

Supplementary Table 1: M&M Individual Participant Demographics

Study Arm	Participant	Sex at birth (Male M, Female F)	Race ¹	Ethnicity ²	HLA Protective Alleles ³	CD4 Nadir	Enrollment data						
							Age (yr)	ART (yr)	CD4 count (/mm ³)	CD8 count (/mm ³)	CD4: CD8	VL (copies/ mL)	
M3	00836	M	AA	NH			33	7.83	758	794	0.95	20	
	01237a	M	U	H			41	10.17	642	766	0.84	20	
	01249	M	C	NH			28	8.11	738	585	1.26	20	
	00937	F	AA	NH			48	9.21	1235	976	1.27	40	
	00835	M	C	NH			43	9.61	789	589	1.34	20	
	01214	M	C	NH			57	16.14	605	464	1.3	20	
	01009	F	AA	NH	B*58:01		60	11.86	515	555	0.93	20	
							Average	44.28	10.42	754.57	675.57	1.23	
M4	00749 ^{4,5}	M	AA	NH			30	5.87	484	332	1.46	20	
	01262	M	C	H			55	12.16	708	938	0.75	20	
	01264 ^{5,6}	M	AA	NH			37	6.03	1002	745	1.3	20 ⁶	
	01276	M	AA	NH			46	11.12	427	493	0.87	20	
	01277 ⁵	M	AA	NH	B*81:01		31	2.47	795	659	1.2	40	
	01095 ⁴	F	C	NH			58	10.41	571	867	0.66	20	
	00115	M	C	NH			59	18.12	821	996	0.82	40	
							Average	45.14	9.45	686.86	718.57	1.01	
M3M4	00631 ⁵	M	AA	NH			29	6.44	944	648	1.46	20	
	00371 ⁴	M	AA	NH	B*57:03, 58:01		30	10.72	566	538	1.05	20	
	01153	M	C	NH			55	12.71	847	942	0.9	40	
	01280	M	C	NH			54	6.55	1086	616	1.76	40	
	01213	M [#]	AA	NH			21	2.45	463	406	1.14	20	
	01295 ⁵	M	AA	NH	B*57:03		41	9.01	1365	876	1.56	20	
	01293 ⁵	M	AA	NH			31	4.34	855	1219	0.7	20	
							Average	37.29	7.46	875.14	749.29	1.22	
Placebo	00818 ⁵	M	B/AA	NH			24	6.68	792	1447	0.55	20	
	01270 ⁵	M	C	NH			28	4.59	1070	1322	0.81	20	
	01220 ⁵	M	C	NH			52	2.37	522	787	0.66	20	
							Average	34.67	4.55	794.67	1185.33	0.67	
							Cohort Average (n=24)	41.29	8.54	775.00	773.33	1.06	

¹C: White/Caucasian, AA: Black/African American, U: Did not fall in NIH categories of American Indian or Alaska Native, Asian, Black or African American, Native Hawaiian or Other Pacific Islander, or White

²H: Hispanic, NH: Non-Hispanic

³ Participant expresses one or more of HLA I alleles, B*57:01/03, B*58:01, B*81:01

⁴ T-cell response against HIV outgrowth virus/s measured

⁵Phenotyped by mass cytometry to assess post-vaccination T-cell activation detailed Figure 4D.

⁶ Virus blip at Day 70 (153 copies/mL), Day 86 (86 copies/mL) detailed Figure 4B

Bolded Participant Identifiers: initiated ART in acute HIV-1 infection, # PID01213 is a transgender woman.

Supplementary Table 2: Number of all solicited¹ and reportable unsolicited adverse events (including recurrences) by treatment arm.

Adverse Event	Treatment Arm				Total
	M3 (n=7)	M4 (n=7)	M3M4 (n=8 ¹)	Placebo (n=3)	
Solicited					
Injection Site Pain and, or tenderness	13	12	13	3	41
Headache	9	4	5	3	21
Fatigue	7	5	5	2	19
Malaise	9	5	3	2	19
Myalgia	5	7	4	1	17
Influenza Like Illness	7	2	4	1	14
Chills	4	4	3	0	11
Decreased Appetite	4	1	3	1	9
Nausea	4	1	4	0	9
Arthralgia	3	0	3	1	7
Hyperhidrosis	3	1	1	1	6
Dizziness	4	1	0	0	5
Abdominal Pain	1	0	2	0	3
Body Temperature Increased	2	1	0	0	3
Injection Site Swelling	3	0	0	0	3
Injection Site Warmth	0	1	2	0	3
Vomiting	1	0	1	0	2
Diarrhea	0	1	0	0	1
Injection Site Discoloration	0	1	0	0	1
Injection Site Erythema	0	0	1	0	1
Injection Site Induration	0	0	1	0	1
Unsolicited					
Procedural Hypertension	12	13	15	5	45
Blood Pressure Increased	0	3	1	0	4
Tissue Infiltration	0	1	3	0	4
Citrate Toxicity	1	0	2	0	3
Presyncope	1	0	0	1	2
Blood Creatinine Increased	1	0	0	0	1
Blood HIV RNA Increased	0	1	0	0	1
Erythema	0	1	0	0	1
Hyperhidrosis	1	0	0	0	1
Post Procedural Contusion	1	0	0	0	1
Post Procedural Hematoma	0	1	0	0	1
Procedural Dizziness	0	0	0	1	1
Procedural Pain	0	0	1	0	1
Pyrexia	0	0	0	1	1
Vessel Puncture Site Hemorrhage	0	0	0	1	1
Vessel Puncture Site Pain	0	0	0	1	1
Total	96	67	77	25	265

¹ In the M3M4 group, one participant was vaccinated with M3M4 and completed study visits but had received a COVID-19 vaccination within two weeks of enrollment. This participant was subsequently replaced.

Supplementary Table 3: Fold-change in Mos-1- and Mos-2-specific T-cell frequencies following vaccination measured by IFN- γ ex vivo ELISpot

