

Supplemental Table 1. List of antibodies used in this study.

Antibodies	Source	Clone No.	Catalog No.
Anti-mouse CD3	BioXCell	145-2C11	BE0001-1
Anti-mouse CD28	BioXCell	37.51	BE0015-1
Anti-mouse IL-4	BioXCell	11B11	BE0045
Anti-mouse IFN- γ	BioXCell	XMG1.2	BE0055
Anti-mouse CD4	BioXCell	GK1.5	BE0003-1
Anti-mouse FoxO4	Santa Cruz	N/A	sc-5221
Anti-mouse DKK3	Proteintech	N/A	10365-1-AP
APC Anti-mouse CD4	BD Biosciences	RM4.5	553051
PerCP-Cyanine5.5 Anti-mouse CD4	eBioscience	RM4.5	45-0042-82
PE Anti-mouse CD8	eBioscience	53-6.7	12-0081-82
PE-Cyanine7 Anti-mouse CD25	eBioscience	PC61.5	25-0251-82
eFluor™ 450 Anti-mouse CD44	eBioscience	IM7	48-0441-82
APC-eFluor™ 780 Anti-mouse CD62L	eBioscience	MEL-14	47-0621-82
PE Anti-mouse IFN- γ	Biologend	XMG1.2	505808
Alexa Fluor® 700 Anti-mouse IFN- γ	BD Biosciences	XMG1.2	557998
PE Anti-mouse IL-4	Biologend	11B11	504104
PE Anti-mouse IL-17A	BD Biosciences	TC11-18H10	559502
FITC Anti-mouse Foxp3	eBioscience	FJK-16s	11-5773-82
APC Anti-mouse B220	BD Biosciences	RA3-6B2	553092
Alexa Fluor® 700 Anti-mouse TCR β	BD Biosciences	H57-597	560705
FITC Anti-Mouse PD-1	eBioscience	RMP1-30	11-9981-82
Biotin Anti-Mouse CXCR5	BD Biosciences	2G8	551960

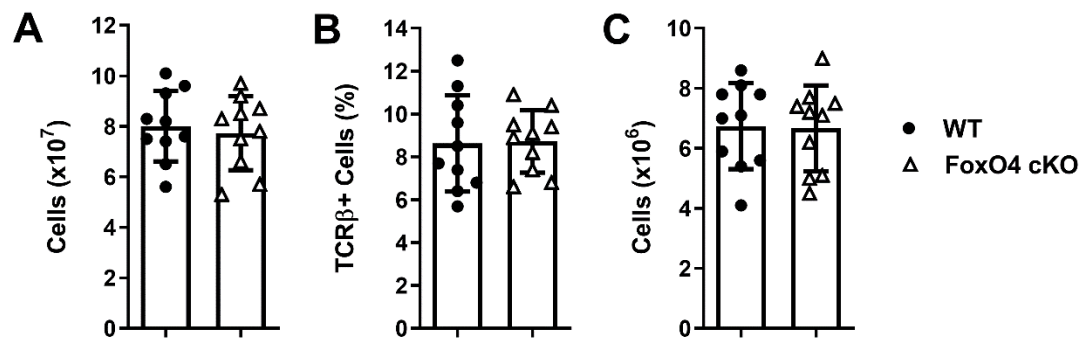
PE Anti-Mouse CD95	BD Biosciences	Jo2	554258
eFluor™ 450 Anti-Mouse GL-7	eBioscience	GL7	48-5902-82

Supplemental Table 2. Primers for Real-Time qPCR and ChIP-qPCR.

