Supplementary Figure Legends

Figure S1

Flowcytometry-based purity analysis of peritoneal macrophage culture.

Thioglycollate elicited peritoneal cells were incubated in culture dish for 2 hours in DMEM supplemented with 10% FBS and antibiotic. After washing of non-adherent cells, adherent macrophages were scraped from dish with rubber policeman and subjected to FITC-labeled CD115 antibody staining. A total of 50,000 or more cells were analyzed on a BD FACSCalibur flow cytometer and CD115 positive cells were recorded as macrophages. Histogram shown is representative of 3 different samples.

Figure S2

Time course of KLF4 gene expression.

Changes in KLF4 mRNA expression levels after treatment with LPS (50ng/ml), IFN_{γ} (5ng/ml), IL4 (10ng/ml), IL13 (10ng/ml) and IL10 (10ng/ml) in (**A**) peritoneal macrophages (PM) and (**B**) bone marrow derived macrophages (BMDM). (**C**) Treatment of peritoneal macrophages with low dose of LPS (10ng/ml). n=3 for each time point. Data presented as mean ± SEM. *p<0.05, Student's t test with Bonferroni correction.

Figure S3

Characterization of myeloid-specific KLF4-deficient mice.

(A) Genotyping analysis of LysM^{cre/cre} (Mye-WT) and LysM^{cre/cre}:KLF4^{flox/flox} (Mye-KO) mice. (**B-D**) Deletion of KLF4 in Mye-KO macrophages as assessed by qPCR (**B** and **C**) and Western blot (**D**). n=3 in each group for qPCR. (**E**) Leukocytes cell populations are comparable in Mye-WT and Mye-KO mice as assayed by CBC (n=25 per genotype). (**F**) Monocyte population distribution (Ly6C^{hi} vs. Ly6C^{lo}) is similar in Mye-WT and Mye-KO mice as analyzed by flow cytometry. Leukocytes were stained with fluorescent anti-CD115-RBE (MCA1898PET, AbDserotec), anti-CD11b-PerCP-Cy5.5 (550993, BD), and anti-Ly6C-FITC (553104, BD) antibodies, and cells gated from CD115+ were further analyzed for CD11b and Ly6C expression. n=3 in each genotype. *p<0.05, Student's *t* test with Bonferroni correction. N.S.: not significant.

Figure S4

KLF4 is involved in the regulation of Arg1 and and PPAR γ expression.

(A) KLF4 enhanced IL4-induced Arg-1 expression. (B) KLF4 and Stat6 synergistically activate PPAR γ promoter in RAW264.7 cells. n=3 in each group. (C) Requirement of KLF binding sites for optimal Arg-1 promoter activity. Schematic diagram of the IL4-responsive enhancer region of the mouse Arg-1 gene showing pertinent transcription factor binding sites. WT: ~4kb wild type mouse arg-1 promoter; Δ KLF#1: arg-1 promoter with 5' KLF-binding site (KLF#1) mutated; Δ KLF#2: arg-1 promoter with 3' KLF-binding site (KLF#1) mutated; Δ KLF#2: arg-1 promoter with 3' KLF-binding site (KLF#1) mutated; Δ KLF#2: arg-1 promoter with 5' KLF-binding sites (KLF#1 mutated; Δ KLF#2: arg-1 promoter with 5' KLF-binding site (KLF#1 mutated; Δ KLF#2: arg-1 promoter with both KLF-binding sites (KLF#1 and KLF#2) mutated. n=3 in each group. (D) ChIP analysis demonstrates that binding of Stat6 to the Arg-1 promoter is not affected by KLF4 deficiency or overexpression. (E) IL4-induced Stat6 phosphorylation is similar in WT and KLF4-deficient BMDM. Western blots of total cell protein were assessed by antibodies against total or phosphorylated Stat6. *p<0.05, Student's *t* test with Bonferroni correction.

Figure S5

Deficiency of KLF4 in macrophages enhanced LPS-induced M1 gene expression.

LPS induced enhanced secretion of MCP-1 (**A**) and TNF α (**B**) in Mye-KO macrophages medium. Cytokine levels were determined by ELISA. n=3 in each group. *p<0.05, Student's *t* test.

Figure S6

LPS-induced pro-inflammation genes expression and NO production in WT and Stat6null macrophages.

