

Figure S1. Identified *SMAD3* methylation site by Mass spectrum (A) whole cell extracts (WCE) of HEK293T cells were collected for immunoprecipitation (IP) with *SMAD3* antibody, followed by Immunoblots (IB) analysis. (B) HEK293T cells were serum-starved, treated with *TGFBI*(5ng/ml) for indicated time points, and whole cell extracts (WCE) were collected for immunoprecipitation (IP) with *SMAD3* antibody, followed by Immunoblots (IB) analysis. (C, D) HEK293T cells transfected with *HA-SMAD3* and treated with *TGFBI*(5ng/ml), WCE were collected for IP with *HA* antibody, followed by IB analysis. (E) HEK293T cells were serum-starved, treated with *TGFBI*(5ng/ml) and Dznep, WCE were collected for IP with *SMAD3* antibody, followed by IB analysis. (F) Showing results from mass spectrometry analysis of *SMAD3* potential methylation sites. (G) HEK293T cells transfected with *HA-SMAD3* WT or K53/333R mutant plasmids, WCE were collected for IB analysis. (H) IB analysis of *SMAD3* expression level in *SMAD3* WT and *SMAD3* KO MDA-MB-231 cells. (I) IB analysis of *SMAD3* expression level in MDA-MB-231^{*SMAD3*^{-/-}} cell expressing ectopic *SMAD3* WT or *SMAD3* K53/333R. All immunoblots are performed three times, independently, with similar results.

Figure S2. The biological function of *SMAD3* K53/333R cells phenocopied *SMAD3* knock out cells. (A) MDA-MB-231 cells transfected with *HA-SMAD3* WT or K53/333R mutant plasmids and treated with *TGFBI*(5ng/ml), quantitative RT-PCR analysis of *TGFBI/SMAD3* signaling pathway downstream genes expression, including *CTGF*, *PAIL*, *PDGFB*, and *SMAD7*; **p<0.05. (B) MCF-7 cells transfected with *HA-SMAD3* WT or K53/333R mutant plasmids and treated with *TGFBI*(5ng/ml), quantitative RT-PCR analysis of *TGFBI/SMAD3* signaling pathway downstream genes expression, including *CTGF*, *PAIL*, *P21*, and *SMAD7*; **p<0.05. (C) Transwell cell migration and invasion assay in MDA-MB-231^{*SMAD3*^{-/-}} cells stably transfected with *SMAD3* WT or *SMAD3* K53/333R plasmids. (D) Immunofluorescence (IF) analysis of EMT marks in indicated cells; Scale bar, 50um. (E, F). Representative lung image (E) and H&E-stained lung sections (F). (G, H) Scatter plot showing lung metastatic nodules (G) and lung weight (H); **p<0.05. (I) Representative image of IHC staining for *SMAD3* K53 and K333 tri-methylation and *SMAD3* S423/S425 phosphorylation for

lung sections; scale bar, 100um. All immunoblots are performed three times, independently, with similar results. Error bars are mean \pm s.d. Statistical significance was assessed using Student's two-tailed t-test.

Figure S3. *EZH2* triggers *SMAD3* methylation. (A) HEK293T cells transfected with *HA-SMAD3* or *Flag-EZH2* plasmids, WCE were collected for IP with *HA* or *Flag* antibody, followed by IB analysis. (B, C) WCE from MDA-MB-231 (B) and MCF-7 (C) cells were collected for IP with *EZH2* antibody, followed by IB analysis. (D) MDA-MB-231 cells were serum-starved, treated with *TGFBI*(5ng/ml), and WCE were collected for IP with *SMAD3* antibody, followed by IB analysis. (E) *EZH2* protein was incubated with GST-*SMAD3* WT or GST protein for the *in vitro* binding assay, followed by immunoprecipitation (IP) of GST-*SMAD3* and IB analysis. (F) HEK293T transfected with *HA-SMAD2* or *HA-SMAD3* and *Flag-EZH2*, WCE were collected for IP with *HA* antibody, followed by IB analysis. (G) HEK293T transfected with *HA-SMAD3* and *Flag-EZH2*, then treated with *TGFBI*(5ng/ml), WCE were collected for IP with *HA* antibody, followed by IB analysis. (H) MCF-7 cells transfected with *HA-SMAD3* and *Flag-EZH2*, then treated with *TGFBI*(5ng/ml), WCE were collected for IP with *HA* antibody, followed by IB analysis. (I) HEK293T (left) and MCF-7 (right) cells transfected with *HA-SMAD3* and silenced with control (ShNC) or *EZH2* ShRNA (#1 and #2), then treated with *TGFBI*(5ng/ml), WCE were collected for IP with *HA* antibody, followed by IB analysis. All immunoblots are performed three times, independently, with similar results.