Study Arm	Participant	Sex ¹	Age ²	Years on ART ³	Protective HLA Alleles ⁴	Fold change from Baseline ⁵					
						Day 7		Day 14		Day 70	
						Mos-1	Mos-2	Mos-1	Mos-2	Mos-1	Mos-2
M3	00836	M	33	7.83		8.877	6.364	8.282	6.277	7.835	5.885
	01237a	M	41	10.17		2.789	2.657	2.770	3.053	1.434	1.197
	01249	M	28	8.11		9.849	2.908	24.590	6.916	20.966	6.543
	00937	F	48	9.21		4.959	3.681	6.727	4.993	3.031	2.204
	00835	M	43	9.61		2.848	2.129	4.857	5.169	2.868	2.114
	01214	M	57	16.14		2.000	1.495	1.444	1.021	1.275	1.027
	01009	F	60	11.86	B*58:01	1.537	1.028	1.434	1.057	1.357	1.102
M4	00749	M	30	5.87		5.242	5.856	4.959	5.736	3.605	3.482
	01262	M	55	12.16		2.114	2.219	2.828	3.182	2.888	2.497
	01264	M	37	6.03		1.007	2.282	1.266	4.347	1.079	2.694
	01276	M	46	11.12		1.569	1.892	1.395	1.778	1.125	1.310
	01277	M	31	2.47	B*81:01	1.516	9.190	2.000	13.929	ND ⁶	ND
	01095	F	58	10.41		1.181	1.602	0.993	1.181	ND	ND
	00115	M	59	18.12		3.272	1.905	7.311	4.500	1.932	1.516
M3M4	00631	M	29	6.44		11.472	3.138	8.340	2.000	4.287	1.357
	00371	M	30	10.72	B*57:03, 58:01	7.674	3.364	3.864	2.549	2.189	1.892
	01153	M	55	12.71		3.364	5.897	3.482	7.062	2.532	4.627
	01280	M	54	6.55		ND	ND	0.722	0.883	0.993	1.165
	01213	M#	21	2.45		6.021	6.589	8.340	9.781	7.062	8.225
	01295	M	41	9.01	B*57:03	1.625	2.657	1.892	3.117	1.395	1.548
	01293	M	31	4.34		2.000	2.585	2.770	4.228	2.732	4.112
Placebo	00818	M	24	6.68		0.889	0.847	1.102	1.102	1.395	0.883
	01270	M	28	4.59		1.057	1.064	0.737	0.920	0.920	0.514
	01220	M	52	2.37		1.050	0.973	1.464	1.414	0.883	1.028
	M3, M4, M3M4 Vaccinees only: Median (IQR)					2.819 (3.579)	2.657 (1.848)	2.828 (3.987)	4.228 (4.013)	2.532 (1.721)	2.114 (2.121)
	Vaccinees with Non-Protective Alleles: Median (IQR)					3.060 (1.824)	2.621 (1.687)	3.482 (2.956)	4.347 (3.596)	2.800 (1.593)	2.350 (1.530)
	Vaccinees with Protective Alleles: Median (IQR)					1.581 (1.842)	3.010 (1.587)	1.946 (2.526)	2.833 (3.068)	1.395 (1.525)	1.548 (1.530)

¹ Sex at birth, M = male, F = female

² age (years) enrollment

³ years from antiretroviral therapy (ART) initiation to enrollment

⁴ Participant expresses one or more of the following HLA I alleles, B*57:01/03, B*58:01, B*81:01

⁵ Fold-change from the average of two pre-vaccination timepoints to indicated visit (see Methods)

⁶ ND – not done. No sample available for testing.

Bolded Participant Identifiers: initiated ART in acute HIV-1 infection # PID01213 is a transgender woman.

Supplementary Table 4: Detection of baseline HIVconsvX-specific T-cells following *in vitro* culture.

Participant	Ex vivo ELISpot				Short term cell lines (STCL) then ELISpot			
	Study Group	Sub-pool tested by ELISpot	Baseline (SFU/10 ⁶ PBMC) ¹	Post-vaccination (SFU/10 ⁶ PBMC) ²	Stimulating Peptide Pool	Sub-pool tested by ELISpot	Day -28 (SFU/10 ⁶ STCL)	Fold-expansion relative to baseline ex vivo frequency ³
836	M3	A (G1)	225	1180	Mos-1	A (G1)	2200	9.8
		C (P3-A)	≤20	69		C (P3-A)	≤20	-
		E (P6)	≤20	71		E (P6)	≤20	-
					Mos-2	A (G1)	2460	11
						C (P3-A)	469	23.4
						E (P6)	≤20	-
1237a	M3	A (G1)	106	300	Mos-1	A (G1)	329	3.1
		C (P3-A)	≤20	27		C (P3-A)	≤20	-
					Mos-2	A (G1)	≤20	-
						C (P3-A)	≤20	-
1249	M3	A (G1)	45	209	Mos-1	A (G1)	700	15.5
		C (P3-A)	≤20	61		C (P3-A)	≤20	-
		D (P3-B)	≤20	105		D (P3-B)	≤20	-
		E (P6)	≤20	143		E (P6)	794	39.7
					Mos-2	A (G1)	1065	24
						C (P3-A)	≤20	-
						D (P3-B)	300	15
						E (P6)	≤20	-
937	M3	A (G1)	76	451	Mos-1	A (G1)	≤20	-
		C (P3-A)	≤20	280		C (P3-A)	≤20	-
		E (P6)	≤20	178		E (P6)	≤20	-
					Mos-2	A (G1)	≤20	-
						C (P3-A)	≤20	-
						E (P6)	≤20	-
835	M3	A (G1)	219	513	Mos-1	A (G1)	3983	18.2
		E (P6)	≤20	160		E (P6)	794	39.7
					Mos-2	A (G1)	1827	8.3
						E (P6)	725	36.3
1214	M3	A (G1)	388	612	Mos-1	A (G1)	8700	22.4
		C (P3-A)	≤20	22		C (P3-A)	≤20	-
					Mos-2	A (G1)	8508	22
						C (P3-A)	150	7.5
1009	M3	A (G1)	1682	1731	Mos-1	A (G1)	1208	-
		C (P3-A)	≤20	45		C (P3-A)	≤20	-
		E (P6)	≤20	71		E (P6)	≤20	-
					Mos-2	A (G1)	1894	1.1
						C (P3-A)	≤20	-
						E (P6)	≤20	-
749	M4	A (G1)		288	Mos-1	A (G1)	≤20	-
		C (P3-A)		85		C (P3-A)	≤20	-
		D (P3-B)		212		D (P3-B)	≤20	-
					Mos-2	A (G1)	200	8.7
						C (P3-A)	≤20	-
						D (P3-B)	142	7.1
1262	M4	A (G1)	150	304	Mos-1	A (G1)	1529	10.2
		D (P3-B)	≤20	32		D (P3-B)	644	32.2
					Mos-2	A (G1)	1479	9.9
						D (P3-B)	300	15.0
1264	M4	A (G1)	296	374	Mos-1	A (G1)	≤20	-
		C (P3-A)	≤20	78		C (P3-A)	≤20	-

