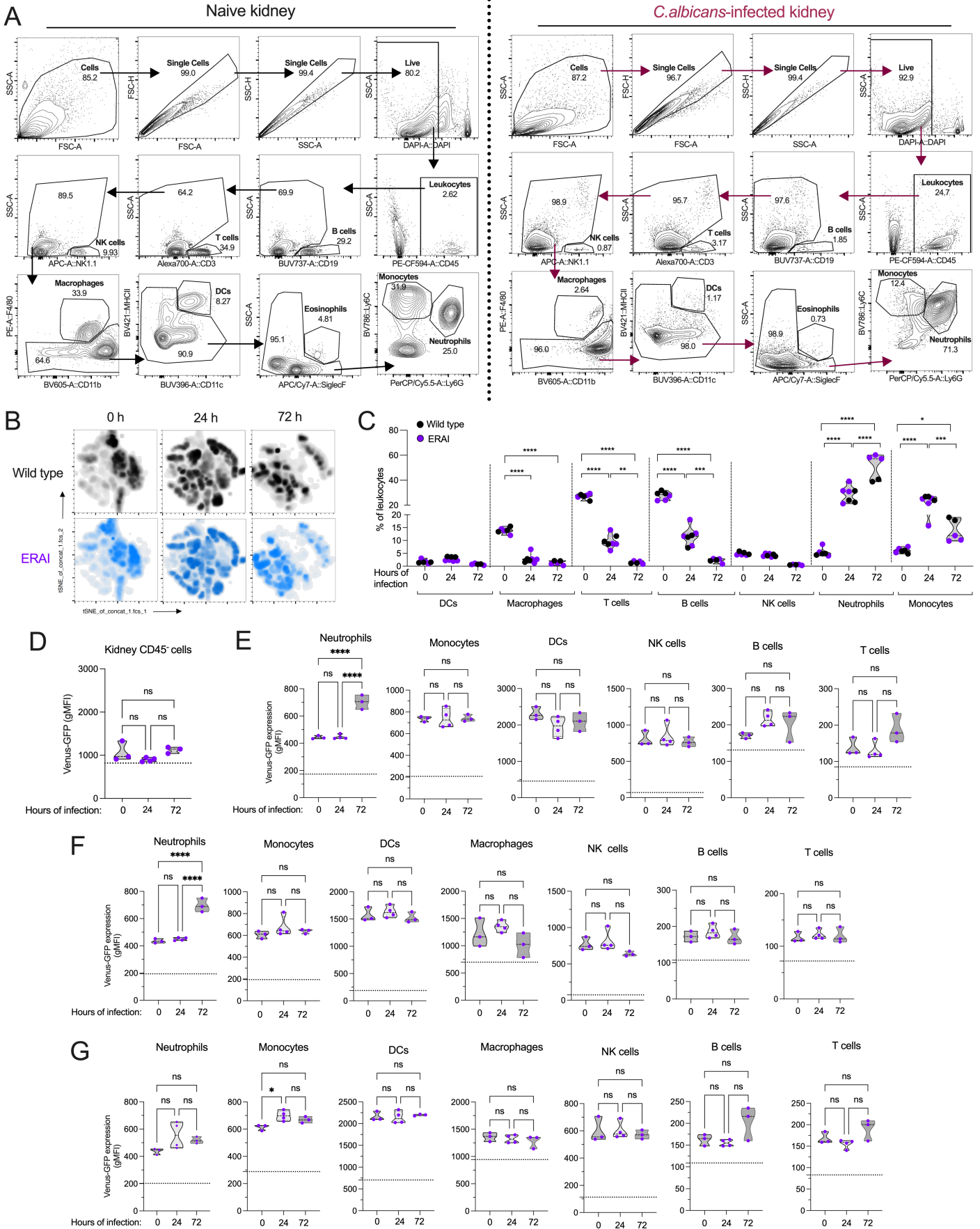
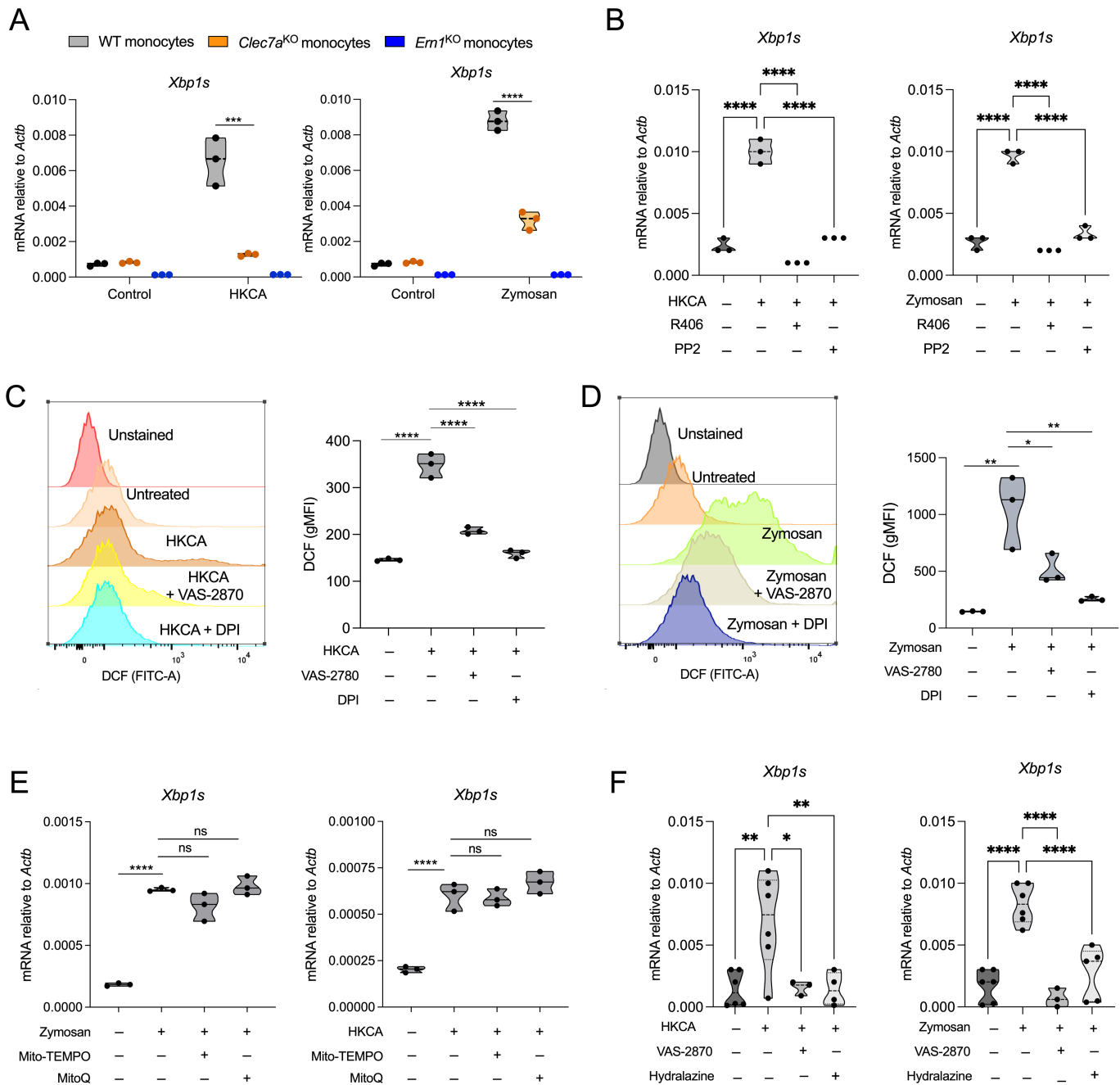


