

Figure S1: *MHC-II^{-/-} mice have normal B cell and CD8⁺ T cell populations, but lack CD4⁺ T cells.* Flow cytometry of the deep cervical lymph nodes of wild type (A) and MHCII^{-/-} (B) mice showing CD4⁺ and CD8⁺ lymphocytes in the TCRβ⁺ population and CD19⁺B220⁺ B cells in the CD45⁺TCRβ⁻ population. Numbers indicate percent CD4⁺ (upper left) and CD8⁺ (lower right), as a percentage of TCRβ⁺ cells and CD19⁺B220⁺ B cells as a percentage of the CD45⁺TCRβ⁻ population.

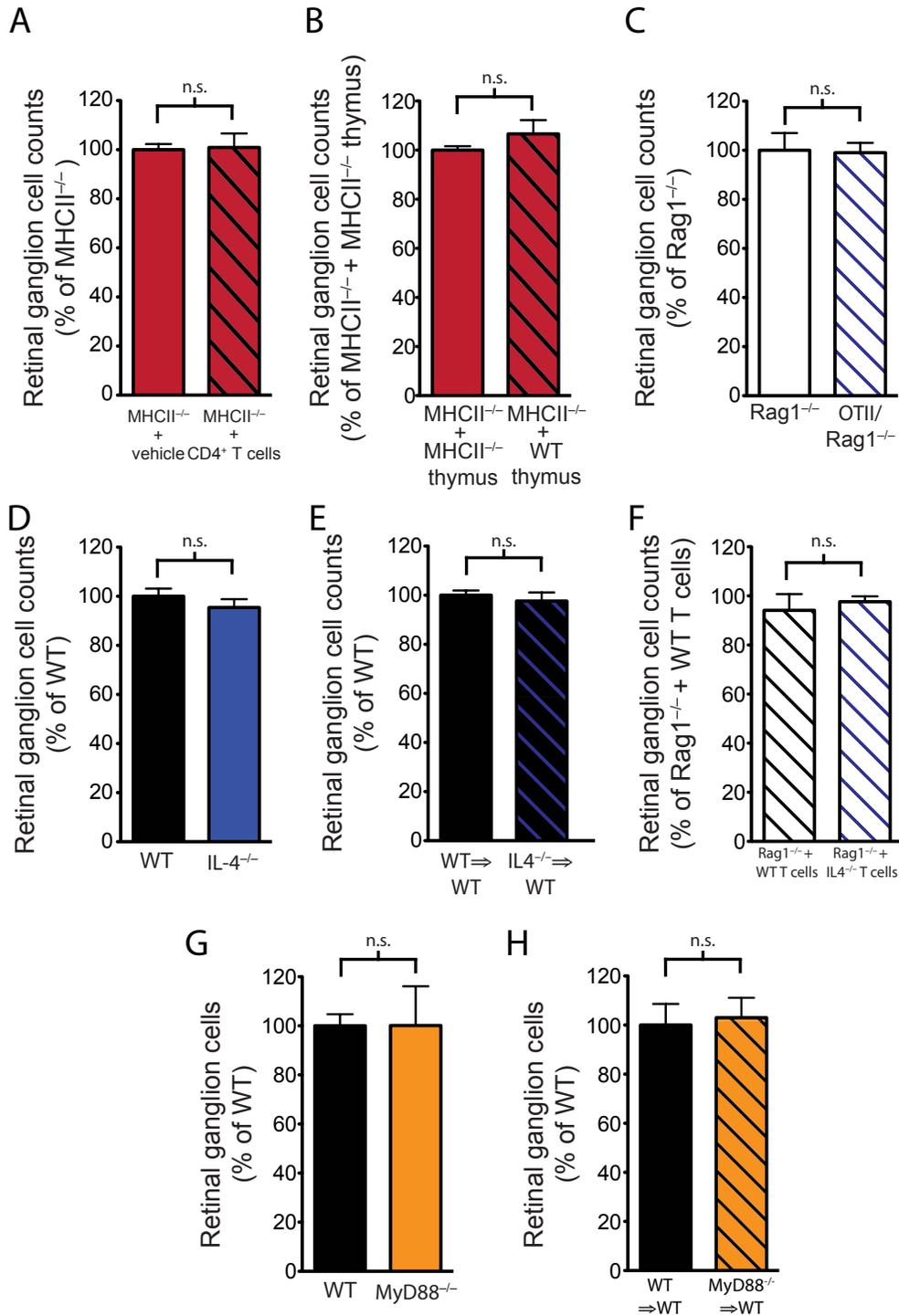


Figure S2: No difference in contralateral, uninjured retinas of all mouse strains and experimental manipulations examined in this manuscript. (A) MHCII^{-/-} mice injected with 3×10^6 wild type CD4⁺ T cells on the day of injury exhibit no difference in neuronal cell counts in the uninjured retinas compared to those injected with saline. Retinal ganglion cell counts (% of MHCII^{-/-}) of MHCII^{-/-} mice and MHCII^{-/-} mice injected with wild type

