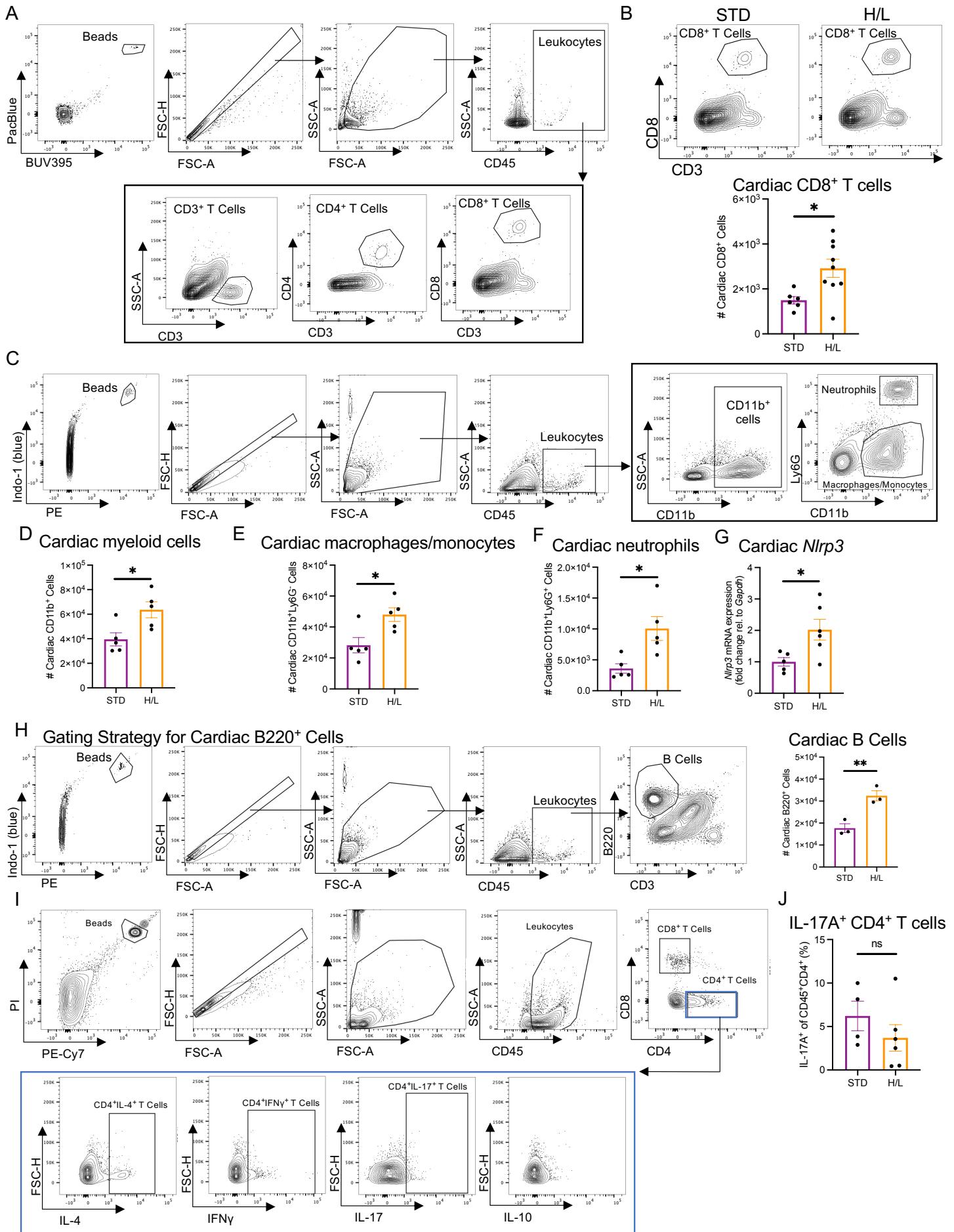
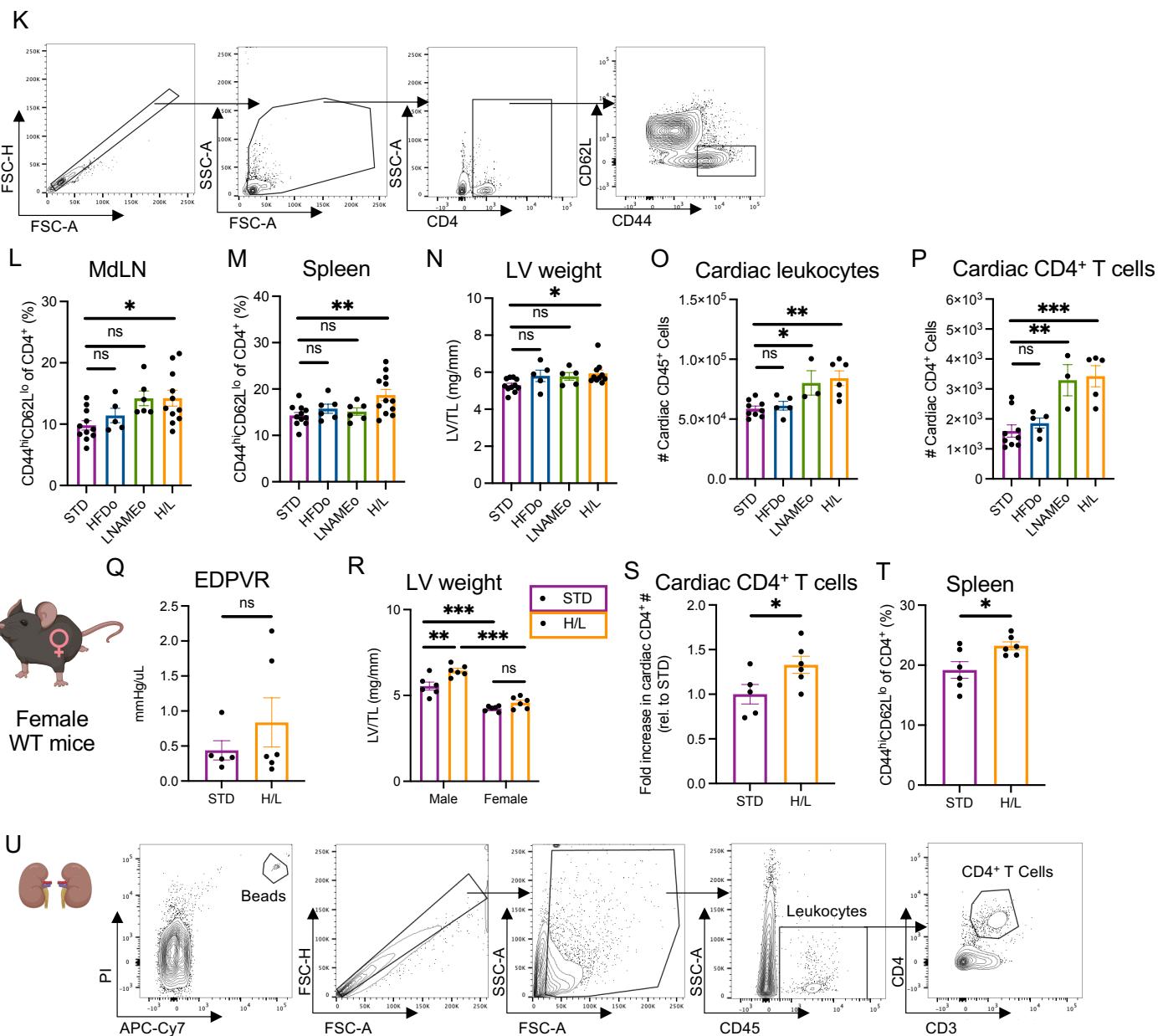


