

Supplemental Materials and Methods and Supplemental Figures

STIM1 and STIM2 control antiviral immunity by CD4 and CD8 T cells

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Supplemental Materials and Methods

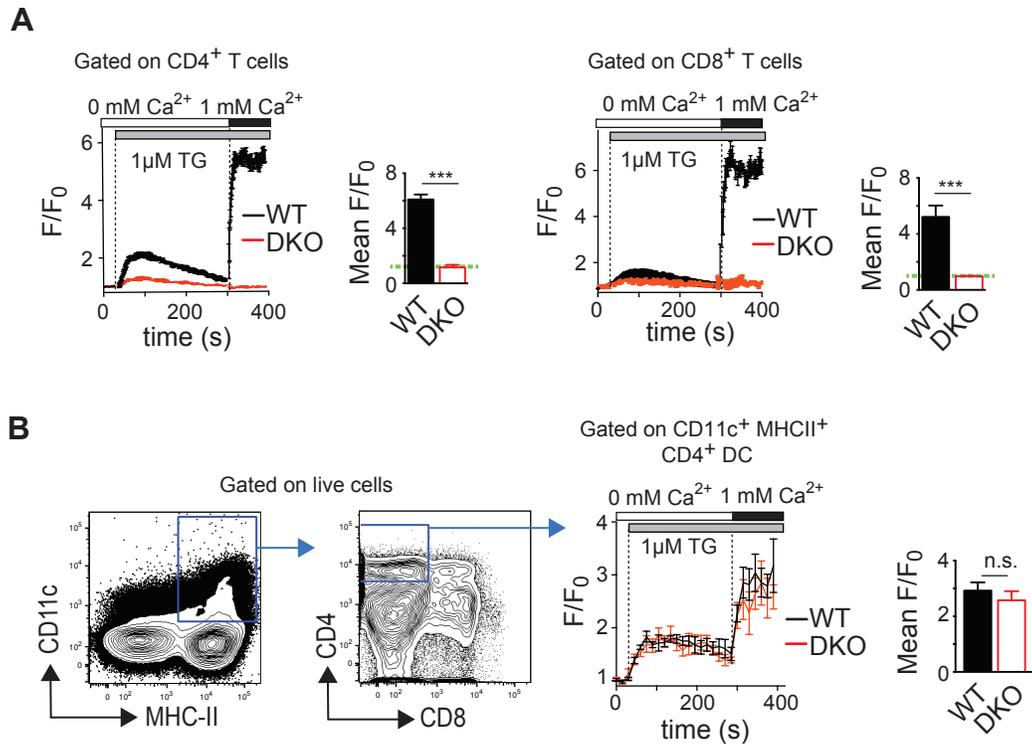
Intracellular Ca²⁺ measurements

Splenocytes were isolated from uninfected *Stim1^{fl/fl} Stim2^{fl/fl} Cd4-Cre* (DKO) or WT littermate mice and labelled with 2 μ M Fluo-4 AM (Invitrogen). Cells were incubated with antibodies against CD4 (GK1.5), CD8 (53-6.7), CD11c (N418) and MHC-II (M5/114.15.2, all from eBioscience) for 30 min at 22–25°C, washed and kept in RPMI 1640 medium until use. Measurements of intracellular [Ca²⁺]_i were conducted using a LSR II flow cytometer (BD Biosciences). Baseline Ca²⁺ levels (F₀) were acquired in nominally Ca²⁺-free Ringer solution containing (in mM) 155 NaCl, 4.5 KCl, 3 MgCl₂, 10 D-glucose, 5 Na-HEPES. After 30 s, cells were stimulated with 1 μ M of the Sarco/endoplasmic reticulum ATPase (SERCA) inhibitor thapsigargin (EMD). After 300 s, Ringer solution containing 2 mM CaCl₂ was added to the cells (final extracellular Ca²⁺ concentration 1 mM). Data were analyzed using FlowJo software (Tree Star) by gating on CD4⁺ and CD8⁺ T cell populations or CD11c⁺ MHC-II⁺ CD4⁺ dendritic cells. Intracellular Ca²⁺ levels were calculated as the F/F₀ ratio (F=Fluorescence intensity; F₀=Fluorescence intensity before addition of TG) over time in seconds (s). SOCE was averaged as the mean \pm SEM of the F/F₀ ratio at 30 s after readdition of Ca²⁺ from 3 independent experiments performed in duplicates; data were plotted using Prism6 software (Graphpad).

