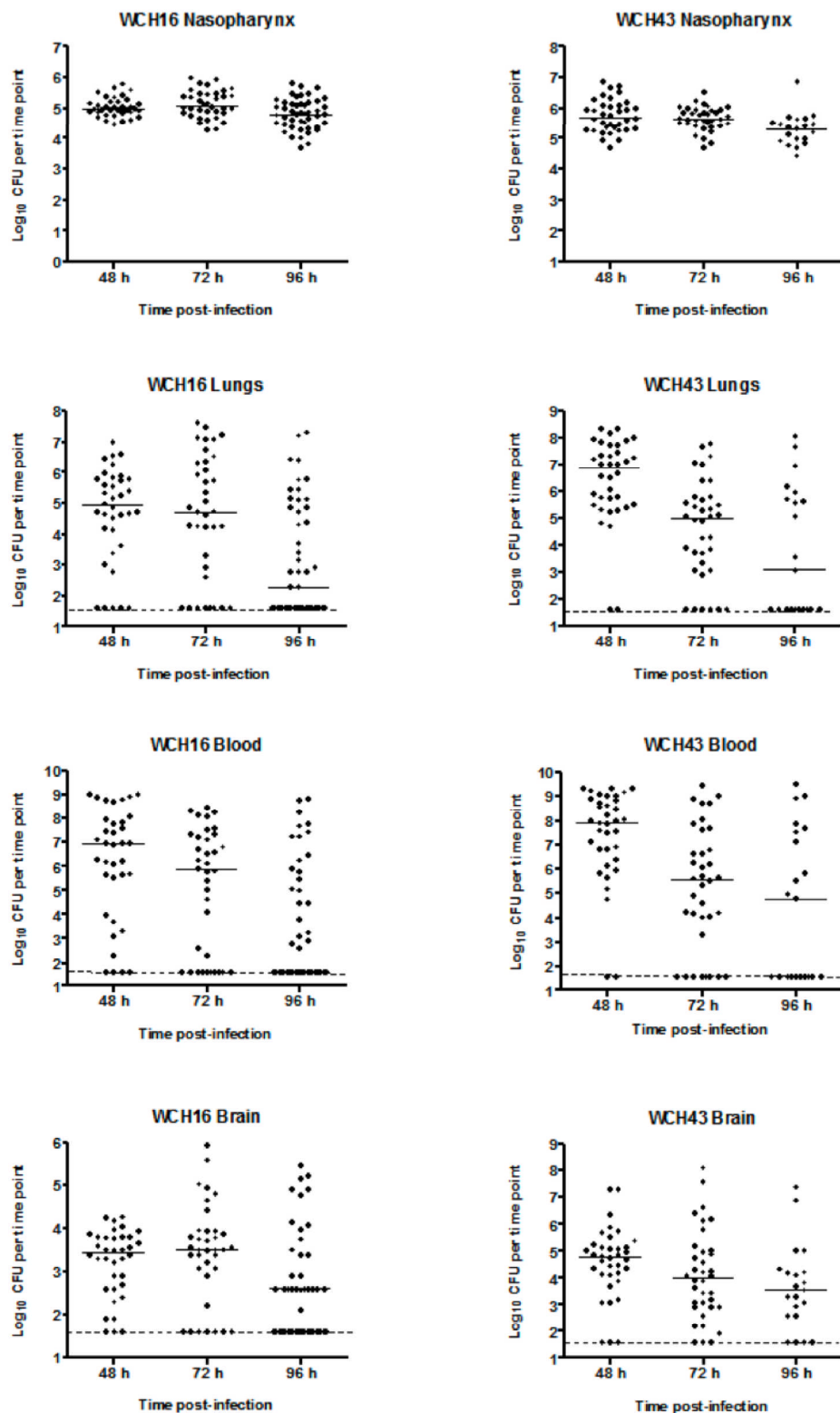
