

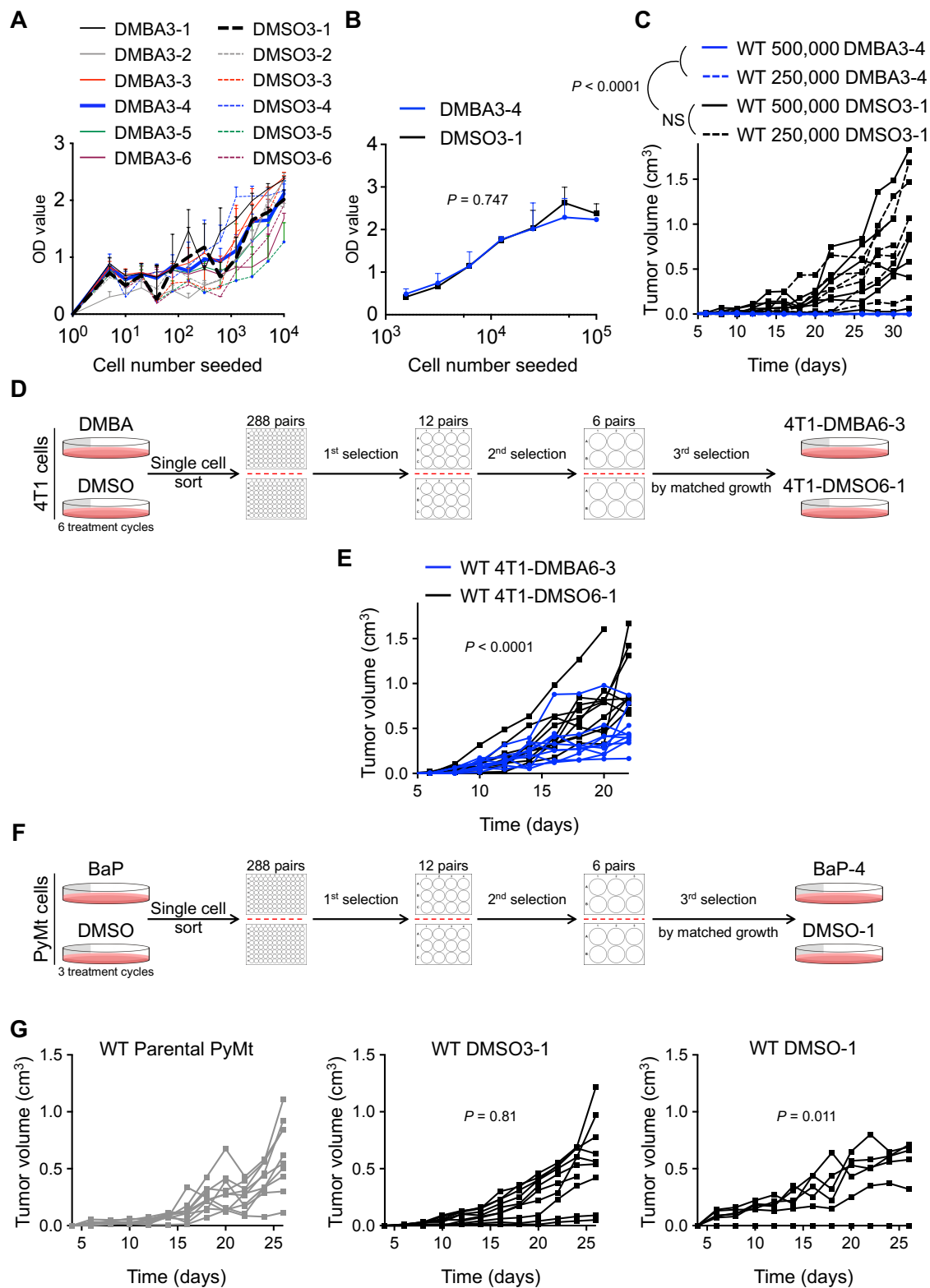
Supplemental Materials for

Carcinogen exposure enhances cancer immunogenicity by blocking the development of an immunosuppressive tumor microenvironment

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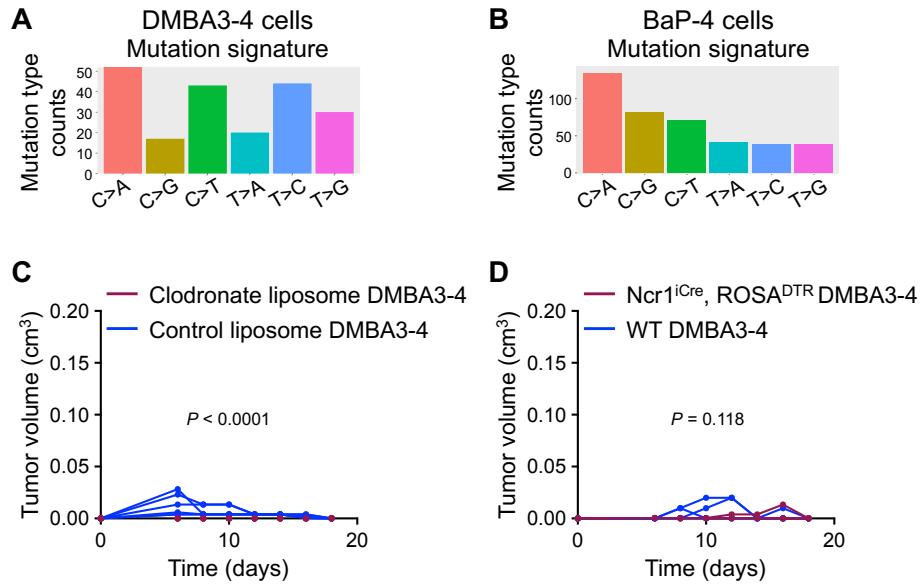
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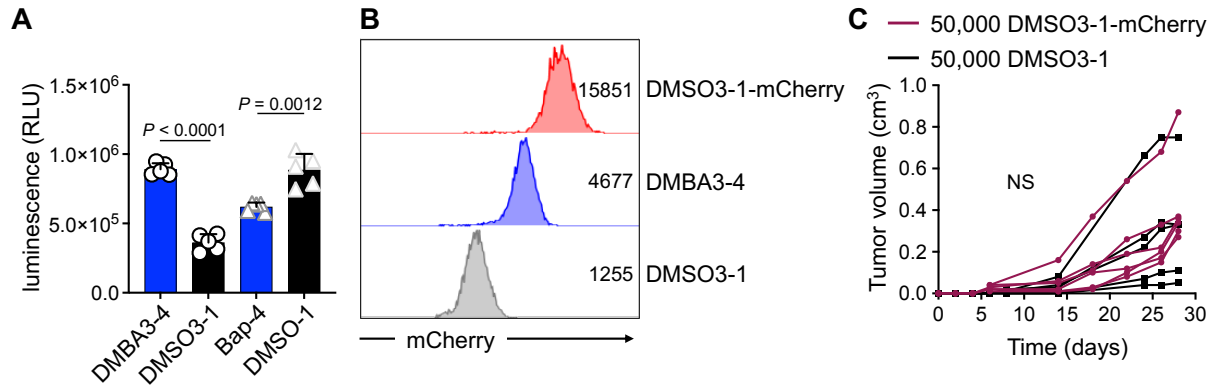
Supplemental Figure 1. Generation of carcinogen-exposed breast cancer cells and examination of their growth kinetics *in vitro* and *in vivo*.