Supplementary Figure 1

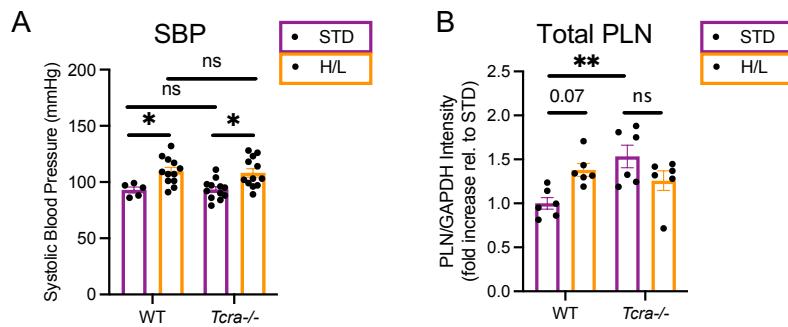


Supplementary Figure 1 continued



1008 **Supplemental Figure 1: Gating strategies for leukocyte characterization by flow**
1009 **cytometry, and characterizing inflammation in female mice and single diet controls.**
1010 WT mice were fed H/L or STD for 5 weeks. Lymphocytes (A) were measured by flow
1011 cytometry, and CD48⁺ T cell infiltration was visualized in LV (B). Leukocytes (C),
1012 CD45⁺CD11b⁺ cells (D), CD45⁺CD11b⁺Ly6G⁻ cells (E), and CD45⁺CD11b⁺Ly6G⁺ cells (F)
1013 in LV of mice were measured directly by flow cytometry. Cardiac gene expression of *Nlrp3*
1014 was measured by qPCR (G). CD45⁺B220⁺ cells in LV of mice were measured directly by
1015 flow cytometry (H). LV CD45⁺CD4⁺ cells expressing IL-4, IFN γ , IL-10, and IL-17 (I-J) were
1016 analyzed by flow cytometry. CD4⁺CD44^{hi}CD62L^{lo} effector T cells were measured in
1017 mediastinal lymph nodes and spleen (K) by flow cytometry. WT mice fed HFD only
1018 (HFDo), L-NAME only (LNAMEo), H/L, or STD chow for 5 weeks were analyzed for
1019 lymphoid expansion of CD4⁺CD44^{hi}CD62L^{lo} effector T cells (L-M). LV hypertrophy was
1020 measured (normalizing LV weight to tibia length, LV/TL, N), and cardiac CD45⁺ cells (O)
1021 and CD45⁺CD3⁺CD4⁺ cells (P) were analyzed directly in LV by flow cytometry. Female
1022 mice were fed H/L or STD for 5 weeks and invasive hemodynamic analysis were used to
1023 characterize end diastolic pressure volume relationship (EDPVR, Q). Normalized LV
1024 weight was used to measure hypertrophy (LV/TL, R), and cardiac CD4⁺ cells (S) and
1025 splenic effector CD4⁺ cells (T) were measured by flow cytometry. In male WT mice (U),
1026 renal CD45⁺ cells and CD4⁺ cells were measured by flow cytometry. n=4-12. B-J, W, S-
1027 T: unpaired t test. L-P: one-way ANOVA with Tukey's multiple comparisons test. R: two-
1028 way ANOVA with Sidak's multiple comparisons test. L-M: HFDo, LNAMEo data are shown
1029 compared to STD, H/L data from Figure 1. *p≤0.05, **p≤0.01, ***p≤0.001.

Supplementary Figure 2



1030 **Supplemental Figure 2: Comparison of WT vs *Tcra*-/- fed HFD/L-NAME or STD diet.**

1031 WT and T cell deficient (*Tcra*-/-) mice were fed STD or H/L for 5 weeks and systolic blood
1032 pressure (SBP, A) was measured by invasive hemodynamic analysis. Total
1033 phospholamban (PLN) protein expression was assessed in left ventricle (LV) samples by
1034 Western blot (B). n=6-12. Two-way ANOVA with Sidak's multiple comparisons test.