Figure S4. *EZH2* triggers *SMAD3* methylation at K53 and K333. (A) HEK293T cells transfected with *HA-SMAD3* WT or mutant plasmids and *Flag-EZH2* plasmid as indicated, WCE were collected for IP with *HA* antibody, followed by IB analysis. (B) Conducting *in vitro* methylation assays using purified *EZH2* protein as the source of methyltransferase, and the synthetic *SMAD3* non methylation peptides containing K53 or K333 as the substrate. *SMAD3* K53 tri-methylation peptides (left) and *SMAD3* K333 tri-methylation peptides (right) as a positive control. (C) MCF-7 silenced with control (ShNC) or *EZH2* ShRNA (#1 and #2), WCE were collected for IP with *SMAD3* antibody, followed by IB analysis. (D) MDA-MB-231(left) and MCF-7(right) cells transfected

with Vector, *EZH2*^{WT} and treated with *TGFBI*(5ng/ml), WCE were collected for IP with *SMAD3* antibody, followed by IB analysis. **(E)** HEK 293T cells transfected with Vector, *EZH2*^{WT}, *EZH2*^{T372A}, WCE were collected for IP with *SMAD3* antibody, followed by IB analysis. All immunoblots are performed three times, independently, with similar results.

Figure S5. *EZH2*-mediated tumor metastasis depended on *SMAD3* methylation.

(A, B) Trans-well cell migration assay in MCF-7 silenced with control (ShNC) or *EZH2* ShRNA (#1 and #2) **(A)**, quantitative cell migration number was shown **(B)**. **(C)** Trans-well cell migration and invasion assay in MDA-MB-231 cells silenced with control (ShNC) or *EZH2* ShRNA (#1 and #2). **(D)** *Flag-EZH2* plasmid was transfected into MDA-MB-231^{*SMAD3*^{-/-}} cell ectopic expressing *SMAD3* WT or *SMAD3* K53/333R, followed by IB analysis. **(E)** *Flag-EZH2* WT or *Flag-EZH2* H689A plasmid transfected into MDA-MB-231 KO or MDA-MB-231 WT cells, WCE was collected for IB analysis. **(F, G)** Trans-well cell migration assay in MDA-MB-231 KO or MDA-MB-231 WT cells **(F)**, quantitative cell migration number was shown **(G)**; **p<0.05. All immunoblots are performed three times, independently, with similar results. Error bars are mean ± s.d. Statistical significance was assessed using Student's two-tailed t-test.

Figure S6. Deletion of *SMAD3* K53 and K333 methylation inhibited its membrane localization.

(A, B) IB analysis of *EZH2* expression level in MDA-MB-231 **(A)** and HEK293T **(B)** cells silenced with control (ShNC) or *EZH2* ShRNA (#1). **(C)** HEK293T cells silenced with control (ShNC) or *EZH2* ShRNA (#1), cellular membrane was extracted, followed by IB analysis of *SMAD3* expression level. **(D)** MDA-MB-231 cells silenced with control (ShNC) or *EZH2* ShRNA (#1) and treated with *TGFBI*(5ng/ml), immunocytochemistry analysis of the cellular localization of *SMAD3*. **(E)** WCE from MDA-MB-231 cells were collected for IP with *SMAD3* antibody, followed by IB analysis. **(F)** WCE from MDA-MB-231^{*SMAD3*^{-/-}} cell ectopic expressing *SMAD3* WT or *SMAD3* K53/333R were collected for IP with *SMAD3* antibody, followed by IB analysis. **(G)** HEK293T cell transfected with *Flag-EZH2* plasmid, WCE was collected for IP with *SMAD3* antibody, followed by IB analysis. **(H)** HEK293T cells silenced with control (ShNC) or *EZH2* ShRNA (#1 and #2), WCE were collected for IP with *SMAD3*

antibody, followed by IB analysis. All immunoblots and immunocytochemistry assays are performed three times, independently, with similar results.

Figure S7. *EZH2* expression level is upregulated in various kinds of cancer and has a negative cancer patients' survival time. (A) MDA-MB-231 cells treated with *TGFBI*(5ng/ml) with or without TAT-peptides, IF showing the cellular localization of *SMAD3*. (B) Trans-well cell migration assay in MDA-MB-231 treated with TAT-peptides. (C) Analysis of *EZH2* expression level in various kinds of cancer using GEPIA database.

Figure S8. *EZH2* expression level has a positive correlation with *SMAD3* K53/K333 methylation level and *SMAD3* S423/S425 phosphorylation level in breast cancer. (A, B) Analysis of patients' survival time in breast cancer patients(A) or breast patients with lymph node positive(B) grouped by *EZH2* expression level using GEPIA database. (C) Representative image of IHC staining for *EZH2*, *SMAD3* K53 and K333 tri-methylation and *SMAD3* S423/S425 phosphorylation in breast cancer; scale bar, 100um. (D) Representative image of IHC staining of *SMAD3* K53 and K333 tri-methylation in non-metastasis primary breast tumors and lung-metastasis primary breast tumors. (E, F) Representative image (E) of IHC staining of *SMAD3* K53 and K333 tri-methylation in breast cancer, quantitative IHC staining score was shown (F); $p=0.0018$. Statistical significance is assessed using Chi-Square test.

