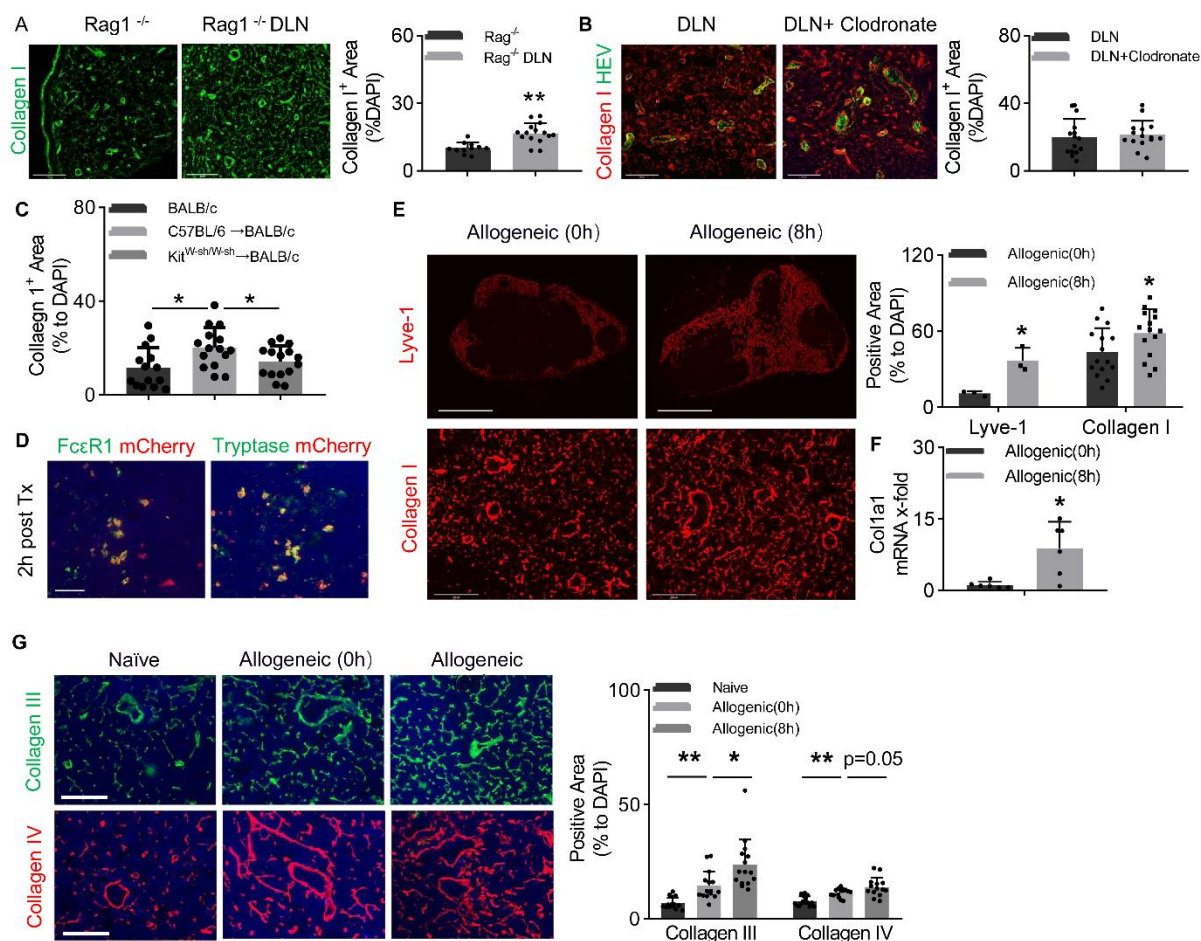


Supplementary Table 1: Sequences of primers used for Real-time PCR.