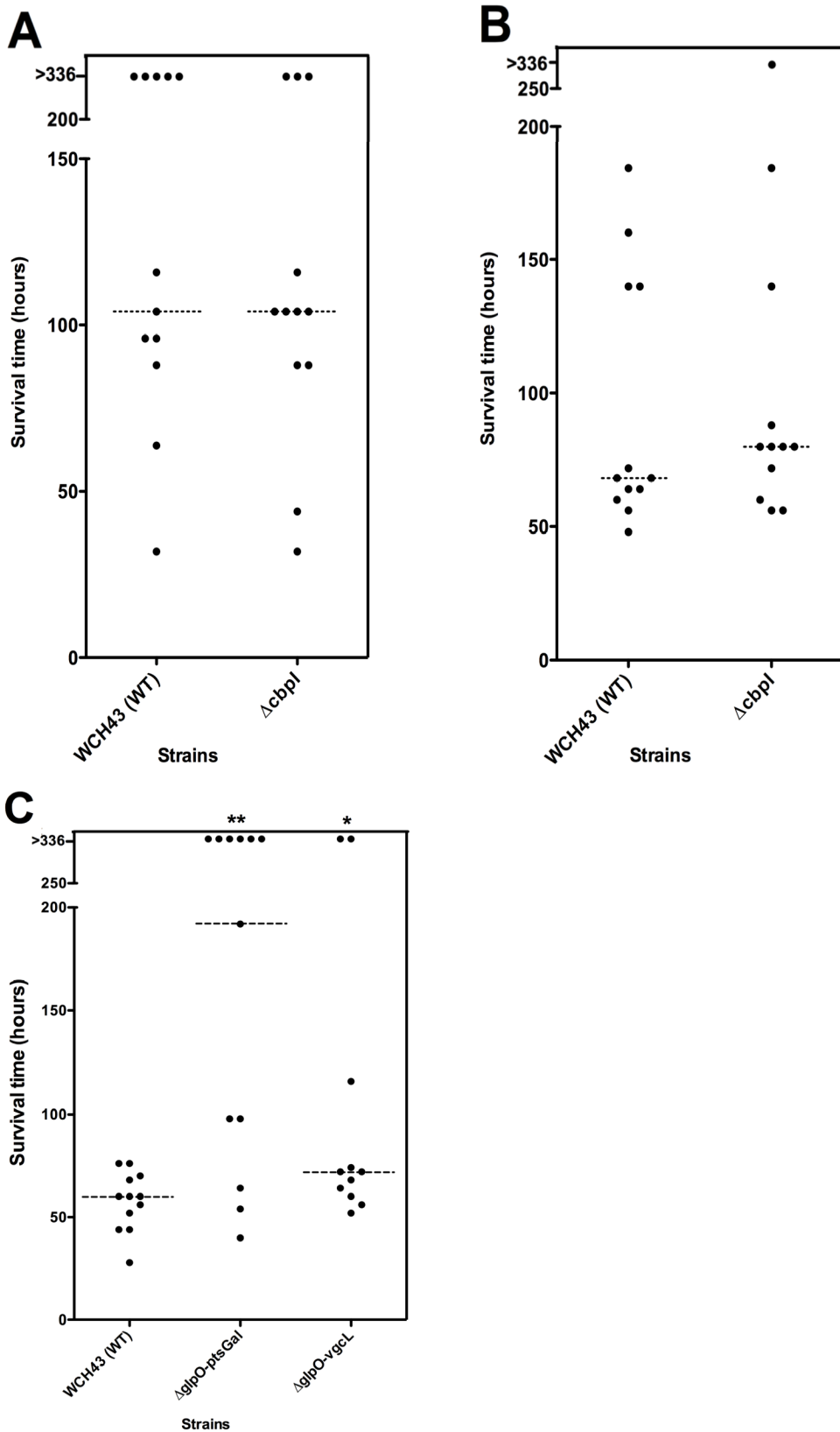