(A) Proliferation assay for the 6 pairs of DMBA- versus DMSO-treated PyMt cell clones *in vitro*. Cells were serially diluted and seeded for MTT assay (OD: optical density). (B) Proliferation assay for DMBA3-4 and DMSO3-1 cells (n=3 per group). (C) DMBA3-4 and DMSO3-1 tumor growth after injection of 250,000 and 500,000 cells into WT C57BL/6 mice (n=8 for 500,000 DMBA3-4, n=8 for 250,000 DMBA3-4, n=6 for 500,000 DMSO3-1 and n=6 for 250,000 DMSO3-1 group). (D) Schematic diagram of 4T1-DMBA6-3 and 4T1-DMSO6-1 cell clones derived from DMBA and DMSO (vehicle control) exposed 4T1 cell line, respectively. (E) 4T1-DMBA6-3 and 4T1-DMSO6-1 tumor growth in syngeneic WT BALB/c mice (n=10 per group). Mice received 100,000 cells per injection site. (F) Schematic diagram of BaP-4 and DMSO-1 cell clones derived from BaP and DMSO (vehicle control) exposed PyMt cell line, respectively. Note that BaP-4 and DMSO-1 cells have similar proliferation rates *in vitro*. (G) Parental PyMt, DMSO3-1 and DMSO-1 tumor growth in WT C57BL/6 mice (n=10 for parental PyMt, n=10 for DMSO3-1 and n=6 for DMSO-1 group). Mice received 100,000 cells per injection site. DMSO3-1 and DMSO-1 tumor growth data are also shown in Figure 1. Two-way ANOVA.



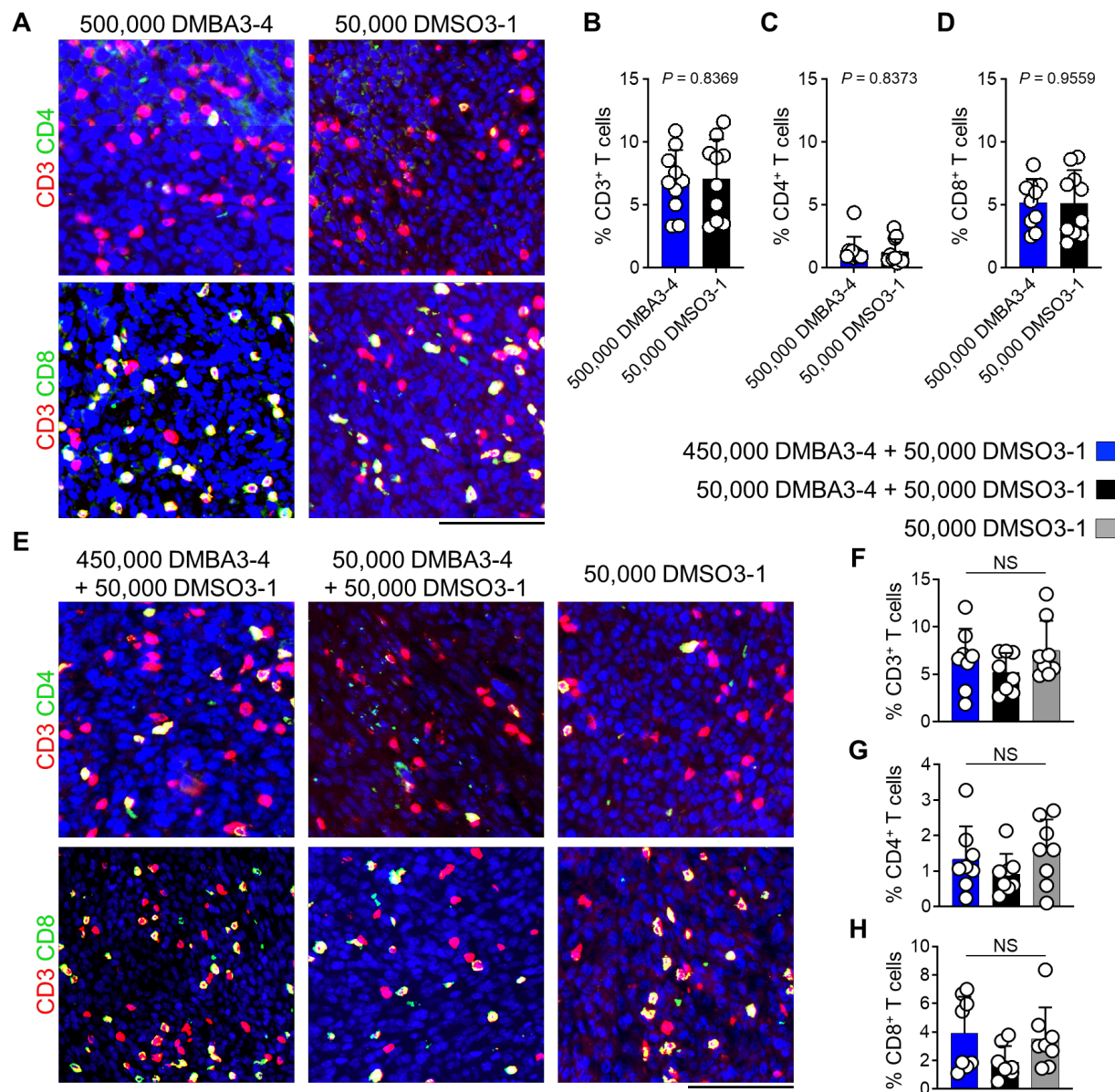
Supplemental Figure 2. Carcinogen-exposed cancer cell mutations and growth in immunodeficient mice.