1035 *p≤0.05, **p≤0.01.

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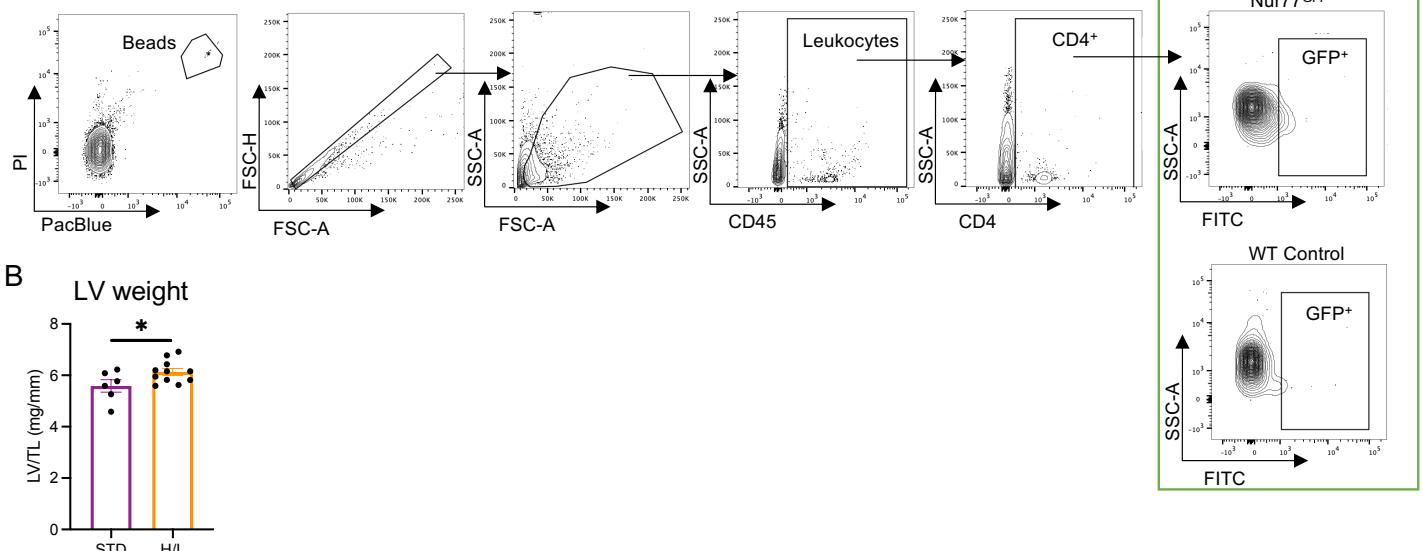
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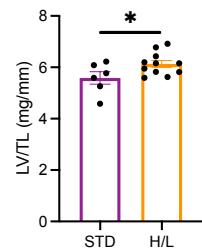
Supplementary Figure 3