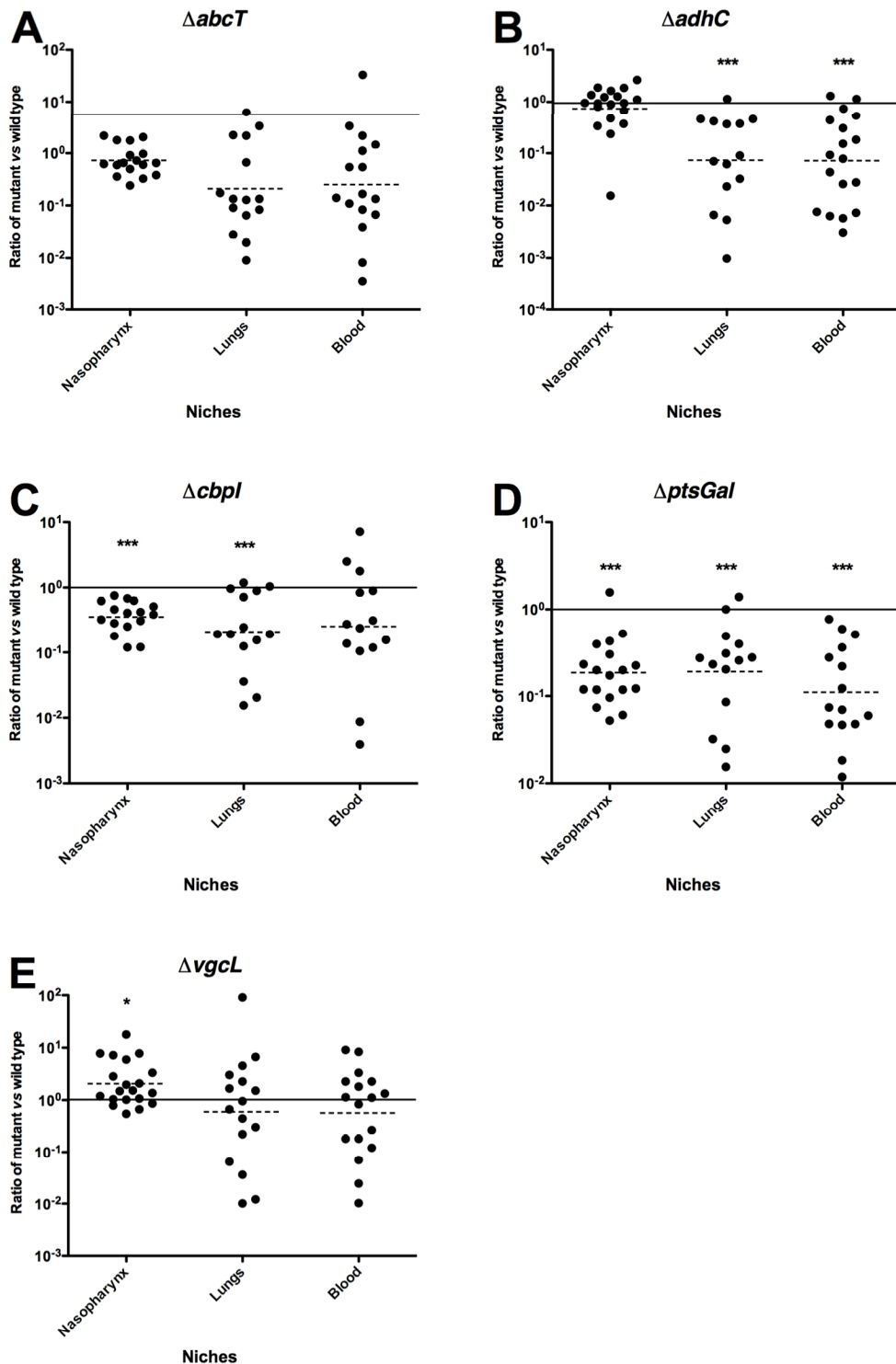
Supplemental Figure 1

Pathogenesis of pneumococcal meningitis in WCH16 and WCH43. Bacteria in the indicated niches of CD1 mice were enumerated at 48, 72, and 96 h after i.n. challenge. Each datum point represents one mouse. The horizontal lines indicate the median CFU recovered from each niche at each time point for the indicated strains. The limit of detection, indicated by dashed lines, was 40 CFU for all niches.