(A) Somatic mutations count in DMBA3-4 cells categorized based on base substitution types. (B) Somatic mutations count in BaP-4 cells categorized based on base substitution types. (C) DMBA3-4 tumor growth in WT mice treated with clodronate liposome or control liposome (n=6 per group). (D) DMBA3-4 tumor growth in Ncr1^{iCre}, ROSA^{DTR} versus WT mice treated with DT (n=4 per group). Mice received 100,000 cancer cells per injection site. Two-way ANOVA.



Supplemental Figure 3. Luciferase and mCherry effect on breast tumor immunogenicity.

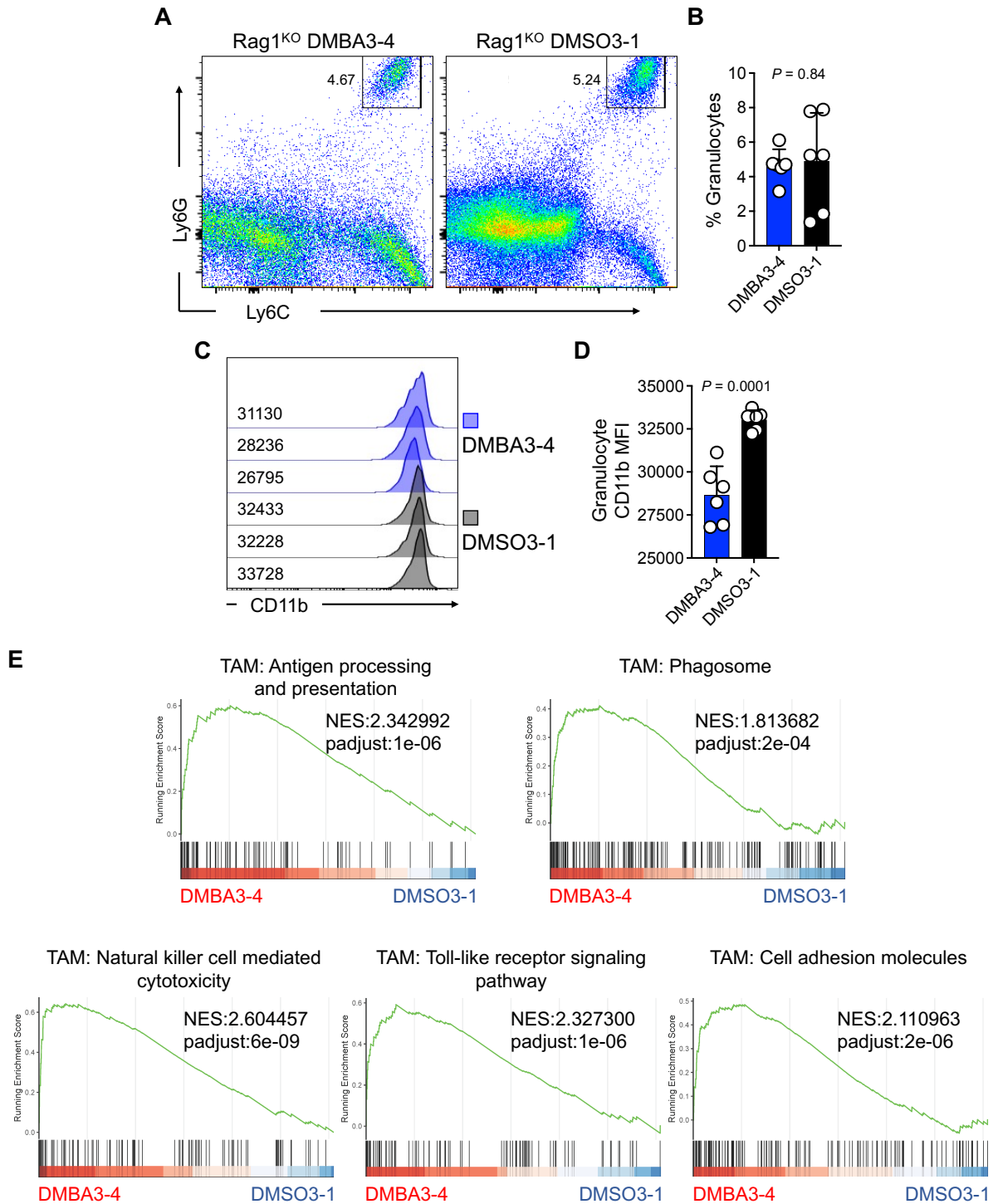
(A) The luciferase enzyme activity levels in DMBA3-4, DMSO3-1, BaP-4 and DMSO-1 cells (n=5 per group, bar graphs show mean + s.d., unpaired *t*-test). (B) mCherry expression in DMBA3-4, DMSO3-1 and DMSO3-1-mCherry cells. Numbers on the flow histograms represent mCherry MFI. (C) DMSO-1-mCherry (test, n=6) and DMSO3-1 (control, n=5) tumor growth in syngeneic WT C57BL/6 mice. NS: not significant, two-way ANOVA.



