## SUPPLEMENTAL DATA



Figure S1. Flow cytometric gating strategy for Tregs, T-cells and dendritic cells. A) Viable cells were identified by first placing a size gate based on forward scatterarea (FSC-A) v side scatter-area (SSC-A). Doublets were then excluded using a FSC-A v FSC-height (FSC-H) gate. For all surface stains dead cells were excluded based on DAPI staining. A time gate was placed in order to exclude clogs or debris at the end of the run. B) For identification of T-cells and Tregs, cells were first gated for size and time as described for A. When identification of adoptively transferred T-cells or Tregs was required a CD45 allelic difference between host mice and donor mice was used. C) For T-cell reconstitution experiments the CD4<sup>+</sup>CD25<sup>-</sup> T-cell fraction following magnetic separation was flow sorted for CD4 expression, then the absence of expression of CD38 and CD44, resulting in a population of naïve T cells with minimal Foxp3-expressing cells. D) For DC analysis cells were first gated according to A, then B220-expressing cells excluded. DC subsets were then identified based on expression of CD11c and MHCII. In the pLN and mLN two major subsets of DCs were present: migratory DCs (MHCII<sup>hi</sup> CD11c<sup>int</sup>) and blood-derived (MHCII<sup>int</sup> CD11c<sup>hi</sup>). Spleen contained only the blood-derived subset, which was further subdivided based on differential expression of CD8 and CD11b.



**Figure S2. Proliferation of reconstituting Treg cells in lymphopenic host mice.**  $2.5 \times 10^6$  CFSE-labeled CD4<sup>+</sup>CD25<sup>+</sup> Tregs were adoptively transferred into lymphopenic  $Rag^{-/-}$  mice and treated with IL-2/JES6-1 second daily. Treg proliferation was assessed on days 1, 3, 5, and 7. Donor cells were identified based on a CD45 allelic difference from the host, then gated based on expression of CD4 and Foxp3. CFSE division profiles of donor CD4<sup>+</sup>FoxP3<sup>+</sup> donor cells from the pLN, mLN and spleen of Treg-reconstituted mice are shown. Plots are from a single experiment. B) Expression of CD25 and FoxP3 on donor CD4<sup>+</sup> T cells in the mLN and spleen of  $Rag^{-/-}$  mice reconstituted with the specified number of Treg (as described in Figure 1A). Data are representative plots from a single experiment with n=4 per group.



Figure S3. Administration of IL-2/JES6-1 complexes to immunodeficient mice does not affect DC expression of co-stimulatory molecules.  $Rag^{-/-}$  mice were treated with IL-2/JES6-1 on days 0, 2, 4 and 6, and lymphoid organs harvested on day 7. MFI of CD80 (upper panel) and CD86 expression (lower panel) by migratory DCs from the pLN, and CD8<sup>+</sup> and CD11b<sup>+</sup> splenic DCs in IL-2/JES6-1-treated versus untreated  $Rag^{-/-}$  mice was examined (n=3/group). Data are from a single experiment. Statistical analysis was performed using two-tailed unpaired t tests. Bars represent mean  $\pm$  SEM with individual values indicated by the open circles. ns=not significant.



Figure S4. Comparison of Treg expression of activation markers and Treg:DC ratio in the pLN and spleen of Treg-reconstituted mice. Rag<sup>-/-</sup> mice were reconstituted with either  $0.25 \times 10^6$  or  $2.5 \times 10^6$  purified Treg as described in Figure 1. Expression of a panel of Treg surface markers including Foxp3, CD25, CTLA-4, ICOS, LAG-3, CD39, CD73, GARP and PD-1 was examined on donor CD4<sup>+</sup>Foxp3<sup>+</sup> cells 7 days post-Treg transfer. Intracellular staining for CTLA-4 expression was also performed. Expression levels of each marker were compared between Treg from untreated WT mice (shaded histograms),  $Rag^{-/-}$  mice reconstituted with 0.25x10<sup>6</sup> Treg (unbroken line), and  $Rag^{-/-}$  mice reconstituted with 2.5x10<sup>6</sup> Treg (broken line). Analysis was performed on pooled samples from 4 mice per group. B) The absolute number of DCs in Treg-reconstituted mice for the experiment described in Figure 2C-D was calculated for the pLN (including migratory DCs and blood-derived DCs) and spleen (including CD8<sup>+</sup> and CD11b<sup>+</sup> blood-derived DCs). C) The ratio of Treg to DC in the pLN and spleen was calculated for the experiment described in Fig 2C-D. Data in B-C are from a single experiment with 4 mice per group. Statistical analysis was performed using a one-way ANOVA with Newman-Keuls post-test. Bars represent mean  $\pm$  SEM with individual values indicated by the open circles. ns=not significant, \*=p<0.05, \*\*=p<0.01.



Figure S5. Treg purity and cell numbers in lymphopenic mice reconstituted with either CTLA-4-sufficient or CTLA-4-deficient Tregs. CTLA-4-deficient Tregs were isolated from mixed bone marrow chimeras, generated as described in Figure 4. A) Upper panels: expression of Foxp3 versus CD25 on CD4<sup>+</sup> T cells before flow sorting. Lower panels: sorted CD4<sup>+</sup>CD25<sup>+</sup> Tregs were stained for Foxp3 expression to assess purity. Numbers refer to the percentage of cells within the gate. B) Representative plots of expression of Foxp3 versus CD25 on CD4<sup>+</sup> T cells from the pLN (upper panels) and spleen (lower panels) of  $Rag^{-/-}$  mice reconstituted with either WT Treg or CTLA-4-deficient Treg (n=4/group). C) Absolute numbers of Foxp3<sup>+</sup> Treg in the pLN and spleen of  $Rag^{-/-}$  mice reconstituted with either WT or CTLA-4-deficient Treg cells. Data in A-C are representative of two independent experiments. Statistical analysis was performed using a two-tailed unpaired t test. Bars represent mean ± SEM with individual values indicated by the open circles. \*\*=p<0.01.