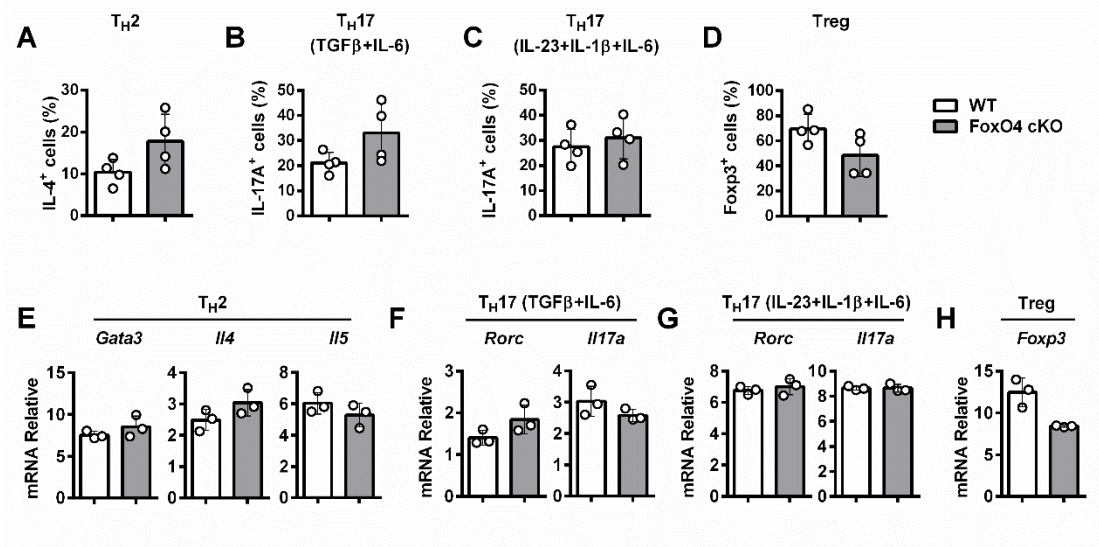
Name	Forward (5'→3')	Reverse (5'→3')
<i>Gapdh</i>	GACAACTTTGGCATTGTGG	ATGCAGGGATGATGTTCTG
<i>Ifng</i>	GATGCATTCATGAGTATTGCCAAGT	GTGGACCACTCGGATGAGCTC
<i>Tbx21</i>	CAACAACCCCTTTGCCAAAG	TCCCCCAAGCAGTTGACAGT
<i>Stat4</i>	TGGCAACAATTCTGCTTCAAAC	GAGGTCCCTGGATAGGCATGT
<i>Runx3</i>	CAGGTTCAACGACCTTCGATT	GTGGTAGGTAGCCACTTGGG
<i>Gata3</i>	AGGGACATCCTGCGCGAACTGT	CATCTTCCGGTTTCGGGTCTGG
<i>Il4</i>	AGATCATCGGCATTTTGAACG	TTTGGCACATCCATCTCCG
<i>Il5</i>	CGCTCACCGAGCTCTGTTG	CCAATGCATAGCTGGTGATTTTT
<i>Rorc</i>	GACCCACACCTCACAAATTGA	AGTAGGCCACATTACACTGCT
<i>Il17a</i>	CTGGAGGATAAACACTGTGAGAGT	TGCTGAATGGCGACGGAGTTC
<i>Foxp3</i>	CCCATCCCCAGGAGTCTTG	ACCATGACTAGGGGCACTGTA
<i>Il3</i>	GGGATACCCACCGTTTAACCA	AGGTTTACTCTCCGAAAGCTCTT
<i>Dkk3</i>	CTCGGGGGTATTTTGCTGTGT	TCCTCCTGAGGGTAGTTGAGA
<i>Csf1</i>	ATGAGCAGGAGTATTGCCAAGG	TCCATTCCCAATCATGTGGCTA
<i>Csf2</i>	GGCCTTGGAAGCATGTAGAGG	GGAGAACTCGTTAGAGACGACTT
<i>Csf3</i>	ATGGCTCAACTTTCTGCCAG	CTGACAGTGACCAGGGGAAC
<i>Bcl6</i>	CCGGCACGCTAGTGATGTT	TGTCTTATGGGCTCTAAACTGCT
<i>Ccr6</i>	CCTGGGCAACATTATGGTGGT	CAGAACGGTAGGGTGAGGACA
<i>Il23r</i>	TTCAGATGGGCATGAATGTTTCT	CCAAATCCGAGCTGTTGTTCTAT
<i>Tcf4</i>	CGAAAAGTTCCTCCGGGTTTG	CGTAGCCGGGCTGATTCAT
<i>Tcf7</i>	AGCTTTCTCCACTCTACGAACA	AATCCAGAGAGATCGGGGGTC
<i>Lef1</i>	TGTTTATCCCATCACGGGTGG	CATGGAAGTGTCGCCTGACAG