<i>Mcpt2</i>	Forward 5'-ATTTCATTGCCTAGTTCCTCTGAC-3'
	Reverse 5'-CAGGATGAGAACAGGCTGGGAT-3'
<i>Mcpt4</i>	Forward 5'-GTAATTCCTCTGCCTCGTCCTTC-3'
	Reverse 5'-GTAATTCCTCTGCCTCGTCCTTC-3'
<i>Mcpt6</i>	Forward 5'-AGTAAGTGGCCCTGGCAGGTGAGCC-3'
	Reverse 5'-GGTCCCCATAGTATAGATACTGCTC-3'
<i>Vegfa</i>	Forward 5'-CACAGCAGATGTGAATGCAG-3'
	Reverse 5'-TTTACACGTCTGCGGATCTT-3'
<i>Fgf2</i>	Forward 5'-GAAACACTCTTCTGTAACACACTT-3'
	Reverse 5'-GTCAAACACTACAACCTCCAAGCAG-3'
<i>Il6</i>	Forward 5'-CTCTGGGAAATCGTGGAAAT-3'
	Reverse 5'-CCAGTTTGGTAGCATCCATC-3'
<i>Colla1</i>	Forward 5'-CCTGGTAAAGATGGTGCC-3'
	Reverse 5'-CACCAGGTTACCTTTTCGCACC-3'
<i>Tgfb1</i>	Forward 5'-CAACAATTCCTGGCGTTACCTTGG-3'
	Reverse 5'-GAAAGCCCTGTATTCCGTCTCCTT-3'
<i>Smad2</i>	Forward 5'-ATGTCGTCCATCTTGCCATTC-3'
	Reverse 5'-AACCGTCCTGTTTTCTTTAGCTT-3'
<i>Acta2</i>	Forward 5'-CTGACAGAGGCACCACTGAA-3'
	Reverse 5'-CATCTCCAGAGTCCAGCACA-3'
<i>Fn1</i>	Forward 5'-CGAGGTGACAGAGACCACAA-3'
	Reverse 5'-CTGGAGTCAAGCCAGACACA-3';
<i>Smad2</i>	Forward 5'-ATGTCGTCCATCTTGCCATTC-3'
	Reverse 5'-AACCGTCCTGTTTTCTTTAGCTT-3'