# Supplementary Figure 1



**Supplementary Figure 1. Analysis of WT or ERAI mice with systemic candidiasis.** (A-F) ERAI or wild type C57BL/6J mice ( $n = 4$  per genotype per time point) were left untouched or injected i.v. with  $10^5$  *C. albicans* cells, and their kidneys, blood, spleen, and bone marrow were collected at the indicated time points. Single-cell suspensions were procured from various organs, as described in the methods, and stained with fluorescently-labeled antibodies specific for CD45, CD19, CD3, NK1.1, F4/80, CD11c, MHC-II, SiglecF, CD11b, Ly6C, Ly6G and DAPI. (A) Gating strategy used to analyze the kidney immune contexture in naïve or *C. albicans*-infected mice. FACS plots are representative of kidney analysis in an uninfected (naïve) mouse or in a mouse systemically infected with *C. albicans* for 72 h. (B) tSNE plots representing time-dependent changes in kidney total CD45<sup>+</sup> immune cells of ERAI or wild type mice. (C) Violin plots showing proportion of the indicated immune cell subsets within total CD45<sup>+</sup> leukocytes infiltrating the kidney at 0, 24, and 72 h after *C. albicans* infection. (D-G) Mean fluorescence intensity (gMFI) of Venus reporter expression in CD45-negative cells in the infected kidney. (E-G) Violin plots for gMFI of Venus reporter expression in the indicated immune cell types in blood (D), spleen (E), and bone marrow (F). (D-G) Dashed lines represent intrinsic autofluorescence in WT mice. (C-G) One-way ANOVA (Tukey's test). \* $P < 0.05$ , \*\* $P < 0.005$ , \*\*\* $P < 0.0005$ , \*\*\*\* $P < 0.0001$ , ns, not significant.