<i>Myc</i>	ATGCCCCTCAACGTGAACTTC	CGCAACATAGGATGGAGAGCA
<i>H19</i>	GCATGGTCCTCAAATTCTGCA	GCATCTGAACGCCCAATTA
<i>IfngCNS1</i>	CACTTCTGTGCAACCCTTGA	AAGCACTCACTGGGTCATTG
<i>IfngCNS2</i>	AACTGGAAAATGGCAGGCTA	CCCGAGATAAATTCCATCCA
<i>IfngCNS22</i>	ATGACAAAATGCAGGGCTTC	CCCACACTAGATGATATATGATTTTCC
<i>IfngCNS34</i>	AAAAGAGTCCAAGATATGAAAGCAA	GGCTTTGGGAATTCTACCTTG
<i>IfngCNS55</i>	TGTCTCGGTGACACATCCTT	GGGAGGCAGGAGGAACTTTA
<i>IfngPromoter</i>	CCCCACCTATCTGTCACCAT	CACCTCTCTGGCTTCCAGTT
<i>EomesFBS1</i>	CGGGGTTTGTTCCTTGCG	GATTGTAGGTGCCCTTTCCT
<i>EomesFBS2</i>	AAGATCAAGGTCTGGGAACCCG	GTGGGGAGTGTAACAAGCCG
<i>EomesFBS3</i>	CACCGATAACAAGCCTCCATT	GGGTTCCAGACCTTGATCTTTAT