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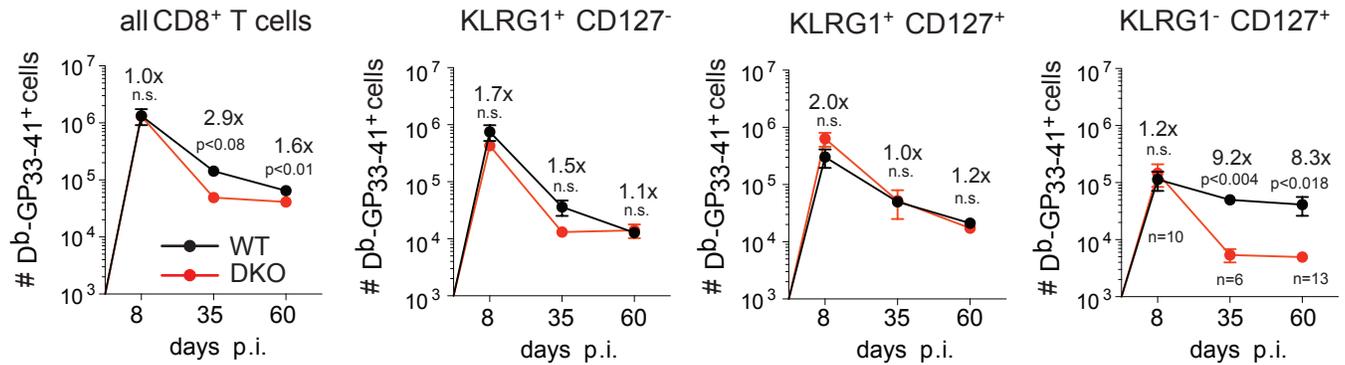
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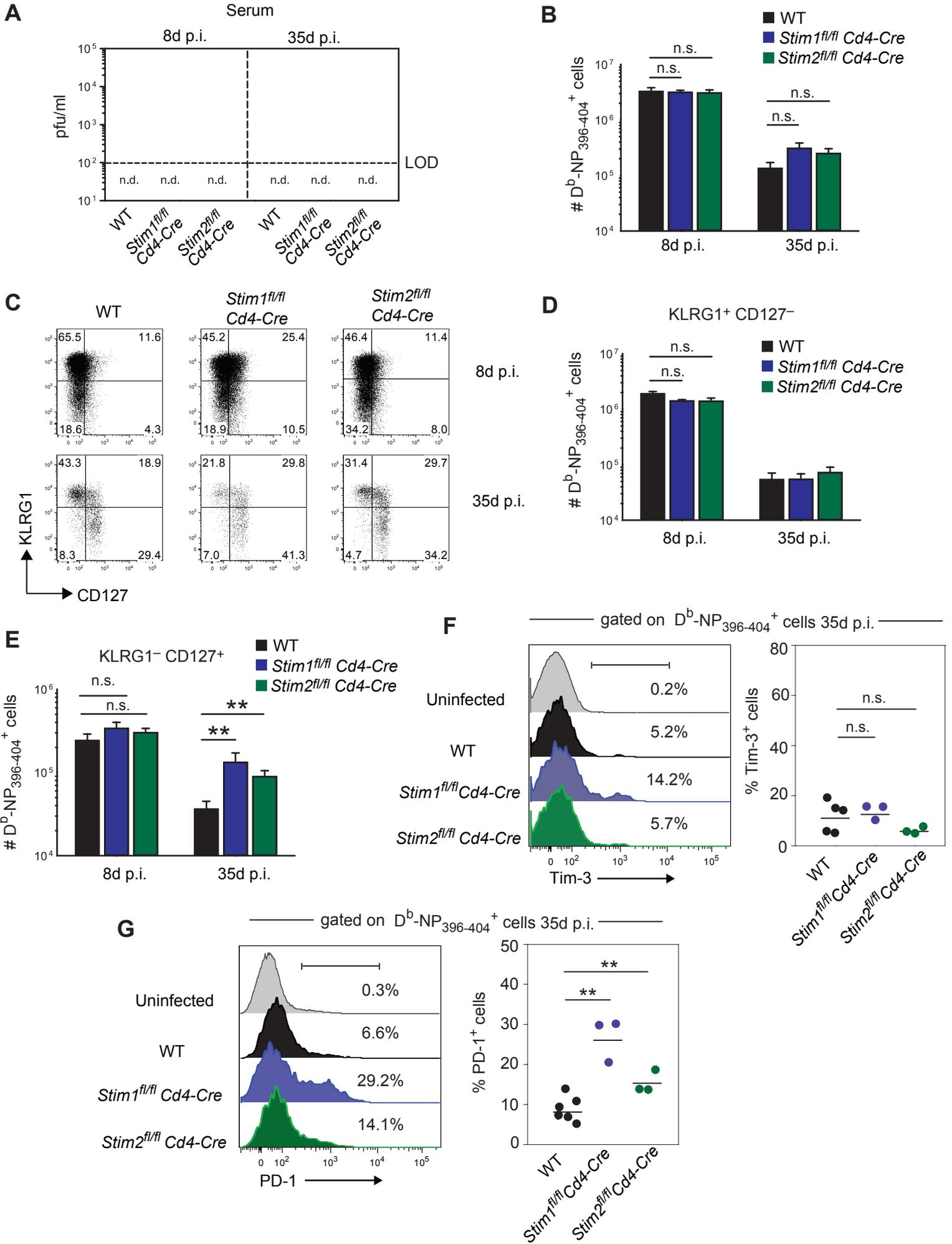
Supplemental Figure 1. Store-operated Ca²⁺ entry (SOCE) is impaired in T cells but not CD4⁺ dendritic cells (DC) from *Stim1^{fl/fl} Stim2^{fl/fl} Cd4-Cre* mice.

(A) SOCE in CD4⁺ and CD8⁺ T cells of DKO (*Stim1^{fl/fl} Stim2^{fl/fl} Cd4-Cre*) or WT (*Stim1^{fl/fl} Stim2^{fl/fl}* Cre-negative littermate). Splenocytes were isolated from uninfected WT or DKO mice, labeled with the Ca²⁺ indicator dye Fluo-4 and stained with antibodies against CD4, CD8, CD11c and MHC-II. Endoplasmic reticulum (ER) Ca²⁺ stores were depleted with 1 μM thapsigargin (TG) in Ca²⁺-free Ringer solution (0 mM Ca²⁺) and SOCE induced by readdition of 2 mM Ca²⁺. SOCE in CD4⁺ and CD8⁺ cells was calculated as the F/F₀ ratio (F=Fluorescence intensity; F₀=Fluorescence intensity before addition of TG) over time in seconds (s). Dotted green line in bar graphs represents F₀. ***, p<0.001, n.s.= non-significant. (B) SOCE in CD11c⁺, MHC-II⁺, CD4⁺ DC from WT and DKO mice stimulated as in (A). Shown are representative graphs and mean ± SEM of Ca²⁺ influx levels after the readdition of Ca²⁺ (at 300 s) from 3 independent experiments performed in duplicates.

