

Clinical investigation of hypoxia-inducible factors: getting there

Gregg L. Semenza

Armstrong Oxygen Biology Research Center, Institute for Cell Engineering, and Department of Genetic Medicine, Johns Hopkins University School of Medicine, Baltimore, Maryland, USA.

As part of the *JCI*'s 100th-anniversary celebration, I reflect here on a few of the many papers published in *JCI* that have advanced our understanding of the physiological and pathological responses to hypoxia, with an emphasis on the hypoxia-inducible factors (HIFs), which mediate transcriptional responses to decreased O₂ availability. HIFs consist of an O₂-labile HIF-1 α , HIF-2 α , or HIF-3 α subunit and a constitutively expressed HIF-1 β (also known as ARNT) subunit. The HIF prolyl hydroxylases use O₂ and α -ketoglutarate as substrates to modify the HIF- α subunits by inserting an oxygen atom into a prolyl residue (either P402 or P564 in human HIF-1 α) under normoxic conditions; the hydroxylated proteins are selectively bound by the von Hippel-Lindau tumor suppressor protein (VHL), which recruits a ubiquitin-protein ligase that ubiquitinates the HIF- α subunits, thereby marking them for proteasomal degradation (1). In contrast, under hypoxic conditions, the hydroxylases are inhibited and HIF- α subunits rapidly accumulate, providing a direct mechanism to transduce changes in O₂ availability to changes in HIF activity (1). I also briefly highlight the role of HIFs in erythropoiesis, pulmonary vascular biology, angiogenesis, metabolism, cancer, and diabetic eye diseases.

In 1999, my group reported that mice, which were heterozygous for a knockout allele at the locus encoding HIF-1 α , manifested impaired responses to chronic hypoxia (exposure to a 10% O₂ ambient environment for 3 weeks) (2). The reason for studying heterozygotes was that the homozygotes, which completely lacked HIF-1 α expression, died at midgestation,

with defects in heart development, tissue vascularization, and red blood cell production, thereby demonstrating that HIF-1 α was required for development of all three components of the circulatory system. Analysis of the heterozygotes provided a connection between the role of HIF-1 in development and its role in physiology. Mice exposed to chronic hypoxia develop erythrocytosis and pulmonary hypertension, and these responses were impaired in the heterozygotes. The effect on erythrocytosis remains unexplained; HIF-2 is now considered the primary regulator of adult erythropoiesis through its regulation of EPO production, based on studies of conditional knockout mice (3) and studies demonstrating that missense mutations in the HIF pathway that increase HIF-2 α expression result in the phenotypic triad of hereditary erythrocytosis, pulmonary hypertension, and thrombosis in humans and mice (4).

Hypoxic pulmonary vasoconstriction associated with lobar pneumonia is an adaptive response to hypoxia, as it shunts blood away from areas of lung that are not ventilated, whereas panlobar vasoconstriction is a maladaptive response to ambient hypoxia, as it leads to decreased blood oxygenation. Mice heterozygous for a HIF-2 α -knockout allele were also protected from the development of pulmonary hypertension (5). The roles of HIF-1 and HIF-2 signaling in the pathogenesis of pulmonary hypertension are broad and essential, with complex feed-forward circuits, both within and between, pulmonary artery endothelial and smooth muscle cells, as delineated in an excellent review

(6), which also discussed pharmacologic targeting of HIFs as a potential therapy in this disorder. The critical role of HIF-1 in the vascular response of lung transplants to chronic rejection was also established by a *JCI* paper; data from a mouse model suggested that adenoviral stimulation of HIF activity could prevent lung transplant rejection by maintaining integrity of the microvasculature and tissue perfusion during chronic rejection (7).

Hypoxia induces the HIF-dependent expression of angiogenic growth factors, such as VEGFA, which stimulate new blood vessel formation, thereby increasing tissue perfusion and oxygenation (8). One of the early studies demonstrating that VEGFA administration could augment ischemia-induced vascularization was published in *JCI* (9). Unfortunately, the study utilized young and previously healthy mice, which are not a suitable model for ischemic cardiovascular disease, which is a disease associated with aging. It took the field a long time to learn this lesson.