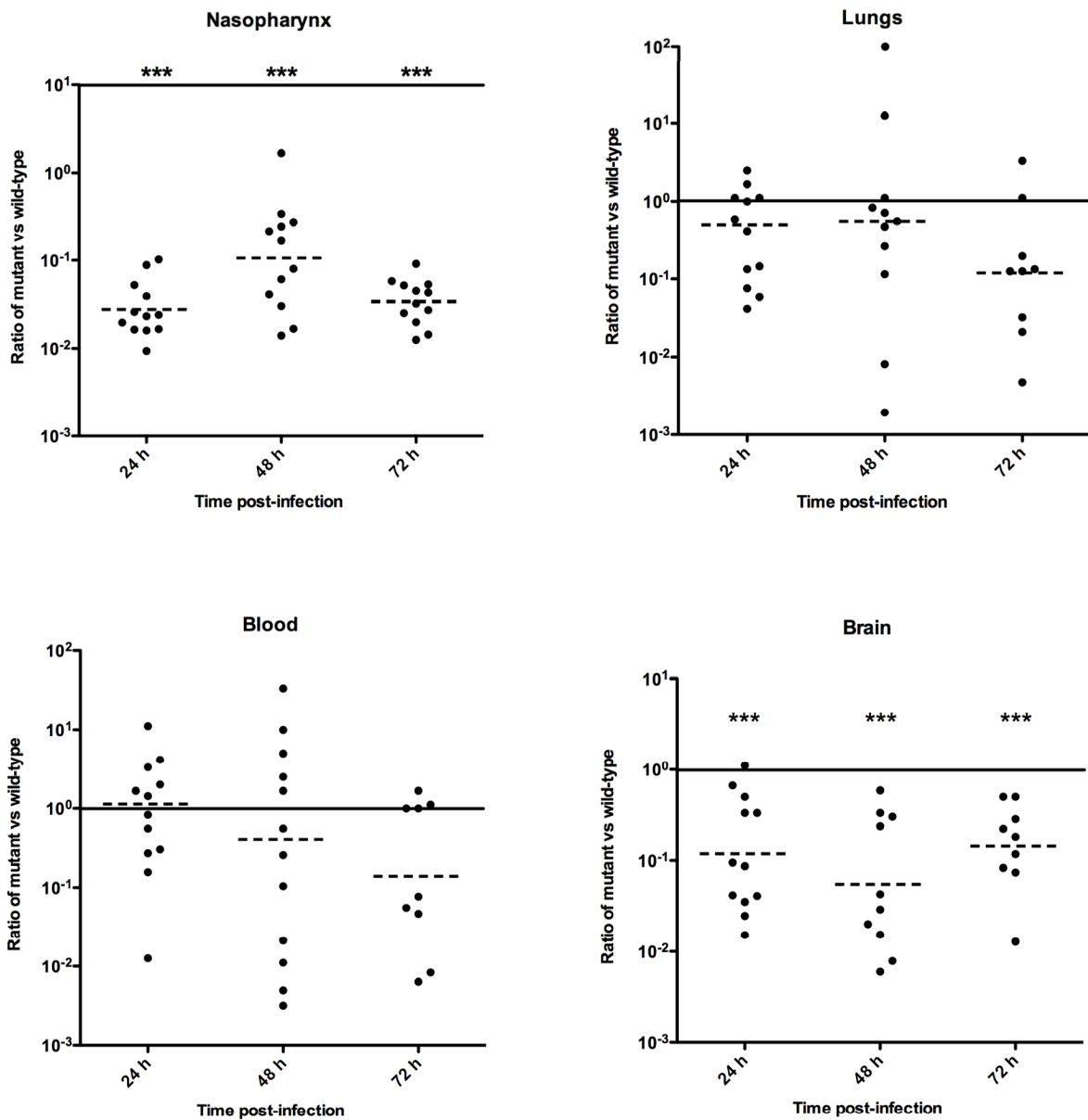
Supplemental Figure 2

Characterization of single $\Delta cbpI$ and double $\Delta glpO + \Delta ptsGal$ and $\Delta glpO + \Delta vgcL$ mutants of *S. pneumoniae* WCH43. Survival times for mice after (A) i.p., and (B) i.n. challenge with $\Delta cbpI$ mutant vs wild type. (C) Survival times after i.n. challenge with $\Delta glpO + \Delta ptsGal$ and $\Delta glpO + \Delta vgcL$ mutants vs wild type. Groups of 12 CD1 mice were challenged i.p. with approx. 