Gated on D^b-GP₃₃₋₄₁⁺ CD8⁺ T cells

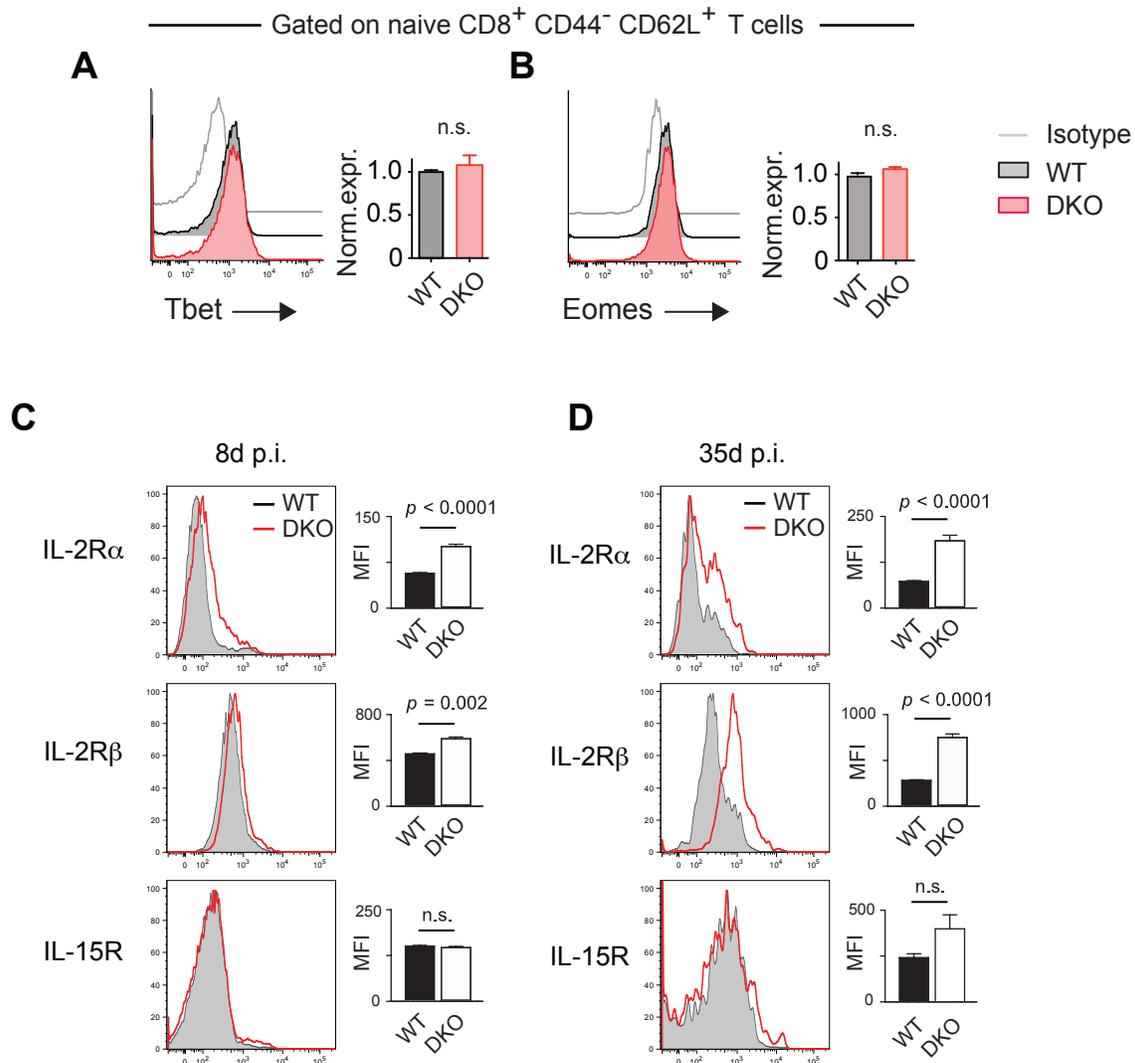
Supplemental Figure 2. Total number of LCMV-specific (D^bGP₃₃₋₄₁ tetramer⁺) CD8⁺ T cells in WT and DKO mice.

WT and DKO mice were infected with 2×10^5 pfu LCMV^{ARM} (i.p.) and splenocytes were isolated at days 8, 35 and 60 p.i. Cells were stained with antibodies against CD8, KLRG1, CD127 and D^bGP₃₃₋₄₁ MHC class I tetramer. The total number of all LCMV-specific CD8⁺ T cells and those of effector (KLRG1⁺ CD127⁻) and memory (KLRG1⁺ CD127⁺ and KLRG1⁻ CD127⁺) subsets were analyzed in 6-13 mice per group. Numbers represent fold differences in LCMV-specific CD8⁺ T cell numbers between WT and DKO mice. Statistical significance was calculated using Student's t-test. The analysis of D^bGP₃₃₋₄₁-specific CD8⁺ T cells complements data shown in **Figure 1C-G**.



