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1 **Serum cAMP levels are increased in patients with asthma**

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23 As a cornerstone in the treatment of asthma, β_2 -agonists prevent or reverse the shortening
24 of human airway smooth muscle (HASM), the pivotal cell regulating bronchomotor tone. β_2 -
25 agonists act upon β_2 -adrenoceptor (β_2 AR)—the cognate G_s -coupled G protein-coupled receptor
26 (G_s -GPCR) expressed on HASM—and activate adenylyl cyclase which generates 3',5'-cyclic
27 adenosine monophosphate (cAMP) (1). Increased intracellular cAMP levels ($[cAMP]_i$)
28 consequently stimulate protein kinase A that in turn modulates multiple downstream targets to
29 promote HASM relaxation and reverse airflow obstruction (2).

30 Classically, the signal transduction evoked by β_2 ARs is short-lived and multiple
31 mechanisms ensure homeostatic regulation of $[cAMP]_i$, with phosphodiesterase (PDE)
32 degradation of cAMP considered to play a dominant role. Using primary HASM cells in culture as
33 a model, we recently reported that β_2 AR activation evokes cAMP egress to the extracellular space
34 ($[cAMP]_e$) that is long-lived in culture, independent of PDE activity, and mediated by ABCC1 (ATP-
35 binding cassette subfamily C member 1) membrane transporter (3). Inhibition of ABCC1 activity
36 or expression decreases cAMP egress, increases $[cAMP]_i$ and enhances HASM relaxation
37 elicited by structurally diverse agonists acting upon G_s -GPCRs (3). These findings suggest a
38 class effect of G_s -GPCR activation and identify ABCC1 as previously unrecognized cAMP signal
39 response modifier in HASM. Of note, in a small cohort of patients with and without asthma, we
40 detected increased cAMP levels in the blood of patients with asthma (3).

41 To further explore the clinical utility of detecting circulating cAMP, we measured cAMP
42 levels in a serum biobank from the Severe Asthma Research Program (SARP)-3 (4). For this
43 study, we obtained 87 serum samples of patients with asthma, of which 39 are characterized as
44 “severe” according to European Respiratory Society / American Thoracic Society criteria for
45 asthma severity (Supplementary Table 1–2) (5). Since SARP-3 did not have a sufficient serum
46 biobank of healthy controls, we leveraged the database from the Rutgers Corona Cohort (RCC)
47 study (6) and obtained 273 serum samples of the study participants without a known history of

48 asthma or other lung diseases shown in Supplementary Table 1 (see detailed inclusion and
49 exclusion criteria in Supplementary Information).

50 We detected a high variability or a wide spread of cAMP levels in 87 serum samples of
51 patients with asthma (Figure 1A), ranging from 0.291 to 563.9 picomole. In contrast, the range of
52 cAMP levels in 273 serum samples of individuals without asthma was markedly smaller (0 to
53 27.72 picomole) (Figure 1A). Compared with non-asthma group (0.520 picomole, median), serum
54 cAMP levels were significantly higher in patients with asthma (6.220 picomole, median)
55 (Supplementary Table 1). To further test the hypothesis that cAMP levels can differentiate asthma
56 severity (severe versus nonsevere) in the SARP-3 samples, we applied linear regression models
57 with age and sex as covariates across clinical groups. There was no significant difference of
58 serum cAMP levels between severe asthma and nonsevere asthma (Supplementary Table 2), but
59 each asthma group showed significantly higher cAMP levels (adjusted $P < 0.00001$) than non-
60 asthma group (Figure 1B).