Figure.S1

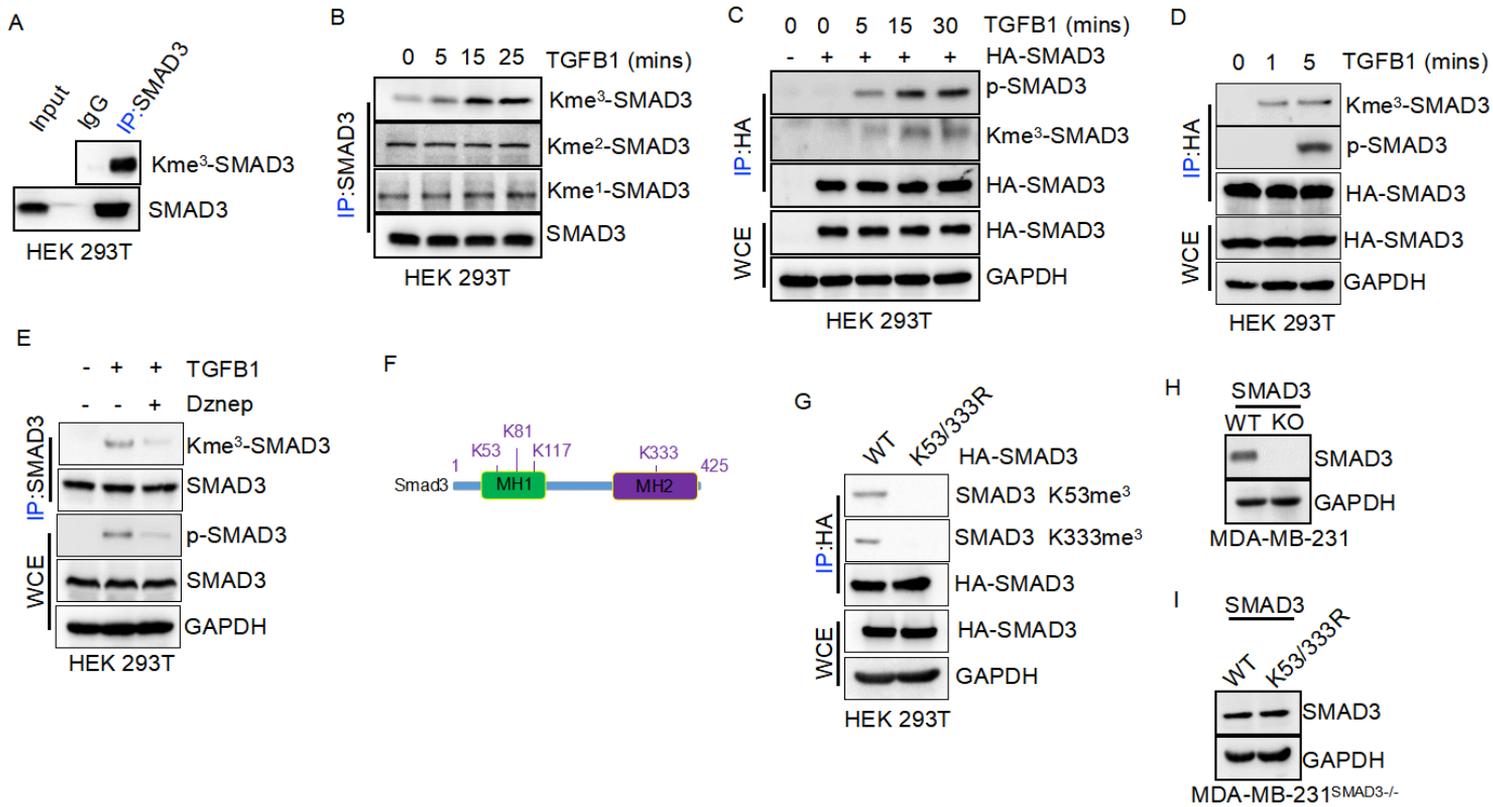


Figure.S2