CD4⁺ T cells as assessed by Fluoro-Gold staining (n = 3, wild type thymus; n = 3 MHCII^{-/-} thymus; Student's t-test). **(B)** MHCII^{-/-} mice implanted with wild type thymi exhibit no difference in neuronal counts in the uninjured retinas compared to those implanted with MHCII^{-/-} thymi. Retinal ganglion cell counts (% of MHCII^{-/-} + MHCII^{-/-} thymus) of MHCII^{-/-} mice implanted with either a MHCII^{-/-} or wild type thymi (six weeks after implantation) as assessed by Fluoro-Gold staining (n = 5, wild type thymus; n = 8 MHCII^{-/-} thymus; Student's t-test). **(C)** OTII/Rag1^{-/-} mice exhibit no difference in retinal ganglion cell number in the uninjured retinas compared to Rag1^{-/-} mice. Retinal ganglion cell counts (% of Rag1^{-/-}) of Rag1^{-/-} and OTII/Rag1^{-/-} mice assessed by Fluoro-Gold staining (n = 3, Rag1^{-/-}; n = 3, OTII/Rag1^{-/-}; Student's t-test). **(D)** IL-4^{-/-} mice exhibit no difference in retinal ganglion cell number in the contralateral retinas compared to wild type mice. Bar graphs represent retinal ganglion cell counts (% of wild type) of IL-4^{-/-} or wild type mice, assessed by Fluoro-Gold staining (n = 5, wild type; n = 5, IL-4^{-/-}; Student's t-test). **(E)** Mice transplanted with IL-4^{-/-} bone marrow exhibit no difference in retinal ganglion cell number in the contralateral retinas compared to those transplanted with wild type bone marrow. Retinal ganglion cell counts (% of wild type ⇒ wild type) of wild type ⇒ wild type or IL-4^{-/-} ⇒ wild type bone marrow chimeras. Bone marrow was allowed to engraft for 6 weeks before optic nerve injury, and retinal ganglion cell counts were assessed by Fluoro-Gold staining (n = 3, wild type bone marrow; n = 3, IL-4^{-/-} bone marrow; Student's t-test). **(F)** Rag1^{-/-} mice receiving IL-4^{-/-} CD4⁺ T cells demonstrate no difference in retinal ganglion cell number in the contralateral retinas compared to Rag1^{-/-} mice receiving wild type T cells. Retinal ganglion cell counts (% of Rag1^{-/-} mice injected with wild type T cells) of Rag1^{-/-} mice injected with either wild type or IL-4^{-/-} CD4⁺ T cells 3 weeks before optic nerve injury. Retinal ganglion cell counts were assessed by Fluoro-Gold staining (n = 7, wild type; n = 7, IL-4^{-/-} T cell injected; Student's t-test). **(G)** MyD88^{-/-} mice exhibit no difference in retinal ganglion cell number in the contralateral retinas compared to wild type mice. Retinal ganglion cell counts (% of wild type) of wild type and MyD88^{-/-} mice were assessed by Fluoro-Gold staining (n = 3, wild type; n = 3, MyD88^{-/-}; Student's t-test). **(H)** Mice receiving MyD88^{-/-} bone marrow exhibit no difference in retinal ganglion cell number in the contralateral retinas compared to those receiving wild type bone marrow. Retinal ganglion cell counts (% of wild type ⇒ wild type) of wild type ⇒ wild type or MyD88^{-/-} ⇒ wild type bone marrow chimeras. Bone marrow was allowed to engraft for 6 weeks before optic nerve injury, and retinal ganglion cell survival was assessed by Fluoro-Gold staining (n = 3, wild type bone marrow recipients; n = 3 MyD88^{-/-} bone marrow recipients; Student's t-test).