Two of the most critical adaptations to localized tissue hypoxia are the increased production of angiogenic growth factors to stimulate new blood vessel formation as a means to increase O₂ delivery and the switch from oxidative to glycolytic metabolism as a means to decrease O₂ consumption (10). The co-option of these physiological responses by cancer cells is best illustrated by the clear cell renal cell carcinoma (ccRCC) in patients with the von Hippel-Lindau syndrome, in which one copy of the *VHL* gene is inactivated by germline mutation and the other by somatic mutation within kidney cells. It is a peculiarity of this particular cancer that during tumor progression many ccRCCs lose HIF-1 α expression and are driven solely by dysregulated HIF-2 α expression; this led to the development of belzutifan, a drug that selectively binds to HIF-2 α and blocks its dimerization with HIF-1 β , thereby causing

Conflict of interest: GLS is an inventor on provisional patent application US/63,231,216 and is a founder of, and holds equity in, HIF Therapeutics Inc. This arrangement has been reviewed and approved by Johns Hopkins University in accordance with its conflict-of-interest policies.

Copyright: © 2024, Semenza. This is an open access article published under the terms of the Creative Commons Attribution 4.0 International License.

Reference information: *J Clin Invest*. 2024;134(3):e176253. <https://doi.org/10.1172/JCI176253>.

Table 1. Approved and potential therapeutic targets for small-molecule HIF inducers and HIF inhibitors

	Approved	Potential
HIF inducers (HIF prolyl hydroxylase inhibitors)		
Anemia due to: chronic kidney disease; impaired iron handling	✓	✓
Cisplatin-induced acute kidney injury		✓
Inflammatory bowel disease: ^A Crohn's disease, ulcerative colitis		✓
Ischemic cardiovascular disease: ^A brain, heart, limb		✓
Neurodegenerative disorders: AD, ALS, PD		✓
Nonhealing wounds ^A		✓
Organ transplantation: ^A kidney, liver, lung		✓
Skeletal muscle injury ^A		✓
HIF inhibitors		
Cancer: ccRCC in VHL syndrome; ^B common cancers	✓	✓
Familial erythrocytosis/pulmonary hypertension/thrombosis ^B		✓
Iron overload disorders ^B		✓
NAFLD/NASH		✓
Obesity-associated metabolic disorders		✓
Ocular neovascularization: ^A DME/PDR, wet AMD		✓
Psoriasis		✓
Pulmonary arterial hypertension		✓
Rheumatoid arthritis		✓
Systemic hypertension due to obstructive sleep apnea ^C		✓
Systemic lupus erythematosus		✓
Thyroid eye disease		✓

AD, Alzheimer dementia; ALS, amyotrophic lateral sclerosis; AMD, age-related macular degeneration; ccRCC, clear cell renal cell carcinoma; DME, diabetic macular edema; NAFLD, nonalcoholic fatty liver disease; NASH, nonalcoholic steatohepatitis; PD, Parkinson disease; PDR, proliferative diabetic retinopathy; VHL, von Hippel–Lindau. ^ADrug would likely be administered locally. ^BHIF-2–selective inhibitor. ^CHIF-1–selective inhibitor.

loss of HIF-2 transcriptional activity and a dramatic therapeutic benefit in patients with advanced ccRCC whose prognosis was previously bleak (11).

Among its many effects in breast cancer, HIF-1 was recently shown to control the production of small extracellular vesicles, which were shown to drive cancer progression via multiple mechanisms (12).

Another major recent advance has been the discovery that HIFs play critical roles in mediating the ability of cancer cells of many types, including breast, colorectal, and liver cancer, to evade killing by the immune system and that small-molecule inhibitors of HIF-1 and HIF-2 dramatically improve the response to immune checkpoint inhibitors (anti-CTLA4, anti-PD-1 or anti-PD-L1 antibody) in mouse models (13, 14). These HIF inhibitors have been remarkably well tolerated in mouse models, with no changes in mouse appearance, behavior, or body weight, suggesting that a

therapeutic window exists for HIF inhibition, but, of course, this issue can only be definitively resolved by clinical trials.