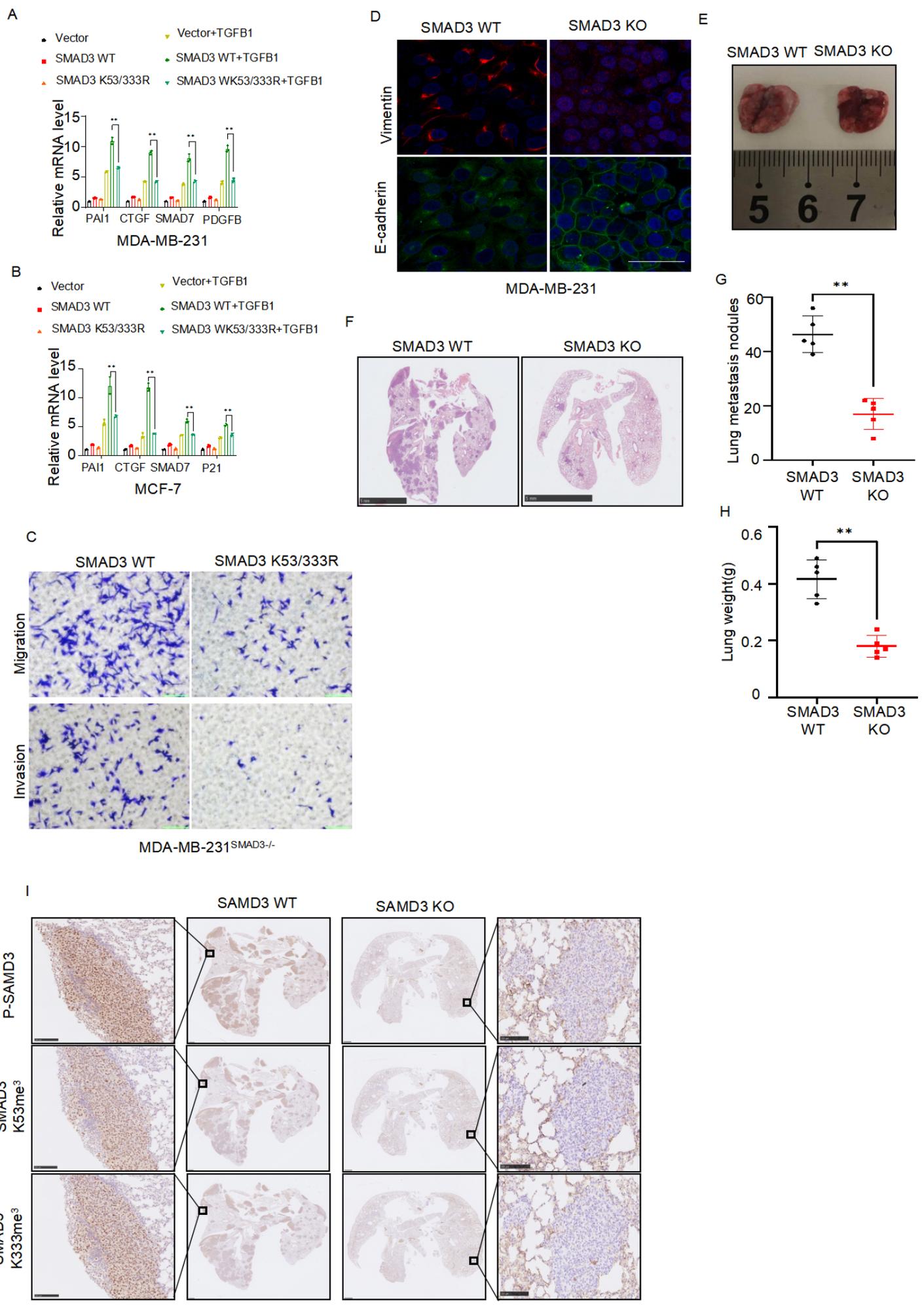


Figure.S3