1×10^5 CFU, and i.n. with approx. 2×10^7 CFU of wild type WCH43 and its isogenic mutant strains. Each datum point represents one mouse. The horizontal broken lines denote the median survival time for each group. * $P < 0.05$; ** $P < 0.01$; Mann Whitney *U*-test, one-tailed.



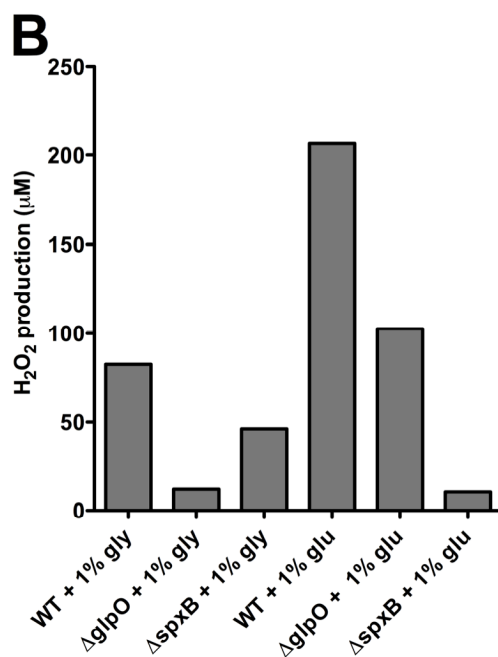
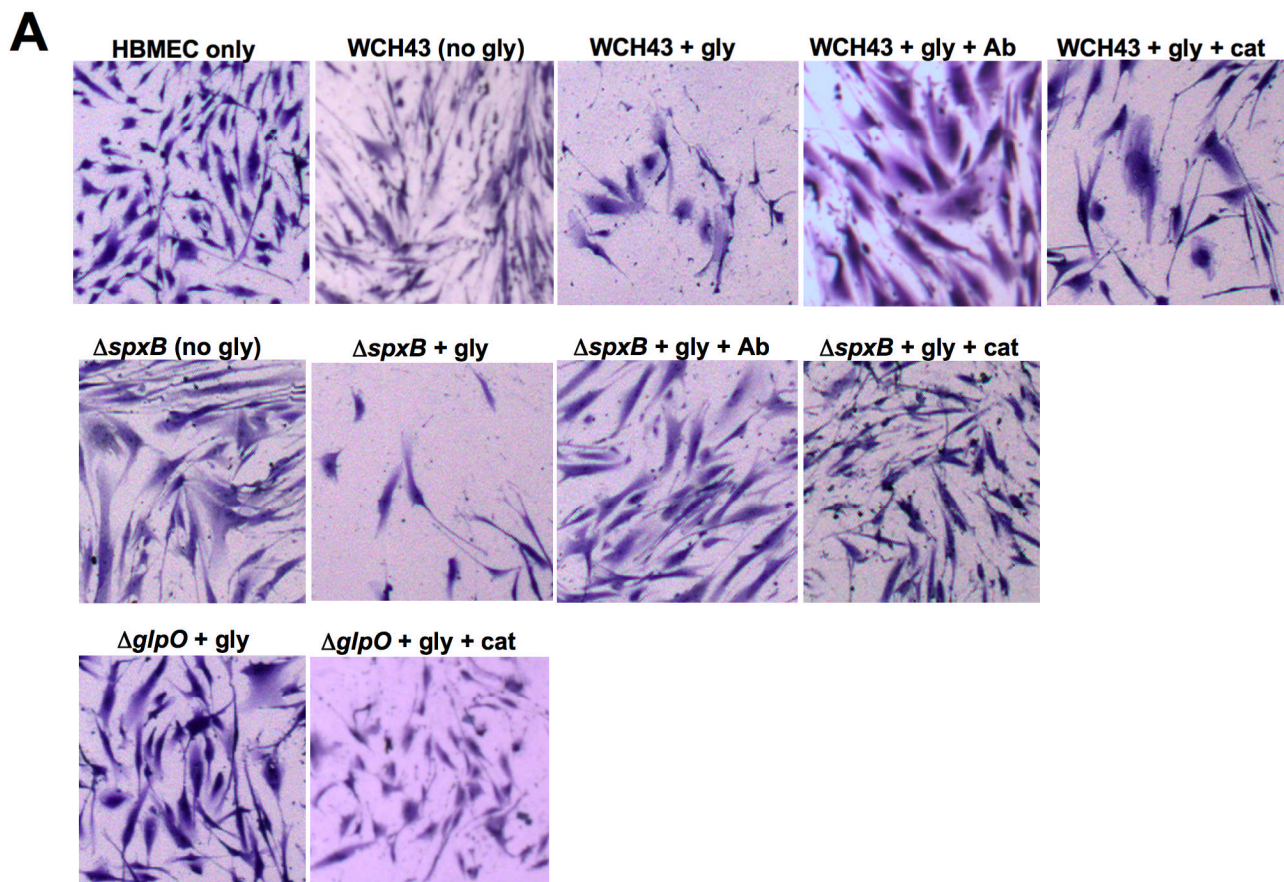
Supplemental Figure 4