Diabetic eye disease is a major and increasing cause of progressive vision loss and blindness in the adult US population. Both HIF-1 and HIF-2 play major roles in retinal neovascularization, and, in mouse models, a small-molecule dual HIF-1/HIF-2 inhibitor safely and effectively blocks the development of macular edema and ischemic retinal neovascularization, which are the causes of vision loss associated with diabetes (15). These studies have highlighted that a large battery of angiogenesis-associated gene products are expressed in a HIF-dependent manner in diabetic eye diseases and that, whereas current therapies target only one of them (VEGFA), HIF inhibitors have a much broader effect on gene expression, which may translate into a higher response rate among patients with diabetic eye diseases,

less than half of whom respond well to anti-VEGFA therapies (15). Because drugs for these conditions are delivered by intraocular injection, systemic adverse effects are not a major consideration.

The last quarter of the *JCI*'s first century has seen dramatic advances in our understanding of the role of HIFs in the pathogenesis of diseases, with major effects on morbidity and mortality. Looking forward, I anticipate that further pharmacologic targeting of HIFs will provide therapeutic benefit in several of these disorders. Table 1 lists the approved and some of the potential therapeutic applications for pharmacologic HIF inducers and HIF inhibitors. Let's hope to see more patients replace mice as the recipients of novel therapies in the *JCI*. Watch this space.

Acknowledgments

GLS is the C. Michael Armstrong Professor at the Johns Hopkins University School of Medicine. Research in GLS's laboratory is supported by the Armstrong Family Foundation.

Address correspondence to: Gregg L. Semenza, Johns Hopkins University School of Medicine, Institute for Cell Engineering, Miller Research Building, Suite 671, 733 North Broadway, Baltimore, Maryland 21205, USA. Email: gsemenza@jhmi.edu.

- Semenza GL. HIF-1 mediates metabolic responses to intratumoral hypoxia and oncogenic mutations. *J Clin Invest.* 2013;123(9):3664–3671.
- Yu AY, et al. Impaired physiological responses to chronic hypoxia in mice partially deficient for hypoxia-inducible factor 1 α . *J Clin Invest.* 1999;103(5):691–696.
- Rankin EB, et al. Hypoxia-inducible factor-2 (HIF-2) regulates hepatic erythropoietin in vivo. *J Clin Invest.* 2007;117(4):1068–1077.
- Hickey MM, et al. The von Hippel-Lindau Chuvash mutation promotes pulmonary hypertension and fibrosis in mice. *J Clin Invest.* 2010;120(3):827–839.
- Brusselmanns K, et al. Heterozygous deficiency of hypoxia-inducible factor-2 α protects mice against pulmonary hypertension and right ventricular dysfunction during prolonged hypoxia. *J Clin Invest.* 2003;111(10):1519–1527.
- Pullamsetti SS, et al. Hypoxia-inducible factor signaling in pulmonary hypertension. *J Clin Invest.* 2020;130(11):5638–5651.
- Jiang X, et al. Adenovirus-mediated HIF-1 α gene transfer promotes repair of mouse airway allograft microvasculature and attenuates chronic rejection. *J Clin Invest.* 2011;121(6):2336–2349.
- Semenza GL. Series introduction: tissue ischemia: pathophysiology and therapeutics. *J Clin Invest.* 2000;106(5):613–614.

9. Takeshita S, et al. Therapeutic angiogenesis. A single intraarterial bolus of vascular endothelial growth factor augments revascularization in a rabbit ischemic hind limb model. *J Clin Invest.* 1994;93(2):662–670.
10. De Heer EC, et al. HIFs, angiogenesis, and metabolism: elusive enemies in breast cancer. *J Clin Invest.* 2020;130(10):5074–5087.
11. Chappell JC, et al. Hypoxia, angiogenesis, and metabolism in the hereditary kidney cancers. *J Clin Invest.* 2019;129(2):442–451.
12. Bertolini I, et al. Intercellular HIF1 α reprograms mammary progenitors and myeloid immune evasion to drive high-risk breast lesions. *J Clin Invest.* 2023;133(8):e164348.
13. Bailey CM, et al. Targeting HIF-1 α abrogates PD-L1-mediated immune evasion in tumor microenvironment but promotes tolerance in normal tissues. *J Clin Invest.* 2022;132(9):e150846.
14. Salman S, et al. HIF inhibitor 32-134D eradicates murine hepatocellular carcinoma in combination with anti-PD1 therapy. *J Clin Invest.* 2022;132(9):e156774.
15. Zhang J, et al. Targeting hypoxia-inducible factors with 32-134D safely and effectively treats diabetic eye disease in mice. *J Clin Invest.* 2023;133(13):e163290.