A

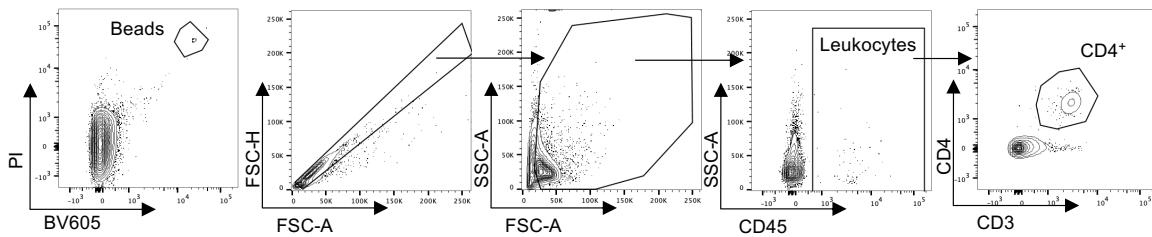


B

LV weight

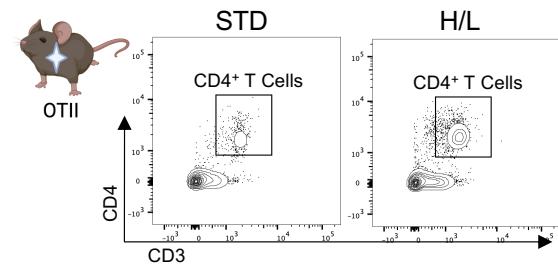


C

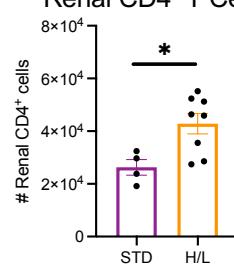


D

Kidneys

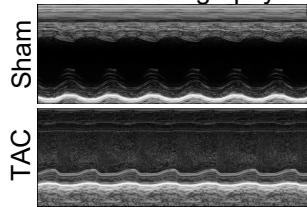


E Renal CD4+ T Cells

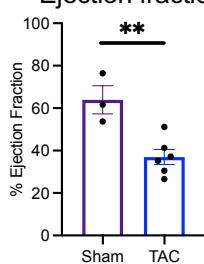


F

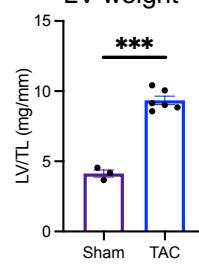
Echocardiography



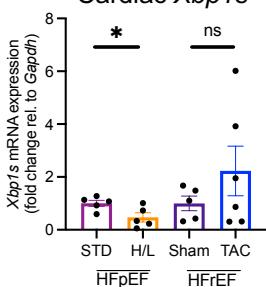
G Ejection fraction



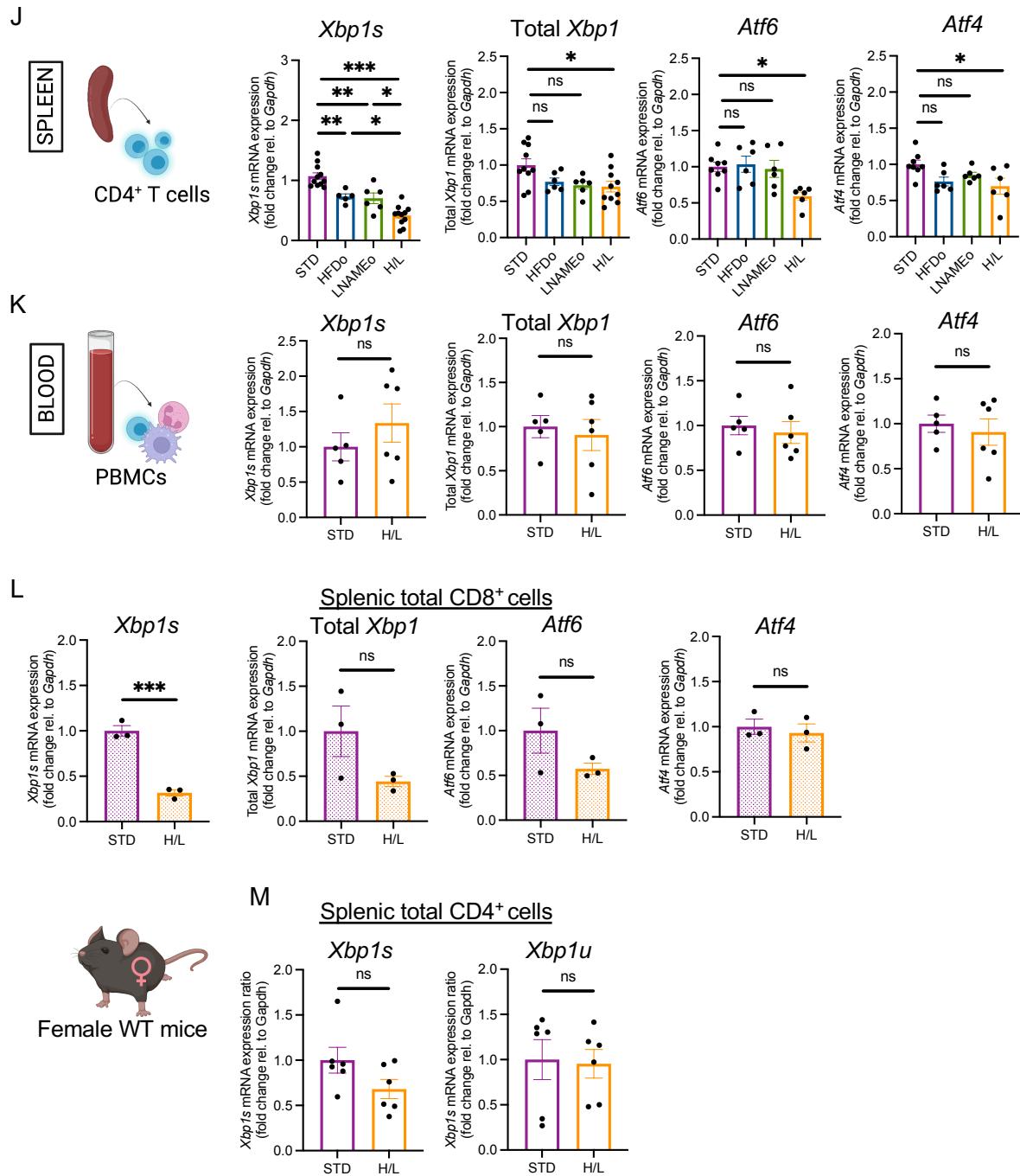
H LV weight



I Cardiac *Xbp1s*

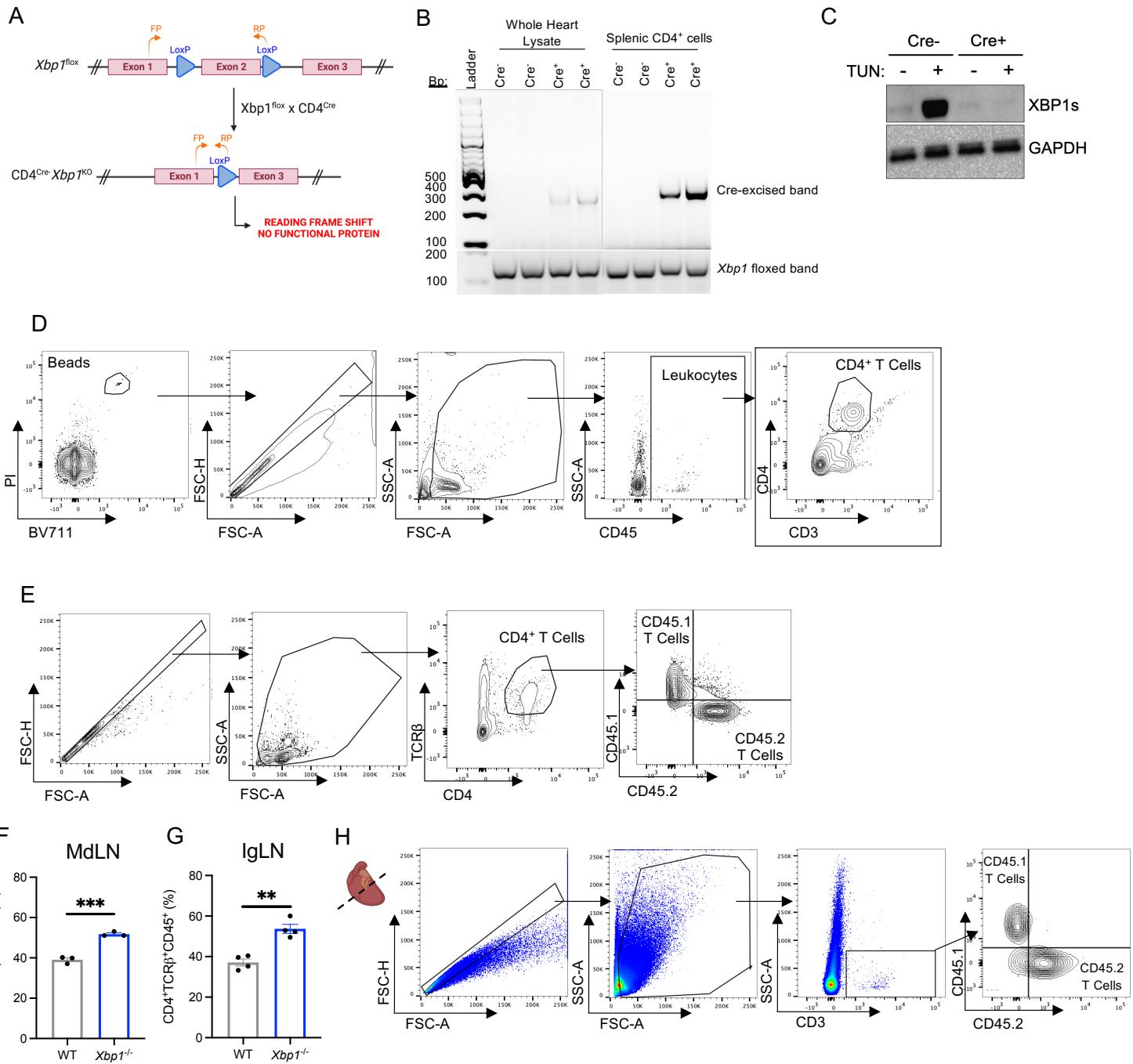


Supplementary Figure 3 continued



1052 **Supplemental Figure 3: Gating strategies for Nur77^{GFP} and OTII mice and leukocyte**
1053 **UPR expression.** CD45⁺CD4⁺GFP⁺ cells were measured in LV of Nur77^{GFP} mice 5 weeks
1054 after H/L, STD, or TAC surgery by flow cytometry, using a WT mouse as a negative
1055 control for gating (A). Left ventricular hypertrophy was measured in Nur77^{GFP} mice by
1056 normalizing LV weight to tibial length (LV/TL, B). CD45⁺CD3⁺CD4⁺ cells in LV (C) or in
1057 kidneys (D-E) of OTII mice were directly measured by flow cytometry. WT mice underwent
1058 TAC or sham surgery, and 5 weeks later, ejection fraction was assessed by
1059 echocardiography (F-G) and LV weight was measured (normalized to tibia length, LV/TL,
1060 H). Cardiac LV expression of *Xbp1s* was measured by qPCR from mice fed H/L or STD,
1061 or receiving TAC or sham surgery, for 5 weeks (I). WT mice were fed HFD only (HFDo),
1062 L-NAME only (LNAMeO), H/L, or STD for 5 weeks, then splenic CD4⁺ T cells were isolated
1063 to measure *Xbp1s*, total *Xbp1*, *Atf6*, and *Atf4* expression by qPCR (J). In WT mice fed
1064 H/L or STD for 5 weeks, the circulating leukocyte population in blood (K) and splenic CD8⁺
1065 T cell population (L) were analyzed for expression of *Xbp1s*, total *Xbp1*, *Atf6*, and *Atf4* by
1066 qPCR. Splenic CD4⁺ T cells were isolated from female WT mice fed H/L or STD for 5
1067 weeks, and expression of *Xbp1s* and total *Xbp1* were analyzed by qPCR (M). n=3-11. K-