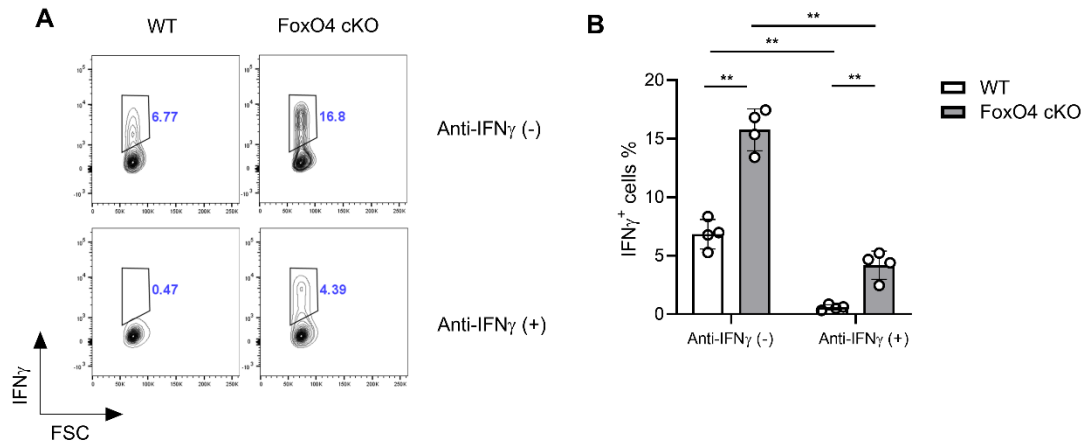
Supplemental Figures



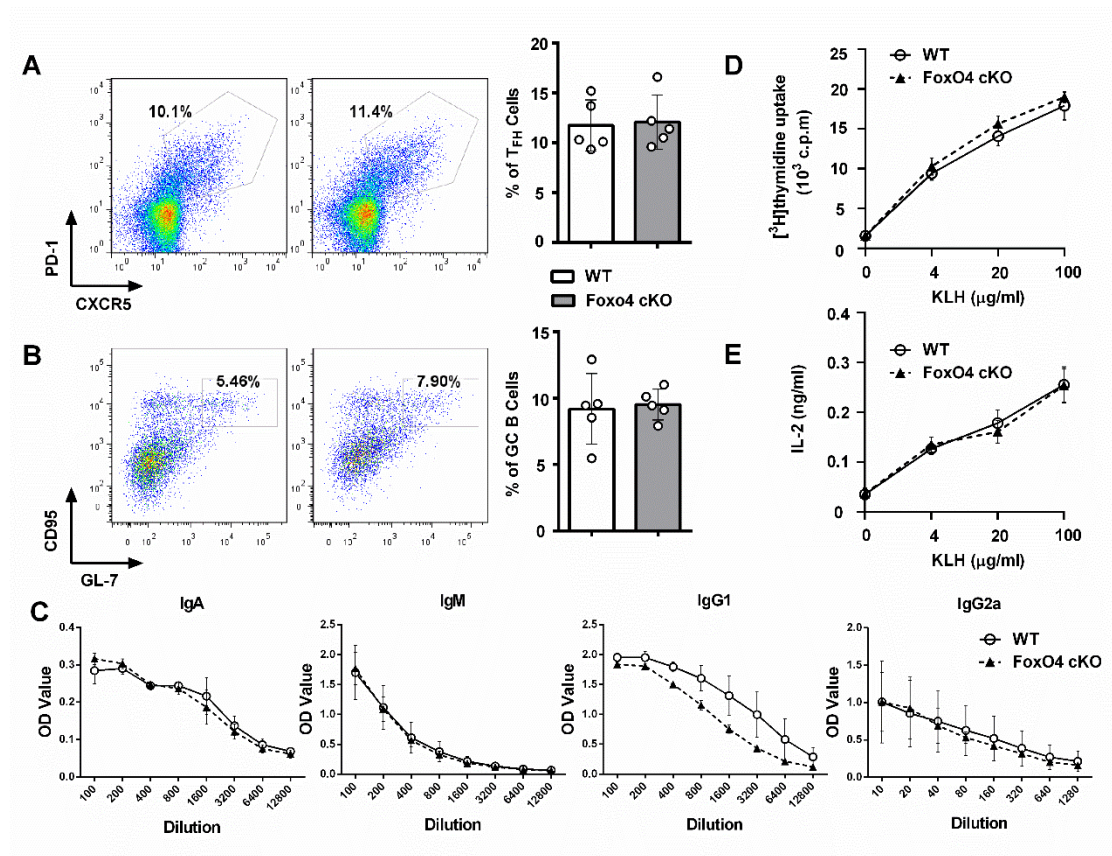
Supplemental Figure 1. Numbers of mature T lymphocytes in thymus are normal in *FoxO4* cKO mice. (A) Absolute thymocyte numbers in WT and *FoxO4* cKO mice ($n=10$). (B and C) Percentages (B) and absolute cell numbers (C) of TCR β^+ T lymphocytes in thymus in WT and *FoxO4* cKO mice ($n=10$). Each symbol (A-C) represents an individual mouse. ns=not statistically significant, unpaired Student's *t*-test. Data are one representative of three independent experiments with similar results (means and SD in A-C).



