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Commentary

Historical perspectives In this issue of the JCI, the elegant publication by Hoff and colleagues (1), by serendipity or on purpose, falls on the 40th anniversary of the discovery of calcitonin (CT) as a hypocalcemic principle from the parathyroid gland (2). The parathyroid origin of calcitonin was revisited shortly thereafter, and a thyroid C cell origin was firmly established (3, 4). Calcitonins from several species — mammals, birds, and fishes — were subsequently purified (5), and the first of many calcitonin receptors was cloned in 1991 (6). Tissue-specific alternative splicing of the calcitonin gene at exon 4 was shown to result in production of a novel 37–amino acid peptide: calcitonin gene–related peptide- α (CGRP- α) (7). Both CGRP and calcitonin date back to their neural origins in protochordates and primitive chordates (8). Nonetheless, while CGRP is an established vasodilator and neuromodulator (8, 9), calcitonin has had a checkered history, to say the least, both as a hormone and as a drug. A novel effect on bone formation (1), direct or indirect, is thus intriguing and, if proven, will surely add another historical dimension to calcitonin's role in skeletal biology. Molecular mechanism of calcitonin action Calcitonin is a 32–amino acid peptide with an N-terminal disulphide bridge and a C-terminal prolineamide residue. Both ends of the molecule contain species-invariant residues that are required for its [...]

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Historical perspectives

In this issue of the *JCI*, the elegant publication by Hoff and colleagues (1), by serendipity or on purpose, falls on the 40th anniversary of the discovery of calcitonin (CT) as a hypocalcemic principle from the parathyroid gland (2). The parathyroid origin of calcitonin was revisited shortly thereafter, and a thyroid C cell origin was firmly established (3, 4). Calcitonins from several species — mammals, birds, and fishes — were subsequently purified (5), and the first of many calcitonin receptors was cloned in 1991 (6). Tissue-specific alternative splicing of the calcitonin gene at exon 4 was shown to result in production of a novel 37-amino acid peptide: calcitonin gene-related peptide- α (CGRP- α) (7). Both CGRP and calcitonin date back to their neural origins in protochordates and primitive chordates (8). Nonetheless, while CGRP is an established vasodilator and neuromodulator (8, 9), calcitonin has had a checkered history, to say the least, both as a hormone and as a drug. A novel effect on bone formation (1), direct or indirect, is thus intriguing and, if proven, will surely add another historical dimension to calcitonin's role in skeletal biology.

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Nonstandard abbreviations used: calcitonin (CT); calcitonin gene-related peptide (CGRP); parathyroid hormone (PTH).

Molecular mechanism of calcitonin action

Calcitonin is a 32-amino acid peptide with an N-terminal disulphide bridge and a C-terminal prolineamide residue. Both ends of the molecule contain species-invariant residues that are required for its binding to high-affinity G protein-coupled receptors on the osteoclast (5). Attempts to map the entire bimolecular surface of the hormone receptor complex through photolabeling studies have yielded important new information on contact sites (10). It has also become clear that conformational flexibility of a given calcitonin molecule is the primary determinant of its biological potency (5). Thus, the more flexible fish calcitonins (eel and salmon), containing amino acids such as Gly with less bulky side chains, are about 40-fold more potent than mammalian (porcine, ovine, and human) homologs (11, 12). The ligand specificity of a particular calcitonin receptor isoform is defined by receptor activity-modifying proteins (RAMPs) that form heterodimers with a pair of receptors (13). Femtomolar calcitonin concentrations inhibit the resorptive function of mature osteoclasts, with quiescence (Q effect) being followed by margin retraction (R effect) (14). These kinetically separable morphological changes, Q and R, are exerted through separate cAMP- and Ca^{2+} -dependent signaling pathways involving distinct G proteins (14–17).

Physiological significance

The physiological significance of calcitonin in skeletal conservation has been challenged, despite clear evidence of osteoclastic sensitivity to the peptide *in vitro* (12). This is because it has

been difficult to explain why thyroidectomy does not result in osteoporosis, and why high circulating calcitonin in medullary thyroid carcinoma fails to trigger overt osteopetrosis. At the cellular level, the mechanism is simple. Calcitonin inhibits extracellular Ca^{2+} sensing (18), a potent antiresorptive signal, and by implication, calcitonin withdrawal should enhance Ca^{2+} sensing and limit resorption. Basal resorption is likewise unaffected in the CT/CGRP-null mouse (1). A key finding, that calcitonin withdrawal sensitizes an osteoclast to parathyroid hormone-induced (PTH-induced) stimulation, witnessed by elevations in serum calcium and urinary deoxypyridinoline cross-links, most certainly establishes its physiological function in resorption control. In humans, serum calcitonin rises during pregnancy, growth, and lactation (19). It is during these periods of calcium stress that a tonic antiresorptive hormone will best exert its effect to limit skeletal loss. That, we are now led to believe (1), may be the primary role of calcitonin in skeletal conservation.

What is impressive, albeit unexpected and difficult to explain, is the osteosclerotic phenotype in the CT/CGRP-null mouse described by Hoff et al. (1) (Figure 1). The dramatic increase in the rate of bone formation, evident from tetracycline labeling studies (1), appears to counteract (and abolish) the bone loss due to ovariectomy. A direct role for calcitonin in this effect, however, seems unlikely. According to the authors, there is little evidence of an osteoblastic calcitonin receptor, and it is unlikely that an osteoclast-derived molecule signals the increased bone formation. Growth factors released from the

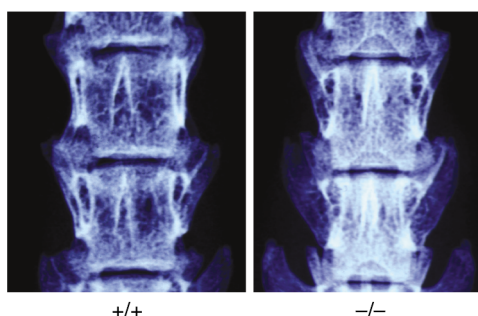


Figure 1

Radiographic analysis of the vertebrae of 3-month-old female wild-type and CT/CGRP-deficient mice (1).

bone matrix could be potential mediators, but again, basal resorption in these animals remains normal. This also rules out the possibility that the increase in bone formation is due to a tight coupling between formation and resorption, as occurs following ovariectomy.