1068 L: each replicate is n=2 mice pooled. B, E-H, K-M: unpaired t test. I: HF groups (H/L or
1069 TAC) are compared against their respective controls (STD and sham, respectively) in
1070 unpaired t test. J: one way ANOVA with Tukey's multiple comparisons test, and HFDo,
1071 LNAMeO groups are shown compared to STD, H/L groups in Figure 3. *p≤0.05, **p≤0.01,
1072 ***p≤0.001.
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Supplementary Figure 4



1075 **Supplemental Figure 4: Genotyping CD4^{Cre}-Xbp1^{flox} mice and gating strategies for**
1076 **cardiac and lymphoid leukocyte populations.** CD4^{Cre}-Xbp1^{flox} mice were generated by
1077 breeding Xbp1^{flox} and CD4^{Cre} mice, generating progeny with nonfunctional XBP1 protein
1078 (A). Cre excision in splenic CD4⁺ T cells was present in Cre⁺ mice, and absent in Cre⁻
1079 mice (B). Splenic CD4⁺ T cells were isolated from CD4^{Cre}-Xbp1^{flox} (T-Xbp1^{KO}) and CD4^{WT}-
1080 Xbp1^{flox} (T-Xbp1^{WT}) mice and expanded into T cell blasts in the presence of
1081 αCD3/αCD28. Cells were either left untreated or treated with tunicamycin (TUN) for 6
1082 hours, then collected for analysis of XBP1s expression by Western blotting (C). Cardiac
1083 CD45⁺CD3⁺CD4⁺ T cells were analyzed in LV of T-Xbp1^{KO} or T-Xbp1^{WT} mice directly
1084 using flow cytometry (D). CD45.2⁺CD4⁺Tcrβ⁺Xbp1^{-/-} and CD45.1⁺CD4⁺Tcrβ⁺WT T cells
1085 were co-transferred into T cell deficient (*Tcra*-/-) recipients fed H/L for 3 weeks, then
1086 respective populations of T cells were directly measured in lymphoid organs using flow
1087 cytometry (E). Relative proportions of each cell type were characterized in mediastinal
1088 lymph nodes (MdLN, F), and inguinal lymph nodes (IgLN, G) of recipient *Tcra*-/- mice by
1089 flow cytometry. Relative proportions of CD3⁺CD45.1⁺ and CD3⁺CD45.2⁺ cells were
1090 measured in left ventricles of *Tcra*-/- mice (H). n=3-4. Unpaired t test. **p≤0.01,
1091 ***p≤0.001