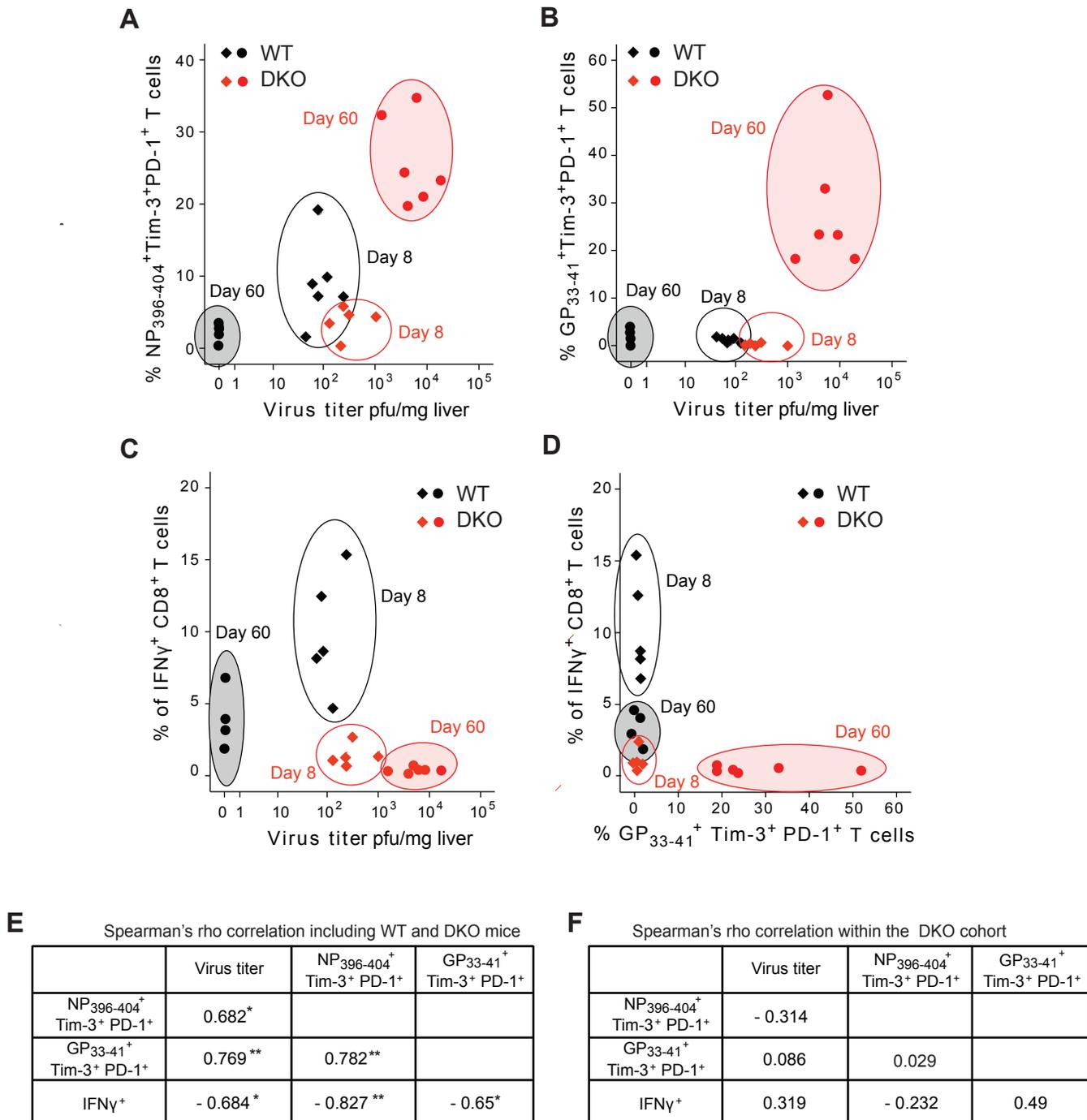
Supplemental Figure 3. Mice with T cell specific deletion of *Stim1* or *Stim2* genes alone mount and maintain normal antiviral immunity.

Stim1^{fl/fl} Cd4-Cre, *Stim2^{fl/fl} Cd4-Cre* and wild-type (WT, i.e. Cre-negative littermates) mice were infected with 2×10^5 pfu LCMV^{ARM} (i.p.). **(A)** Undetectable viral titers in the serum at days 8 and 35 p.i. (n.d., not detectable; LOD, level of detection). **(B)** Comparable numbers of D^bNP₃₉₆₋₄₀₄⁺ CD8⁺ T cells in the spleen of WT, STIM1- and STIM2-deficient mice at days 8 and 35 p.i. **(C)** Expression of KLRG1 and CD127 (IL-7R) on D^bNP₃₉₆₋₄₀₄⁺ CD8⁺ T cells at days 8 and 35 p.i. Numbers represent percentages of cells in gates. **(D,E)** Comparable numbers of D^bNP₃₉₆₋₄₀₄⁺ terminal effector (KLRG1⁺ CD127⁻ in D) and memory (KLRG1⁻ CD127⁺ in E) CD8⁺ T cells in the spleen at days 8 and 35 p.i. (mean \pm SEM). **(F,G)** Frequencies of Tim-3⁺ **(F)** and PD-1⁺ **(G)** D^bNP₃₉₆₋₄₀₄⁺ CD8⁺ T cells were determined in the spleens of WT, STIM1 or STIM2-deficient mice at day 35 p.i. 3-5 mice per group and time point were used. Horizontal lines in **F,G** represent mean percentages. n.s. = non-significant; ** p<0.01.



