO) were generated by mating Foxp1/4DKO mice with Npy–/– mice, which do not have a significant AHR phenotype (data not shown). Foxp1/4 NpyTKO mutants were grossly normal and had normal breeding habits. ISH and Q-PCR for Npy expression showed loss of Npy expression in Foxp1/4 NpyTKO mutants (Figure 3, A and B). Foxp1/4 NpyTKO mutants were challenged with methacholine, and the enhanced AHR observed in the Foxp1/4DKO mutants was attenuated in the Foxp1/4 NpyTKO mice (Figure 3, C). BAL cell counts from control, Foxp1/4DKO, and Foxp1/4 NpyTKO mutants are significantly different (Figure 3D). Expression of the proinflammatory cytokine IL-6 was significantly lower in Foxp1/4 NpyTKO mutants compared with Foxp1/4DKO mutants, whereas expression of KC was significantly increased (Supplemental Figure 4).

The airway epithelium of Foxp1/4DKO mutants contains large goblet-like cells with reduced expression of markers of differentiated club secretory cells, including Scgb1a1 (ref. 3 and Supplemental Figure 5). Interestingly, we found that expression of Scgb1a1 was partially rescued in the Foxp1/4 NpyTKO mutants (Supplemental Figure 6). In contrast, expression of multiple goblet cell markers that were elevated in the Foxp1/4DKO mutants was not altered by the additional loss of Npy expression (Supplemental Figure 6).

Current therapies for asthma target the inflammatory response or promote relaxation of airway smooth muscle shortening. While these treatments are effective in the short term, the underlying cause of asthma remains elusive, and many patients are insensitive to conventional therapy. Approximately half of asthma patients exhibit eosinophilic inflammation with increased levels of IL-4, IL-5, IL-13, and IgE (23, 24). In contrast, about half of asthma patients lack a strong eosinophilic inflammatory response, but exhibit a variable neutrophilic inflammatory response. Foxp1 and Foxp4 double mutants represent a model for non eosinophilic asthma, which remains poorly understood. In addition, patients with this disorder tend to be the least responsive to current therapies.

Previous studies have reported increased expression of Npy in animal models of allergen-induced AHR (19, 25). Moreover, Npy polymorphisms are associated with increased risk for asthma in overweight subjects (18, 25). Other reports have demonstrated that plasma levels of NPY were increased in asthmatics at rest and in acute exacerbations (26–28). Our study shows that increased expression of Npy in airway epithelium can induce an AHR phenotype in a paracrine manner (Supplemental Figure 6). This phenotype is recapitulated in a human lung explant assay, demonstrating that NPY can act on human small airways in a similar manner. Importantly, our studies are among the first, to our knowledge, to mechanistically link a critical transcription pathway in airway epithelium that activates a paracrine response to the promotion of AHR in the absence of a Th2-induced eosinophilic inflammatory response.

Methods

Animals. Generation and genotyping of Foxp1fl/fl, Foxp4fl/fl, Scgb1a1-Cre, and Npy–/– mouse lines have been previously reported (3, 29). The mouse lines were kept on a mixed background consisting of C57BL/6 and 129SvJ. Control animals were derived from the same background and, when possible, the same litters as the mutant animals.

Microarray studies. All original microarray data were deposited in the NCBI’s Gene Expression Omnibus (GEO GSE64774).
**Statistics.** We used the R package lme4 (1, 30) to perform a linear mixed-effects analysis of the relationship between lung resistance and genotype. As fixed effects, we entered dose and genotype with an interaction term into the model. As random effects, we used a random intercept model for the subjects. P values were obtained by comparing the full model and reduced model without the genotype fixed effect using an F test with Kenward-Roger approximation (31). Other data are reported as mean ± SEM of at least 3 independent assays unless otherwise noted. Unpaired Student’s t test (2-tailed) was used for single comparisons and 1-way ANOVA for multiple comparisons. A P value of less than 0.05 was considered significant.

**Study approval.** All animal procedures were approved by the Institute for Animal Care and Use Committee at the University of Pennsylvania.

See Supplemental Methods for additional information regarding these studies.

**Author contributions**

SL, RAP, and EEM designed and developed the assays and experiments. SL, CKW, JJ, MJ, HZ, GC, EY, WJ, SZ, YY, and MML performed experiments. MPM performed bioinformatics and statistical analysis. SL, RAP, and EEM wrote the manuscript.

**Acknowledgments**

These studies were supported by funding from the NIH to E.E. Morrisey (HL071589 and HL110942) and R.A. Panettieri Jr. (HL110942).

Address correspondence to: Edward E. Morrisey, University of Pennsylvania, Translational Research Center, Room 11-124, 3400 Civic Center Boulevard, Building 421, Philadelphia, Pennsylvania 19104-5129, USA. Phone: 215.573.3010; E-mail: emorisse@mail.med.upenn.edu.

---


14. Dimitrijevic M, Stanojevic S. The intriguing misfolding of the oxytocin receptor in airway smooth muscle: regulation by TNF