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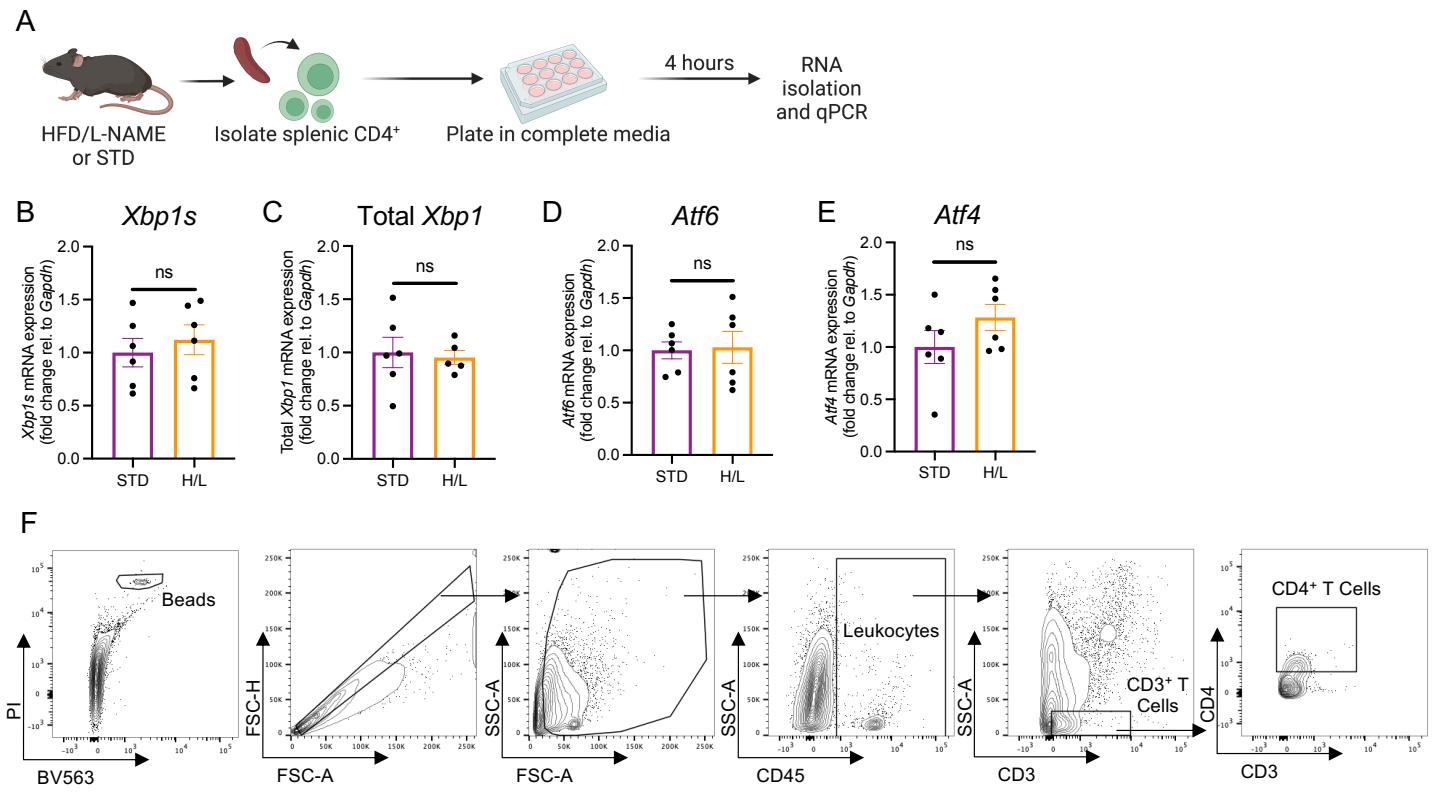
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Supplementary Figure 5



1098 **Supplemental Figure 5: UPR recovery in vitro and in vivo.** Splenic CD4⁺ T cells were
1099 isolated from WT mice fed H/L or STD for 5 weeks, plated in complete media for 4 hours
1100 in 37°C, then measured for gene expression of *Xbp1s*, total *Xbp1*, *Atf6* and *Atf4* by qPCR
1101 (A-E). Cardiac CD45⁺CD3⁺CD4⁺ T cells were directly measured by flow cytometry in WT
1102 mice were fed STD or H/L, or H/L for 5 weeks then reverted to STD for 2 weeks (H/L->S)
1103 prior to analysis. n=5-6. Unpaired t test.

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1122 **Supplemental Tables**

1123 **Supplementary Table 1: Characterization of left ventricular function of female WT**

1124 **mice by echocardiography or pressure volume loops after 5 week treatment with**

1125 **STD or H/L**

	Ejection Fraction (%)	Fractional Shortening (%)	Anterior Wall Thickness s (mm)	Internal Diastolic Diameter (mm)	Internal Systolic Diamete r (mm)	Posterior Wall Thickness (mm)	Heart Rate (bpm)	Systolic Blood Pressure (mm Hg)	dP/dt min (mmHg/s)	dP/dt max (mmHg/s)	EDPVR	ESPVR	PRSW
WT	59.23 ± 2.048	30.76 ± 1.418	0.793 ± 0.029	3.518 ± 0.046	2.437 ± 0.067	1.174 ± 0.037	478.3 ± 12.2	95.4 ± 1.66	-6142 ± 437.2	6759 ± 333.1	0.439 ± 0.138	2.489 ± 0.357	38.73 ± 6.363
STD													
WT	62.25 ± 1.216	32.64 ± 0.806	0.925 ± 0.068	3.319 ± 0.074*	2.237 ± 0.068	1.295 ± 0.048	492.3 ± 13.26	120 ± 2.817***	-7048 ± 620.9	7377 ± 730.1	0.837 ± 0.351	2.674 ± 0.511	39.47 ± 8.692
H/L													

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1127 Values are means ± SEM. n=7 for echocardiography; n=7-9 for pressure volume loops.