		D (P3-B)	≤20	105				
					Mos-2	D (P3-B)	≤20	-
						A (G1)	3225	10.9
						C (P3-A)	≤20	-
						D (P3-B)	≤20	-
1277	M4	A (G1)	859	1295	Mos-1	A (G1)	≤20	-
		D (P3-B)	≤20 ¹	275		D (P3-B)	≤20	-
		E (P6)	≤20 ¹	164		E (P6)	≤20	-
					Mos-2	A (G1)	≤20	-
						D (P3-B)	≤20	-
						E (P6)	≤20	-
1095	M4	A (G1) ¹	348	759	Mos-1	A (G1)	2771	8.0
		C (P3-A)	≤20	118		C (P3-A)	≤20	-
		E (P6)	≤20	198		E (P6)	1113	55.6
					Mos-2	A (G1)	2260	6.5
						C (P3-A)	≤20	-
						E (P6)	1760	88.0
631	M3M4	A (G1)	91	645	Mos-1	A (G1)	≤20	-
		B (G2+P4+P5)	≤20	263		B (G2+P4+P5)	≤20	-
		C (P3-A)	≤20	67		C (P3-A)	≤20	-
		D (P3-B)	≤20	116		D (P3-B)	≤20	-
		E (P6)	≤20	125		E (P6)	≤20	-
					Mos-2	A (G1)	1656	18.2
						B (G2+P4+P5)	≤20	-
						C (P3-A)	≤20	-
						D (P3-B)	≤20	-
						E (P6)	≤20	-
371	M3M4	A (G1)	759	2498	Mos-1	A (G1)	2331	3.1
		B (G2+P4+P5)	≤20	144		B (G2+P4+P5)	≤20	-
					Mos-2	A (G1)	2929	3.9
						B (G2+P4+P5)	≤20	-
1153	M3M4	A (G1)	46	192	Mos-1	A (G1)	250	5.4
		E (P6)	≤20	50		E (P6)	213	10.6
					Mos-2	A (G1)	310	6.7
						E (P6)	≤20	-
1213	M3M4	A (G1)	81	414	Mos-1	A (G1)	1581	19.5
		C (P3-A)	≤20	92		C (P3-A)	≤20	-
					Mos-2	A (G1)	1825	22.5
						C (P3-A)	≤20	-
1295	M3M4	A (G1)	153	401 (D14)	Mos-1	A (G1)	3135	20.5
		B (G2+P4+P5)	≤20	48 (D14)		B (G2+P4+P5)	≤20	-
		D (P3-B)	≤20	27 (D14) ⁶		D (P3-B)	≤20	-
					Mos-2	A (G1)	3129	20.5
						B (G2+P4+P5)	≤20	-
						D (P3-B)	≤20	-
1293	M3M4	A (G1)	645	2113 (D14)	Mos-1	A (G1)	11223	17.4
		C (P3-A)	≤20	73 (D14) ⁵		C (P3-A)	94	4.7
					Mos-2	A (G1)	12567	19.5
						C (P3-A)	369	18.4

¹ Baseline T-cell frequency is the average of two pre-vaccination visits. ² Day 7 post-MVA.HIVconsVX visit unless otherwise specified in parentheses. Note: A HIVconsVX sub-pool was assigned positive post-vaccination only if positivity criteria for ex vivo ELISpot (Methods) were met at 2 or more post-vaccination visits. ³ Fold-change calculated from baseline T-cell frequency (the average of two pre-vaccination visits, details Methods). For sub-pools <20 SFU/10⁶ PBMC and/or had a replicate value of '0', a value of 20 SFU was assigned to calculate fold change. ⁴ In all participants, a positive ex vivo T-cell response (details Methods) was detected against sub-pool A, G1. ⁵ PID1293, T-cell response in ex vivo ELISpot to sub-pool C was negative at Day 7 but positive at Day 14 (shown) ⁶ PID1295, T-cell response in ex vivo ELISpot to sub-pool D was negative at Day 7 but positive at Day 14 (shown). **Bold** = Positive HIV-specific T-cell response to HIVconsVX in cultured ELISpot. Criteria: > 200 SFU/10⁶ cells, > 3-fold above mock-stimulated cells. For sub-pools B-E, a > 3.5-fold expansion to sub-pool A (G1) in the same parent STCL was required as a positive control for HIV-specific expansion of T-cells.

Supplementary Table 5: T-cell frequencies measured by ex vivo IFN-γ ELISpot against autologous HIV sequences.

Participant - group	HXB2 Region	Autologous Virus Sequence (predicted HLA restriction) ¹	# autologous viruses in which sequence occurs	Sequence in HIVCONSX immunogen ² (Y/N)	%Seq. Identity with Mos-1 peptide ²	%Seq. Identity with Mos-2 peptide ²	%Seq. Identity with Clade B peptide ³	Ex vivo ELISpot SFU/10 ⁶ , (SEM) ⁴	
								Pre-vaccination Visit 3 - Baseline	Post-MVA.HIVconsvX Vaccination Visit 8 - Day 28
00371-M3M4	Gag 240→249	TSTLQEQIGW ⁴ (HLA-B*57:03, B*58:01)	3/3	Y	90.00%	100.00%	100.00%	321 (21)	882 (10)
	Gag 244→261	QEQIGWMTNNPPIPVGEI	3/3	Y	88.88%	100.00%	100.00%	276 (15)	875 (31)
	Gag 300→317	FYKTLRAEQASQDVKNWM	3/3	Y	94.44%	83.33%	94.44%	23 (3)	13 (5)
	Gag 196→213	AAMQMLKETINEEAAEWD	3/3	Y	94.44%	100.00%	100.00%	21 (4)	12 (3)
	Pol 873→890	LKKIIGQVRDQAEHLKTA	3/3	Y	94.44%	100.00%	100.00%	38 (8)	75 (3)
00749-M4	Pol 481→498	IAEIQKQGQGWYQIYQ	15/15	Y	94.44%	100.00%	100.00%	146 (15)	395 (23)
	Pol 929→946	QKQISKIQNFRVYYRDNR	15/15	Y	88.88%	88.88%	88.88%	60 (9)	183 (12)
	Pol 969→986	NSEIKVVP RRKAKIIRDY	15/15	Y	88.88%	100.00%	94.44%	53 (4)	183 (10)
	Vif 32→40, _{36K}	SKKCKGWFY ⁴	14/16	N	N/A	N/A	77.77%	44 (8)	146 (4)
	Vif 32→40, _{36N}	SKKCKGWFY ⁴ (HLA-B*15:03)	2/16	N	N/A	N/A	66.66%	0	4 (2)
01095-M4	Pol 313→321, _{320T}	AIFQSSMTK ⁴	32/35	Y	100.00%	88.88%	100.00%	559 (20)	795 (12)
	Pol 313→321, _{320I}	AIFQSSMTK ⁴ (HLA-A*03:01, A*11:01)	3/35	Y	88.88%	77.77%	88.88%	411 (32)	593 (27)
	Pol 273→290	VPLDKDFRKYTAFTIPSI	35/35	Y	88.88%	94.44%	100.00%	393 (15)	498 (19)
	Gag 397→414, _{406K,411K}	REGHIAKNCKAPRKKG CW	8/35	Y	83.33%	77.77%	88.88%	68 (15)	52 (7)
	Gag 397→414, _{406R,411R}	REGHIAKNCKAPRKRG CW	5/35	Y	94.44%	77.77%	88.88%	77 (10)	61 (14)
	Gag 397→414, _{406R,411K}	REGHIAKNCKAPRKKG CW	19/35	Y	88.88%	83.33%	94.44%	55 (4)	58 (7)
	Pol 353→370, _{355A,359E,366K}	HRAKIEELRQHLLK WGFT	11/35	Y	94.44%	88.88%	88.88%	76 (14)	105 (7)
	Pol 353→370, _{355I,359E,366K}	HRKIEELRQHLLK WGFT	6/35	Y	88.88%	88.88%	88.88%	76 (14)	108 (8)
	Pol 353→370, _{355I,359E,366R}	HRKIEELRQHLLR WGFT	2/35	Y	83.33%	94.44%	94.44%	89 (10)	103 (7)
	Pol 353→370, _{355A,359K,366K}	HRAKIEELRQHLLK WGFT	1/35	Y	88.88%	83.33%	83.33%	4 (SEM)	5 (SEM)
	Pol 353→370, _{355A,359E,366R}	HRAKIEELRQHLLR WGFT	15/35	Y	88.88%	94.44%	94.44%	84 (5)	82 (9)