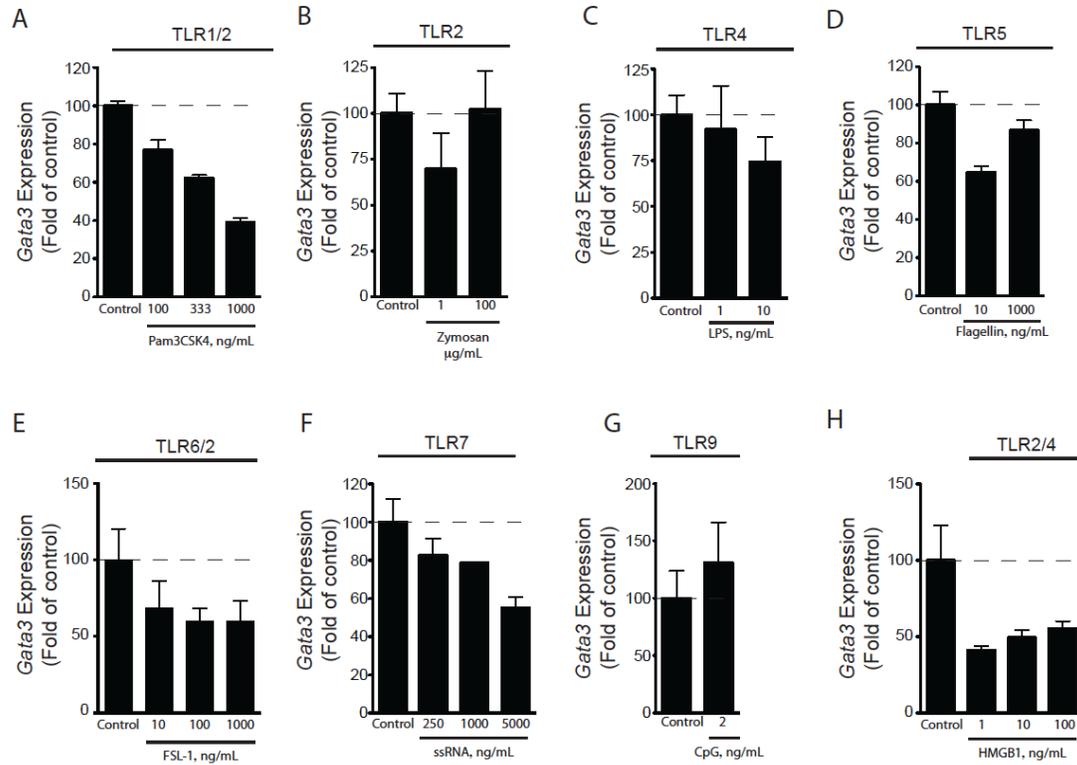


Figure S3: Classical TLR ligands do not increase *Gata3* mRNA expression in CD4⁺ T cells. *Gata3* expression (mean ± s.e.m.) in sorted wild type CD4⁺ T cells co-cultured with Pam3CSK4 (A), zymosan (B), LPS (C), flagellin (D), FSL-1 (E), ssRNA (F), CpG (G), or HMGB1 (H) at the indicated concentrations for 3 days (n = 3 per group, representative of two experiments).