1128 Unpaired t-test. *** ≤ 0.001 comparing STD vs H/L

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1139 **Supplementary Table 2: Characterization of left ventricular function of WT or *Tcra*-**
 1140 **/- mice by echocardiography or pressure volume loops after 5 week treatment with**
 1141 **STD or H/L**

	Fractional Shortening (%)	Anterior Wall Thickness (mm)	Internal Diastolic Diameter (mm)	Internal Systolic Diameter (mm)	Posterior Wall Thickness (mm)	Heart Rate (bpm)	dP/dt min (mmHg/s)	dP/dt max (mmHg/s)	ESPVR	PRSW
WT	36.831 ± 3.020	0.822 ± 0.061	3.592 ± 0.13	2.29 ± 0.166	0.768 ± 0.39	473.111 ± 17.501	-8172.2 ± 497.635	8878 ± 551.957	2.508 ± 0.302	59.618 ± 8.555
WT	35.266 ± 1.749	0.976 ± 0.047	3.578 ± 0.1	2.331 ± 0.121	0.854 ± 0.028	520.667 ± 10.761*	-8964.9 ± 562.069	8880.9 ± 441.473	5.343 ± 1.699	51.915 ± 5.105
<i>Tcra</i>-/-	35.880 ± 1.444	0.852 ± 0.046	3.86 ± 0.117	2.483 ± 0.119	0.806 ± 0.027	496.778 ± 11.351	-5523.2 ± 232.392\$\$	6190.8 ± 293.030\$\$	2.713 ± 0.589	47.458 ± 6.893
<i>Tcra</i>-/- H/L	33.534 ± 1.595	0.925 ± 0.04	3.544 ± 0.091	2.364 ± 0.107	0.854 ± 0.022	488.7 ± 8.234	-8458.44 ± 359.889	8622.889 ± 420.918	1.597 ± 0.207	52.726 ± 6.001

1142 Values are means ± SEM. n=9-12 for echocardiography; n=5-10 for pressure volume
 1143 loops. Two-way ANOVA with Tukey's multiple comparisons test. * ≤ 0.05 comparing WT
 1144 STD vs H/L; \$\$ ≤ 0.01 comparing WT vs *Tcra*-/- STD

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1150 **Supplementary Table 3: Characterization of left ventricular function of OTII mice by**
 1151 **echocardiography or pressure volume loops after 5 week treatment with STD or**
 1152 **H/L**

	Ejection Fraction (%)	Fractional Shortening (%)	Anterior Wall Thickness (mm)	Internal Diastolic Diameter (mm)	Internal Systolic Diameter (mm)	Posterior Wall Thickness (mm)	Heart Rate (bpm)	Systolic Blood Pressure (mm Hg)	dP/dt min (mmHg/s)	dP/dt max (mmHg/s)	EDPVR	ESPVR	PRSW
OTII STD	62.60 ± 1.402	33.33 ± 0.978	0.8090 ± 0.038	3.884 ± .07338	2.592 ± 0.2104	0.6859 ± .05153	442.3 ± 42.31	97.14 ± 2.849	-6850 ± 447.2	7322 ± 576.2	0.427 ± 0.164	2.722 ± 0.43	42.13 ± 6.575
OTII H/L	66.41 ± 2.037	36.11 ± 1.636	0.9237 ± 0.1378	3.665 ± 0.0668*	2.345 ± 0.2338	0.8344 ± 0.03616*	495.6 ± 51.68*	113.9 ± 2.201***	-8094 ± 303.6*	7895 ± 334.8	0.371 ± 0.132	2.501 ± 0.4371	50.57 ± 5.849

1153 Values are means ± SEM. n=7 for echocardiography; n=7-9 for pressure volume loops.

1154 Unpaired t-test. * ≤ 0.05, *** ≤ 0.001 comparing STD vs H/L

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1165 **Supplementary Table 4: Characterization of left ventricular function of WT mice by**
1166 **echocardiography 5 weeks after TAC or sham surgery**

	Fractional Shortening (%)	Anterior Wall Thickness (mm)	Internal Diastolic Diameter (mm)	Internal Systolic Diameter (mm)	Posterior Wall Thickness (mm)	Heart Rate (bpm)
WT Sham	34.28 ± 5.016	0.8529 ± 0.088	3.252 ± 0.1696	2.141 ± 0.3881	0.7008 ± 0.0323	488.3 ± 59.5
WT TAC	17.80 ± 1.941**	1.086 ± 0.058*	4.409 ± 0.0936***	3.63 ± 0.3586***	0.9833 ± 0.0548*	520.8 ± 49.49

1167 Values are means ± SEM. n=3-6 for echocardiography. Unpaired t-test. * ≤ 0.05, ** ≤ 0.01,
1168 *** ≤ 0.001 comparing sham vs TAC.