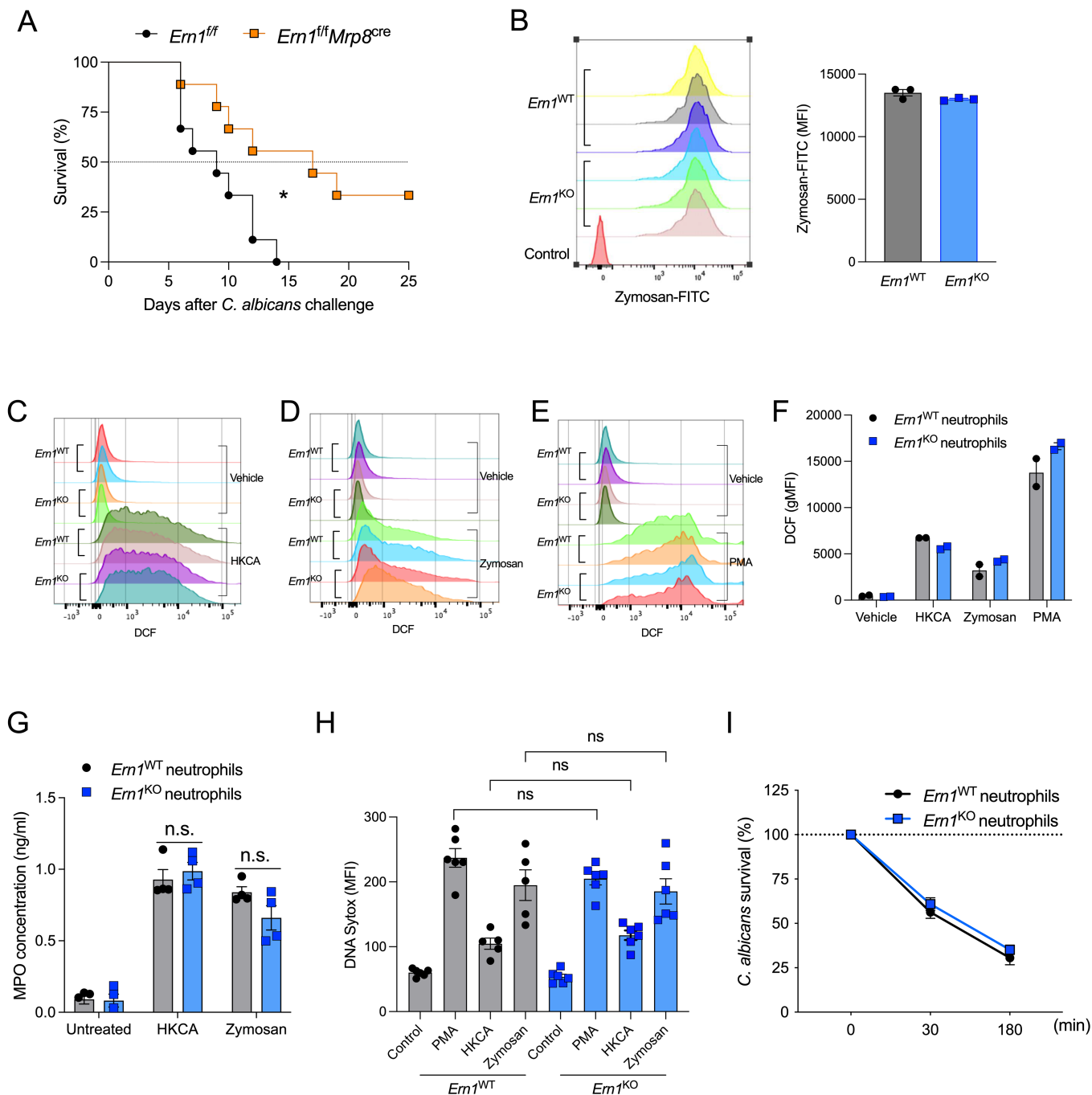
## Supplementary Figure 2



**Supplementary Figure 2. The Dectin-1-Syk-NOX axis also mediates IRE1 $\alpha$  activation in monocytes responding to zymosan or *C. albicans*.** (A) Bone marrow-resident monocytes were isolated from mice of the indicated genotypes ( $n = 3$  per genotype) and then stimulated for 6 h with HKCA (MOI=10) or zymosan (25  $\mu$ g/ml). *Xbp1s* transcript levels were measured using quantitative RT-PCR. (B) Bone marrow-resident monocytes from WT C57BL/6J mice ( $n = 3$ ) were pretreated for 1 h with vehicle control or the Syk inhibitor R406 (10  $\mu$ M), and cells were then stimulated for 6 hours with HKCA (MOI=10) or zymosan (25  $\mu$ g/ml). *Xbp1s*

transcript levels were measured using quantitative RT-PCR. **(C and D)** WT bone marrow neutrophils ( $n = 3$  mice) were pretreated for 30 min with vehicle control or ROS inhibitors DPI (10  $\mu$ M) or VAS-2870 (10  $\mu$ M) and then stimulated with either **(C)** HKCA (MOI=10) or **(D)** zymosan (25  $\mu$ g/ml) for 1 hour. ROS production was measured by flow cytometry as described in the methods. **(E)** WT bone marrow neutrophils ( $n = 3$  mice) were pretreated for 30 min with vehicle control or mitochondrial ROS scavengers, Mito-TEMPO (10  $\mu$ M) or MitoQ (2  $\mu$ M), and then stimulated with either zymosan (25  $\mu$ g/ml) or HKCA (MOI=10) for 6 hours. *Xbp1s* transcript levels were measured using quantitative RT-PCR. **(F)** Bone marrow-resident monocytes isolated from WT mice ( $n = 3-6$  mice) were pretreated with VAS-2870 or hydralazine and then stimulated for 6 hours with HKCA (MOI=10) or zymosan (25  $\mu$ g/ml). Representative violin plots are shown from at least 2 independent experiments with similar results. *Xbp1s* transcript levels were measured using quantitative RT-PCR. Data are shown as violin plots. One-way ANOVA (Tukey's test) was used for statistical analysis; \* $P < 0.05$ , \*\* $P < 0.005$ , \*\*\*\*  $P < 0.0001$ . gMFI, geometric mean fluorescence intensity. ns, not significant.