¹ HIV outgrowth assays were performed at limiting dilution in each participant prior to vaccination. Near full-length virus sequences were produced using PacBio sequencing (see Methods). 00749 and 01095 sequences were previously published in (42). Sequences deposited GenBank.² HIVconsvX sequences published in (7) ³ Clade B sequence (www.lanl.gov)⁴ Each participant was first mapped against overlapping peptides spanning HIV Clade B. The corresponding 18-mer autologous outgrowth reservoir virus peptide/s for all previously defined reactive 18-mers from the initial mapping ELISpot were also synthesized. In addition, optimal CD8 T-cell peptides predicted through LANL's epitope location finder ('ELF') software based on HLA-prediction software (https://www.hiv.lanl.gov/content/sequence/ELF/epitope_analyzer.html), and 18-mer peptides spanning regions where > 40% of the sequences contain non-synonymous mutations in the same position were synthesized. These autologous sequences were then tested in ex vivo ELISpot pre- and post-vaccination to produce the final list of reactive autologous epitopes within each participant. 'Negative' data to variant sequences are also shown.

⁵ Empirically confirmed optimal CD8 T-cell epitope (restricting HLA allele).

Supplementary Table 6: Mos-1- and Mos-2-specific CD4+ and CD8+ T cells detected by intracellular cytokine staining (ICS) measured post-vaccination.

Study Arm	Participant	Set at birth	Race ¹	Ethnicity ²	Days from vaccination ³	ICS (background -subtracted frequency)							
						CD4 T cells				CD8 T cells			
						IFN-g +		IFN-g+CD107a+		IFN-g+		IFN-g+CD107a+	
						Mos-1	Mos-2	Mos-1	Mos-2	Mos-1	Mos-2	Mos-1	Mos-2
M3	00836	M	AA	NH	7	0.158%	0.174%	0.074%	0.089%	<i>0.021%</i>	<i>0.016%</i>	0.026%	0.017%
	01237a	M	U	H	56	0.045%	0.128%	<i>0.012%</i>	<i>0.016%</i>	0.160%	0.123%	0.150%	0.093%
	01249	M	C	NH	14	0.258%	0.252%	0.033%	0.036%	0.024%	0.034%	<i>0.011%</i>	<i>0.007%</i>
	00937	F	AA	NH	56	0.106%	0.058%	0.017%	<i>0.009%</i>	0.203%	0.118%	0.209%	0.119%
	00835	M	C	NH	7	0.104%	0.064%	0.015%	0.015%	0.100%	0.026%	0.098%	0.028%
	01214	M	C	NH	7	0.086%	<i>0.051%</i>	<i>0.022%</i>	<i>0.016%</i>	<i>0.0%</i>	<i>0.005%</i>	<i>0.006%</i>	<i>0.020%</i>
	01009 ¹	F	AA	NH	7	0.041%	<i>0.027%</i>	0.007%	<i>0.005%</i>	0.100%	0.100%	0.098%	0.108%
M4	00749	M	AA	NH	7	<i>0.043%</i>	<i>0.037%</i>	<i>0.003%</i>	<i>0.004%</i>	0.123%	0.477%	0.065%	0.246%
	01262	M	C	H	56	0.045%	0.047%	<i>0.014%</i>	<i>0.015%</i>	<i>0.023%</i>	<i>0.008%</i>	0.056%	0.036%
	01264	M	AA	NH	56	0.088%	0.126%	<i>0.001%</i>	<i>0.000%</i>	0.432%	0.081%	0.278%	0.056%
	01276	M	AA	NH	7	0.015%	0.026%	<i>0.006%</i>	<i>0.011%</i>	0.182%	0.217%	0.186%	0.216%
	01277 ¹	M	AA	NH	14	0.264%	0.298%	0.017%	0.019%	0.369%	0.272%	0.368%	0.268%
	01095	F	C	NH	14	0.113%	0.085%	0.012%	0.018%	0.622%	0.058%	0.621%	0.061%
	00115	M	C	NH	7	0.165%	0.139%	0.040%	0.039%	<i>0.0%</i>	0.133%	0.018%	0.127%
M3M4	00631	M	AA	NH	7	0.068%	0.092%	0.039%	0.050%	0.049%	0.096%	0.050%	0.085%
	00371 ¹	M	AA	NH	7	0.082%	0.095%	<i>0.0%</i>	<i>0.000%</i>	0.050%	0.424%	<i>0.013%</i>	0.180%
	01153 ²	M	C	NH	7	<i>0.037%</i>	<i>0.049%</i>	<i>0.004%</i>	<i>0.008%</i>	<i>0.011%</i>	<i>0.005%</i>	<i>0.003%</i>	<i>0.003%</i>
	01280	M	C	NH	56	0.081%	0.088%	0.030%	0.024%	<i>0.0%</i>	<i>0.0%</i>	<i>0.003%</i>	<i>0.005%</i>
	01213	F	AA	NH	14	0.090%	0.086%	<i>0.006%</i>	<i>0.007%</i>	<i>0.008%</i>	<i>0.009%</i>	<i>0.0%</i>	0.009%
	01295 ¹	M	AA	NH	14	<i>0.021%</i>	0.055%	<i>0.001%</i>	<i>0.000%</i>	0.049%	0.065%	0.058%	0.055%
	01293	M	AA	NH	56	0.438%	0.427%	<i>0.013%</i>	<i>0.010%</i>	0.642%	0.378%	0.460%	0.290%

¹ Participant expresses one or more of the following HLA I alleles, B*57:01/03, B*58:01, B*81:01 (detailed Supplementary Table 1)

² 001153 ICS performed on Day 7 post-vaccination. Average T-cell responses measured by IFN- γ ELISpot at baseline (average two pre-vaccination visits) was Mos-1 105 SFU/10⁶ and Mos-2 39 SFU/10⁶ and average on Day 7 for Mos-1 was 354 SFU/10⁶ and Mos-2 was 221 SFU/10⁶

³ To maximize assay sensitivity, ICS was performed at or near the peak measured HIVconsvX-specific T cell response by IFN- γ ELISpot.

Bold = Mos-1- or Mos-2-specific T-cell response defined by $\geq 2 \times$ mock-stimulated cells and functional gate of Mos-1- or Mos-2-stimulated cells had ≥ 25 events than corresponding functional gate of mock stimulated cells. Negative measurements indicated in *gray italic*.