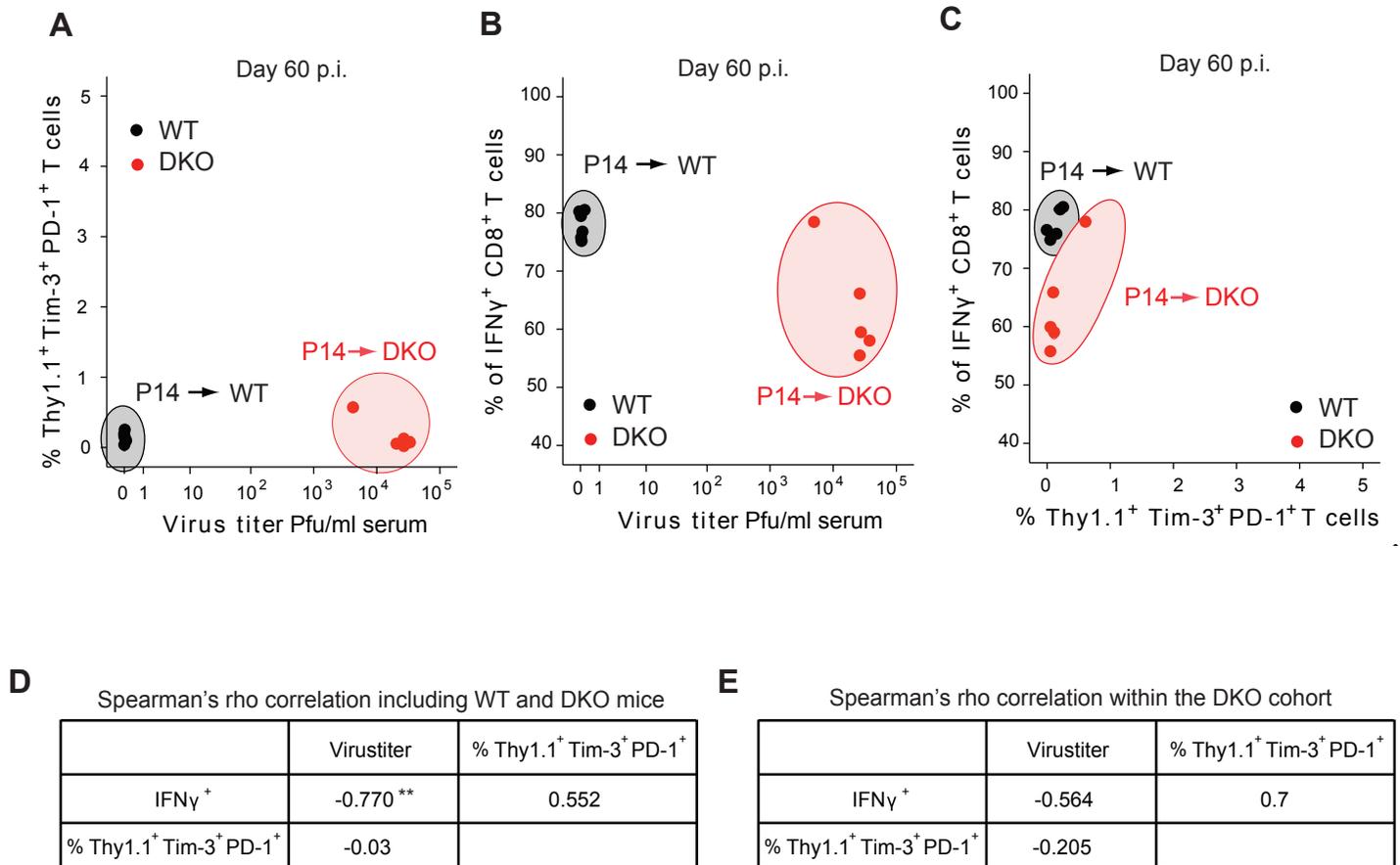
Supplemental Figure 4. Expression levels of T-bet, Eomes and cytokine receptors in STIM1/STIM2-deficient CD8⁺ T cells.

(A-B) Splenocytes from uninfected *Stim1^{fl/fl}Stim2^{fl/fl}Cd4-Cre* (DKO) and wild-type (WT) mice were isolated and the expression of T-bet (A) and Eomes (B) was determined in CD8⁺ CD44⁻ CD62L⁺ T cells by flow cytometry. Representative histogram plots (left) and bar graphs (right) showing the means \pm SEM of relative T-bet and Eomes expression from 3 independent experiments (one WT and DKO mouse per experiment performed in duplicates). To correct for variation in T-bet and Eomes levels between experiments, expression in DKO cells was normalized to that in WT cells for each experiment. (C-D) WT and DKO mice were infected with 2×10^5 pfu of LCMV^{ARM} (i.p.). LCMV-specific CD8⁺ T cells were isolated from the spleen and analyzed 8 (C) and 35 (D) days p.i. for the expression of cytokine receptors required for memory CD8⁺ T cell survival. Representative histogram plots (left) and bar graphs showing the means \pm SEM of IL-2R α , IL-2R β and IL-15R expression from 6 mice per group.



Supplemental Figure 5. Correlation analysis of viral titers and CD8⁺ T cell exhaustion.

