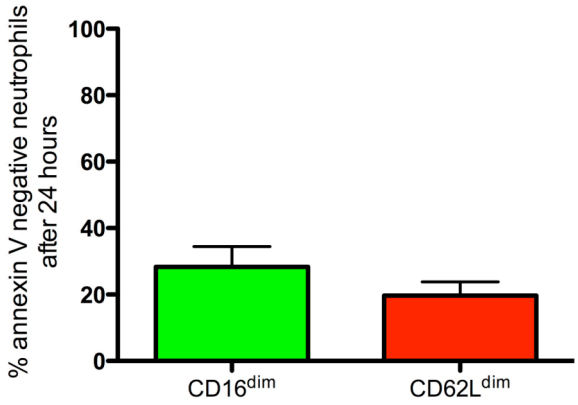
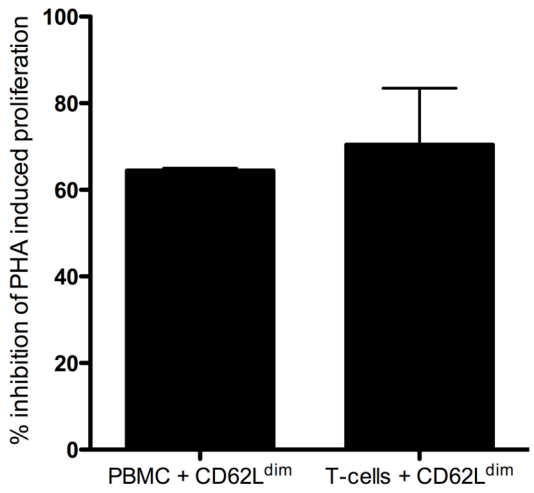
Supplementary Information

Supplemental table 1. Neutrophil phenotype after LPS administration. Measurements performed in erythrocyte-lysed whole blood. Leukocytes were stained with CD16, CD62L and different third stains. Neutrophil subsets were gated using CD16 and CD62L. Data are expressed as mean ± SEM of n=5.

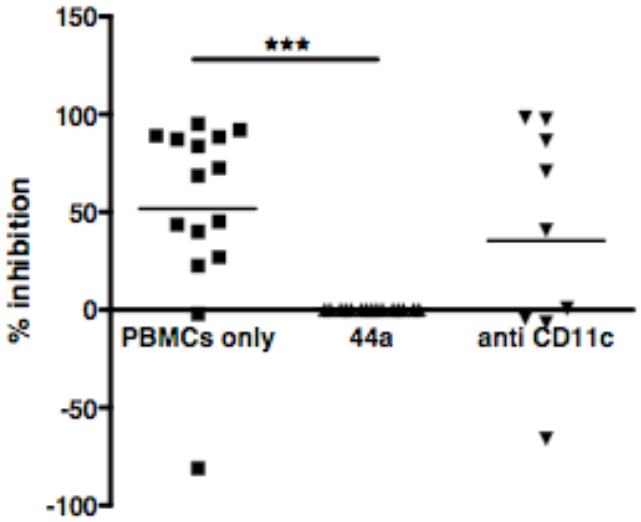
Antigen	*$CD16^{dim}$/ $CD62L^{bright}$*	*$CD16^{bright}$/ $CD62L^{bright}$*	*$CD16^{bright}$/ $CD62L^{dim}$*	*Clone*	*Antibody supplier*
CD11b	269.5 ± 35.6	364.1 ± 43.2	848.1 ± 179.4	2LPM19c	DAKO
CD11c	33.6 ± 10.0	83.4 ± 23.0	161.1 ± 3804	s-HCL-3	BD
CD29	43.0 ± 5.8	54.0 ± 7.8	60.0 ± 8.9	4B7R	Serotec
CD49d	1.9 ± 0.1	1.5 ± 0.1	3.9 ± 0.2	9F10	eBioscience
CD32	99.9 ± 6.0	185.2 ± 14.2	246.8 ± 17.7	FL18.26	BD
CD64	16.8 ± 2.3	13.8 ± 4.3	12.0 ± 0.9	10.1	Serotec
CD89	14.1 ± 3.2	11.2 ± 2.9	24.2 ± 4.6	A3	Santa cruz
CD33	24.6 ± 3.9	18.0 ± 4.1	34.5 ± 4.1	WM53	Serotec
CD66b	190.0 ± 28.6	118.5 ± 18.8	212.1 ± 23.5	80H3	Gene Tex
CD63	1.8 ± 0.1	1.5 ± 0.1	4.2 ± 0.2	H5C6	BD
CD14	9.9 ± 1.2	11.4 ± 0.9	15.3 ± 1.1	M5E2	BD
TLR4	8.3 ± 0.8	9.2 ± 0.7	11.7 ± 2.3	HTA 125	Imgenex
CD88	382.0 ± 169.9	424.1 ± 182.7	377.4± 165.7	P1 2/1	Serotec
CXCR1	33.7 ± 15.5	49.1 ± 22.2	47.7 ± 20.5	42705	R&D systems
CXCR2	18.8 ± 4.7	36.8 ± 8.7	39.4 ± 10.3	48311	R&D systems
CD54	10.7 ± 1.6	14.9 ± 3.4	20.0 ± 5.4	MEM-111	Invitrogen
CD35	25.4 ± 3.2	83.0 ± 22.4	201.0 ± 21.2	E11	BD
CD95	36.5 ± 1.1	37.2 ± 1.1	35.2 ± 0.9	DX2	DAKO
CD83	2.2 ± 0.5	2.0 ± 0.6	5.6 ± 1.5	HB15a	Santa cruz
CD45	5.5 ± 0.8	8.8 ± 1.9	36.4 ± 2.9	2D1	R&D systems