Competition experiments between wild type WCH43 and isogenic mutants in the nasopharynx, lungs, and blood of mice at 48 post-infection. 14-20 CD1 mice were infected i.n. with equal numbers (approx. 1×10^7 CFU each) of wild type and mutant. Each datum point represents the ratio of mutant bacteria to wild type for each mouse. The horizontal solid line denotes a 1:1 ratio. The horizontal broken line denotes the geometric mean ratio of mutant:wild type for each comparison. * $P < 0.05$; *** $P < 0.001$; one sample t -test, two-tailed.



Supplemental Figure 5

Competition experiments between wild type WCH43 and its isogenic $\Delta glpO$ mutant in the nasopharynx, lungs, blood and brain of mice at 24, 48, and 72 h post-infection. 10-12 CD1 mice were infected i.n. with equal numbers (approx. 1×10^7 CFU each) of wild type and mutant. Each datum point represents the ratio of mutant:wild type for each mouse. The horizontal solid line denotes a 1:1 ratio. The horizontal broken line denotes the geometric mean ratio for each comparison. *** $P < 0.001$; one sample t -test, two-tailed.



Supplemental Figure 6

HBMEC cytotoxicity and H₂O₂ production. (**A**) Crystal violet-stained HBMEC monolayers after infection with the strain and supplements, as indicated on top of each panel and detailed in the Methods. (**B**) H₂O₂ production. The indicated strains were cultured in medium supplemented with glycerol (gly) or glucose (glu) and assayed for H₂O₂ production as detailed in the Methods. WT = Wild type WCH43; cat = catalase; Ab = polyclonal anti-GlpO serum.

Wild-type



***ΔglpO*
mutant**



Uninfected



Supplemental Figure 7

Blood-brain permeability. Mice were infected with WCH43 or *ΔglpO* and after 48 h, injected i.p. with Evans Blue. After a further 2 h, mice were euthanased and perfused as described in the Methods. Brains were then removed and photographed.

Table S1. *S. pneumoniae* strains used in this study.