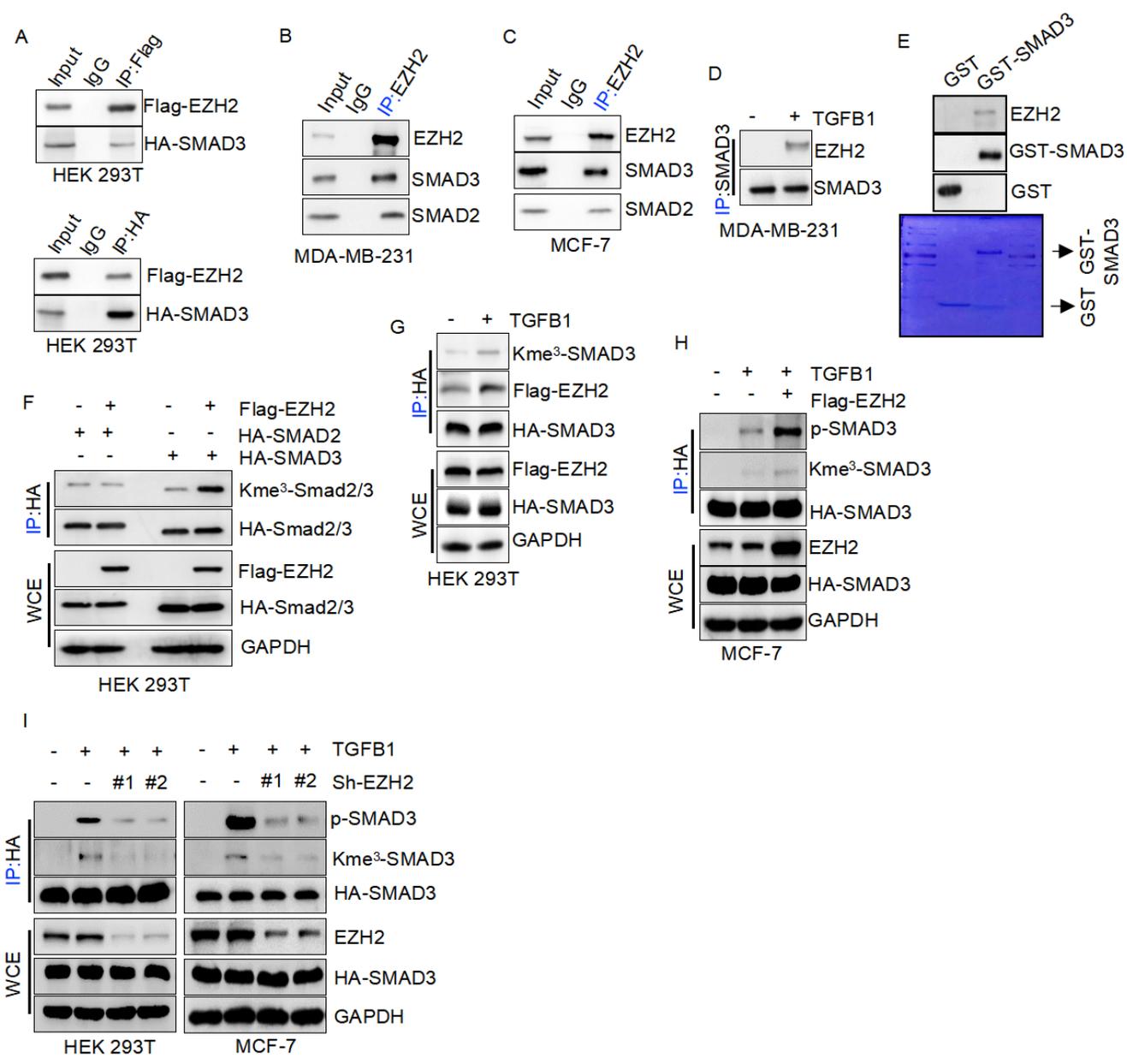
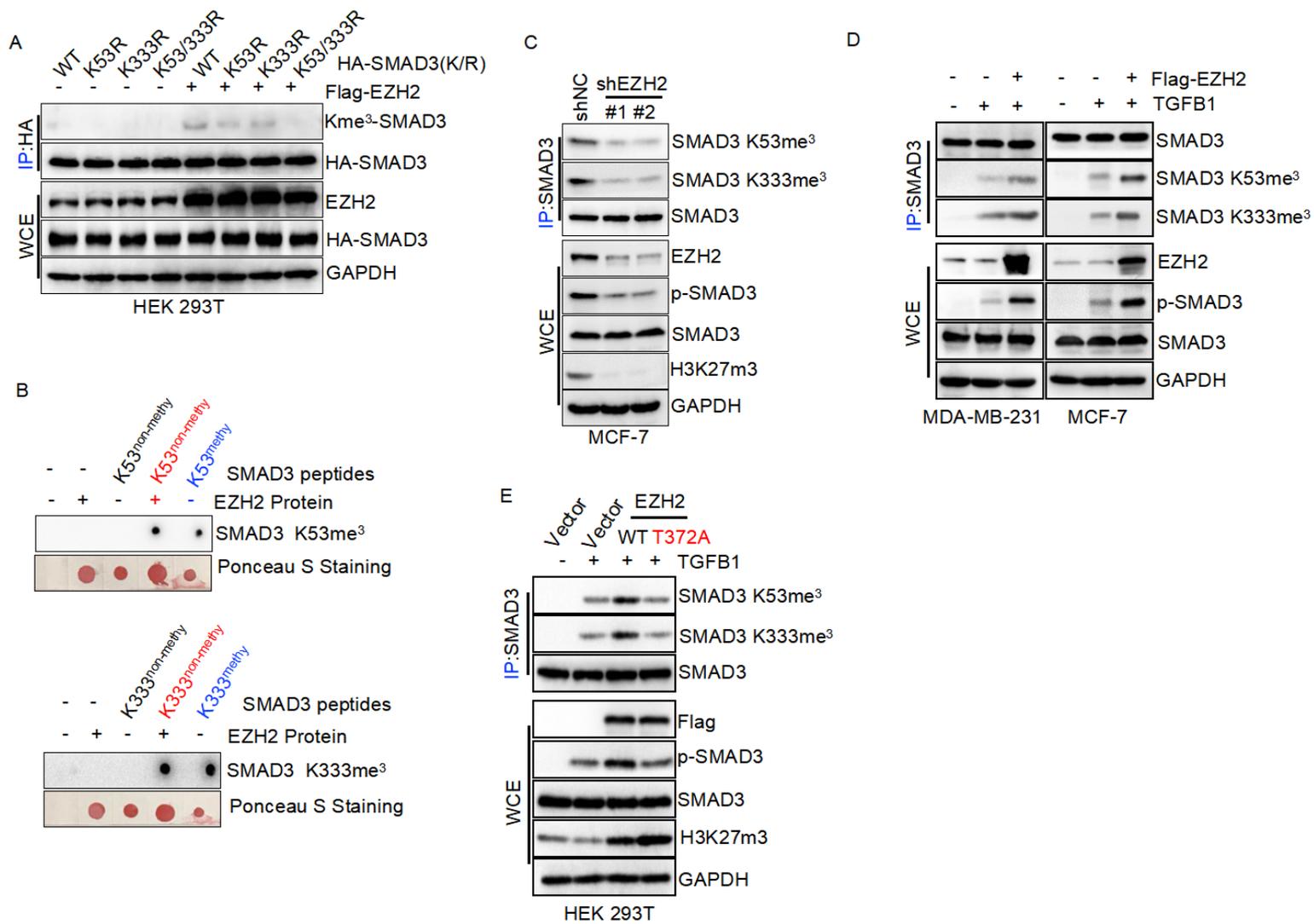


