

SUPPLEMENTARY TABLES

Supplementary Table 1. Baseline characteristics of patients and controls.

Characteristic	Patients with DNA (n=439)	Controls (n=302)
Age (median- IQR ^a)	59 (41-68)	58 (45-66)
Male sex	210 (48%)	148 (49%)
Ethnicity		
White	415 (94%)	287 (95%)
African	17 (4%)	13 (4%)
Asian	7 (2%)	2 (1%)

^aIQR – interquartile range

Supplementary Table 2. Baseline characteristics of included patients with and without DNA.^a

Characteristic	Patients with DNA (n=439)	Patients without DNA (n=197)	P value
Age	59 (41-68)	62 (49-72)	0.001
Male sex	210 (48%)	94 (48%)	0.430
Immunocompromise	96/436 (22%)	37/165 (20%)	0.915
Distant focus of infection	200/436 (46%)	73/165 (44%)	0.721
Clinical signs and symptoms			
Headache	340/394 (86%)	111/134 (83%)	0.327
Fever	326/396 (83%)	118/146 (81%)	0.687
Neck stiffness	325/421 (77%)	115/152 (76%)	0.528
Glasgow Coma Scale score ^b	11 (9-14)	10 (8-14)	0.039
Indices of cerebrospinal fluid inflammation ^c			
Leukocyte count - cells/mm ³	3232 (793-8675)	1700 (281-6538)	0.001
Glucose level – mmol/L	0.50 (0.00-2.60)	0.55 (0.20-2.21)	<0.001
Protein level – g/L	0.15 (0.00-1.40)	4.18 (2.49-6.05)	0.046
Causative microorganism			
<i>S. pneumoniae</i>	314 (72%)	150 (76 %)	0.867
<i>N. meningitidis</i>	63 (14%)	18 (9%)	0.363
Other	62 (14%)	29 (15%)	0.784
Mortality	35/435 (8%)	69/164 (42%)	<0.001
Unfavorable outcome	114/435 (25%)	98/164 (60%)	<0.001

^a Data are number/number evaluated (percentage), continuous data are median (interquartile range) ^b Score on Glasgow Coma Scale was known in 434/439 (99%) patients with DNA and 162/197 (82%) patients without. ^c CSF leukocyte count was reported in 409/439 (93%) patients with DNA and 157/197 (80%) without, CSF glucose level was reported in 415/439 (95%) patients with DNA and 156/197 (79%) without, CSF protein level was reported in 412/439 (94%) patients with DNA and 154/197 (77%) without DNA.

Supplementary Table 3. Allele frequency, Hardy-Weinberg equilibrium and genotyping success rate of evaluated common complement component polymorphisms in 287 white controls.

Gen	SNP ID	A %	B %	A	B	AA	AB	BB	HWE ^a P - value	Success rate
C3	rs1047286	78,6%	21,4%	451	123	179	93	15	0,816	99,4%
C3	rs2230199	77,1%	22,9%	438	130	169	100	15	0,999	98,9%
C5	rs17611	43,2%	56,8%	247	325	53	141	92	0,997	99,7%
C6	rs1801033	69,3%	30,7%	398	176	138	122	27	1,000	99,3%
C7	rs1063499	35,2%	64,8%	202	372	40	122	125	0,514	99,6%
C7	rs13157656	23,0%	77,0%	129	433	13	103	165	0,831	97,7%
C7	rs60714178	16,4%	83,6%	94	480	8	78	201	0,991	99,9%
C8B	rs12067507	6,3%	93,7%	36	538	6	24	257	<0,001	99,6%
C8B	rs12085435	94,4%	5,6%	540	32	254	32	0	0,605	98,6%
C9	rs700233	61,7%	38,3%	343	213	106	131	41	0,999	95,7%
C9	rs34882957	94,3%	5,7%	532	32	250	32	0	0,600	98,7%
CFH	rs505102	70,4%	29,6%	404	170	143	118	26	0,973	99,3%
CFH	rs1065489	17,4%	82,6%	99	471	14	71	200	0,083	99,3%
CFH	rs1410996	54,9%	45,1%	315	259	83	149	55	0,715	99,7%
CFH	rs3753396	16,6%	83,4%	95	479	10	75	202	0,659	99,7%
CFH	rs6677604	80,5%	19,5%	459	111	187	85	13	0,710	99,1%
CFH	rs3753394	26,0%	74,0%	148	422	20	108	157	0,971	99,4%

^aHardy Weinberg equilibrium.