Strain	Description (Sequence type)	Source/Reference
D39	Capsular serotype 2 (595)	NCTC 7466
D39:: Δfhs	<i>fhs</i> deletion mutant of D39 [Spec ^R]	Present study
WCH16	Capsular serotype 6A clinical isolate (4966)	Women's and Children's Hospital, North Adelaide, Australia
WCH16:: Δfhs	<i>fhs</i> deletion mutant of WCH16 [Spec ^R]	Present study
WCH43	Capsular serotype 4 clinical isolate (205)	Women's and Children's Hospital, North Adelaide, Australia
WCH37	Capsular serotype 33 clinical isolate	Women's and Children's Hospital, North Adelaide, Australia
WCH163	Capsular serotype 35F clinical isolate	Women's and Children's Hospital, North Adelaide, Australia
$\Delta abcT$	<i>abcT</i> deletion mutant of WCH43 [Ery ^R]	Present study
$\Delta adhC$	<i>adhC</i> deletion mutant of WCH43 [Ery ^R]	Present study
$\Delta cbpI$	<i>cbpI</i> deletion mutant of WCH43 [Ery ^R]	Present study
Δfhs	<i>fhs</i> deletion mutant of WCH43 [Spec ^R]	Present study
$\Delta glpO$	<i>glpO</i> deletion mutant of WCH43 [Spec ^R]	Present study
$\Delta ptsGal$	<i>ptsGal</i> deletion mutant of WCH43 [Spec ^R]	Present study

$\Delta spxB$	<i>spxB</i> deletion mutant of WCH43 [Ery ^R]	Present study
$\Delta vgcL$	<i>vgcL</i> deletion mutant of WCH43 [Spec ^R]	Present study
$\Delta glpO + \Delta ptsGal$	Double <i>glpO</i> and <i>ptsGal</i> deletion mutants of WCH43 [Ery ^R + Spec ^R , respectively]	Present study
$\Delta glpO + \Delta vgcL$	Double <i>glpO</i> and <i>vgcL</i> deletion mutants of WCH43 [Ery ^R + Spec ^R , respectively]	Present study

Table S2. Primers used in this study.

Primers for real time RT-PCR ^A	Sequence (5' → 3')
16S F	AAGCAACGCGAAGAACCTT
16S R	GTCTCGCTAGAGTGCCCAAC
<i>abcT</i> F	TTTGTGATGAACCGACTGGA
<i>abcT</i> R	GCACCACATCCTTGACACTG
<i>adhC</i> F	TGTTCAAAAAGGGGACAAGG
<i>adhC</i> R	CACGCACCTTGGTAATTCCT
<i>cbpA</i> F	GTGTTGGAGTAGCTAGTGTAGTTG
<i>cbpA</i> R	TCAACTATTTTATTCAAATACTCGTTC
<i>cbpI</i> F	ATGGTTCTACTTTGACGGCTCAGGAGC
<i>cbpI</i> R	TACCCATTACCATTTACACG
<i>fhs</i> F	CTGTTGGTCTTGGTGGTCCT
<i>fhs</i> R	ACCCTCAACCTGCAAATCAC
<i>glpO</i> F	TCCATCTGCAGTTTCTCGTG
<i>glpO</i> R	TTTGCTGGGTTCAATTCTCC

<i>ply</i> F	GATGGCAAATAAAGCAGTAAATGACT
<i>ply</i> R	GCTACTTGCCAAACCAGGCAAATC
<i>priA</i> F	GTTGGAGAGGCCAAAAACAA
<i>priA</i> R	CCATGTTCTCCACCAATTCC
<i>pspA</i> F	CAAGTCTAGCCAGCGTCGCTATCT
<i>pspA</i> R	GAACAACAAGCGCCACATCATTC
<i>ptsGal</i> F	TGATTCCTCACACAGATATTGTTCA
<i>ptsGal</i> R	CCATTTCCTGTGAAGAAGTCCAT
<i>sodA</i> F	GCATTTGCACCATGACAAAC
<i>sodA</i> R	AGGCTGCTTGGAATTCTTCA
<i>spxB</i> F	CAACATGTGCTACCCAGACG
<i>spxB</i> R	CGAGCATCGATGACAACAGT
<i>vgcL</i> F	ATTGAAAAGGGCTTCCACCT
<i>vgcL</i> R	ACCAGTTGAGGGACATGAGG

**Primers for
constructing
mutants^A**

Sequence (5' → 3')