Figure.S4



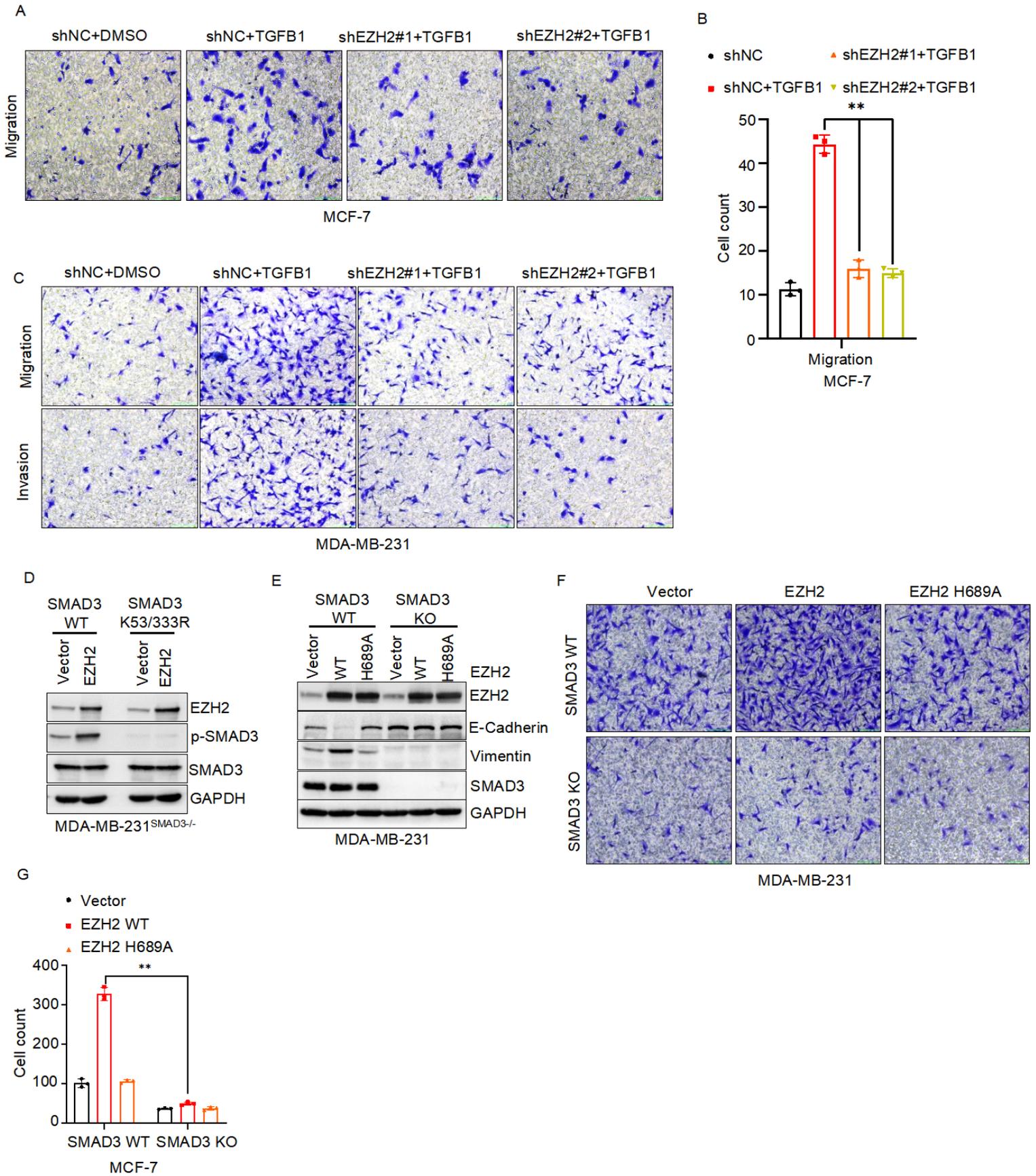


Figure.S6

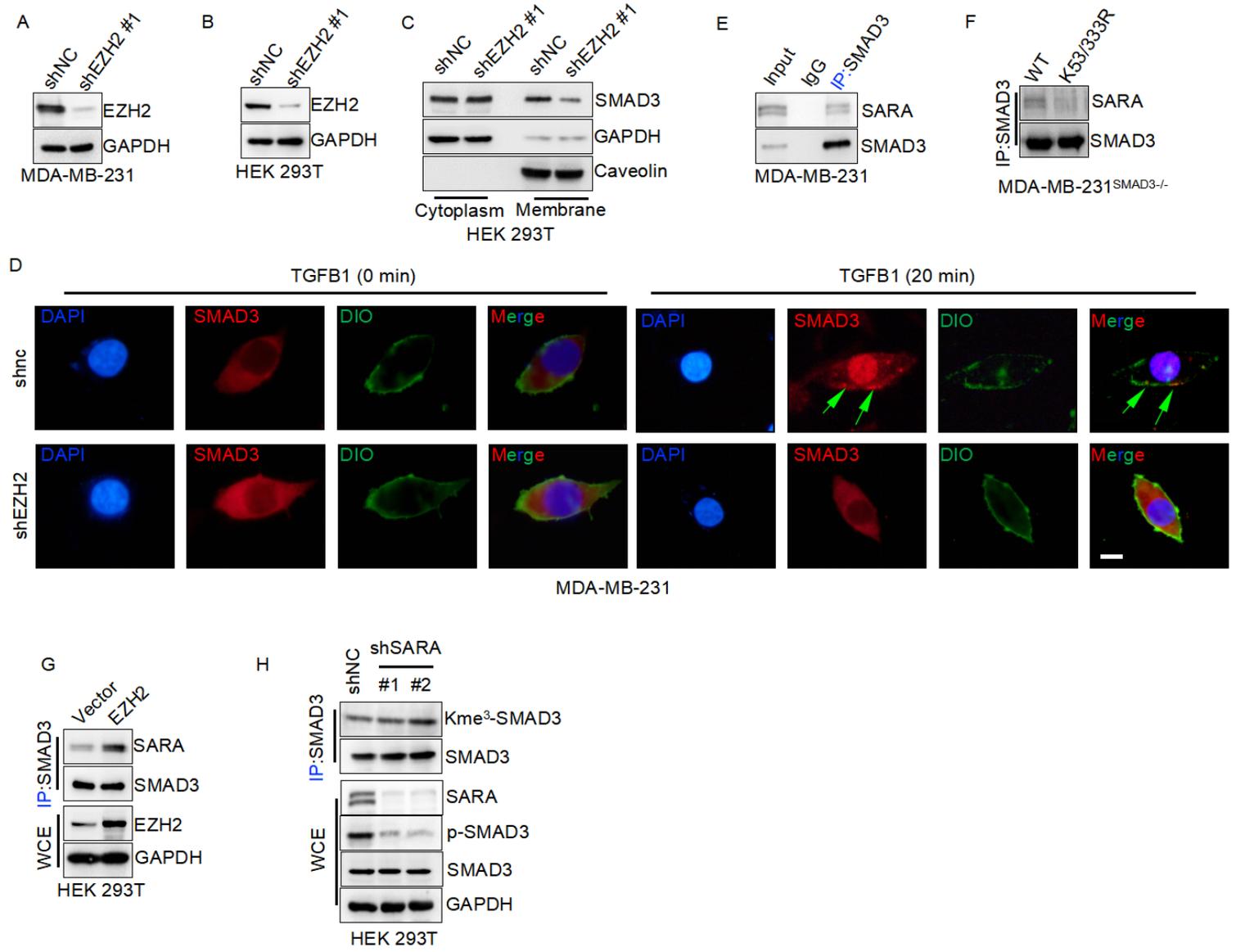