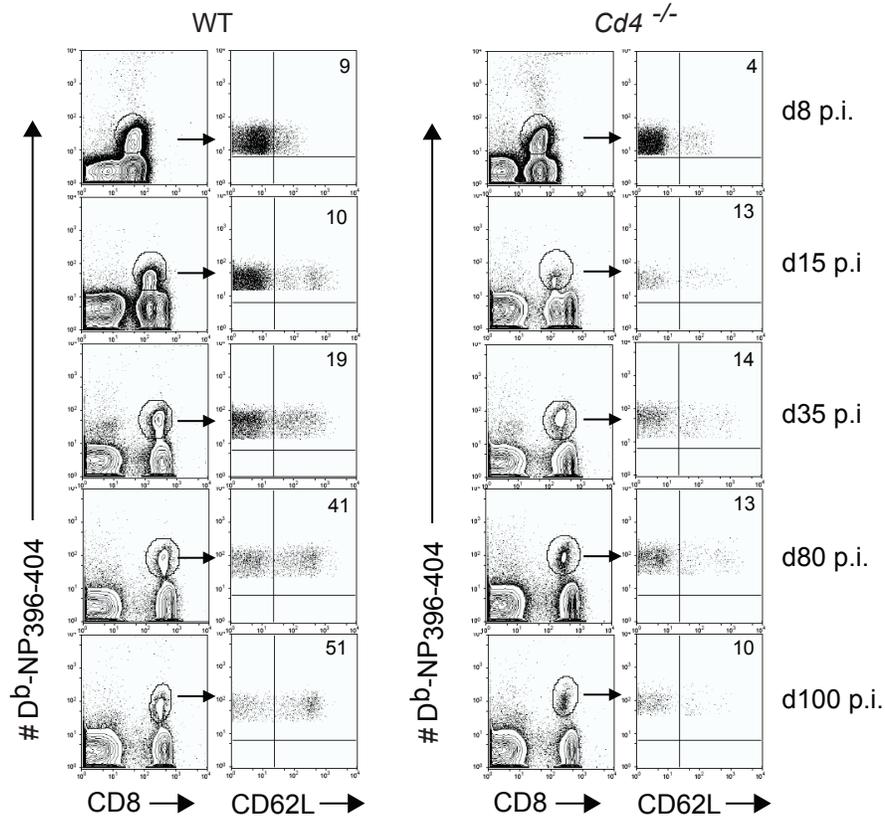
(A-D) WT (n=4-6) and *Stim1^{fl/fl} Stim2^{fl/fl} Cd4-Cre* (DKO) (n=5-6) mice were infected with LCMV^{ARM} and analyzed at days 8 and 60 p.i. for viral titers in the liver as well as expression of exhaustion markers (PD-1, Tim-3) and IFN γ production by GP₃₃₋₄₁ or NP₃₉₆₋₄₀₄ specific CD8⁺ T cells. The frequency of IFN γ producing CD8⁺ T cells was measured after restimulation with GP₃₃₋₄₁ peptide for 6h in vitro. Each symbol represents one mouse. (E,F) Correlation coefficients were calculated for samples 60 days p.i. using Spearman's Rho test. (* p<0.05; ** p<0.01). (E) Both WT and DKO cohorts were included in the analysis. (F) Only DKO samples were included in the analysis. Correlation analyses were performed using primary data shown in **Figures 1, 2 and 3**.



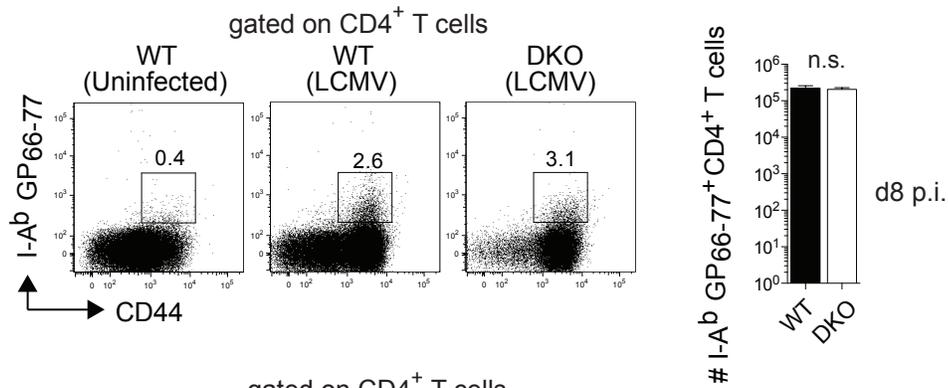
Supplemental Figure 6. Correlation analysis of viral titers and CD8⁺ T cell exhaustion in mice after transfer of WT P14 cells.

(A-C) WT (n=6) and *Stim1^{fl/fl} Stim2^{fl/fl} Cd4-Cre* (DKO, n=5) mice were infected with LCMV^{ARM} and at the same time injected (i.v.) with WT CD8⁺ P14 cells that express a GP₃₃₋₄₁ specific TCR. Mice were sacrificed 60 days p.i. and analyzed for viral titers in the serum as well as expression of exhaustion markers (PD-1, Tim-3) and IFN γ production by Thy1.1⁺ P14 cells after restimulation with GP₃₃₋₄₁ peptide for 6h in vitro. Each symbol represents one mouse. (D-E) Correlation coefficients were calculated for samples 60 days p.i. using Spearman's Rho test. * p<0.05; ** p<0.01. (D) Analysis including WT and DKO cohorts. (E) Analysis including only DKO samples. Correlation analyses were performed using primary data shown in **Figure 4E-L**.