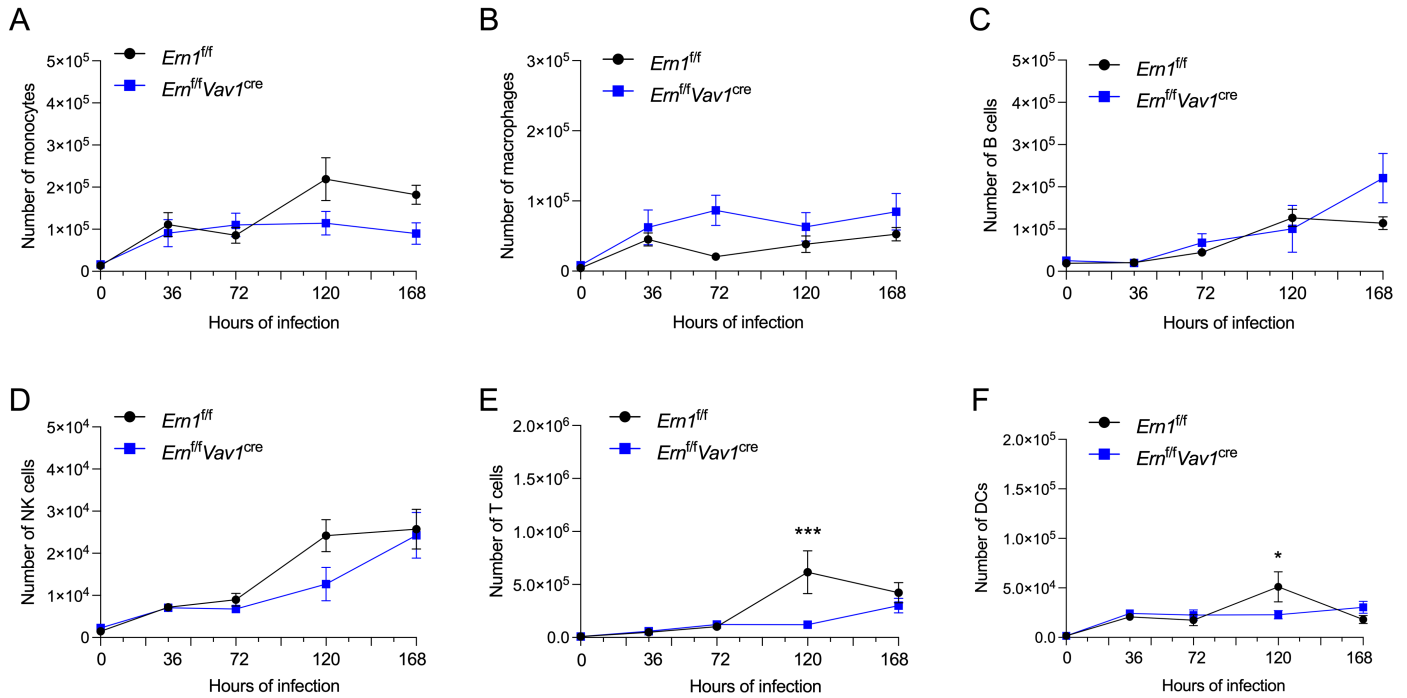
### Supplementary Figure 3



**Supplementary Figure 3. Loss of IRE1 $\alpha$  in neutrophils increases overall survival in mice with systemic candidiasis without altering their anti-*C. albicans* effector functions.** (A) *Em1<sup>fl/fl</sup>* ( $n = 9$ ) or *Em1<sup>fl/fl</sup>Mrp8<sup>cre</sup>* ( $n = 9$ ) mice were infected i.v. with  $10^5$  *C. albicans* and overall host survival was monitored. (B) *Em1<sup>WT</sup>* or *Em1<sup>KO</sup>* neutrophils ( $n = 3$  per genotype) were incubated with FITC-labeled zymosan for 30 min and their phagocytic capacity was assessed by FACS. (C-F) *Em1<sup>WT</sup>* or *Em1<sup>KO</sup>* neutrophils ( $n = 2$  per genotype) were stimulated with (C) HKCA (MOI=5), (D) zymosan (25  $\mu$ g/ml) or (E) PMA (50 nM) for 1 h. (F) ROS production was then quantified by FACS using the intensity of DCF signal generated. (G) *Em1<sup>WT</sup>* or *Em1<sup>KO</sup>* neutrophils ( $n = 4$  per genotype) were

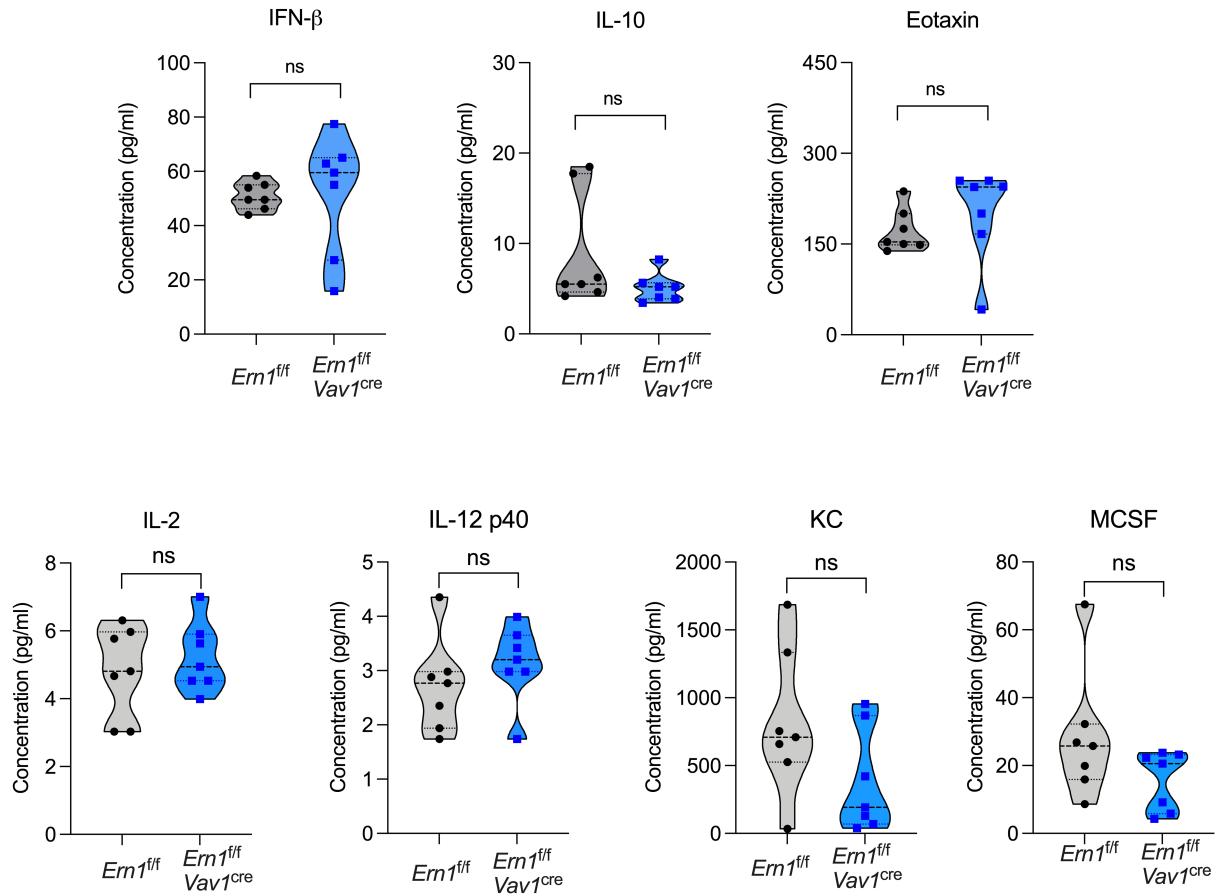
stimulated with HKCA (MOI=5) or zymosan (25  $\mu$ g/ml) for 6 h and myeloperoxidase (MPO) production was measured in supernatants by ELISA. **(H)** *Em1*<sup>WT</sup> or *Em1*<sup>KO</sup> neutrophils ( $n = 5-6$  per genotype) were stimulated with HKCA (MOI=20), zymosan (25  $\mu$ g/ml) or PMA (100 nM) for 8 hours and DNA release was measured as a marker for NETosis using Sytox green. **(I)** *Em1*<sup>WT</sup> or *Em1*<sup>KO</sup> neutrophils ( $n = 4$ ) were isolated and co-cultured with the yeast form of *C. albicans* for the indicated time points and CFUs were determined by serial dilutions on YPD agar. Percent survival was determined by normalization to *C. albicans* cultured without neutrophils. Data are shown as mean  $\pm$  SEM. **(A)** Log-rank test \* $P < 0.05$ . **(G and H)** One-way ANOVA (Tukey's test) was used for statistical analysis; ns, not significant; MFI, mean fluorescence intensity; gMFI, geometric mean fluorescence intensity.