Supplemental Figure 4. T cell quantification in DMBA3-4 tumors that developed in WT mice.

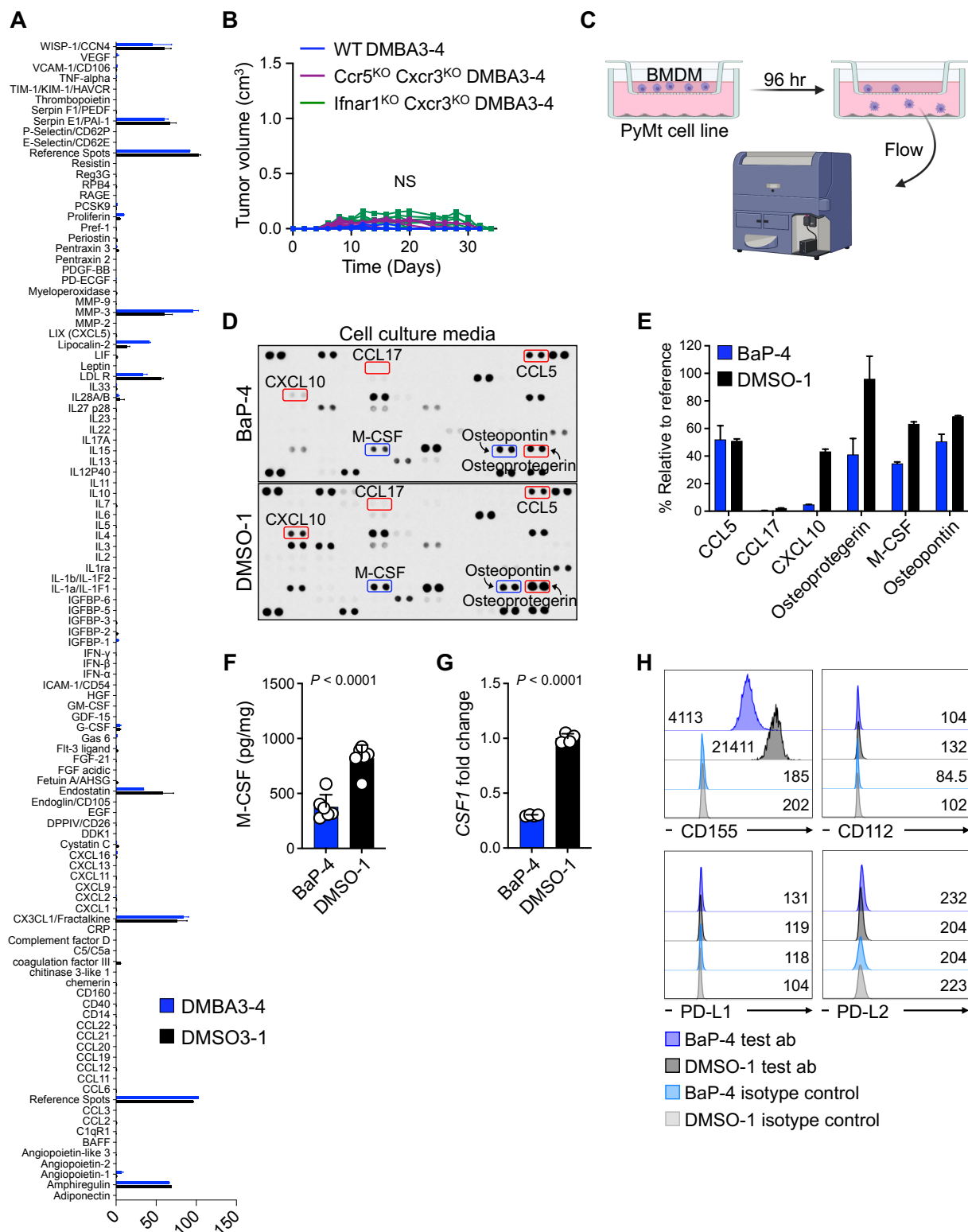
(A) Representative immunofluorescence (IF) images of CD3/CD4- and CD3/CD8-stained DMBA3-4 and DMSO3-1 tumors that grew in a WT mouse highlighted in Figure 3C (scale bar: 100 μ m). (B-D) Quantification of (B) CD3⁺ T cells, (C) CD4⁺ T cells and (D) CD8⁺ T cells in DMBA3-4 and DMSO3-1 tumors in a WT mouse highlighted in Figure 3C. T cells were counted as percent DAPI⁺ cells in 10 randomly selected high-power field (hpf) images of each tumor. Each dot represents a hpf image (unpaired *t*-test). (E) Representative IF images of CD3/CD4- and

CD3/CD8-stained DMBA3-4 plus DMSO3-1 mixed and DMSO3-1 alone tumors in WT mice (scale bar: 100 μm). **(F-H)** Quantification of (F) CD3⁺ T cells, (G) CD4⁺ T cells and (H) CD8⁺ T cells in DMBA3-4 plus DMSO3-1 mixed versus DMSO3-1 alone tumors in WT mice. T cells were counted as percent DAPI⁺ cells and averaged across 10 randomly selected hpf images per tumor. Each dot represents a tumor (n=8 per group, NS: not significant, one-way ANOVA). Bar graphs show mean + s.d.