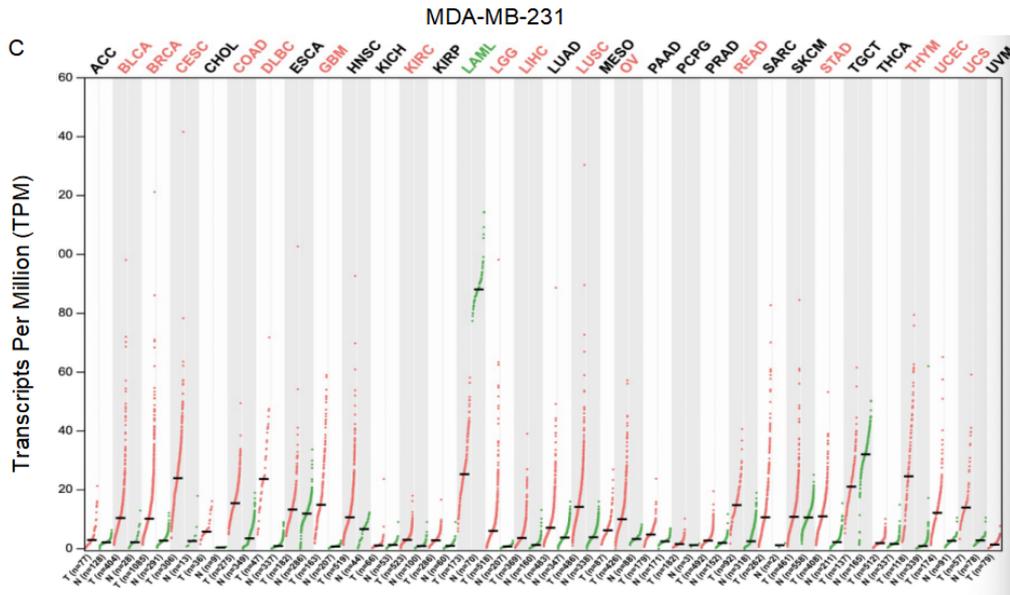
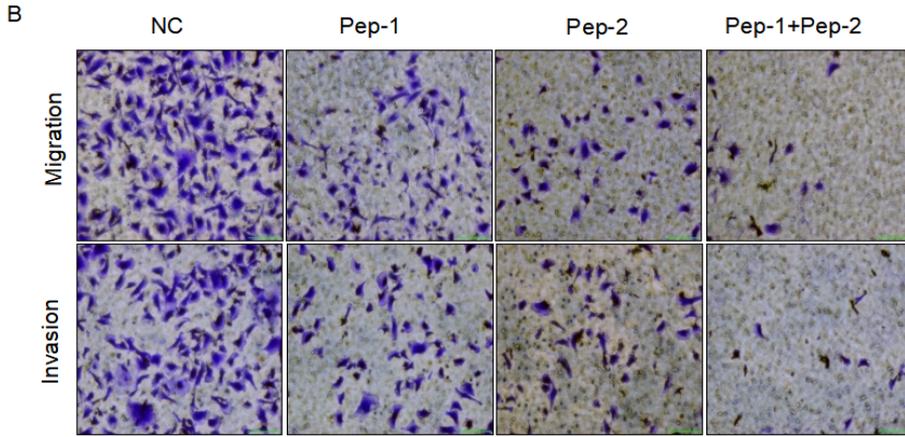
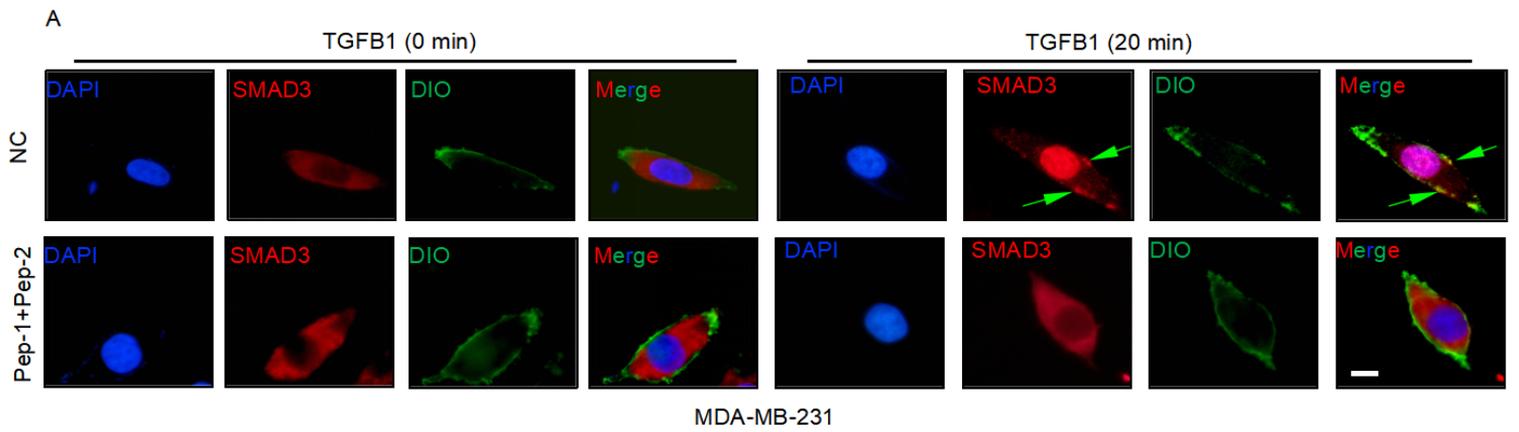