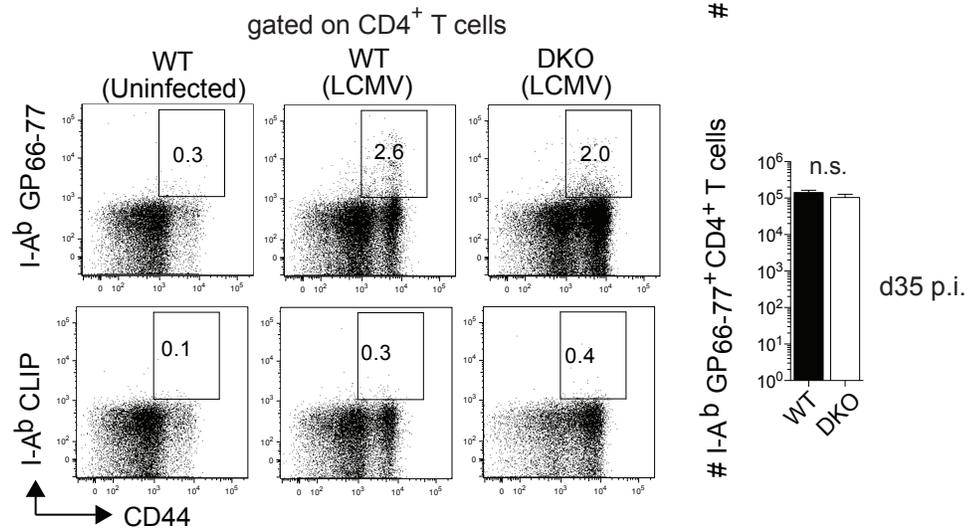
A



B

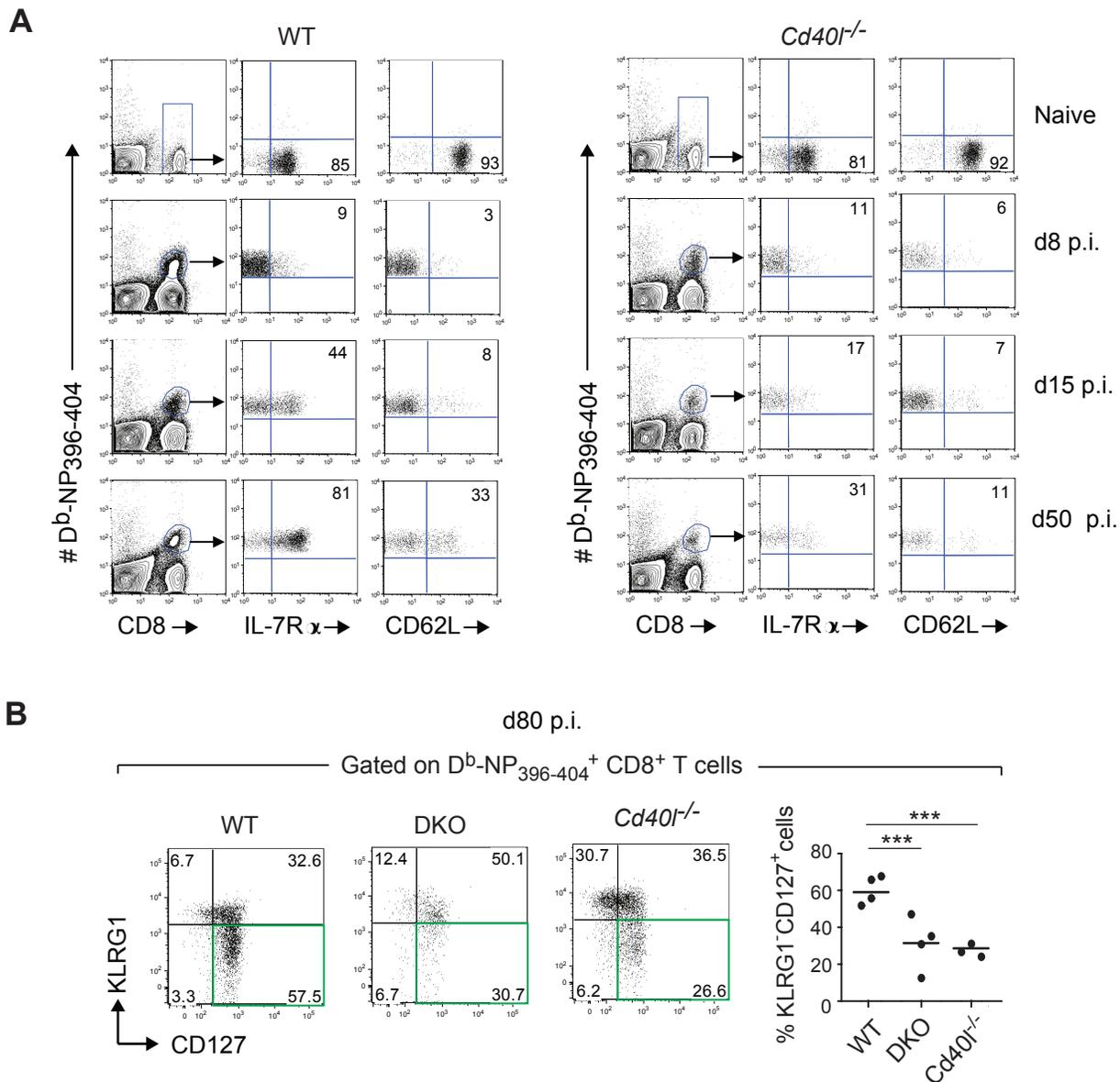


C



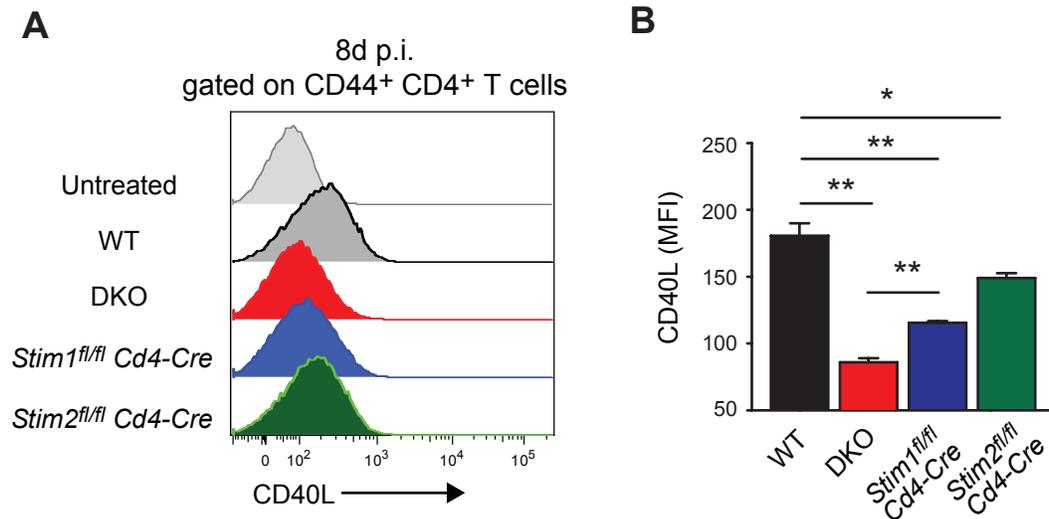
Supplemental Figure 7. CD4⁺ T cells are required to maintain memory CD8⁺ T cells but the development of LCMV-specific CD4⁺ T cells is independent of STIM1 and STIM2.

(A) Impaired maintenance of LCMV-specific memory CD8⁺ T cells in the absence of CD4⁺ T cell help. WT and *Cd4*^{-/-} mice were infected with 2 x 10⁵ pfu of LCMV^{ARM} (i.p.) and analyzed at the indicated time points post-infection (p.i.) for the frequencies of D^bNP₃₉₆₋₄₀₄ tetramer⁺ CD62L⁺ memory CD8⁺ T cells. (B,C) WT and DKO mice were infected with 2 x 10⁵ pfu of LCMV^{ARM} and splenocytes were stained with I-A^b GP₆₆₋₇₇ MHC class II tetramers and analyzed for the total number of LCMV specific CD4⁺ T cells 8 days (B) and 35 days p.i. (C). Representative dot plots and the mean ± SEM absolute number of LCMV-specific CD4⁺ T cells from 4 WT and 4 DKO mice per time point are shown. Controls included CD4⁺ T cells from non-infected mice and CD4⁺ T cells stained with APC-conjugated I-A^b tetramers loaded with CLIP peptide. Statistical significance was calculated by Student's t-test. n.s., non-significant. Numbers in FACS plots in A-C represent percentages of cells in gates.



Supplemental Figure 8. Impaired maintenance of virus-specific memory CD8⁺ T cells in *Cd40l*^{-/-} and DKO mice.