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1180 **Supplementary Table 5: Characterization of left ventricular function of T-Xbp1^{WT}**
 1181 **and T-Xbp1^{KO} mice by echocardiography or pressure volume loops after 5 week**
 1182 **treatment with STD or H/L**

	Fractional Shortening (%)	Anterior Wall Thickness (mm)	Internal Diastolic Diameter (mm)	Internal Systolic Diameter (mm)	Posterior Wall Thickness (mm)	Heart Rate (bpm)	dP/dt min (mmHg/s)	dP/dt max (mmHg/s)	ESPV R	PRS W
T-Xbp1 ^{WT} H/L	29.63 ± 1.217	0.946 ± 0.0534	3.76 ± 0.1118	2.645 ± 0.1536	0.9080 ± 0.061	526.1 ± 34.1	-8412 ± 812.0	8554 ± 492.3	1.126 ± 0.524	42.65 ± 5.707
T-Xbp1 ^{KO} H/L	34.55 ± 1.931	1.060 ± 0.1067	3.557 ± 0.1050	2.330 ± 0.2337	1.059 ± 0.0579	508.5 ± 37.8	-7414 ± 276.1	7307 ± 172.2*	0.731 ± 0.238	48.18 ± 9.814

1183 Values are means ± SEM. n=4 for echocardiography; n=4-6 for pressure volume loops.

1184 Unpaired t-test. * ≤ 0.05 comparing Xbp1^{WT} vs Xbp1^{KO}

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1192 **Supplementary Table 6: Characterization of left ventricular function of WT mice by**
 1193 **echocardiography or pressure volume loops after treatment with STD, H/L, or H/L**
 1194 **then STD for 2 weeks (2 STD) or 3 weeks (3 STD).**

	Ejection Fraction (%)	Fractional Shortening (%)	Anterior Wall Thickness (mm)	Internal Diastolic Diameter (mm)	Internal Systolic Diameter (mm)	Posterior Wall Thickness (mm)	Heart Rate (bpm)	dP/dt min (mmHg/s)	dP/dt max (mmHg/s)	ESPVR	PRSW
WT STD	63.71 ± 1.675	34.10 ± 1.160	0.9438 ± 0.043	3.822 ± 0.1462	2.525 ± 0.3251	0.7998 ± 0.007	498.5 ± 16.23	-7743 ± 596.5	7671 ± 450.7	1.820 ± 0.2144	34.52 ± 6.813
WT H/L	63.45 ± 2.883	33.80 ± 2.037	0.9304 ± 0.077	3.472 ± 0.142	2.306 ± 0.3539	0.8369 ± 0.0203	482.5 ± 10.23	-7406 ± 249.8	8122 ± 116.3	9.353 ± 3.344	43.28 ± 5.316
WT H/L -> 2 STD	59.74 ± 2.861	32.07 ± 2.443	0.8820 ± 0.066	3.728 ± 0.092	2.556 ± 0.214	0.8142 ± 0.0238	474.5 ± 20.02	-5403 ± 340.3\$	5928 ± 345.5\$	5.662 ± 2.917	33.96 ± 11.83
WT H/L -> 3 STD	60.04 ± 2.130	31.55 ± 1.397	0.898 ± 0.057	3.877 ± 0.105	2.659 ± 0.116	0.924 ± 0.061	524.2 ± 8.94%%, ^^^	-7805 ± 686.7^	8867 ± 687.5^^	1.961 ± 0.5288	43.7 ± 9.448

1195 Values are means ± SEM. n=6 for echocardiography; n=5-6 for pressure volume loops.

1196 One-way ANOVA with Tukey's multiple comparisons test. \$ ≤ 0.05 comparing H/L vs H/L->2STD; %% ≤ 0.01 comparing H/L vs H/L->3STD; ^ ≤ 0.05, ^^ ≤ 0.01, ^^^ ≤ 0.001 comparing H/L->2STD vs H/L->3STD.

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1203 **Supplementary Table 7: Antibodies used for flow cytometry**

Target	Clone	Biolegend Catalog #
CD45.2	104	109841
CD3	145-2C11	100330
CD4	GK1.5	100412
CD8	53-6.7	100738
CD11b	M1/70	101230
Ly6G	1A8	127628
IL-4	11B11	504109
IFN γ	XMG1.2	505809
IL-17	TC11-18H10.1	506904
IL-10	JES5-16E3	505022
CD44	IM7	103012
CD62L	MEL-14	161204
CD45.1	A20	110708
TCR β	H57-597	109220
CD69	H1.2F3	104506

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1205 **Supplementary Table 8: Primer sequences used for qPCR**

Gene	Forward (5' – 3')	Reverse (5' – 3')
<i>Xbp1s</i>	GGTCTGCTGAGTCCGCAGCAGG	GAAAGGGAGGGCTGGTAAGGAAC
Total <i>Xbp1</i>	GACAGAGAGTCAAACTAACGTGG	GTCCAGCAGGCAAGAAGGT
<i>Atf6</i>	AGAAAGCCCCGCATTCTCCAG	ACTCCCAGAACCTCTACTGATGC
<i>Atf4</i>	ATGGCCGGCTATGGATGAT	CGAAGTCAAACTCTTCAGATCCATT
<i>Nlrp3</i>	ATTACCCGCCGAGAAAGG	CATGAGTGTGGCTAGATCCAAG
<i>Gapdh</i>	TGGCAAAGTGGAGATTGTTGCC	AAGATGGTGATGGCTTCCCG
<i>Xbp1</i> (splicing assay)	ACACGCTTGGGAATGGACAC	CCATGGGAAGATGTTCTGGG
<i>18s</i> (splicing assay)	AAACGGCTACCACATCCAAG	CCTCCAATGGATCCTCGTTA

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1211 **Supplementary Table 9: Antibodies used for Western blot**

Target	Catalog #
Phospho-I κ E1 α (Ser724)	Novus NB100-2323
I κ E1 α	Cell Signaling 3294T
Phospho-PERK (Thr980)	Cell Signaling 3179S
PERK	Cell Signaling 3192S
Phospho-EIF2 α	Cell Signaling 9721
EIF2 α	Cell Signaling 9722
ATF6	Cell Signaling 65880
ATP2A2/SERCA2	Cell Signaling 4388
Phospho-Phospholamban (Ser16/Thr17)	Cell Signaling 8496
Phospholamban	Cell Signaling 14562
GAPDH	Cell Signaling 2118S

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