<i>Smad7</i>	Forward 5'-GGGCTTTCAGATTCCTCAACTT-3'
	Reverse 5'-CACGCGAGTCTTCTCCTCC-3'
<i>Bmp7</i>	Forward 5'-CAAGCAGCGCAGCCAGAATCG-3'
	Reverse 5'-CAATGATCCAGTCCTGCCAGCCAA-3';
<i>Cdkn2a</i>	Forward 5'-CCCCAGTGTCTTACAGAGTG-3'
	Reverse 5'-GTGCCAGAGTGGATGTCT-3'

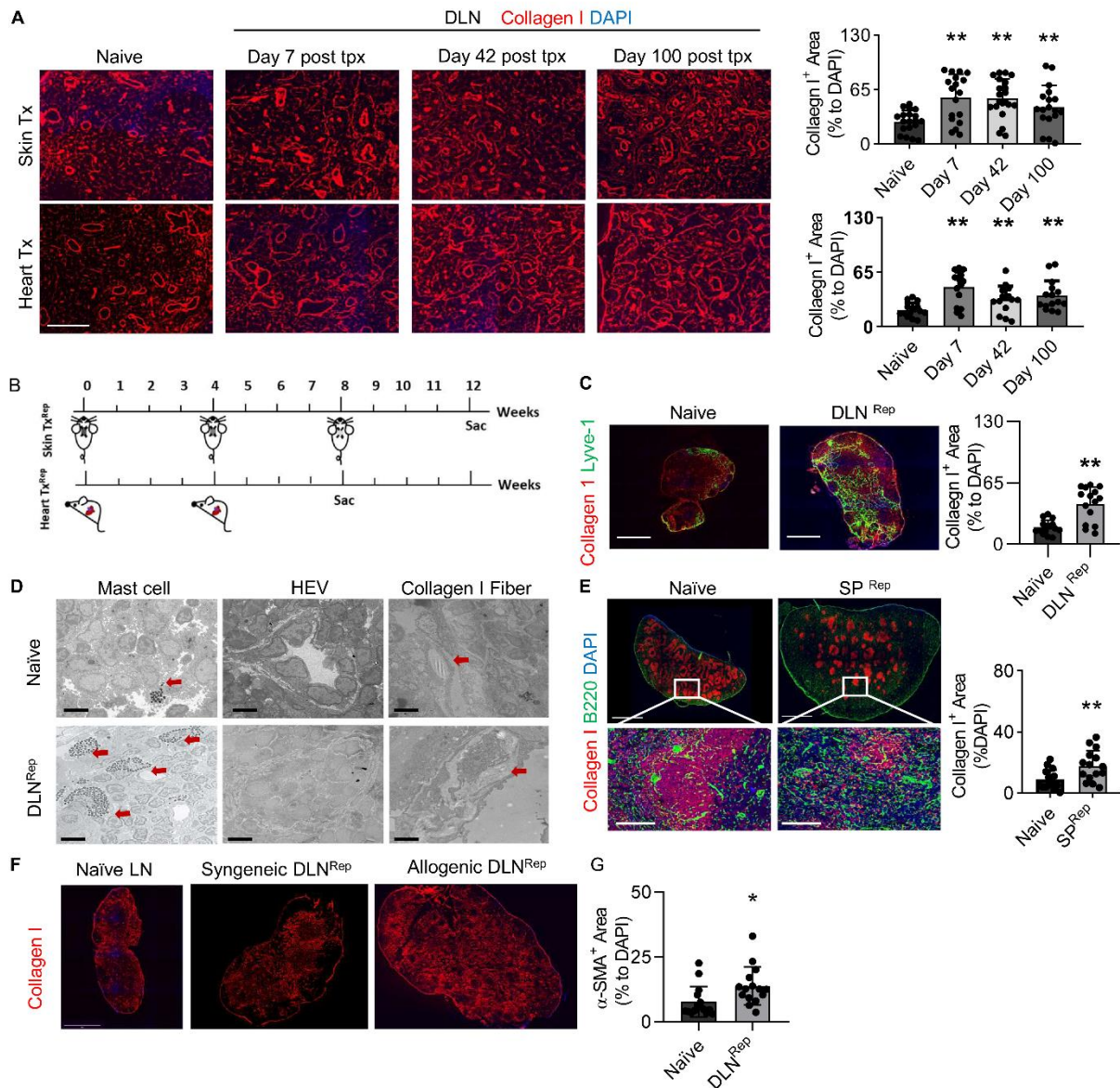
<i>Cdkn1a</i>	Forward 5'-TGGT- GTCATCCCTACCTTCA-3'
	Reverse 5'-TTCTCTCTATCCTC TCCCCAG-3'
<i>Trp53</i>	Forward 5'-TGTGTTACACACTAAGGGG-3'
	Reverse 5'-CCTTTGTTCTTGGCAGAAGACT-3'
<i>Cdkn1c</i>	Forward 5'-CTCAAGCTTCAAGATGTGGACCGTGCCAGT-3'
	Reverse 5'-GAGGAATTCGGGCGAGAACCTTCCAGAA-3'
<i>Gadph</i>	Forward 5'-AGCCACATCGCTCAGACAC-3'
	Reverse 5'-AGGCAGGTTTGATCTCCGTT-3'

Supplementary Figure and Figure legends



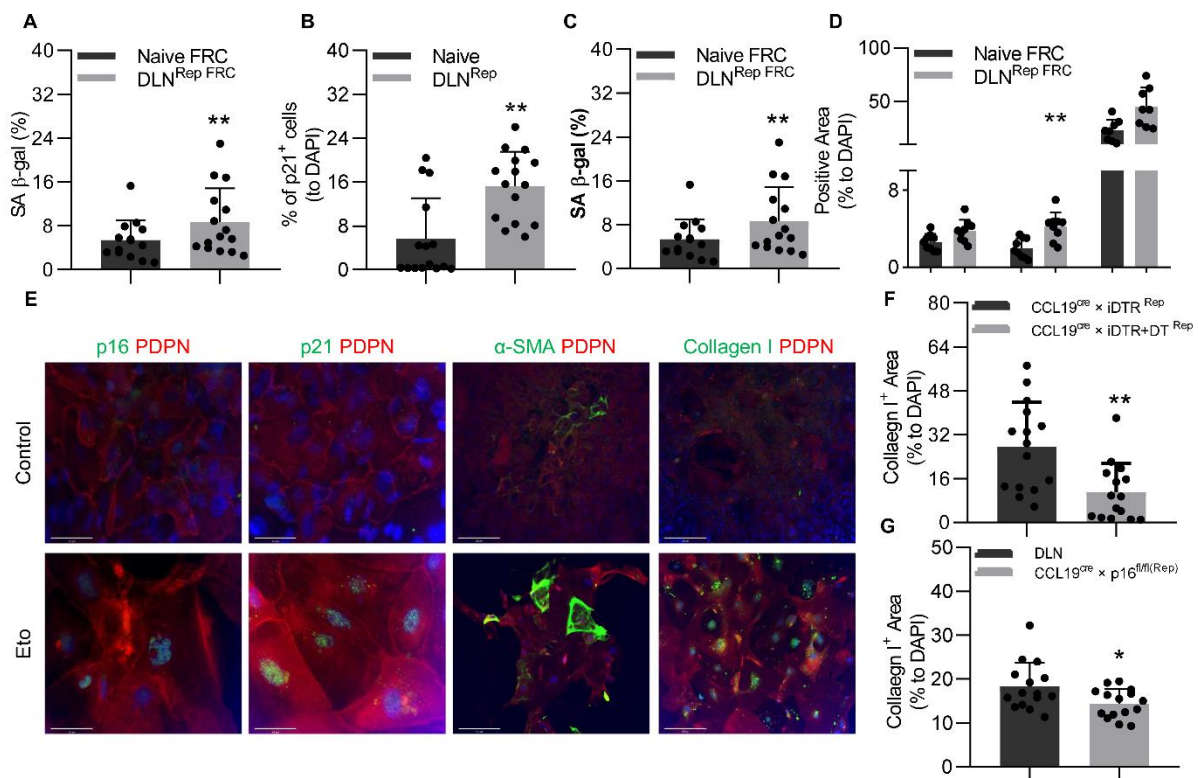
Supplementary Figure 1: Ischemia activates mast cell-induced collagen 1 accumulation and lymphangiogenesis in DLNs following skin transplantation. (A) Comparison and semi-