Supplementary Table 4. Multivariate logistic regression analysis for unfavorable outcome in pneumococcal meningitis.

Patient characteristic	Odds ratio (95% confidence interval)	<i>P</i> value
Age	1.017 (0.996 – 1.039)	0.115
Glasgow coma scale score	1.152 (1.277 – 1.041)	0.006
Thrombocyte count	1.000 (0.997 – 1.003)	0.976
CSF leukocyte count <1000/mm ³	3.623 (1.976 – 6.623)	<0.001
Immunocompromise	1.686 (0.873 – 3.257)	0.120
Otitis/Sinusitis	0.513 (0.283 – 0.930)	0.028
Rs17611	1.920 (1.057 – 3.487)	0.032

Supplementary Table 5. Effects of different antibody treatment modalities on CSF leukocyte counts, clinical status, neuroscore, and cerebellar titers 24 hours after induction of pneumococcal meningitis

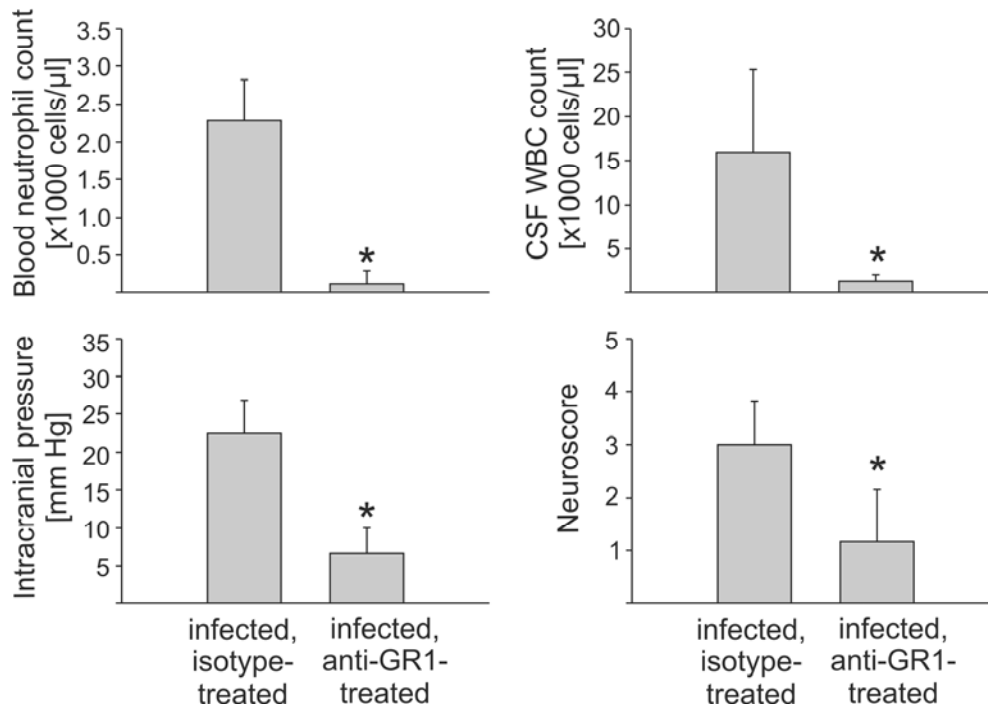
Antibody	Route of administration	Dosage [µg/mouse]	Number of mice [n]	CSF WBC [cells/µl]	Clinical score	Neuroscore	Bacterial titer [log₁₀ CFU/organ]
Anti-CXCL2	i.p.	100	3	8,717 ± 1,537	5.00 ± 2.65	n.d.	6.49 ± 0.46
Isotype control (anti-CXCL2)	i.p.	100	3	14,600 ± 5370	6.33 ± 0.58	n.d.	6.42 ± 0.40
Anti-CXCL1/CXCL2	i.p.	100 each	4	5,338 ± 1,244*	3.75 ± 0.50*	n.d.	6.50 ± 0.37
Isotype controls (anti-CXCL1/2)	i.p.	100 each	4	14,367 ± 3,202	7.25 ± 1.71	n.d.	6.52 ± 0.52
Anti-C5	i.p.	30	3	15,917 ± 5,328	7.33 ± 1.53	n.d.	6.28 ± 0.44
Anti-C5	i.c.	30	4	6,388 ± 2,225*	3.50 ± 1.29*	n.d.	6.39 ± 0.25
IgG controls (anti-C5)	i.c.	30	4	14,275 ± 5,013	6.75 ± 1.71	n.d.	6.20 ± 0.24
Anti-TLR2/4	i.p.	750 each	5	7,740 ± 3,583*	7.80 ± 0.84*	2.25 ± 0.69*	7.01 ± 0.32*
IgG controls (anti-TLR2/4)	i.p.	1500	5	15,200 ± 5,504	6.20 ± 0.45	4.00 ± 1.22	6.03 ± 0.36