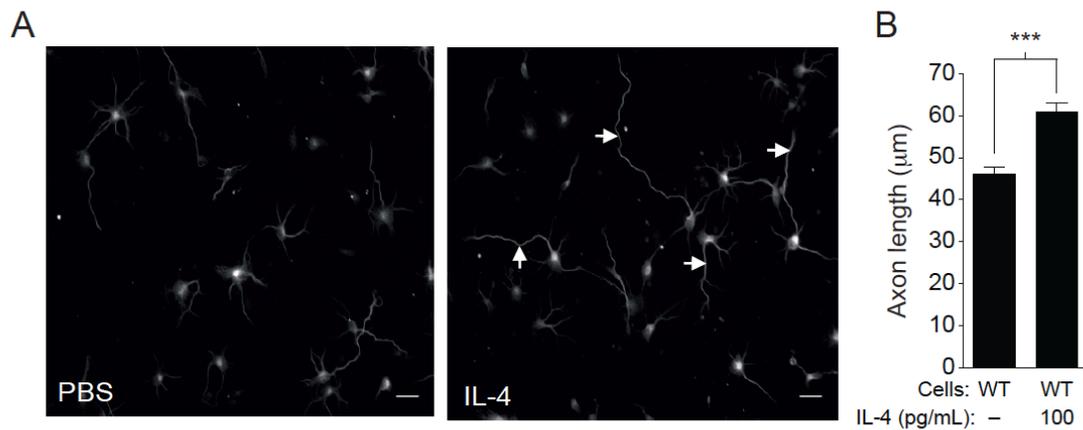


Figure S4: IL-4 induces axon elongation in cultured neurons (A, B) The application of a single dose of 100 pg/ml recombinant IL-4 significantly increased the length of axons (mean \pm s.e.m.) of isolated primary neurons. Representative microphotographs (A) and quantification of axonal length (B) of isolated neurons treated with PBS or a single dose of 100 pg/ml recombinant IL-4. Scale bar: 20 μ m. (n[well] = 4 per group. ***, $p < 0.001$; One-way ANOVA with Bonferroni's post-test).

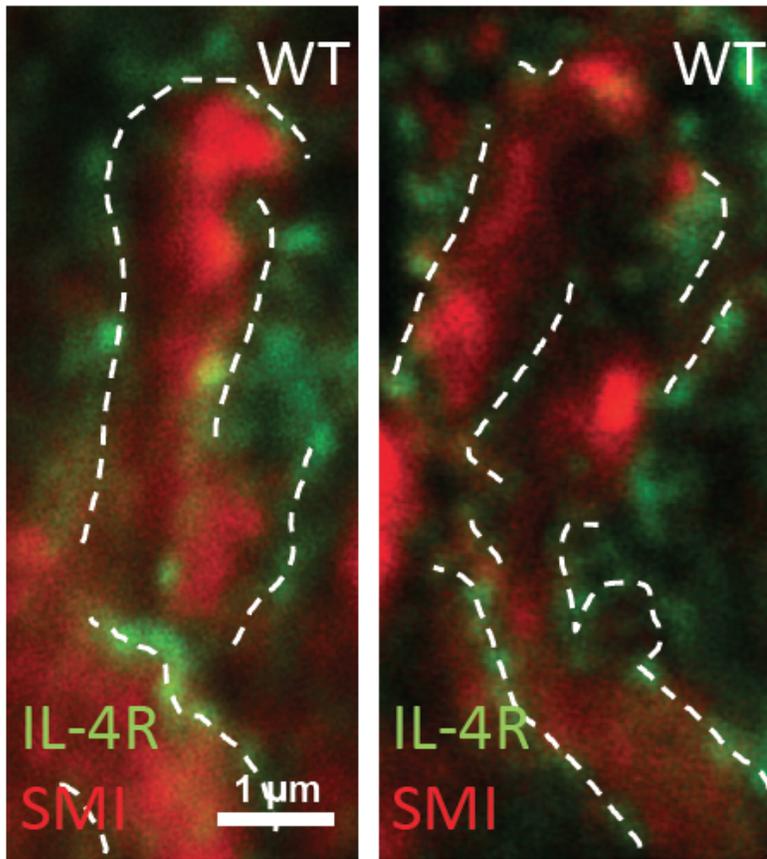


Figure S5: *IL-4R* expression on spinal cord axons. Axons in longitudinal sections of the spinal cord show tubular, membrane-related *IL-4R* expression (green) pattern, adjacent to the stained axonal neurofilament (SMI, red). The presumed membrane is indicated by dotted lines. Two representative axons (left and right panels) are provided. Scale bar: 1 μm .

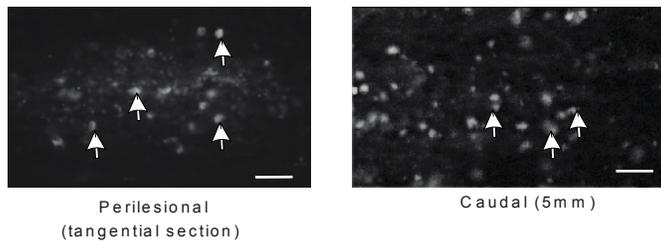


Figure S6: *T cells migrate caudally from the lesion site after injection.* CFDA_{SE} labeled T cells were injected into the site of injury immediately after spinal cord injury. Spinal cords were isolated and visualized for CFDA_{SE} labeled T cells 6 days after the lesion. (Scale bars: 15 μ m). T cells were found at least 5 mm away from the site of injection, where axonal regrowth was observed.