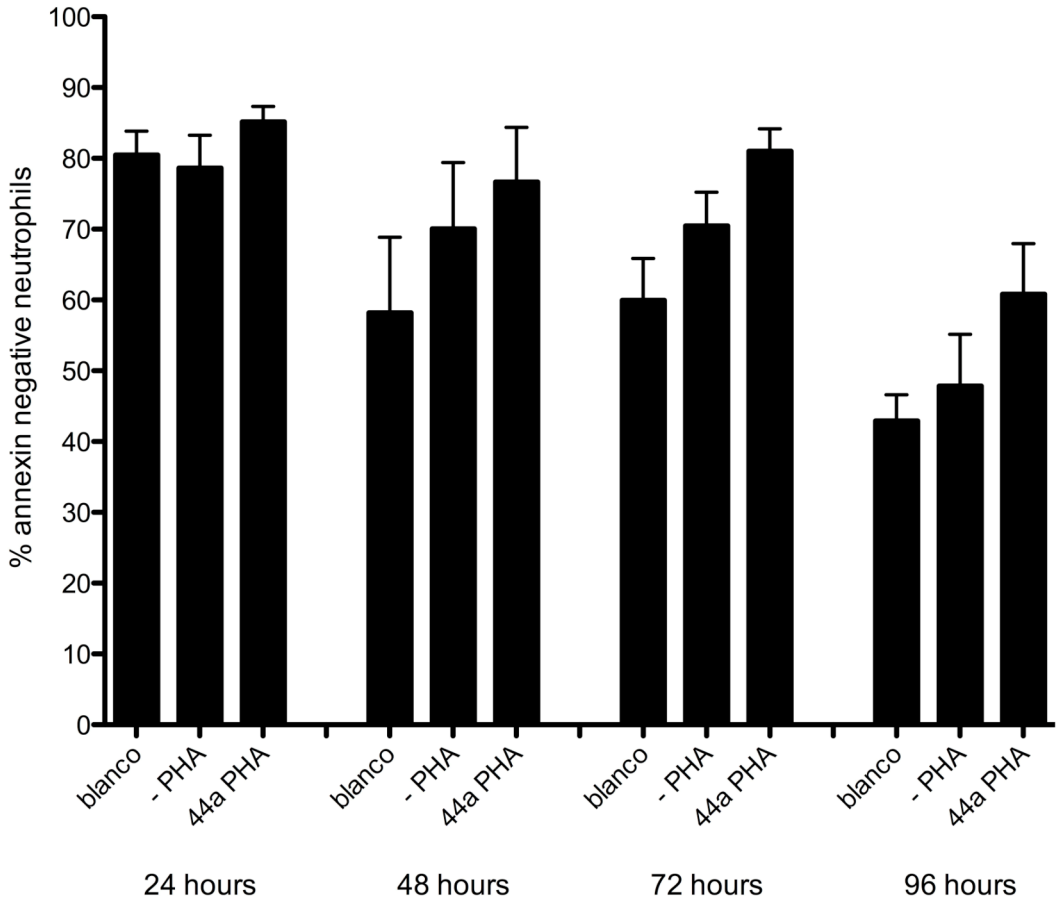
Supplemental figure 1. Neutrophil apoptosis after 24 hours in coculture. Neutrophils were added to PBMCs in a 2:1 ratio. Culture conditions were as described in the Material and methods. After 24 hours co-culture cells were stained with CD3 FITC, Annexin V PE, 7-AAD and CD16 Alexa647. Neutrophils were gated on forward and sideward scatter and CD16. Survival is defined as the percentage Annexin, 7-AAD negative cells. Data are expressed as mean ± SEM of n=6.