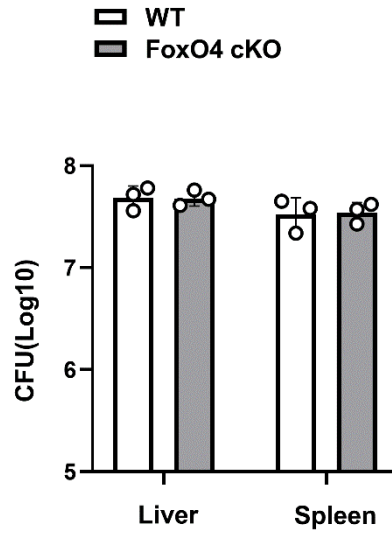
Supplemental Figure 2. FoxO4 does not affect differentiation of T_H2 , T_H17 , and iTreg subsets. (A-D) Percentage of IL-4 (A), IL-17A (B and C), Foxp3 (D) expressing WT and *FoxO4* cKO CD4⁺ T cells, which were differentiated from naïve T cells in various polarizing conditions for 3 days (n=4). (E-H) Real-time qPCR analysis of signature genes of T_H2 (E), T_H17 (induced by TGFβ/IL-6, F), T_H17 (induced by IL-23/IL-1β/IL-6, G) and iTreg (H) from WT and *FoxO4* cKO mice, by stimulating naïve CD4⁺ T cells for 3 days with plate-bound anti-CD3 and anti-CD28 in various polarizing conditions, and assessing after re-stimulation by plate-bound anti-CD3 for 5 hours (n=3). Results were presented relative to the expression *Gapdh* mRNA. ns=not statistically significant, unpaired Student's *t*-test. Data are one representative of two independent experiments with similar results (means and SD in A-H).



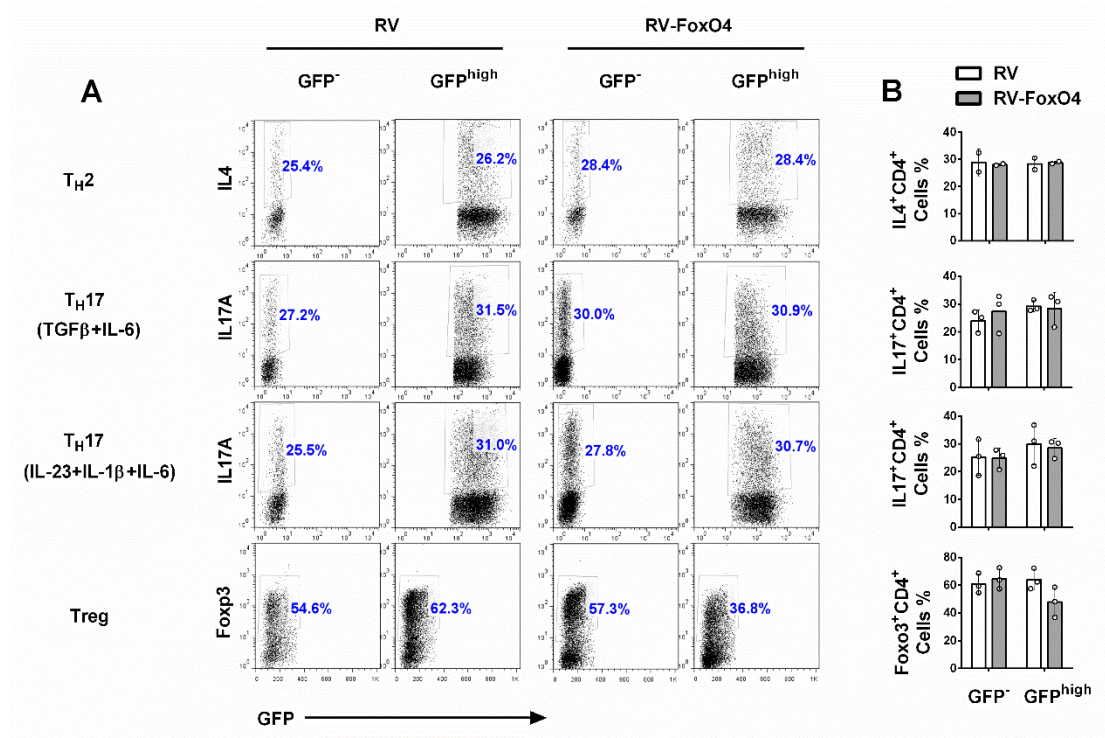
Supplemental Figure 3. FoxO4 negatively regulates IFN- γ expression in T_H1 polarizing condition with or without anti-IFN- γ . (A) Flow cytometry of WT and FoxO4 cKO Naïve CD4⁺ T cells after 2 days of polarization toward the T_H1 lineage, with or without anti-IFN- γ (n=4). (B) Frequencies of IFN- γ -expressing cells as in (A) (n=4). **P<0.01; ***P<0.001, by two-way ANOVA with Tukey's multiple-comparison test (B). Data are one representative of two independent experiments (means and SD in B).