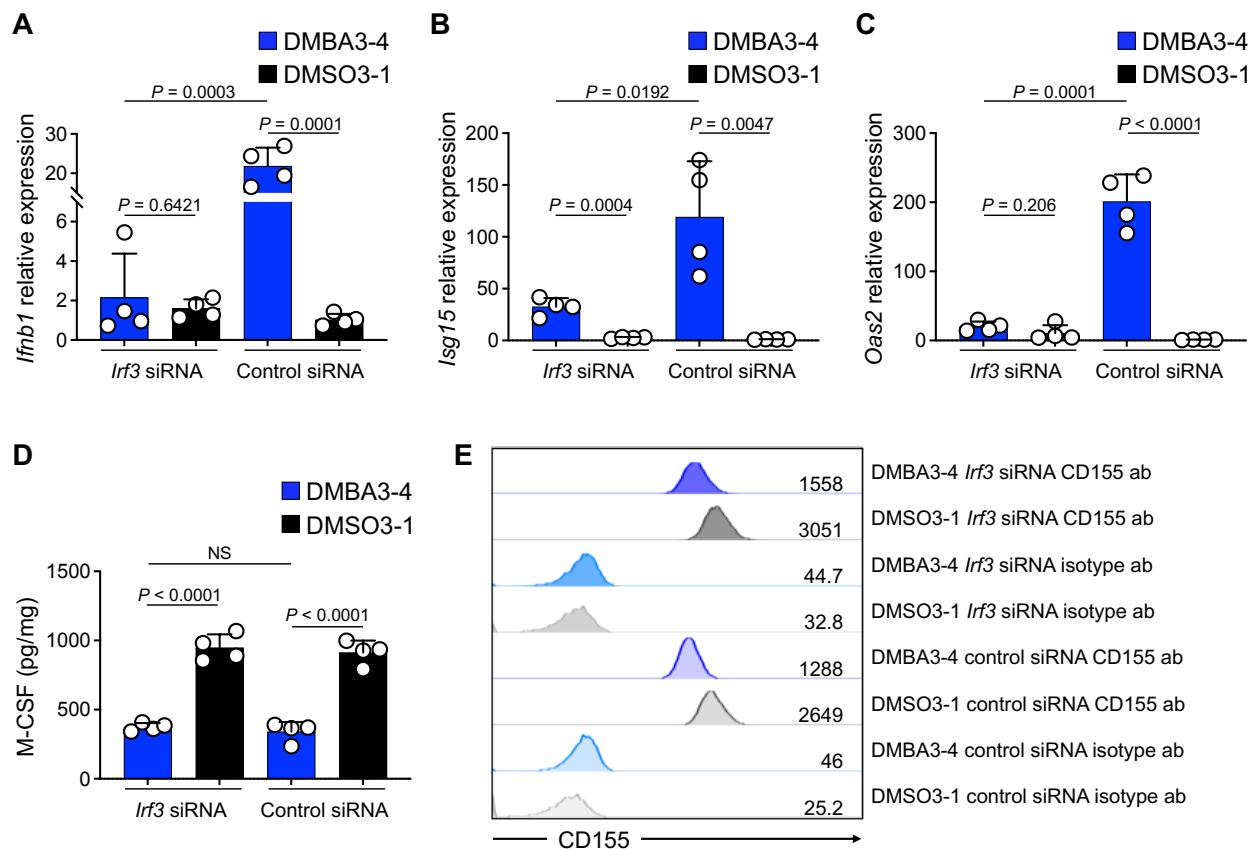
Supplemental Figure 5. Granulocyte and TAM characterization in TME of carcinogen-exposed cancer cells.

(A) Representative flow cytometric analysis of granulocytes in DMBA3-4 and DMSO3-1 tumors from Rag1^{KO} mice. Numbers on the dot plots represent the percent Ly6G⁺ Ly6C⁺ granulocytes of total CD45⁺ CD11b⁺ leukocytes. (B) Ly6G⁺ Ly6C⁺ granulocyte frequencies of total CD45⁺ CD11b⁺ leukocytes in DMBA3-4 and DMSO3-1 tumors (n=6 per group). (C) CD11b expression on Ly6G⁺ Ly6C⁺ granulocytes in DMBA3-4 and DMSO3-1 tumors. Numbers on the flow histograms represent CD11b MFI. (D) Quantification of CD11b MFI on granulocytes in DMBA3-4 and DMSO3-1 tumors (n=6 per group). (E) GSEA analysis of TAMs in DMBA3-4 compared with DMSO3-1 tumors from Rag1^{KO} mice. Unpaired *t*-test (Supplemental Figure 4, B and D), bar graphs show mean + s.d.



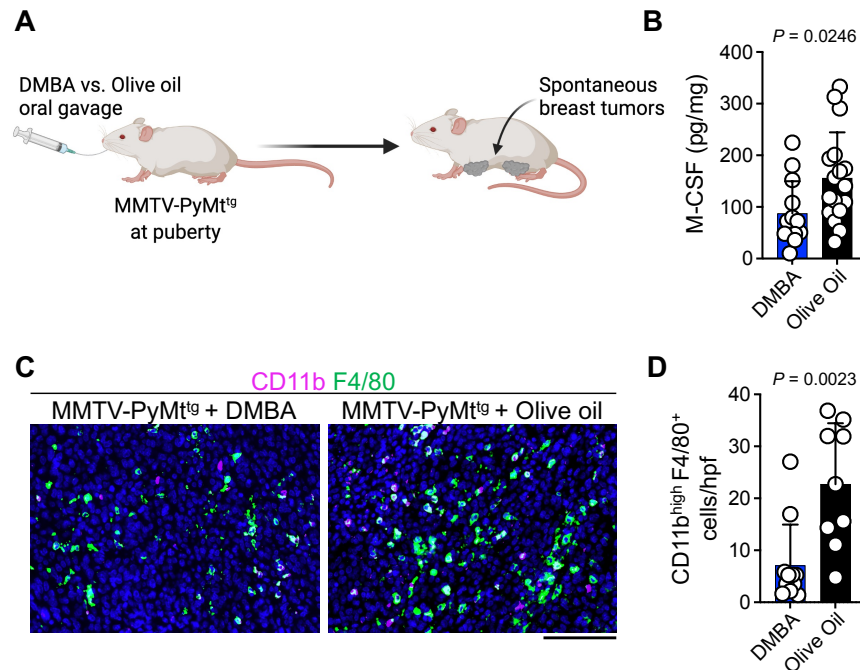
Supplemental Figure 6. Secretome and inhibitory ligand profiling of carcinogen-exposed cancer cells.