(A) WT and *Cd40l*^{-/-} mice were infected with LCMV and analyzed at the indicated days post-infection (p.i.) for the frequencies of LCMV-specific (D^bNP₃₉₆₋₄₀₄ tetramer⁺) memory cells expressing CD62L and IL-7R α (CD127). (B) WT, *Stim1*^{fl/fl}*Stim2*^{fl/fl}*Cd4-Cre* (DKO) and *Cd40l*^{-/-} mice were infected i.p. with 2 x 10⁵ pfu of LCMV^{ARM} and analyzed for memory CD8⁺ T cell and antibody responses 80 days p.i. Plots show the expression of KLRG1 and CD127 on D^bNP₃₉₆₋₄₀₄ tetramer⁺ CD8⁺ T cells. Green boxes emphasize KLRG1⁻ CD127⁺ memory CD8⁺ T cells. The scatter plot (right panel) shows frequencies of KLRG1⁻ CD127⁺ memory CD8⁺ T cells from 3-4 mice per group. Horizontal lines represent mean percentages. Statistical significance was calculated using Student's t-test. ***, p<0.001. Numbers in FACS plots in A,B represent percentages of cells in gates.



Supplemental Figure 9. Partial decrease of CD40L expression in CD4⁺ T cells lacking either STIM1 or STIM2.

WT, *Stim1^{fl/fl} Cd4-Cre*, *Stim2^{fl/fl} Cd4-Cre* and *Stim1^{fl/fl} Stim2^{fl/fl} Cd4-Cre* (DKO) mice were infected with LCMV^{ARM} and sacrificed 8 days p.i. Total CD40L expression was measured by flow cytometry after restimulation of splenocytes with LCMV GP₆₁₋₈₀ peptide in the presence of anti-CD40L antibody LCMV for 2 h. Representative histogram plots (**A**) and mean \pm SEM of the mean fluorescence intensity (MFI) of CD40L expression on CD4⁺ CD44⁺ T cells from 4 mice per group (**B**). Statistical significance was calculated using Student's t-test. * $p < 0.05$; ** $p < 0.01$.