## Supplementary Figure 4



**Supplementary Figure 4. Additional immune cells infiltrating the kidney during systemic *C. albicans*-infection.** *Em1<sup>fl/fl</sup>* or *Em1<sup>fl/fl</sup> Vav1<sup>cre</sup>* mice ( $n = 3-4$  per genotype per time point) were infected i.v. with  $10^5$  *C. albicans* cells and the number of (A) monocytes, (B) macrophages, (C) B cells, (D) NK cells, (E) T cells, and (F) DCs in the kidney were determined by flow cytometry at the indicated times. Data are shown as mean  $\pm$  SEM. Two-way ANOVA (Šidák's multiple comparisons test). \* $P < 0.05$ , \*\*\* $P < 0.0005$ .

## Supplementary Figure 5



**Supplementary Figure 5. Additional cytokines in kidney homogenates from *C. albicans*-infected mice.** *Em1<sup>fl/fl</sup>* ( $n = 7$ ) or *Em1<sup>fl/fl</sup> Vav1<sup>cre</sup>* ( $n = 7$ ) mice were infected i.v. with  $10^5$  *C. albicans* and expression of the indicated factors was determined by ELISA in total kidney homogenates 3 days post-infection. Data are shown as violin plots with dots representing independent mice. Two-tailed Student's *t*-test was used for statistical analysis. ns, not significant.



### Supplementary Table 1.

Differentially expressed genes in kidney-infiltrating neutrophils and monocytes isolated from *Ern1<sup>fl/fl</sup>* vs. *Ern1<sup>fl/fl</sup>Vav1<sup>cre</sup>* mice systemically infected with *C. albicans* for 36 hours.

P.adj<0.05, Log2FoldChange >=1.0. UP, upregulated; DN, downregulated

Gene	DEG	Log2FoldChange	P.adj	Downstream pathways - Hallmark gene sets
<i>Oas1g</i>	UP	3.232091379	2.95E-12	
<i>H2-DMb2</i>	UP	3.203485546	4.15E-09	
<i>Ly6a</i>	UP	3.389266255	1.71E-08	
<i>ligp1</i>	UP	3.981503671	4.58E-07	
<i>Apol9a</i>	UP	5.139129155	1.14E-06	
<i>Gm4951</i>	UP	5.215875903	1.88E-06	
<i>Gbp8</i>	UP	3.52817572	4.16E-06	
<i>Ifi2712a</i>	UP	2.40344031	4.79E-06	
<i>Slc4a8</i>	UP	2.726860312	1.02E-05	
<i>Oas1a</i>	UP	1.306207703	1.46E-05	
<i>Gm4841</i>	UP	3.922026056	4.71E-05	
AC125149.3	UP	2.216308261	5.01E-05	
<i>Hist3h2a</i>	UP	1.358142636	5.82E-05	
<i>Commd10</i>	UP	2.500131721	6.78E-05	
<i>Adi1</i>	UP	1.446226755	0.000180868	
<i>Casp12</i>	UP	2.776045245	0.000180868	
<i>Ccnd1</i>	UP	2.076062651	0.000214936	
BC023105	UP	3.902767585	0.000309536	
<i>Bckdhb</i>	UP	1.63961766	0.000309536	
<i>Smim24</i>	UP	1.523782808	0.000309536	
<i>Gbp4</i>	UP	2.155434105	0.000401785	
<i>Ramp1</i>	UP	1.285300689	0.000698441	
<i>Scimp</i>	UP	2.049182673	0.000801816	
AC132444.2	UP	2.257859962	0.001080291	
AC132444.4	UP	2.257859962	0.001080291	
AC133103.6	UP	2.257859962	0.001080291	
<i>Ubd</i>	UP	4.344593784	0.001095723	
<i>Cd200r4</i>	UP	1.476408397	0.001110358	
<i>Khk</i>	UP	1.237857803	0.001376305	
<i>Grap2</i>	UP	3.108341882	0.001785902	
<i>Fcgrt</i>	UP	1.067513624	0.002470116	
<i>Ifitm3</i>	UP	1.159285309	0.003885756	
<i>Nme3</i>	UP	1.18402526	0.003969311	
<i>Cryab</i>	UP	2.75255852	0.004053168	
<i>Tcea3</i>	UP	1.406040878	0.004065582	
<i>Camp</i>	UP	4.701632212	0.007555553	
<i>Hint2</i>	UP	1.464080466	0.007566823	
<i>Tmem205</i>	UP	1.439318102	0.007767448	