(A) Relative protein levels from the cytokine array on supernatant from DMBA3-4 and DMSO3-1 cells. (B) DMBA3-4 tumor growth in WT (n=6), *Ccr5*^{KO} *Cxcr3*^{KO} (n=8) and *lfnar1*^{KO} *Cxcr3*^{KO} mice (n=8). (C) Schematic diagram of BMDM migration assay protocol. (D) Cytokine array on supernatant from BaP-4 and DMSO-1 cells. Red and blue boxes highlight the upregulated and downregulated proteins from secretome analysis of DMBA3-4 compared with DMSO3-1 cells in Figure 5B, respectively. (E) Relative levels of the select proteins from the BaP-4/DMSO-1 cytokine array. (F) M-CSF protein levels in BaP-4 compared with DMSO-1 cell lysates (n=6 per group). (G) *Csf1* mRNA expression levels in BaP-4 compared with DMSO-1 cells (n=4 per group). (H) CD155, CD112, PD-L1 and PD-L2 expression on BaP-4 and DMSO-1 cells. Numbers on the flow histograms represent the ligands' MFI. Two-way ANOVA (NS: not significant, Supplemental Figure 5B) and unpaired *t*-test (Supplemental Figure 5, F and G), bar graphs show mean + s.d.



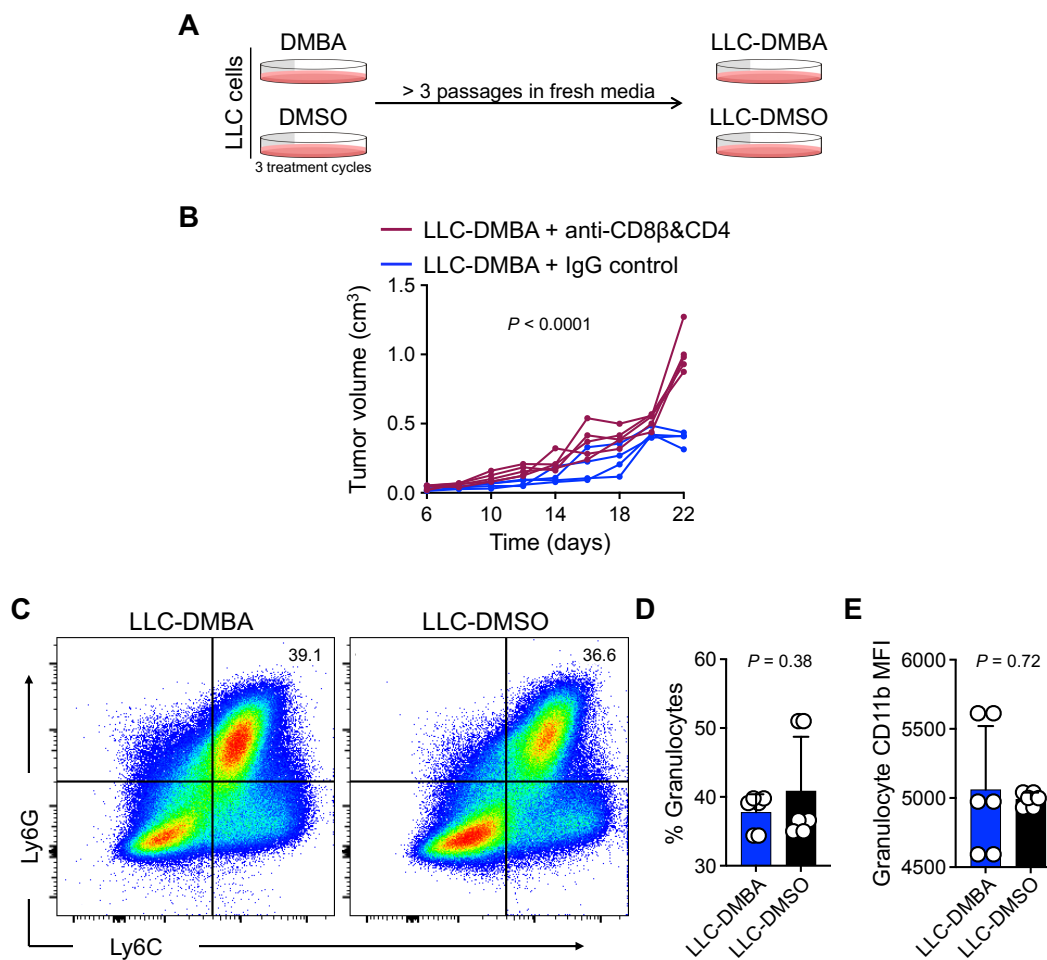
Supplemental Figure 7. Type-I interferon signaling influence on M-CSF and CD155 expression in DMBA3-4 tumor cells.