CSF = cerebrospinal fluid; WBC = white blood cell; CFU = colony forming units; i.p. = intraperitoneally; i.c. = intracisternally; n.d. = not determined.

* p < 0.05, compared to the respective control groups .

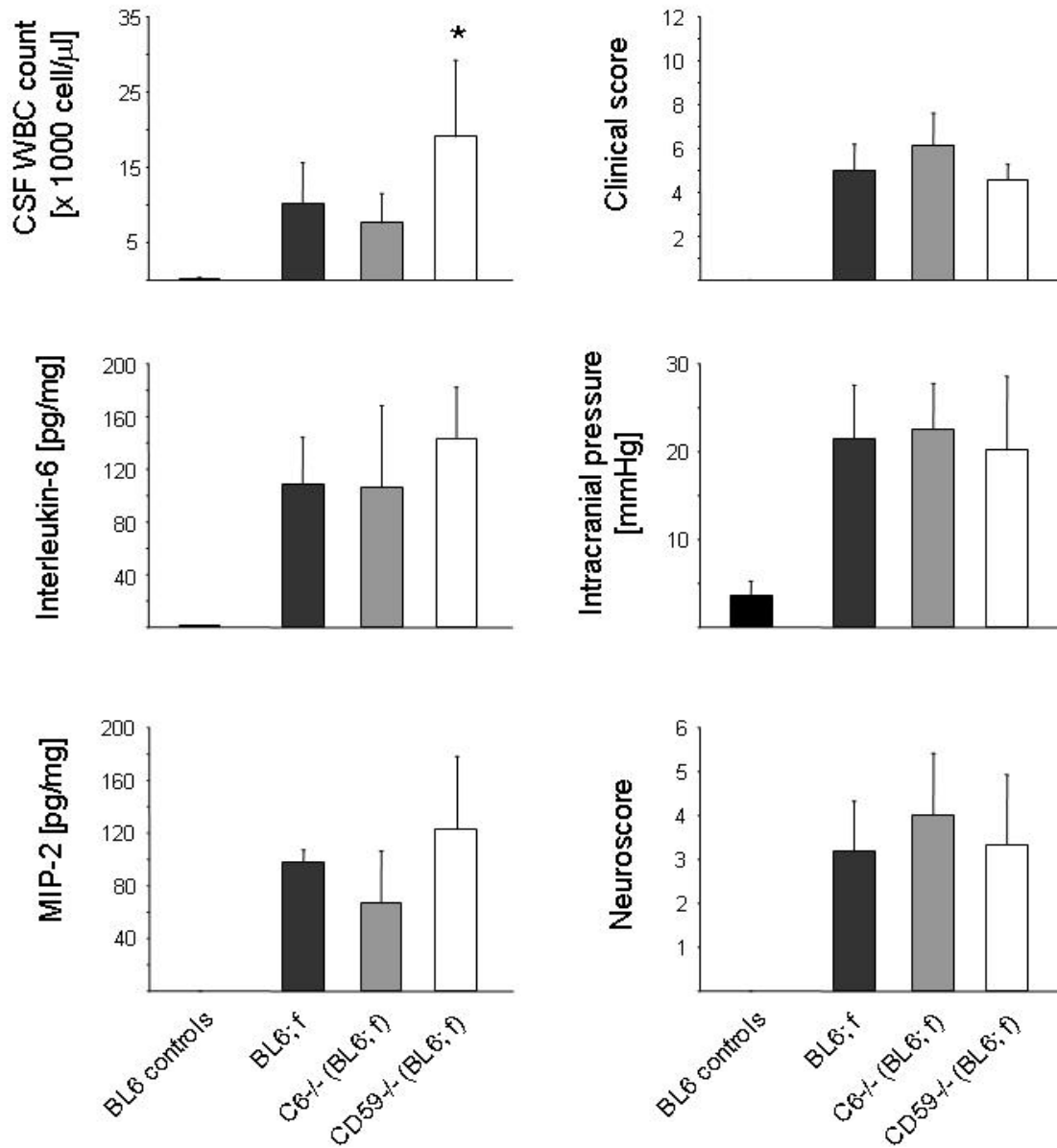
SUPPLEMENTARY FIGURES

Supplementary Figure 1: Effect of neutropenia in experimental pneumococcal meningitis.



In order to assess the role of neutrophils, wild type mice were treated with 250 μ g of either anti-GR1-antibody or rat IgG2b isotype control antibody (n=8 per group) 24 hours before disease induction. Then, animals were infected with *S. pneumoniae* and evaluated 