Supplementary Table 7: The association of age at enrollment on T-cell response to MVA.HIVconsvX vaccination in PWH on ART¹ estimated from simple linear regression

	Age	Mos-1	Mos-2
Fold-change per 10 person years from baseline to D14 post-vaccination	20	7.30	7.64
	30	5.06	5.32
	40	3.50	3.70
	50	2.43	2.58
	60	1.68	1.79
Fold-change per 10 person years from baseline to D70 post-vaccination	20	5.95	4.61
	30	4.05	3.37
	40	3.50	2.46
	50	1.88	1.80
	60	1.68	1.31

Supplementary Table 8: Age at enrollment and years on ART predict post-vaccination T-cell response following MVA.HIVconsvX vaccination

Antigen	log2FC from baseline to day	Predictors	Slope Estimate ¹	p-value ²	Multiple R2
Mos-1	7	Age	-0.046	1.74 ×10 ⁻²	0.28
	14	Age	-0.053	1.64 ×10 ⁻²	0.27
	70	Age	-0.056	6.90 ×10 ⁻³	0.36
	7	Age Years on ART	-0.109 0.236	5.03 ×10 ⁻⁴ 7.63 ×10 ⁻³	0.53
	14	Age Years on ART	-0.110 0.233	6.24 ×10 ⁻⁴ 1.15 ×10 ⁻²	0.49
	70	Age Years on ART	-0.075 0.082	1.79 ×10 ⁻² ns	0.39
	7	Age Pre-vax logIPDA	-0.030 -0.501	ns ns	0.39
	14	Age Pre-vax logIPDA	-0.034 -0.526	ns ns	0.35
	70	Age Pre-vax logIPDA	-0.047 -0.381	ns ns	0.42
	7	Age	-0.042	2.54 ×10 ⁻³	0.41
	14	Age	-0.052	5.52 ×10 ⁻³	0.34
	70	Age	-0.045	8.13 ×10 ⁻³	0.35
	7	Age Years on ART	-0.028 -0.052	ns ns	0.43
	14	Age Years on ART	-0.055 0.011 ns	ns ns	0.34
	70	Age Years on ART	-0.038 -0.030	ns ns	0.35
Mos-2	7	Age Pre-vax logIPDA	-0.040 0.035	0.018 ns	0.36
	14	Age Pre-vax logIPDA	-0.050 0.142	0.024 ns	0.32
	70	Age Pre-vax logIPDA	-0.043 -0.067	ns ns	0.31

¹ Models are simple or multiple linear regression estimates

² ns - non-significant p>0.05

FC=fold change, ART antiviral therapy

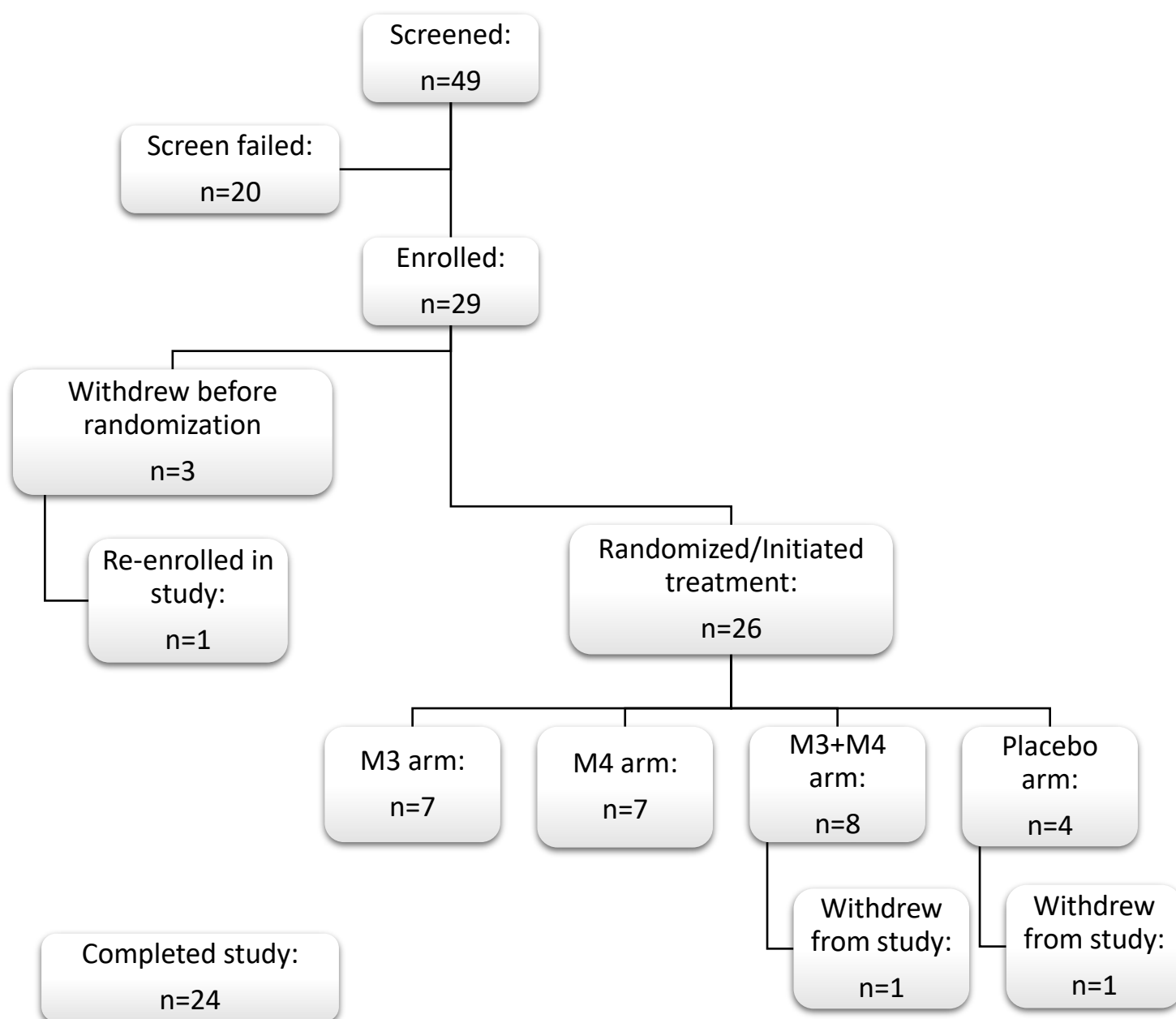
Supplementary Table 9: Key Reagents and Resources