<i>abcT</i> Flank F	CGGACTATCACTTAAATGTCAATCT
<i>abcT</i> Ery F	TTGTTTCATGTAATCACTCCTTCATCAAACAAATTATACTCCTCGAAAATC
<i>abcT</i> Flank R	TGAAGTTCCTCCAAGCCACCAG
<i>abcT</i> Ery R	CGGGAGGAAATAATTCTATGAGGCGAAAACTTATTGGAAGGACTTA
<i>adhC</i> Flank F	GAATGAGAAATATGAAGGCAAAATATG
<i>adhC</i> Ery F	TTGTTTCATGTAATCACTCCTTCTCATTTCACTGAATTCACGCT
<i>adhC</i> Flank R	TGCTCTTTTTCAAACCTTGAGGTC
<i>adhC</i> Ery R	CGGGAGGAAATAATTCTATGAGAGGAGTTTTAGAACTCTATTCGTTTT
<i>cbpI</i> Flank F	GCGGGTCGTATTGCAGGTGCG
<i>cbpI</i> Ery F	TTGTTTCATGTAATCACTCCTTCGGTTAATTAGCGACTCATAGTGAC
<i>cbpI</i> Flank R	CGGGTCCTTGTGTACCTTTGGT
<i>cbpI</i> Ery R	CGGGAGGAAATAATTCTATGAGATAGGTAAAGCATTATCTTACTTAG
<i>glpO</i> Flank F	TGGCTGAAAACGGTGGCCAGC
<i>glpO</i> Spec X	TATGTATTCATATATATCCTCCTCCTAAATAAATCGCTTTATTCTGCCAG
<i>glpO</i> Flank R	GTCTAGCATTTCCTCCGTCGCTTG
<i>glpO</i> Spec Y	AAATAACAGATTGAAGAAGGTATAAGAAAAATAAAAGAGGTGGAGGGCA
<i>fhs</i> Flank F	CAATGGCTAGGGTTAGAATTGCAG

<i>fhs</i> Spec X	TATGTATTCATATATATCCTCCTCAACCTCTTATATGATAATTCATTATATC
<i>fhs</i> Flank R	TCTACTGGCATAGGTTGAGAACG
<i>fhs</i> Spec Y	AAATAACAGATTGAAGAAGGTATAATGGATTGAAATTGACTTTAAGTGAGT
<i>ptsGal</i> Flank F	CTTCTCAATGGAGATAAGGTGGAT
<i>ptsGal</i> Spec X	TATGTATTCATATATATCCTCCTCTCAAAAAGTTATTCTTGATTATCCGC
<i>ptsGal</i> Flank R	ATGGTTATAGGCTTTATCTTCAACTG
<i>ptsGal</i> Spec Y	AAATAACAGATTGAAGAAGGTATAATAGTTCTATGAATGATGAAGCAAGTA
<i>spxB</i> Flank F	GTTGCAGGTAAGCCATATATCCAG
<i>spxB</i> Ery X	TTGTTTCAATGTAATCACTCCTTCTTCAATTTTTTTTAACTTGGAGAATACG
<i>spxB</i> Flank R	AACCCCGTCTTTGTAAATGGCATC
<i>spxB</i> Ery R	CGGGAGGAAATAATTCTATGAGCCGAAAATCAAATATGAACTTGTAG
<i>vgcL</i> Flank F	GACCAAGATTGTTGGTGATAAACCA
<i>vgcL</i> Spec X	TATGTATTCATATATATCCTCCTCAATTAACCTTTCTTTTTTACCTAAAACCAG
<i>vgcL</i> Flank R	TGGGGTGTTACACGCACTTTAAAAT
<i>vgcL</i> Spec Y	AAATAACAGATTGAAGAAGGTATAATATGAGTATTTACATAATTTATGTTATGTA

Primers for	Sequence (5' → 3')
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cloning^A

glpO F TTAAAAAAATGCATGCACGTACCCTGG

glpO ΔFADF GTGTAGCCTTGCATGCGGCAGCTAGCG

glpO R CTGTCTCCATTAAAAAGAAGGGACGGTCGACAAGGAATGC

^A Primers were derived from the *S. pneumoniae* TIGR4 (serotype 4) genome as deposited in the Kyoto Encyclopedia of Genes and Genomes (KEGG) database.