24 h later for blood neutrophil counts, CSF leukocyte counts (CSF WBC count), intracranial pressure (ICP), blood brain barrier-breaching and intracerebral hemorrhage combined in the neuroscore. Anti-GR1-treatment resulted in markedly lower blood neutrophil and CSF leukocyte numbers compared to isotype control-treated mice which was also paralleled by a significant reduction in ICP and neuroscore values (unpaired Student's test; data are shown as means \pm SD).

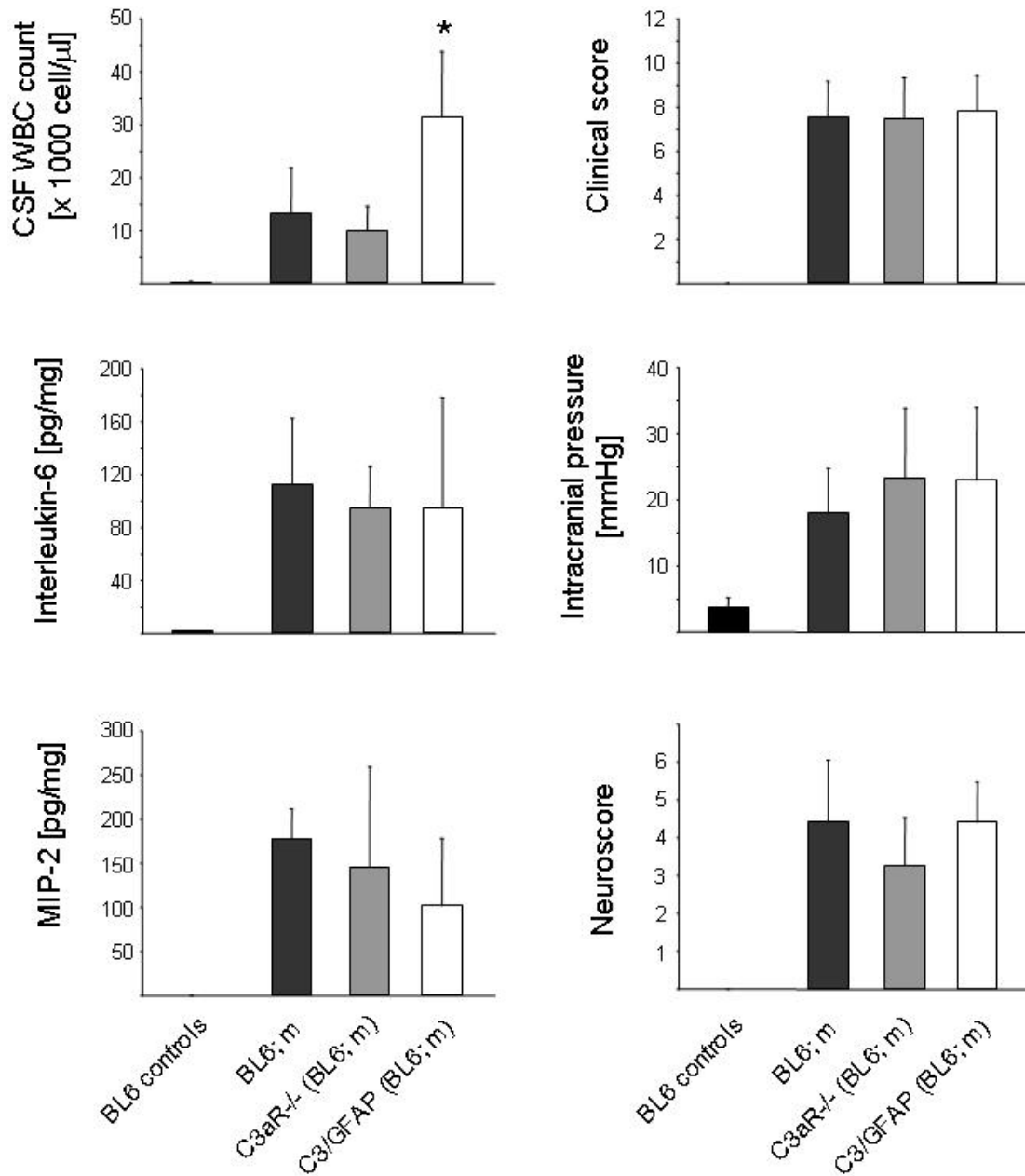
Supplementary Figure 2: Role of the membrane attack complex (MAC) in the mouse model of pneumococcal meningitis.