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Antibodies – flow cytometry		
CD107a-APC	Biolegend	Cat# 328620, RRID:AB_1279055
CD3-PerCP-Cy5.5	Biolegend	Cat# 300430, RRID:AB_893299
CD4-PE-Cy5	Biolegend	Cat# 317412, RRID:AB_571957
CD8-BV510	Biolegend	Cat# 344732, RRID:AB_2564624
CD14-BV650	Biolegend	Cat# 301836, RRID:AB_2563799
CD16-BV650	Biolegend	Cat# 302042, RRID:AB_2563801
CD19-BV650	Biolegend	Cat# 302238, RRID:AB_2562097
CD56-BV650	Biolegend	Cat# 318344, RRID:AB_2563838
CD45RO-BV605	Biolegend	Cat# 304238, RRID:AB_2562153
IFN- γ -PE	Biolegend	Cat# 502509, RRID:AB_315234
TNF α -PE-Dazzle 594	Biolegend	Cat# 502946, RRID:AB_2564173
MIP-1 β -PE-Cy7	BD Biosciences	Cat# 560687, RRID:AB_1727566
Perforin-BV421	Biolegend	Cat# 353307, RRID:AB_11149688
Granzyme B-FITC	Biolegend	Cat# 372206, RRID:AB_2687030
Zombie NIR	Biolegend	Cat# 423106
Antibodies – mass cytometry		
CD45 HI30 111Cd	Standard BioTools	Cat# 3111001B
CCR6 11A9 141Pr	Standard BioTools	Cat# 3141014A
CD19 HB19 142Nd	Standard BioTools	Cat# 3142001B
CD45RA HI100 143Nd	Standard BioTools	Cat# 3143006B
CD4 RPA-T4 145Nd	Standard BioTools	Cat# 3145001B
CCR4 L291H4 149Sm	Standard BioTools	Cat# 3149029A
CD14 M5E2 151Eu	Standard BioTools	Cat# 3151009B
CXCR5 RF8B2 153Eu	Standard BioTools	Cat# 3153020B
PD-1 EH12.2H7 155Gd	Standard BioTools	Cat# 3155009B
CXCR3 G025H7 156Gd	Standard BioTools	Cat# 3156004B
CD27 L128 158Gd	Standard BioTools	Cat# 3158010B
FoxP3 259D/C7 159Tb	Standard BioTools	Cat# 3159028A
CD28 CD28.2 160Gd	Standard BioTools	Cat# 3160003B
Ki-67 B56 162Dy	Standard BioTools	Cat# 3162012B
CD95 DX2 164Dy	Standard BioTools	Cat# 3164008B
CD127 A019D5 165Ho	Standard BioTools	Cat# 3165008B
CCR7 G043H7 167Er	Standard BioTools	Cat# 3167009A
CD8 SK1 168Er	Standard BioTools	Cat# 3168002B
CD25 2A3 169Tm	Standard BioTools	Cat# 3169003B
HLA-DR L243 170Er	Standard BioTools	Cat# 3170013B
CCR5 NP-6G4 171Yb	Standard BioTools	Cat# 3171017A
CD38 HIT2 172Yb	Standard BioTools	Cat# 3172007B
CD56 N901 176Yb	Standard BioTools	Cat# 3172007B
CD16 3G8 209Bi	Standard BioTools	Cat# 3209002B
CD45 HI30 89Y	Standard BioTools	Cat# 3089003B
CD31 WM59 144Nd	Standard BioTools	Cat# 3144023B
CD3 UCHT1	Biolegend	Cat# 300443
CD99 3B2/TA8	Biolegend	Cat# 371302
Bcl-2 100	Biolegend	Cat# 658702
TCR $\gamma\delta$ B1	Biolegend	Cat# 331202
CD57 HCD57	Biolegend	Cat# 359602

CCR10 314305	R&D systems	Cat# MAB3478
CD3 UCHT1 AF488	Biolegend	Cat# 300415, RRID: AB_389310
CD4 OKT4 BV650	Biolegend	Cat# 317436, RRID: AB_2563050
CD8 SK1 BV510	Biolegend	Cat# 344732, RRID: AB_2564624
CD14 M5E2 PerCP Cy5.5	Biolegend	Cat# 325622, RRID: AB_893250
CD19 HIB19 PerCP Cy5.5	Biolegend	Cat# 302230, RRID: AB_2073119
CD56 HCD56 PerCP Cy5.5	Biolegend	Cat# 318322, RRID: AB_893389
CD16 3G8 PerCP Cy5.5	Biolegend	Cat# 302028, RRID: AB_893262
Biological samples		
Human PBMCs	Human	Assigned de-identifying code
Buffy Coats	Human	Not applicable
Tissue Culture		
Fetal Calf Serum (ex vivo studies)	Corning	Cat# 35-079-CV
Human AB serum (short term cell lines)	GemCell™	SKU# 100-512-100
IL-2	PeptoTech	Cat# 200-02-50UG
RPMI-1640	Gibco	Cat# 61870036
L-Glutamine	VWR	Cat # 97068-088
Sodium Pyruvate	Gibco	Cat# 11360070
Chemicals, peptides, and recombinant proteins		
Phytohemagglutinin	ThermoFisher	Cat# 10576015
Cell-ID Intercalator-Ir	Standard BioTools	Cat# 201192A
Cell Stimulation Cocktail (500X) (PMA / Ionomycin)	eBioscience™	00497093
Peptides	Sigma-Aldrich	VC80023
Cell-ID Cisplatin-195Pt	Standard BioTools	Cat# 201195
Critical commercial reagents		
Maxpar X8 Antibody Labeling 147Sm	Standard BioTools	Cat# 201147A
Maxpar X8 Antibody Labeling 152Sm	Standard BioTools	Cat# 201152A
Maxpar X8 Antibody Labeling 173Yb	Standard BioTools	Cat# 201173A
Maxpar X8 Antibody Labeling 175Lu	Standard BioTools	Cat# 201175A
Maxpar X8 Antibody Labeling 166Er	Standard BioTools	Cat# 201166A
Ca ²⁺ Mg ²⁺ free PBS	Rockland	Cat# MB008
Maxpar Cell Staining Buffer	Standard BioTools	Cat# 201068
Human TruStain FcX	Biolegend	Cat# 422301
FoxP3 Fixation/Permeabilization kit	ThermoFisher	Cat# 00-5523-00
Zombie NIR Fixable Viability Kit	Biolegend	Cat# 423106
CD3/28 Human Dynabeads	Gibco™	Cat# 11131D
CD4+ T cell isolation kit	Miltenyi Biotec	Cat# 130-096-533
Deposited data		
Preprocessed mass cytometry data	Goonetilleke Group, UNC	Cytobank Premium, Beckman Coulter
GenBank Sequencing data	Swanstrom /Zhou Groups, UNC.	Gen Bank under PRJNA666896, MT307344-MT308415 and MW054719-MW054856 All data generated (raw)
Software and Algorithms		
Prism v9	Graphpad	https://www.graphpad.com/
FlowJo v10.8.1	BD Biosciences	https://www.flowjo.com/

Python v3.6	Python Software Foundation	https://www.python.org/
Mixcr v2.1.9.6	Bolotin et al. (43, 44)	https://github.com/milaboratory/mixcr/tree/v2.1.9
R v4.2.2	R Core Team	https://cran.r-project.org/
Cytobank v9.3	Cytobank	https://unc.cytobank.org/cytobank
Muscle v5.1	(45)	https://github.com/rcedgar/muscle/releases
Ninja v1.2.2	https://wheelerlab.org/software/ninja/files/NINJA_preprint.pdf	https://github.com/ninja-build/ninja/releases