<i>Apoh</i>	UP	2.740138305	0.011044951	
<i>Sectm1b</i>	UP	2.582254962	0.011044951	
<i>Ccdc34</i>	UP	1.75117771	0.011261551	
<i>Ifit1bl1</i>	UP	2.274692913	0.011963773	
<i>H2-Eb1</i>	UP	2.939587697	0.012318104	
<i>Gbp6</i>	UP	2.192091354	0.013063743	
<i>Pigr</i>	UP	3.93665134	0.015344319	
<i>Epsti1</i>	UP	1.007335245	0.017079724	
<i>Nxn</i>	UP	1.544503478	0.018479486	
<i>Maf</i>	UP	2.059623179	0.018857227	
<i>Hoxa7</i>	UP	2.701686827	0.019271808	
<i>Gbp10</i>	UP	3.507591017	0.0207083	
<i>Apol7c</i>	UP	4.05184554	0.021165601	
<i>AC168977.1</i>	UP	1.463842895	0.023255764	
<i>Xaf1</i>	UP	1.654432913	0.023426286	
<i>Rnf227</i>	UP	2.304538187	0.024141237	
<i>Apol9b</i>	UP	3.961197664	0.026003455	
<i>Mitd1</i>	UP	1.20634796	0.026290847	
<i>Hdac11</i>	UP	5.218413566	0.027243081	
<i>AC132444.1</i>	UP	2.081988305	0.028039066	
<i>Ifi208</i>	UP	1.890164083	0.028039066	
<i>Rps27</i>	UP	1.087105144	0.03078014	
<i>Atp5l</i>	UP	1.007489686	0.034475582	
<i>Cpq</i>	UP	1.287278066	0.034551724	
<i>Dctpp1</i>	UP	1.20302006	0.035038027	
<i>C4b</i>	UP	1.557325163	0.035226493	
<i>Pla2g16</i>	UP	1.903003856	0.035226493	
<i>Ccdc107</i>	UP	1.32751598	0.037582108	
<i>Tgtp2</i>	UP	1.562238402	0.037582108	
<i>Climn</i>	UP	2.209363388	0.038025183	
<i>H2-Aa</i>	UP	2.556981724	0.040420453	
<i>Cisd1</i>	UP	1.134485956	0.043011749	
<i>Gm16026</i>	UP	1.665547866	0.044366098	
<i>Timm10b</i>	UP	1.155789109	0.04455419	
<i>Ighm</i>	UP	1.035307197	0.044576796	
<i>Smim4</i>	UP	1.202240535	0.046055339	
<i>Mcts2</i>	UP	2.539776615	0.046181067	
<i>Cryl1</i>	UP	1.129176672	0.047943986	
<i>Rab13</i>	UP	2.040784748	0.047943986	
<i>Vill</i>	UP	1.108867264	0.047943986	
<i>Fau</i>	UP	1.079821271	0.048564021	
<i>Crim1</i>	UP	2.256496654	0.049153842	
<i>H2-Ab1</i>	UP	2.400859347	0.049504026	
<i>Ceacam1</i>	DN	-1.988805987	4.15E-09	
<i>Nr4a1</i>	DN	-2.152063916	7.05E-09	HALLMARK_TNFA_SIGNALING_VIA_NFKB