To evaluate the role of MAC, mice deficient in C6 (*C6*^{-/-}, n = 14) and thus unable to form MAC and mice deficient in CD59 (*Cd59a*^{-/-}, n = 11), the in vivo inhibitor of MAC, were examined. Animals were infected with *S. pneumoniae* and evaluated at 24 h after infection for CSF leukocyte count (CSF WBC count), clinical score, intracranial pressure (ICP), blood brain barrier-breaching and intracerebral hemorrhage combined in the neuroscore and expression of proinflammatory mediators and cytokines, namely Interleukin-6 and MIP-2. Infected mouse

mutants were compared to infected wt mice (C57BL/6 (BL6), f = female, n = 20). C57BL/6 mice intracisternally injected with PBS served as controls (BL6 controls, n = 8). Compared to infected wt mice and infected *C6*^{-/-} mice, *Cd59a*^{-/-} had significantly increased CSF WBC count. There was no difference in any of the other evaluated parameters (unpaired Student's test; data are shown as means \pm SD).

Supplementary Figure 3: Role of C3a in the mouse model of pneumococcal meningitis.



To evaluate the role of C3a, mice deficient in the C3a-receptor (*C3ar1*^{-/-}, n = 12) and mice with selective expression of C3a in the CNS (C3a/GFAP, n = 11) were examined. Animals were infected with *S. pneumoniae* and evaluated at 24 h after infection for CSF leukocyte count (CSF WBC count), clinical score, intracranial pressure (ICP), blood brain barrier-breaching and intracerebral hemorrhage combined in the neuroscore and expression of proinflammatory

mediators and cytokines, namely Interleukin-6 and MIP-2. Infected mouse mutants were compared to infected wt mice (C57BL/6 (BL6), m = male, n = 12). C57BL/6 mice intracisternally injected with PBS served as controls (BL6 controls, n = 8). Compared to infected wt mice and infected *C3ar1*^{-/-} mice, C3/GFAP mice had significantly increased CSF WBC count. There was no difference in any of the other of the evaluated parameters (unpaired Student's test; data are shown as means \pm SD).

SUPPLEMENTAL METHODS

Participating hospitals, local investigators (number of patients included).

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