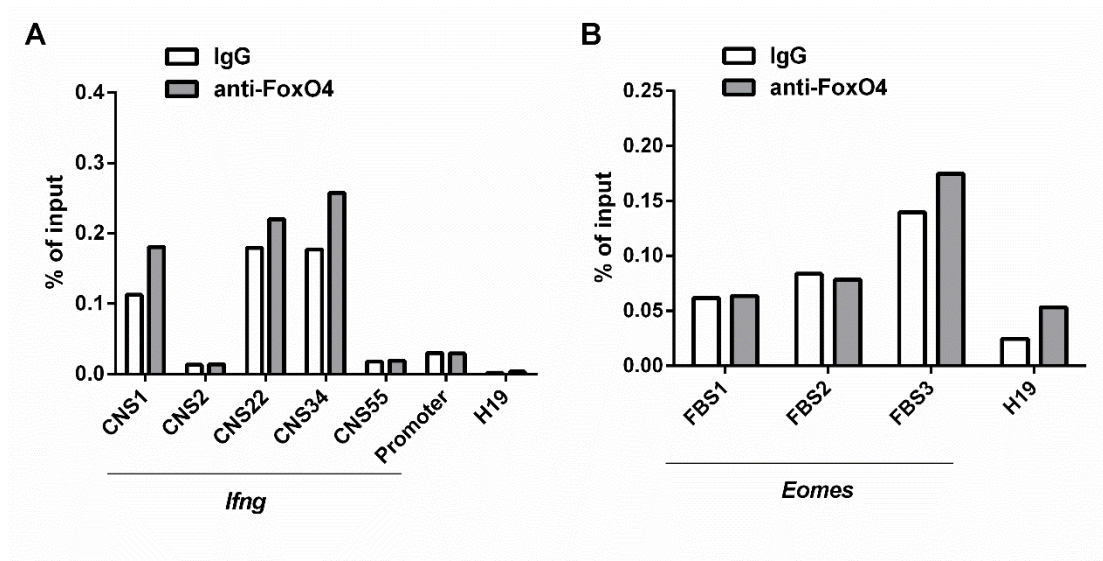
Supplemental Figure 4. *FoxO4* cKO mice have normal germinal center responses and antigen specific Ig production. (A and B) Flow cytometry analysis (left) and quantification (right) of T_{FH} cells (left in A, PD-1⁺CXCR5⁺, gating on CD4⁺CD44^{hi}) and germinal center B cells (left in B, GL-7⁺CD95⁺, gating on B220⁺) in dLNs from WT and *FoxO4* cKO mice which received subcutaneous KLH/CFA immunization for 7 days (n=5). (C) ELISA analysis was performed to determine the concentrations of KLH specific IgA, IgM, IgG1 and IgG2a in serum from WT and *FoxO4* cKO mice which were immunized as in A and B. Cells from draining lymph nodes were isolated 7 days later and restimulated with KLH (n=5). Proliferation was measured by [³H] thymidine incorporation assay (D) and IL-2 expression was determined by ELISA (E) (n=5). ns=not statistically significant, unpaired Student's *t*-test. Data are one representative of three independent experiments with similar results (means and SD in A-E).