(A-E) qPCR analysis of pro-inflammation genes in control and LPS-treated peritoneal macrophages from WT and Stat6-null mice. n=3 in each group. (F) LPS-induced NO production in WT and Stat6-null macrophages as assayed by nitrate concentration in conditioned medium. n=3 for each data point. *p<0.05, Student's *t* test. N.S.: not significant.

Figure S7

Non-specific PCR for Cox-2 promoter using non-targeting primers.

PCR was performed with sense-primers: ATTCAAGCAGCAGAAGAGGGCAG and antisense-primer: CTGGGATGCCAGAGCACACTG.

Figure S8

IL4-pretreatment inhibits subsequent LPS-mediated induction of M1 genes in Mye-WT macrophages but not in Mye-KO macrophages. n=3 for each data point. *p<0.05, Student's *t* test with Bonferroni correction. N.S.: not significant.

Figure S9

KLF4 deficient macrophage exhibit enhanced bactericidal activity.

(**A** and **B**) KLF4-deficient macrophages exhibit enhanced bactericidal activity against *S*. *aureus* ex vivo. n=3 in each genotype. (**C**) NOX1 expression is augmented in KLF4 deficient macrophages after incubation with S. aureus. n=3 for each data point. *p<0.05, Student's *t* test with Bonferroni correction.

Figure S10

Myeloid KLF4 deficiency does not affect bacterial uptake ability of macrophages.

WT and KLF4-deficient macrophages take up similar amounts of K-12 E. coli bioparticles as revealed by fluorescent microscopy (**A**) and flow cytometry assay (**B**). Image shown are representative from 3 sets of samples.

Figure S11

Infiltration of macrophages in skin wound and in vitro migration assay of macrophages.

(**A**) No difference is observed in the numbers of infiltrated macrophages in skin wound between Mye-WT and Mye-KO mice as analyzed by immunohistochemistry. n=3 in each group. (**B**) No difference in MCP-1 (CCL2) induced migration between WT and KLF4-deficient macrophages as assessed by Boyden chamber assay. n=3 in each group. N.S.: not significant as determined by Student's *t* test.

Figure S12

KLF2 and KLF4 expression levels in different human adipose tissues and KLF4 expression in different cells of adipose tissues.

(A) Compared with subcutaneous fat, visceral fat has lower expression of KLF4 but not of KLF2. Data represented were from 24 OB patient samples. (B) KLF4 is predominantly expressed by macrophages in adipose tissue. Cells were isolated from 4 OB patients. *p<0.05, Kruskal–Wallis test.</p>

Figure S13

Metabolic changes in Mye-WT and Mye-KO mice after HFD.

(A) Representative MRI images of body composition. (B) Mye-KO mice demonstrate higher level of total triglycerides in plasma (n=6 per genotype). (C and D) HDL and LDL levels in plasma remain similar between Mye-WT and Mye-KO after HFD. *p<0.05, Student's *t* test. N.S.: not significant.

Figure S14

Food intakes and energy expenditure in Mye-WT and Mye-KO mice.

(A) Food intake during a 24h period. (B) Respiration and heat production analysis. n=5 in each group. *p<0.05, Student's *t* test with Bonferroni correction. N.S.: not significant.

Figure S15

Impaired insulin signaling in HFD-fed Mye-KO mice.

(A) Serum insulin levels in Mye-WT and Mye-KO mice. n=6 in each group. (B) Baseline glucose tolerance test in Mye-WT and Mye-KO mice. n=6 in each group. (C) Impaired insulin signaling in Mye-KO mice after 12-week HFD as determined by phosphorylation of Akt. One animal in Saline group and two animals in Insulin-stimulated group were shown for each genotype.*p<0.05, Student's *t* test.

Figure S16

Myeloid deficiency of KLF4 results in alteration of expression of select metabolic and inflammatory genes in HFD-fed mice.

(A) Quadriceps muscle. (B) Liver tissue. n=6 per genotype. p value by Student's *t* test.





0 2 4 16 LPS (h)

0

BMDM

ΡM











С













Figure S2

Α







Ε





С













Ε







D



Ε



F

























A











Α



Α



В

Adipose tissue fractions





С

Mye-WT

Mye-KO



Α







D







С





Β