<i>Stk10</i>	DN	-1.625835001	7.05E-09	
<i>Fam20c</i>	DN	-2.526752492	1.62E-07	
<i>Tbc1d8</i>	DN	-2.48423362	3.30E-07	
<i>Tiam2</i>	DN	-2.008482429	6.33E-06	
<i>Adgb</i>	DN	-4.213969269	8.69E-06	
<i>Cxcl14</i>	DN	-4.050168573	9.83E-06	
<i>Gm38431</i>	DN	-3.912335866	2.18E-05	
<i>Dock5</i>	DN	-1.776524228	6.42E-05	
<i>Tbc1d2b</i>	DN	-1.757772647	9.58E-05	
<i>Zmiz1</i>	DN	-1.09977427	0.00013969	
<i>Itgb3</i>	DN	-2.698024074	0.000180868	HALLMARK_INFLAMMATORY_RESPONSE, HALLMARK_IL6_JAK_STAT3_SIGNALING
<i>Slc6a12</i>	DN	-4.315717071	0.000180868	
<i>Arg1</i>	DN	-4.025086764	0.000198969	
<i>Fkbp5</i>	DN	-2.433406704	0.000198969	
<i>Hic1</i>	DN	-3.524529567	0.00025309	
<i>Tnfrsf9</i>	DN	-3.845073558	0.000309536	HALLMARK_TNFA_SIGNALING_VIA_NFKB, HALLMARK_INFLAMMATORY_RESPONSE, HALLMARK_IL2_STAT5_SIGNALING
<i>Plagl2</i>	DN	-1.769699281	0.000330641	
<i>Fabp4</i>	DN	-3.261477057	0.000357474	
<i>Tg</i>	DN	-3.012118635	0.000357474	
<i>Itgax</i>	DN	-2.511488968	0.000485695	
<i>Naip1</i>	DN	-4.829189672	0.000511224	
<i>lkbke</i>	DN	-2.204227521	0.000524476	
<i>Tmem119</i>	DN	-3.021863873	0.000620004	
<i>Tnip1</i>	DN	-1.658014742	0.000629866	HALLMARK_TNFA_SIGNALING_VIA_NFKB
<i>Nos2</i>	DN	-2.93310251	0.000716616	
<i>Ace</i>	DN	-1.87376367	0.00080052	
<i>Adam17</i>	DN	-1.54609317	0.001080291	
<i>Nfatc3</i>	DN	-1.296041476	0.001080291	
<i>Sema4d</i>	DN	-1.857170531	0.001096826	HALLMARK_INFLAMMATORY_RESPONSE
<i>Itgb1</i>	DN	-1.404221175	0.001141832	
<i>Trem14</i>	DN	-2.172595734	0.001173512	
<i>Nfkb2</i>	DN	-1.409977429	0.001184046	HALLMARK_TNFA_SIGNALING_VIA_NFKB
<i>Nfe2l1</i>	DN	-1.305218519	0.001232878	
<i>St6gal1</i>	DN	-2.715715758	0.001246543	
<i>Tnfrsf1b</i>	DN	-1.379691198	0.001376305	HALLMARK_INFLAMMATORY_RESPONSE, HALLMARK_IL2_STAT5_SIGNALING, HALLMARK_IL6_JAK_STAT3_SIGNALING
<i>Ibtk</i>	DN	-1.575353545	0.001784011	
<i>Rap1gds1</i>	DN	-1.009878284	0.001900872	
<i>Col5a1</i>	DN	-4.065328662	0.002710904	
<i>Ifnar1</i>	DN	-1.310049699	0.002710904	HALLMARK_INFLAMMATORY_RESPONSE, HALLMARK_IL6_JAK_STAT3_SIGNALING
<i>Ptprj</i>	DN	-1.395249055	0.003120474	

<i>Adgre4</i>	DN	-2.296789497	0.003525844	
<i>Malt1</i>	DN	-1.283531565	0.003758597	
<i>Slc7a2</i>	DN	-3.428305246	0.004210174	HALLMARK_INFLAMMATORY_RESPONSE
<i>Il4i1</i>	DN	-2.093601792	0.004431651	
<i>Acp5</i>	DN	-1.396752531	0.004453897	
<i>Cdc42ep4</i>	DN	-1.345361876	0.004623762	
<i>Jchain</i>	DN	-4.561664262	0.004762463	
<i>Pdgfb</i>	DN	-2.603551704	0.005190962	
<i>Myo1c</i>	DN	-1.527898096	0.00580679	HALLMARK_IL2_STAT5_SIGNALING
<i>Ltbp3</i>	DN	-4.080768727	0.006122409	
<i>Has1</i>	DN	-3.712505146	0.006393246	
<i>Creb3l2</i>	DN	-2.741073315	0.006400908	
<i>Sacm1l</i>	DN	-1.325465789	0.006400908	
<i>Kdm5c</i>	DN	-1.342010108	0.010276852	
<i>Adora2a</i>	DN	-2.246079757	0.011152543	
<i>P4ha1</i>	DN	-1.696623023	0.011152543	HALLMARK_IL2_STAT5_SIGNALING
<i>Tmc6</i>	DN	-1.164999867	0.011152543	
<i>Gpr84</i>	DN	-1.714093539	0.011963773	
<i>Spata13</i>	DN	-1.963945301	0.011963773	
<i>Ndufaf7</i>	DN	-1.086323477	0.012305316	
<i>Gm21985</i>	DN	-7.189605336	0.012318104	
<i>Ero1l</i>	DN	-3.074328375	0.012817858	
<i>Shc1</i>	DN	-1.03335912	0.013307052	
<i>Wnt6</i>	DN	-2.085610045	0.013307052	
<i>Orai2</i>	DN	-1.013940594	0.013928597	
<i>Tmc8</i>	DN	-1.464213739	0.014054157	
<i>Bhlhe40</i>	DN	-2.382643831	0.014752568	HALLMARK_TNFA_SIGNALING_VIA_NFKB, HALLMARK_IL2_STAT5_SIGNALING
<i>Cyth3</i>	DN	-1.970592431	0.015636755	
<i>Pan2</i>	DN	-1.279272234	0.017079724	
<i>Nfkbie</i>	DN	-1.231331098	0.019271808	HALLMARK_TNFA_SIGNALING_VIA_NFKB
<i>Plekhm3</i>	DN	-1.110640588	0.019439476	
<i>Smpdl3b</i>	DN	-1.896573552	0.020132829	
<i>Spg11</i>	DN	-1.506075171	0.020543885	
<i>Adam10</i>	DN	-1.278695278	0.0207083	
<i>Fam135a</i>	DN	-3.138066263	0.021080179	
<i>Arfgef2</i>	DN	-1.452255668	0.021803462	
<i>Kpna1</i>	DN	-1.045169458	0.021993366	
<i>C3</i>	DN	-1.37115169	0.022220393	
<i>Abca1</i>	DN	-2.579011637	0.022729768	HALLMARK_TNFA_SIGNALING_VIA_NFKB, HALLMARK_INFLAMMATORY_RESPONSE
<i>Plpp5</i>	DN	-1.702498899	0.023431519	
<i>Ubp1</i>	DN	-3.167163779	0.024141237	
<i>Elf4</i>	DN	-1.188220314	0.024988497	
<i>Tgm3</i>	DN	-2.440030591	0.025067011	