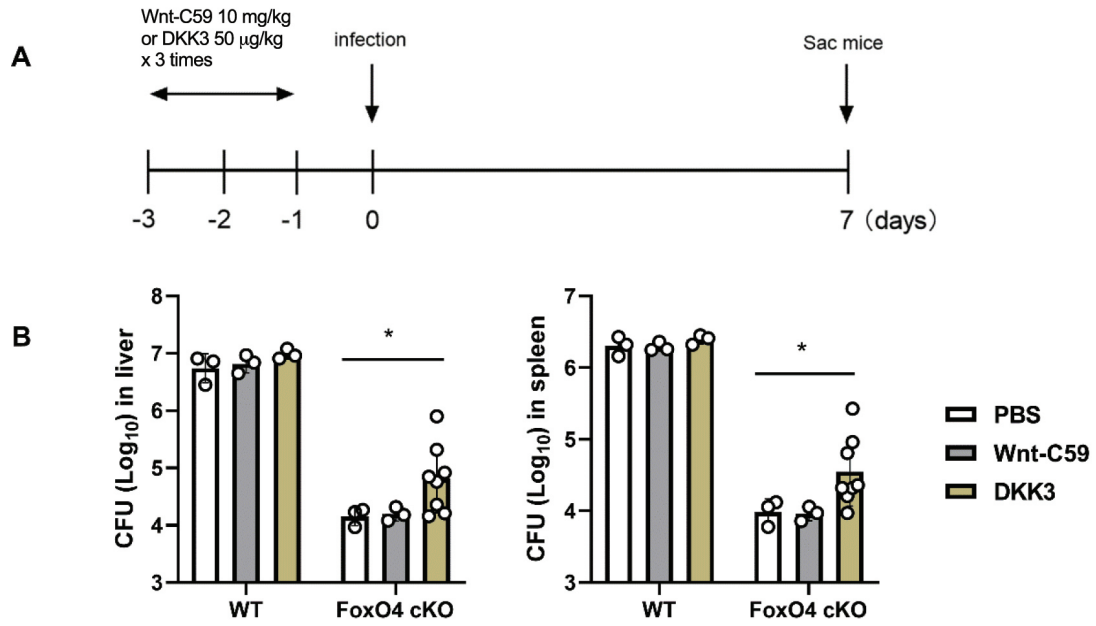
Supplemental Figure 5. FoxO4 deficiency in CD4⁺ T cells provides protection from listeria infection. Infected mice were treated with anti-CD4 antibody (GK1.5, BioXcell) intraperitoneally at day -1 and +1 respective to challenge listeria infection. Livers and spleens were harvested at day 4 post infection for enumeration of bacterial burdens (n=3). ns=not statistically significant, unpaired Student's *t*-test. Data are one representative of two independent experiments with similar results (means and SD).



Supplemental Figure 6. Ectopic expression of FoxO4 doesn't affect differentiation of T_H2, T_H17, and iTreg subsets. (A) Flow cytometry of various polarized CD4⁺ T cells, which were transduced with empty vector (RV) or vector encoding FoxO4 (RV-FoxO4). Data were analyzed in GFP⁻ and GFP^{high} populations, respectively (n=3). (B) Frequency of specific marker-expressing cells as in A (n=3). ns=not statistically significant, unpaired Student's *t*-test. Data are one representative of three independent experiments with similar results (means and SD in B).



Supplemental Figure 7. FoxO4 doesn't bind to *Ifng* and *Eomes* gene loci. (A and B) Chromatin immunoprecipitation analysis of FoxO4 binding to *Ifng* (A) and *Eomes* (B) gene loci in purified CD4⁺ T cells stimulated for 24 hours with plate-bound anti-CD3. Results were expressed as percentage of input. Data are one representative of two independent experiments with similar results.



Supplemental Figure 8. FoxO4-DKK3 axis functions in anti-listeria immunity. (A) Schematic of animal experimental setup. (B) *L. monocytogenes* titers in the spleens and livers of WT and *FoxO4* cKO mice infected with Lm-OVA for 7 days, with treatment of PBS, Wnt-C59 and DKK3, respectively, showed as CFUs (n=3-8). *P<0.05, by two-way ANOVA with Tukey's multiple-comparison test. Data are one representative of two independent experiments with similar results (means and SD in B).