(A-C) The expression of (A) *Ifnb1*, (B) *Isg15*, and (C) *Oas2* in DMBA3-4 and DMSO3-1 cells after transfection with IRF3 versus control siRNA for 24 hours (n=4 per group). (D) M-CSF protein levels in DMBA3-4 and DMSO3-1 cell lysates after transfection with IRF3 versus control siRNA for 24 hours (n=4 per group). (E) CD155 expression on DMBA3-4 and DMSO3-1 cells after transfection with IRF3 versus control siRNA for 24 hours. Numbers on the flow histograms represent CD155 MFI. Unpaired *t*-test, bar graphs show mean + s.d.



Supplemental Figure 8. The impact of DMBA exposure on the TME of spontaneous breast tumors in MMTV-PyMt^{tg} mice.

(A) Schematic illustration of DMBA and olive oil administration to MMTV-PyMt^{tg} mice at puberty and prior to tumor initiation. (B) M-CSF protein levels in spontaneous breast tumors of DMBA- (n=13) versus olive oil-treated (n=17) MMTV-PyMt^{tg} mice. Each dot represents a tumor sample. (C) Representative IF images of CD11b- and F4/80-stained breast tumors from DMBA- and olive oil-treated MMTV-PyMt^{tg} mice (scale bar: 100 μ m). (D) CD11b^{high} F4/80⁺ TAM counts in breast tumors from DMBA- (n=11) and olive oil-treated (n=9) MMTV-PyMt^{tg} mice. TAMs were quantified in 10 randomly selected hpf images per sample. Each dot represents a tumor sample. Unpaired *t*-test, bar graphs show mean + s.d.



Supplemental Figure 9. Development and characterization of DMBA-exposed LLC cells.

(A) Schematic diagram of LLC-DMBA and LLC-DMSO cells derived from DMBA and DMSO (vehicle control) treated Lewis lung carcinoma (LLC) cells. Note that LLC-DMBA and LLC-DMSO cells were generated as cell pools (i.e., not single cell clones), which showed a similar proliferation rate *in vitro*. (B) LLC-DMBA tumor growth in WT mice treated with anti-CD8 β and anti-CD4 combination antibodies (n=5) versus IgG control antibody (n=4). (C) Representative flow cytometric analysis of granulocytes in LLC-DMBA and LLC-DMSO tumors from WT mice. Numbers on the dot plots represent the percent Ly6G⁺ Ly6C⁺ granulocytes of total CD45⁺ CD11b⁺ leukocytes. (D) Ly6G⁺ Ly6C⁺ granulocyte frequencies of total CD45⁺ CD11b⁺ leukocytes in LLC-DMBA and LLC-DMSO tumors (n=6 per group). (E) Quantification of CD11b MFI on granulocytes

in LLC-DMBA and LLC-DMSO tumors (n=6 per group). Two-way ANOVA (Supplemental Figure 6B) and unpaired *t*-test (Supplemental Figure 6, D and E), bar graphs show mean + s.d.

Supplemental Table 1. Summary of somatic SNV and InDel mutations in DMBA3-4 compared with DMSO3-1 cells.

Mutations	DMBA3-4 vs. DMSO3-1
CDS	41
synonymous_SNV	7
missense_SNV	0
frameshift_deletion	1
frameshift_insertion	1
Nonframeshift_deletion	1
Nonframeshift_insertion	0
stopgain	0
stoploss	0
unknown	0
intronic	74
UTR3	12
UTR5	4
splicing	0
ncRNA_exonic	23
ncRNA_intronic	43
ncRNA_UTR3	0
ncRNA_UTR5	0
ncRNA_splicing	0
upstream	3
downstream	5
intergenic	29
SNV	224
InDel	13
Total	237

Supplemental Table 2. Summary of somatic SNV and InDel mutations in BaP-4 compared with DMSO-1 cells.

Mutations	BaP-4 vs. DMSO-1
CDS	137
synonymous_SNV	27
nonsynonymous_SNV	98
frameshift_deletion	1
frameshift_insertion	0
nonframeshift_deletion	1
nonframeshift_insertion	0
stopgain	9
stoploss	0
unknown	1
intronic	163
UTR3	9
UTR5	10
splicing	3
ncRNA_exonic	32
ncRNA_intronic	63
ncRNA_UTR3	0
ncRNA_UTR5	0
ncRNA_splicing	0
upstream	15
downstream	12
intergenic	26
SNV	413
InDel	57
Total	470

Supplemental Table 3. Antibodies used in the study.