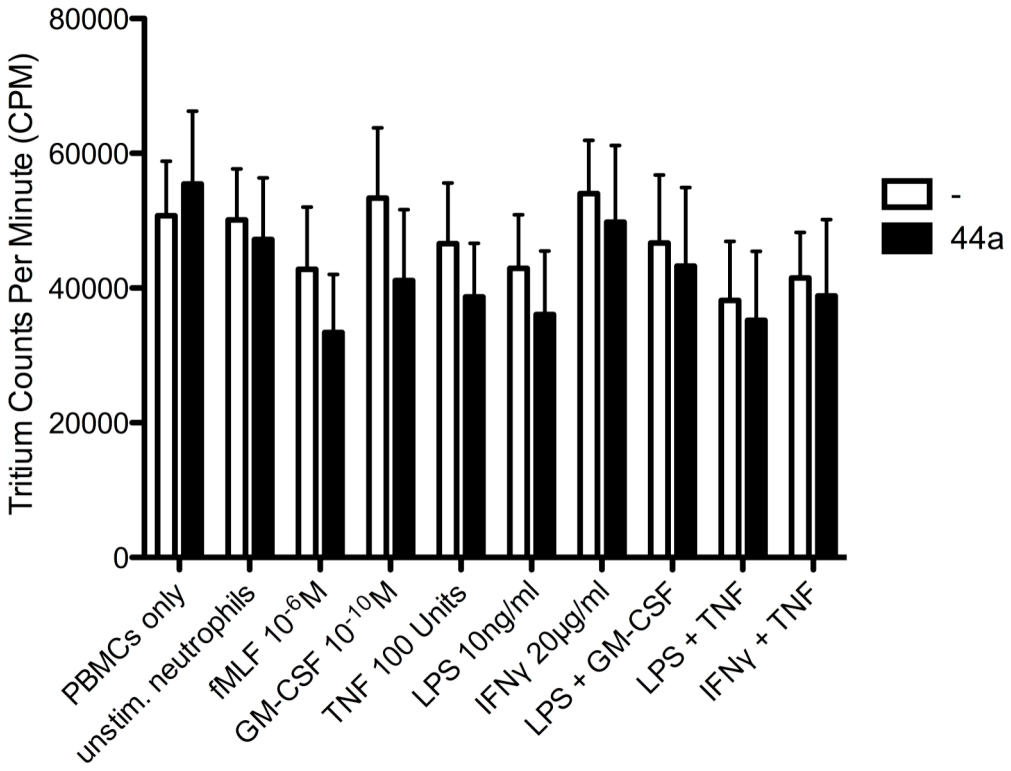
Supplemental figure 2. Inhibition of proliferation after monocyte depletion. Monocytes were depleted by magnetic beads coupled to CD14. A negative selection was performed, which resulted in >95% depletion of monocytes (not shown). Proliferation of T-cells was induced by PHA (10µg/ml). CD62Ldim neutrophils were added in a 2:1 ratio. Data are expressed as mean ± SEM of n=3.



Supplemental figure 3. T-cell suppression assay in whole blood. Erythrocytes were lysed and total leukocytes from LPS administrated donors were incubated with or without Mac-1 blocking antibody 44a, or control antibody CD11c. Cells were stimulated with PHA. After four days proliferation was determined by a ^{3}H-thymidine incorporation assay. Proliferation of 44a treated cells was set to zero % inhibition to determine the % inhibition in samples without 44a present. Data are expressed as mean ± SEM of n=15.



Supplemental figure 4. Neutrophil survival in whole blood suppression assay. Erythrocytes were lysed and total leukocytes were incubated with or without Mac-1 blocking antibody 44a. They were stimulated with PHA. After four days the leukocytes were stained for CD3, CD16, Annexin V, and 7-AAD, neutrophils were gated using forward sideward scatter and CD16. Neutrophil survival was determined as Annexin V negative CD16 positive cells. Data are expressed as mean ± SEM of n=6.



Supplemental figure 5. T-cell proliferation with in vitro stimulated neutrophils. Neutrophils of healthy controls were incubated with stimuli for 1 hour at 37°C. After stimulation these neutrophils were added to PBMCs, with or without Mac-1 blocking antibody 44a, for a T-cell proliferation assay. Assays were performed as described in the material and methods. Data are expressed as mean ± SEM of n=6.

Supplemental movie 1. Time lapse movie of neutrophil- lymphocyte interactions. $CD62L^{dim}$ sorted neutrophils were stained with CD16 FITC were added in a 2:1 ratio to unlabeled PBMCs stimulated with PHA (10µg/ml) were incubated in culture medium containing Amplex Red (20mM) and HRP (200 U/ml). Images were acquired using a Zeiss LSM510 Meta microscope. One representative example is shown of 4 independent experiments.