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eFluor™ 450 Anti-mouse CD44	eBioscience	IM7	48-0441-82
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<i>Stat4</i>	TGGCAACAATTCTGCTTCAAAC	GAGGTCCCTGGATAGGCATGT
<i>Runx3</i>	CAGGTTCAACGACCTTCGATT	GTGGTAGGTAGCCACTTGGG
<i>Gata3</i>	AGGGACATCCTGCGCGAACTGT	CATCTTCCGTTTTCGGGTCTGG
<i>Il4</i>	AGATCATCGGCATTTTGAACG	TTTGGCACATCCATCTCCG
<i>Il5</i>	CGCTCACCGAGCTCTGTTG	CCAATGCATAGCTGGTGATTTTT
<i>Rorc</i>	GACCCACACCTCACAAATTGA	AGTAGGCCACATTACACTGCT
<i>Il17a</i>	CTGGAGGATAAACTGTGAGAGT	TGCTGAATGGCGACGGAGTTC
<i>Foxp3</i>	CCCATCCCCAGGAGTCTTG	ACCATGACTAGGGGCACTGTA
<i>Il3</i>	GGGATACCCACCGTTTAACCA	AGGTTTACTCTCCGAAAGCTCTT
<i>Dkk3</i>	CTCGGGGGTATTTTGCTGTGT	TCCTCCTGAGGGTAGTTGAGA
<i>Csf1</i>	ATGAGCAGGAGTATTGCCAAGG	TCCATTCCCAATCATGTGGCTA
<i>Csf2</i>	GGCCTTGGAAGCATGTAGAGG	GGAGAACTCGTTAGAGACGACTT
<i>Csf3</i>	ATGGCTCAACTTTCTGCCAG	CTGACAGTGACCAGGGGAAC
<i>Bcl6</i>	CCGGCACGCTAGTGATGTT	TGTCTTATGGGCTCTAAACTGCT

<i>Ccr6</i>	CCTGGGCAACATTATGGTGGT	CAGAACGGTAGGGTGAGGACA
<i>Ii23r</i>	TTCAGATGGGCATGAATGTTTCT	CCAAATCCGAGCTGTTGTTCTAT
<i>Tcf4</i>	CGAAAAGTTCCTCCGGGTTTG	CGTAGCCGGGCTGATTCAT
<i>Tcf7</i>	AGCTTTCTCCACTCTACGAACA	AATCCAGAGAGATCGGGGGTC
<i>Lef1</i>	TGTTTATCCCATCACGGGTGG	CATGGAAGTGTCGCCTGACAG
<i>Myc</i>	ATGCCCTCAACGTGAACTTC	CGCAACATAGGATGGAGAGCA
<i>H19</i>	GCATGGTCCTCAAATTCTGCA	GCATCTGAACGCCCAATTA
<i>IfngCNS1</i>	CACTTCTGTGCAACCCTTGA	AAGCACTCACTGGGTCATTG
<i>IfngCNS2</i>	AACTGGAAAATGGCAGGCTA	CCCGAGATAAATTCCATCCA
<i>IfngCNS22</i>	ATGACAAAATGCAGGGCTTC	CCCACACTAGATGATATATGATTTTCC
<i>IfngCNS34</i>	AAAAGAGTCCAAGATATGAAAGCAA	GGCTTTGGGAATTCTACCTTG
<i>IfngCNS55</i>	TGTCTCGGTGACACATCCTT	GGGAGGCAGGAGGAACTTTA
<i>IfngPromoter</i>	CCCCACCTATCTGTCACCAT	CACCTCTCTGGCTTCCAGTT
<i>EomesFBS1</i>	CGGGGTTTGTTCCTTGCG	GATTGTAGGTGCCCTTTCT
<i>EomesFBS2</i>	AAGATCAAGGTCTGGGAACCCG	GTGGGGAGTGTAACAAGCCG
<i>EomesFBS3</i>	CACCGATAAACAAGCCTCCATT	GGGTTCCAGACCTTGATCTTTAT