<u>Flow cytometry Antibodies</u>	<u>Conjugate</u>	<u>Clone</u>	<u>Manufacturer</u>	<u>Cat #</u>	<u>Isotype</u>	<u>RRID</u>
CD45	Brilliant Violet 605	30-F11	BioLegend	103155	Rat IgG2b, κ	AB_2650656
CD11b	BUV395	M1/70	BD biosciences	563553	Rat IgG2b, κ	AB_2738276
F4/80	PE-Cy7	BM8	BioLegend	123114	Rat IgG2a, κ	AB_893478
NK1.1	APC	PK136	BioLegend	108710	Mouse IgG2a, κ	AB_313397
MHCII	Alexa Fluor® 488	M5/114.15.2	BioLegend	107616	Rat IgG2b, κ	AB_493523
Arginase 1	PE		R&D Systems	IC5868P	Polyclonal Sheep IgG	AB_10718118
CD206	PerCP-Cy5.5	C068C2	BioLegend	141716	Rat IgG2a, κ	AB_2561992
CD86	Alexa Fluor® 700	GL-1	BioLegend	105024	Rat IgG2a, κ	AB_493721
Ly6C	Brilliant Violet 785	HK1.4	BioLegend	128041	Rat IgG2c, κ	AB_2565852
Ly6G	Alexa Fluor® 647	1A8	BioLegend	127610	Rat IgG2a, κ	AB_1134159
CD155	APC	TX56	BioLegend	131510	Rat IgG2a, κ	AB_10645507
CD112	APC	829038	R&D Systems	FAB3869A	Rat IgG2A	
PD-L1	Brilliant Violet 421	10F.9G2	BioLegend	124315	Rat IgG2b, κ	AB_10897097
PD-L2	APC	TY25	BioLegend	107210	Rat IgG2a, κ	AB_2566345
<u>Immunofluorescence Antibodies</u>	<u>Conjugate</u>	<u>Clone</u>	<u>Manufacturer</u>	<u>Cat #</u>	<u>Isotype</u>	<u>RRID</u>
CD3	Purified	CD3-12	Abcam	Ab11089	Rat IgG1	AB_2889189
CD4	Purified	EPR19514	Abcam	Ab183685	Rabbit IgG	AB_2686917
CD8a	Purified	D4W2Z	Cell Signaling Technologies	98941	Rabbit IgG	AB_2756376
CD11b	Purified	EPR19387	Abcam	Ab184308	Rabbit IgG	AB_2889154
F4/80	Purified	D2S9RI	Cell Signaling Technologies	70076S	Rabbit IgG	AB_2799771
<u>Depleting/Blocking Antibodies</u>	<u>Conjugate</u>	<u>Clone</u>	<u>Manufacturer</u>	<u>Cat #</u>	<u>Isotype</u>	<u>RRID</u>
<i>InVivo</i> MAb anti-mouse CD4	Purified	GK1.5	BioXCell	BE0003	Rat IgG2b, κ	AB_1107636
<i>InVivo</i> MAb anti-mouse CD8β	Purified	53-5.8	BioXCell	BE0223	Rat IgG1, κ	AB_2687706
<i>InVivo</i> MAb anti-mouse NK1.1	Purified	PK136	BioXCell	BE0036	Mouse IgG2a, κ	AB_1107737
<i>InVivo</i> MAb anti-mouse CSF1R	Purified	AFS98	BioXCell	BE0213	Rat IgG2a, κ	AB_2687699
<i>InVivo</i> MAb anti-mouse TIGIT	Purified	1G9	BioXCell	BE0274	Rat IgG1, κ	AB_10950522

Supplemental Table 4. Primers used for mouse genotyping and quantitative PCR.

<u>Gene</u>	<u>Forward primer</u>	<u>Reverse primer</u>
<u>PCR genotyping</u>		
Batf3 ^{WT}	CCAGACCTTTGAATGTGTGAGG	GTTGTGAGTCGAAACCACGC
Bat3 ^{KO}	CCAGACCTTTGAATGTGTGAGG	AGATGAAAAGGCAGCAAGTGT
Ifng ^{WT}	AGAAGTAAGTGGAAGGGCCCAAG	AGGGAAACTGGGAGAGGAGAAATAT
Ifng ^{KO}	CCTTCTATCGCCTTCTTGACG	AGGGAAACTGGGAGAGGAGAAATAT
Klrk1 ^{WT}	CATAAAGTCCGCTTTGATGTTA	GCAGATTCCCAAATTCTTG
Klrk1 ^{KO}	CATAAAGTCCGCTTTGATGTTA	ATGAACTTCAGGGTCAGCTT
Ifnar ^{WT}	CGAGGCGAAGTGGTAAAAAG	ACGGATCAACCTCATTCCAC
Ifnar ^{KO}	CGAGGCGAAGTGGTAAAAAG	AATTCGCCAATGACAAGA CG
Ccr5 ^{WT}	CAGGCAACAGAGACTCTTGG	TCATGTTCTCCTGTGGATCG
Ccr5 ^{KO}	CTTGGGTGGAGAGGCTATTC	AGGTGAGATGACAGGAGATC
Cxcr3 ^{WT}	GCCTTCCTGCTGGCTTGTAT	AGCAGTGCATGTACCCCATG
Cxcr3 ^{KO}	CTTGGGTGGAGAGGCTATTC	AGCAGTGCATGTACCCCATG
Sting ^{WT}	AGAACGGACAGCCAGTAAGTATACAG	CAATGCTCTCATAGCCTTCACTATC
Sting ^{KO}	AACTTCCTGACTAGGGGAGGAGTAG	CAATGCTCTCATAGCCTTCACTATC
Ticam ^{WT}	AGATGGTTCAGCTGGGTGTC	GGTTCTCCGAACACTCAGTC
Ticam ^{KO}	AGATGGTTCAGCTGGGTGTC	GGTTCTCCGAACACTCAGTT
Myd88 ^{WT}	GTTGTGTGTGTCGACCGT	GTCAGAAACAACCACCACCATGC
Myd88 ^{KO}	CCACCCTTGATGACCCCTA	GTCAGAAACAACCACCACCATGC
Ncr1 ^{iCre}	GACCATGATGCTGGGTTTGGCCAGATG	ATGCGGTGGGCTCTATGGCTTCTG
Cd11c-DTR ^{WT}	ACAACAGAAATCACCTGGA	TGGCAGTGTTAAAATGCAGA
Cd11c-DTR ^{Mut}	ACAACAGAAATCACCTGGA	CGAGAGGACCTCAGACTGCT
Rosa iDTR ^{WT}	AAAGTCGCTCTGAGTTGTTAT	GGAGCGGGAGAAATGGATATG
Rosa iDTR ^{Mut}	AAAGTCGCTCTGAGTTGTTAT	GCGAAGAGTTTGTCTCAACC
<u>qPCR</u>		
Csf1	CGGGCATCATCCTAGTCTTGCTGACTGT	ATAGTGGCAGTATGTGGGGGGCATCCTC
Ifnb1	CAGCTCCAAGAAAGGACGAAC	GGCAGTGTAACCTTTCTGCAT
Isg15	GGTGTCCGTGACTAACTCCAT	TGGAAGGGTAAGACCGTCCT
Oas2	TTGAAGAGGAATACATGCGGAAG	GGGTCTGCATTACTGGCACTT
Gapdh	AATGTGTCCGTCGTGGATCTGA	GATGCCTGCTTACCACCTTCT