Indeed, the increased bone formation appears to be predominant and independent of resorption (1). Speculatively, we suggest that the effects of CT/CGRP withdrawal are exerted centrally via leptin. The enhanced bone formation in the leptin receptor-deficient *db/db* mouse is identical to that seen in the CT/CGRP-null mouse (20). Even more interesting is the recent revelation that the sympathetic nervous system, regulated by leptin, may control bone formation through adrenergic receptors on osteoblasts (21). Furthermore, several neuropeptides, such as neuropeptide Y, have recently been shown to exert unexpected but powerful effects on the skeleton (22). CGRP is localized to cells in close proximity to leptinergic neurons (23), and it would not be surprising, therefore, if the leptin axis were modulated by CGRP directly or indirectly. CGRP- β , the product of a second CGRP gene (5), should, however, at least in part, compensate for the loss of CGRP- α , and this requires further investigation using double knockout approaches.

Calcitonin as a drug?

While the CT/CGRP-null phenotype points out that there is still much about the physiological effects of calcitonin and its role in bone remodeling that we do not understand, decades of application in humans

have taught us a great deal about its therapeutic utility. Calcitonin has been used for Paget bone disease, hypercalcemia of malignancy, osteogenesis imperfecta, and postmenopausal osteoporosis. Both subcutaneous and nasal calcitonin formulations have been developed and are in use worldwide. There is rich anecdotal evidence that subcutaneous calcitonins, namely salmon, eel, porcine, and human calcitonins, all significantly increase bone mineral density and, in doing so, likely reduce fracture risk (24). However, to the contrary, there is evidence that the effects of the currently available nasal salmon-calcitonin formulation in reducing fracture risk are minimal, mainly due to issues of bioavailability (25). This is not unexpected, because like other highly charged peptides, including insulin and PTH, calcitonin

is unlikely to cross multicellular mucous membranes effectively in the absence of a vehicle to aid its delivery.

The widespread use of calcitonin as a nasal spray is thus likely to be re-evaluated (26). Evolving therapies, including newer bisphosphonates, have impinged and would continue to impinge upon calcitonin's success as an antiosteoporosis therapeutic (Table 1). Nonetheless, two key advantages of calcitonin would remain should the desired bioavailability be achieved. One is its stabilizing effect on bone microarchitecture, which precedes changes in bone mineral density (25). This may account for its rapid effect in reducing fracture risk, also seen with the newer bisphosphonates (27). Second and perhaps more important is the use of the peptide in achieving analgesia in vertebral crush syndromes and other types of osteogenic pain.

Finally, with the revelation that low-dose intermittent PTH reduces fracture risk through a dramatic increase in bone formation (28), the possibility of sequential therapy with the two hormones, PTH and calcitonin, makes solid therapeutic sense. The paper by Hoff and colleagues (1), however, raises an important question: Can systemically administered calcitonin inhibit bone formation and thus counteract the anabolic effect of PTH in humans? Although evidence suggests that this is not the case, future systematic clinical studies should clarify this one way or another.

Table 1

Osteoporosis therapeutics on the horizon

Antiresorptive

Bisphosphonates^A, including new potent intravenous analogues
 Selective sex steroid (estrogen^A and androgen) receptor modulators
 Calcitonin^A, calcitonin analogues, and calcitonin secretagogues
 Future therapies targeted at inhibiting osteoclast formation and function
 Osteoprotegerin and its analogues
 RANK and RANK-ligand inhibitors
 Specific cytokine inhibitors and receptor modulators
 Cathepsin K inhibitors
 Integrin inhibitors
 Vacuolar H⁺ATPase (proton pump) inhibitors

Bone forming

Parathyroid hormone and its analogues
 Parathyroid hormone secretagogues (calcium receptor antagonists)
 Anabolic vitamin D analogues
 Modifiers of the bone morphogenetic protein cascade, including new statins
 Centrally acting bone formation modulators

^AEstablished therapies.

The future

The report by Hoff et al. (1) not only establishes a role for calcitonin in skeletal conservation, expected since its discovery four decades ago, but also opens new areas of investigation. These investigations extend from examining a role for calcitonin and CGRP in the central control of bone remodeling to defining other, perhaps more novel functions of both peptides, considering that their receptors have a widespread distribution (5).

On the therapeutic front, calcitonin has stood the test of time. Nonetheless, several new approaches are being used to enhance calcitonin access to the osteoclast, and, for its longer-term use in humans, to prevent receptor down-regulation. The next decade should thus witness the development of oral formulations, new peptide delivery systems, and membrane-permeant analogs with in vivo potencies suggested by calcitonin's powerful cellular effects (29, 30). A full characterization of the calcitonin receptor has allowed the synthesis of isoform-specific and even domain-specific calcitonin analogs. Likewise, organic molecules that could allosterically enhance receptor sensitivity to circulating calcitonin constitute possible future therapeutics. Finally, one could envisage the development of a calcitonin secretagogue — a small-molecule agonist targeted to the C cell Ca^{2+} receptor that would activate calcitonin release, as has been achieved successfully for PTH (31, 32).

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