61 Using the SARP-3 data, we next assessed whether measured serum cAMP levels are
62 associated with any clinical traits of asthma. Specifically, we asked whether serum cAMP levels
63 had associations with: 1) asthma endotypes; 2) poor control indicators; and 3) post-bronchodilator
64 airflow reversibility. There were no significant differences in cAMP levels among or between
65 groups stratified by eosinophilic or neutrophilic asthma (Supplementary Figure 1) and any of the
66 poor control indicators (Supplementary Figure 2). In addition, we did not detect significant
67 differences in serum cAMP levels with maximum FEV₁ reversibility with albuterol (Supplementary
68 Figure 3A). Of note, serum cAMP levels arithmetically increased with the number of inhaled
69 corticosteroids (ICS) puffs and controllers used (Supplementary Figure 1) and, in nonsevere
70 asthma group, increased with the increases of post-bronchodilator lung function (Supplementary
71 Figure 3B-C). Further studies are necessary to explore the link between serum cAMP levels with:
72 1) bronchodilator or treatment responses by asthma severity; 2) ABCC1 expression and activity
73 in health and disease, including specific cell types of origin; and, 3) whether these physiological

74 outcomes and clinical phenotypes are affected by variations in ABCC1 genotypes in a large cohort
75 of patients with and without asthma.

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86 These companies had no role in study design or data analysis, and the only restriction on the
87 funds was that they be used to support the SARP initiative.

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90 **References**

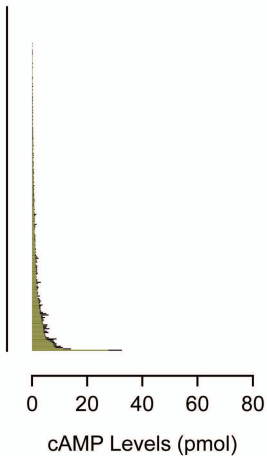
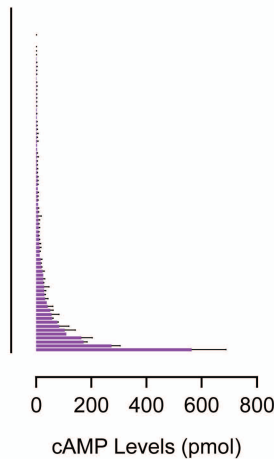
- 91 1. Limbird LE, Lefkowitz RJ. Resolution of beta-adrenergic receptor binding and adenylate
92 cyclase activity by gel exclusion chromatography. *J Biol Chem.* 1977;252(2):799–802.
- 93 2. Morgan SJ, et al. β -agonist-mediated relaxation of airway smooth muscle is protein kinase A-
94 dependent. *J Biol Chem.* 2014;289(33):23065–23074.
- 95 3. Cao G, et al. Inhibition of ABCC1 decreases cAMP egress and promotes human airway
96 smooth muscle cell relaxation. *Am J Respir Cell Mol Biol.* 2022;66(1):96–106.
- 97 4. Teague WG, et al. Baseline features of the severe asthma research program (SARP III)
98 cohort: differences with age. *J Allergy Clin Immunol Pract.* 2018;6(2):545–554.
- 99 5. Chung KF, et al. International European Respiratory Society/American Thoracic Society
100 guidelines on definition, evaluation and treatment of severe asthma. *Eur Respir J.*
101 2014;43(2):343–373.
- 102 6. Barrett ES, et al. Prevalence of SARS-CoV-2 infection in previously undiagnosed health care
103 workers in New Jersey, at the onset of the U.S. COVID-19 pandemic. *BMC Infect Dis.*
104 2020;20(1):853.

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107 **FIGURE LEGENDS**

108 **Figure 1: Serum cAMP levels are increased in patients with asthma.** (A) Histogram of cAMP
109 levels detected in the blood of RCC study participants without a known history of asthma or other
110 lung diseases (N=273) and patients with asthma in the SARP-3 (N=87). For each sample, cAMP
111 levels (pmol per 60 μ l serum) were measured in duplicates by using cAMP-Screen System ELISA
112 kit (Applied Biosystem) and presented as Mean \pm SD. (B) Serum cAMP levels by asthma severity.
113 Asthma severity was defined as “nonsevere” (N=48) and “severe” (N=39) according to ERS/ATS
114 criteria. Linear regression models were used with age and sex as covariates across clinical
115 groups. To satisfy the normalization assumption necessary for linear regression testing, cAMP
116 levels underwent log transformation and the analysis was conducted using R version 4.4.1.

ARutgers Corona Cohort
(Non-Asthma Samples)SARP III Cohort
(Asthma Samples)**B**