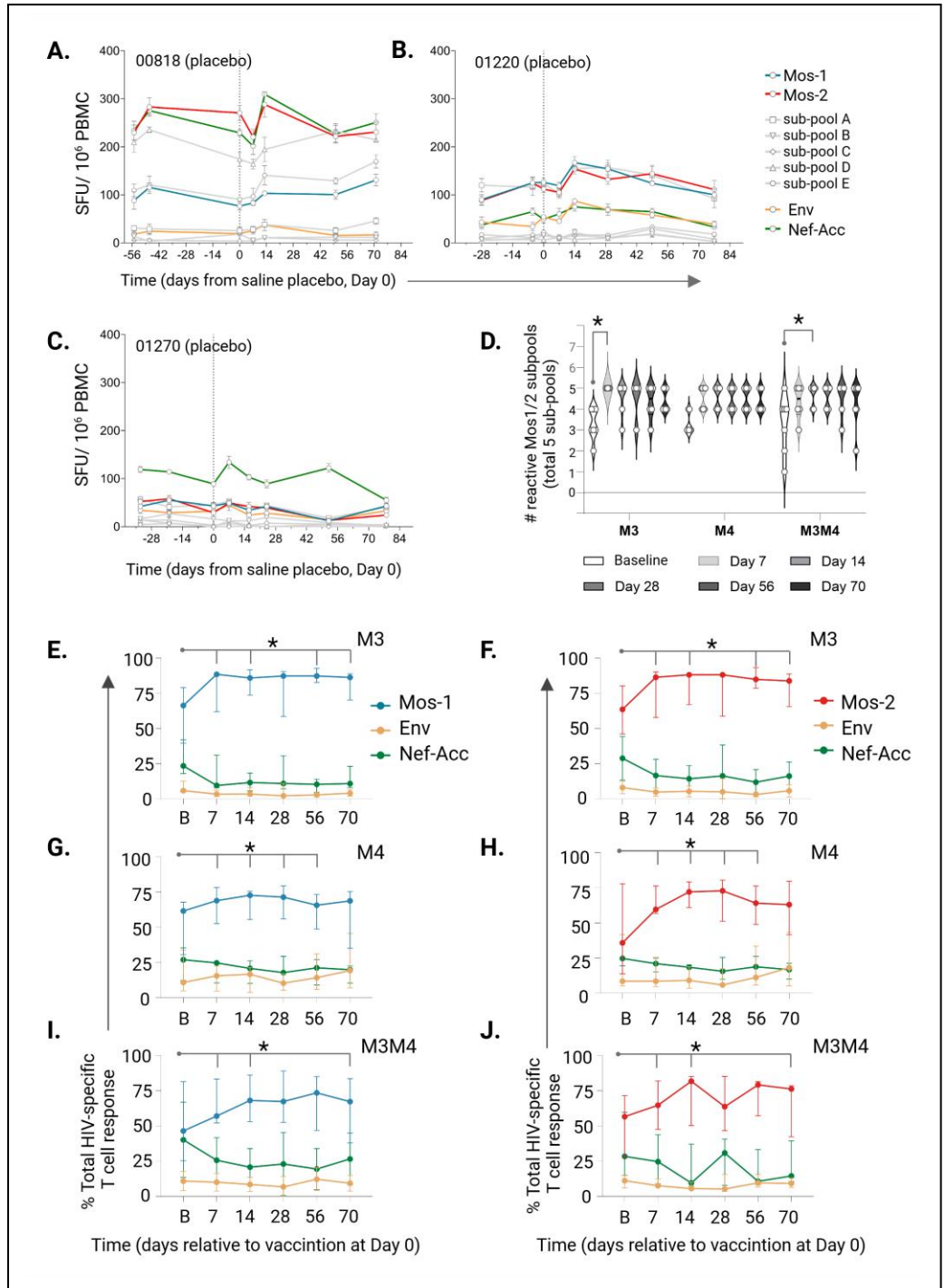
Supplementary Figure 1: M&M Study Flowchart of participants enrollment



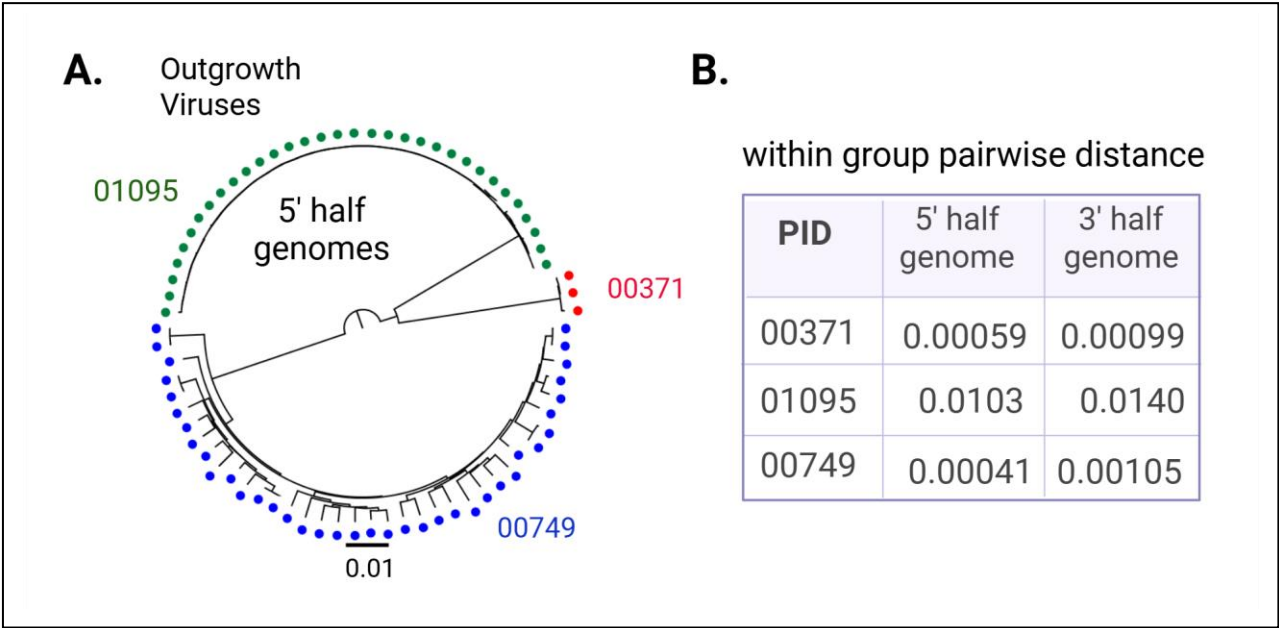
Supplementary Figure 2:
Vaccination with MVA-
vectored HIVconsVX
immunogens produce
significant increases in the
frequency of HIV-1-specific
T-cells. (A-C) Frequency of
 HIV-1-specific T-cells (mean
 +/- SEM) measured by ex
 vivo IFN- γ ELISpot in
 participants **(A)** 00818, **(B)**
 01220 and **(C)** 01270
 receiving saline placebo at
 day 0. **(D)** Within vaccine
 arm (n=7/group) changes in
 T-cell breadth from baseline.
 Average and 95% CI. **(E-J)**
 Median and IQR of %Mos-1
 (left, E, G, I) and %Mos-2
 (right, F, H, J) T-cell
 frequencies relative to total
 measured HIV-1-specific T-
 cell frequency in each
 vaccine group (n=7/group).
 %Env and %NEF-ACC also
 shown. Total frequency =
 sum of (Mos-1 or 2), Env,
 NEF and ACC.