Figure S6. Conventional T cells undergo rapid division in immunodeficient hosts independent of IL-2/JES6-1 treatment.  $1\times10^6$  CFSE-labeled CD4 T cells were adoptively transferred into  $Rag^{-/-}$  mice, with or without second daily treatment with IL-2/JES6-1 complexes. Representative CFSE division profiles in the pLN and spleen at day 7 post-transfer from a single experiment (n=3 per group) are shown (left). The ratio of fully divided (CFSE<sup>-</sup>) cells to undivided or slowly dividing T cells (CFSE<sup>+</sup>) cells was calculated (right). Statistical analysis was performed using a two-tailed unpaired t test. Bars represent mean ± SEM with individual values indicated by the open circles. \*=p<0.05, \*\*=p<0.01.



Figure S7. Expression of 4-1BBL, ICOSL, OX40L and CD70 on pLN migratory DCs from Treg-reconstituted mice.  $Rag^{-/-}$  mice were reconstituted with  $2.5 \times 10^6$  Tregs or naïve conventional T-cells at a dose of  $1.25 \times 10^6$  or  $2.5 \times 10^6$  as described in Figure 7 (n=4 per group). Expression of a range of co-stimulatory molecules including 4-1BBL, ICOSL, OX40L and CD70 was examined on pLN migratory DCs at day 7. Data are from a single experiment. Statistical analysis was performed using a one-way ANOVA with Newman-Keuls post-test. Bars represent mean  $\pm$  SEM with individual values indicated by the open circles. ns=non-significant.



**Figure S8.** Phenotype of Tregs and DCs following acute Treg depletion in DEREG mice. Flow cytometric analysis of CD4 T cells and DCs following acute Treg depletion of DEREG mice, as described for Figure 8. A) Representative plots of Foxp3 versus CD25 expression on pLN or spleen cells gated for CD4<sup>+</sup>. Numbers indicate the percentage of cells within the gate on days 0-3. B) Expression of CD80 (left panel) and CD86 (right panel) by pLN migratory DCs and splenic DCs at days 1-3 (open histograms) compared with untreated mice at d0 (shaded histograms). Data in A-B are from a single experiment with 3-4 mice per group. C) Representative plots of Foxp3 versus CD25 expression on pLN or spleen cells gated for CD4<sup>+</sup>. Numbers indicate the percentage of cells within the gate on days 0 and 3-7. D) Expression of CD80 (left panel) and CD86 (right panel) by pLN migratory DCs and splenic DCs at days 3-7 (open histograms) compared with untreated mice at day 0 (shaded histograms). Data in C-D are from a single experiment with 3-4 mice per group.



Figure S9. Effect of starting weight and IL-2/JES6-1 treatment in a GVHD model. A) Relationship between starting weight at the time of irradiation (x-axis) and the weight maintained following recovery from BMT (y-axis). Included are all mice that survived >50 days in the experiments described in Figures 10E-F (BMT group: n=9; T cell control: n=1, co-transferred Tregs: n=2; B10.BR Treg-reconstituted: n=9; [B6.BR]F1 Treg-reconstituted: n=10). The weight maintained following recovery from BMT was calculated based on weight at day 51, and is expressed as a percentage of the initial weight at time of irradiation. A line of best fit with  $R^2$  value is shown (p<0.0001). Data are pooled from 2 independent experiments. B-C) Mice were irradiated and transplanted with allogeneic BM cells as described in Figure 10. Mice received either BM cells alone (n=7), BM cells with allogeneic T cells at day 7 (n=10), or BM cells with allogeneic T cells at day 7 and IL-2/JES6-1 treatment at days 0, 2, 4, 6, and 8 (n=8). Shown are weights (B) and Kaplan-Meier survival analysis (C) from a single experiment. Median survival time for T-cell group was 20.5 days, while the median survival time for IL-2-treated T cell recipients was 21.5 days. Survival significantly differed between BMT controls and both the T-cell group (p=0.0003) and IL-2-treated T-cell recipients (p=0.0004). There was no significant difference between the T-cell group and the IL-2-treated T cell group (p=0.5313).



Figure S10. Relationship between Treg:DC ratio and DC expression of CD80/CD86 after Treg-reconstitution, Treg expansion, and Treg depletion. MFI of CD80 (left panel) and CD86 (right panel) expression on migratory DCs in the pLN and mLN, and blood-derived DCs in the pLN, mLN, and spleen was examined in Treg-reconstituted mice at day 7 post Treg-transfer (n=30, from 5 independent experiments including those described in Figures 2C-D, 3A, and 7A), WT mice at day 5 after IL-2/JES6-1 treatment at days 0, 1, and 2 (n=12, from 3 independent experiments including Figure 6A-C), or DEREG mice at day 1-7 following DT treatment at days 0 and 1 (n=30, from 3 independent experiments including those described in CD80/CD86 for each subset was normalized against the mean WT expression level in the relevant experiment, and is plotted against the Treg:DC ratio in the lymphoid organ (normalized against the WT ratio). Data were fitted to a one-phase decay model (CD80:R<sup>2</sup>=0.3444; CD86: R<sup>2</sup>=0.2812). The WT level of CD80/CD86 and the WT Treg:DC ratio are both indicated by broken lines.