Figure.S7



ACC, Adrenocortical carcinoma; BLCA, Bladder Urothelial Carcinoma; BRCA, Breast invasive carcinoma; CESC, Cervical squamous cell carcinoma and endocervical adenocarcinoma; CHOL, Cholangio carcinoma; COAD, Colon adenocarcinoma; DLBC, Lymphoid Neoplasm Diffuse Large B-cell Lymphoma; ESCA, Esophageal carcinoma; GBM, Glioblastoma multiforme; HNSC, Head and Neck squamous cell carcinoma; KICH, Kidney Chromophobe; KIRC, Kidney renal clear cell carcinoma; KIRP, Kidney renal papillary cell carcinoma; LAML, Acute Myeloid Leukemia; LGG, Brain Lower Grade Glioma; LIHC, Liver hepatocellular carcinoma; LUAD, Lung adenocarcinoma; LUSC, Lung squamous cell carcinoma; MESO, Mesothelioma; OV, Ovarian serous cystadenocarcinoma; PAAD, Pancreatic adenocarcinoma; PCPG, Pheochromocytoma and Paraganglioma; PRAD, Prostate adenocarcinoma; READ, Rectum adenocarcinoma; SARC, Sarcoma; SKCM, Skin Cutaneous Melanoma; STAD, Stomach adenocarcinoma; TGCT, Testicular Germ Cell Tumors; THCA, Thyroid carcinoma; THYM, Thymoma; UCEC, Uterine Corpus Endometrial Carcinoma; UCS, Uterine Carcinosarcoma; UVM, Uveal Melanoma