Baseline was defined as the
 average of two pre-
 vaccination visits, at Day 0
 and a previous visit mostly
 occurring between days -28
 to -7. ACC in NEF/ACC
 peptide pool spans clade B
 Rev, Tat, Vif, Vpu, Vpr. (D-J)

Statistically significant
 comparisons (*) between baseline (B) and post-vaccination visits are identified with downward ticks, Wilcoxon
 signed-ranked test. * = $p < 0.05$



Supplementary Figure 3: (A) Phylogenetic trees displaying 5' half genome sequences for 00749 (blue), 01905 (green) and 00371 (red). The genetic distance is shown by scale bar (indicated with an asterisk), 1 substitution out of 1000 bp. Trees are midpoint-rooted. (B) Group pairwise distance for 5' and 3' genomes.



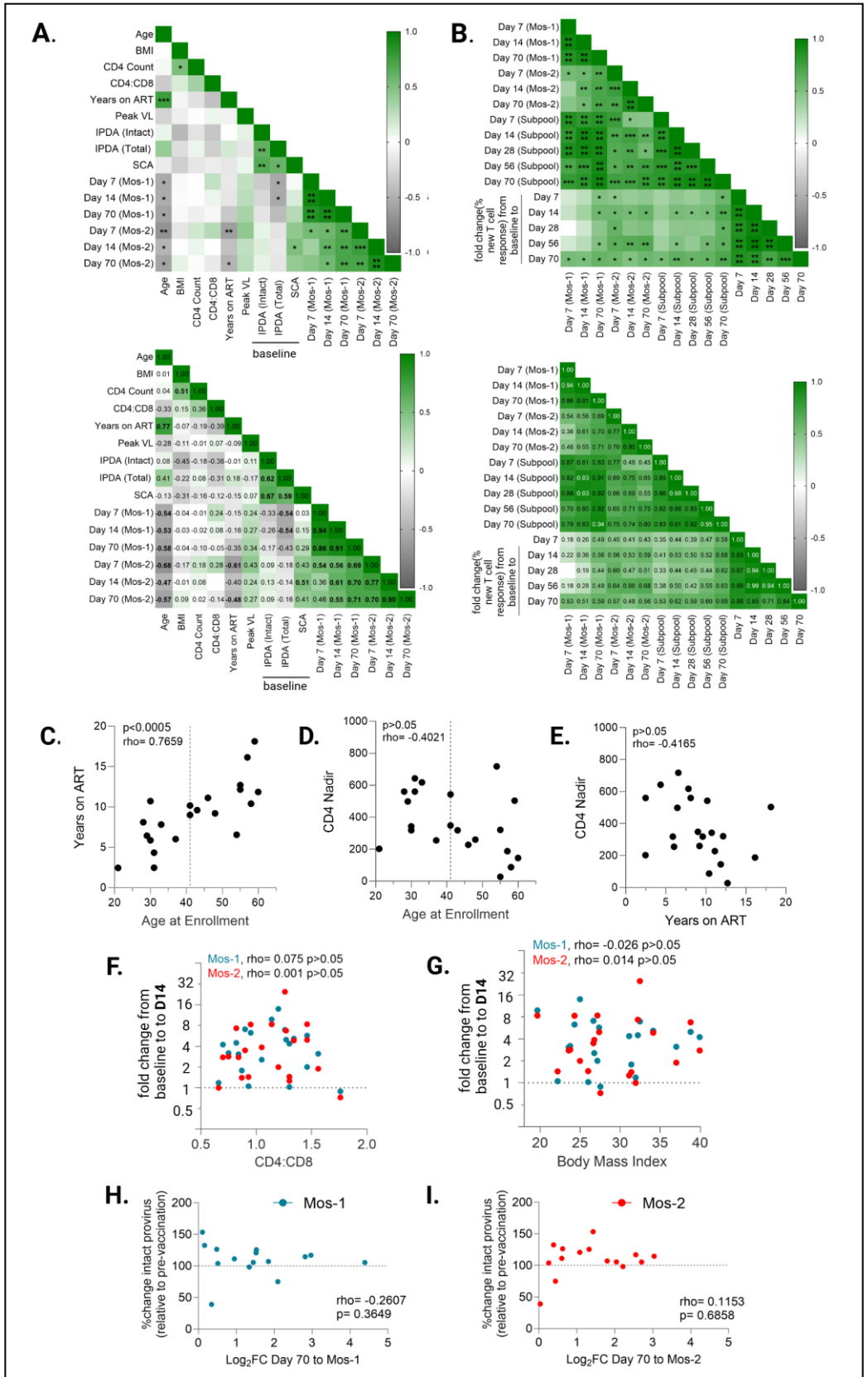
**Supplementary Figure 4:
Demographic and clinical
data, particularly age,
were associated with T-
cell response to
vaccination with
MVA.HIVconsVX vaccines.**

(A-I) All associations
Spearman Rank, two-tailed.

(A) Correlation matrix
between fold-change in T-
cell response to Mos-1 and
Mos-2 from baseline
(average of two pre-
vaccination visits) to
indicated post-vaccination
visit, demographic data and
baseline measurements of
persistent viremia. Upper
panel shows significance
levels, lower panel rho
values. Age = age at
enrolment, BMI = body mass
index, SCA = single copy
assay, IPDA = integrated
proviral DNA assay. CD4
count and CD4:CD8 =
values at enrolment
(B) Correlation matrix
between fold-change in T-
cell response to Mos-1 and
-2, Mos-1/2 sub-pools and
%new T cells detected
following vaccination across
all vaccinees (n=21). Upper
panel shows significance
levels, lower panel rho
values. New T-cells were
defined by T-cell response
to Mos1/2 sub-pools not
detected at baseline
timepoints but detected at
two or more post-vaccination
timepoints. **(C-G)**

Associations between
selected clinical and or
demographic data in
vaccinees (n=21). **(H-I)**
%Change in intact IPDA®
from pre-vaccination

(median day -7) to post-vaccination (median day 28) versus log2FC in frequency of Mos-1 **(H)** and Mos-2 **(I)** T-
cell frequency for vaccinees (n=17). * = $p < 0.05$, ** = $p < 0.005$, *** = $p < 0.0005$, **** = $p < 0.00005$



Supplementary Figure 5: Gating strategy for intracellular cytokine staining. Fluorescence minus one (FMO) controls were used to inform gates for functional markers.