<i>Acp2</i>	DN	-1.943235457	0.026662889	
<i>Rps6ka4</i>	DN	-1.235922004	0.027779287	
<i>Gys1</i>	DN	-2.385319182	0.028039066	
<i>Il10</i>	DN	-3.732763622	0.028039066	HALLMARK_INFLAMMATORY_RESPONSE, HALLMARK_IL2_STAT5_SIGNALING
<i>S1pr5</i>	DN	-2.030749978	0.028150631	
<i>Sec24d</i>	DN	-2.764348104	0.028986659	
<i>Tbc1d9b</i>	DN	-1.111551993	0.03078014	
<i>Ubqln1</i>	DN	-1.12434983	0.03078014	
<i>Frs2</i>	DN	-1.280505667	0.032400753	
<i>Jag1</i>	DN	-2.304506625	0.032400753	HALLMARK_TNFA_SIGNALING_VIA_NFKB
<i>Cd300e</i>	DN	-2.518290701	0.033139761	
<i>Tank</i>	DN	-1.294686928	0.03383381	HALLMARK_TNFA_SIGNALING_VIA_NFKB
<i>Eno3</i>	DN	-1.869092324	0.035226493	HALLMARK_IL2_STAT5_SIGNALING
<i>Dennd4b</i>	DN	-1.146758397	0.035383651	
<i>Itch</i>	DN	-1.085504096	0.035383651	
<i>Tnfai3</i>	DN	-2.247852235	0.035579275	HALLMARK_TNFA_SIGNALING_VIA_NFKB
<i>Hip1</i>	DN	-1.142316141	0.037582108	
<i>Pygo2</i>	DN	-1.048828733	0.037582108	
<i>Jade3</i>	DN	-2.421967462	0.039336806	
<i>Phf2</i>	DN	-1.953464473	0.044214599	HALLMARK_IL2_STAT5_SIGNALING
<i>Cnot1</i>	DN	-1.318001649	0.046407323	
<i>Dhx38</i>	DN	-1.22416948	0.046538442	
<i>Irak3</i>	DN	-2.166800432	0.046760382	
<i>Pdpr</i>	DN	-2.45034046	0.048239154	HALLMARK_INFLAMMATORY_RESPONSE

**Supplementary Table 2.** Primers used in this study

Species	Transcript	Primer direction	Sequence 5'-3'	Purpose
mouse	<i>Xbp1</i>	Forward	ACACGCTTGGGAATGGACAC	Splicing assay
		Reverse	CCATGGGAAGATGTTCTGGG	
mouse	<i>Actb</i>	Forward	CTCAGGAGGAGCAATGATCTTGAT	RT-qPCR
		Reverse	TACCACCATGTACCCAGGCA	
mouse	<i>Xbp1s</i>	Forward	AAGAACACGCTTGGGAATGG	RT-qPCR
		Reverse	CTGCACCTGCTGCGGAC	
mouse	<i>Ddit3</i>	Forward	GTCCTAGCTTGGCTGACAGA	RT-qPCR
		Reverse	TGGAGAGCGAGGGCTTTG	
mouse	<i>Hspa5/BiP</i>	Forward	TCATCGGACGCACTTGGAA	RT-qPCR
		Reverse	CAACCACCTTGAATGGCAAGA	
mouse	<i>Dnajb9/ERdj4</i>	Forward	TAAAAGCCCTGATGCTGAAGC	RT-qPCR
		Reverse	TCCGACTATTGGCATCCGA	
mouse	<i>Sec61a1</i>	Forward	CTATTTCCAGGGCTTCCGAGT	RT-qPCR
		Reverse	AGGTGTTGTACTGGCCTCGGT	
mouse	<i>Atf4</i>	Forward	GAGCTTCCTGAACAGCGAAGTG	RT-qPCR
		Reverse	TGGCCACCTCCAGATAGTCATC	
mouse	<i>Erp44</i>	Forward	GCTGAAACGACACCAGTCAG	RT-qPCR
		Reverse	CAGATGCTCCTTGCTGCTC	
mouse	<i>Rpn1</i>	Forward	GTTTCCACAACGACCGAGAT	RT-qPCR
		Reverse	CCTAGGCGTGACAGATAAAGG	
mouse	<i>HGSNAT</i>	Forward	CTGATGACTGTTACCAATGCACC	RT-qPCR
		Reverse	GCACCAAAAAGGAATAGTTTCCA	
mouse	<i>Tapbp</i>	Forward	GGAGGGTGTCTACCTGGCTA	RT-qPCR
		Reverse	AACGGGTGCTGGTGTTAGAG	
mouse	<i>Bloc1s1</i>	Forward	GAAGCGTTGGTGGATCACCT	RT-qPCR
		Reverse	TCACCTCATGGTCCAGCTTTC	

mouse	<i>Ii6</i>	Forward	GAACAACGATGATGCACTTGC	RT-qPCR
		Reverse	TCCAGGTAGCTATGGTACTCC	
mouse	<i>Tnf</i>	Forward	AATGGCCTCCCTCTCATCAGTT	RT-qPCR
		Reverse	CCACTTGGTGGTTTGCTACGA	
mouse	<i>Ii1b</i>	Forward	CTCCACCTCAATGGACAGAA	RT-qPCR
		Reverse	GCCGTCTTTCATTACACAGG	
mouse	<i>Ptgs2</i>	Forward	TGGGTGTGAAGGGAATAAGGAG	RT-qPCR
		Reverse	ATTTGAGCCTTGGGGGTCAG	