quantitative analysis of collagen I staining in DLNs from BALB/c → Rag1^{-/-} skin transplant recipient mice in comparison to the axillary lymph nodes of Rag1^{-/-} mice at day 1 after skin transplantation. Scale bar 100μm. (B) Comparison and analysis of collagen I staining and between DLNs of BALB/c→C57BL/6J skin transplant recipient mice with and without clodronate treatment. Scale bar 100μm. (C) Assessment of collagen 1⁺ region in the DLNs of naïve BALB/c mice, and those that received skin allografts from C57BL/6J and Kit^{W-sh/W-sh} mice (n=4). (D) Fluorescence micrographs of DLNs of BALB/c recipients of skin allografts from C57BL/6-Tg(UBC-mCherry) donor mice demonstrate presence of mast cells within 2 hours following transplantation, as demonstrated by co-staining of the mast cell markers FcεR1 (green, left) and tryptase (green, right) with mCherry (red). (E) IF staining of Lyve-1⁺ lymphatic vessels and collagen 1⁺ region and semi-quantitative assessment in the DLNs following transplantation of WT and ischemic organs (~8 hours cold ischemia time, scale bars 1000μm and 100μm, n=4). (F) Gene expression level of *Colla1* in the DLNs following transplantation of WT and ischemic organs (n=6). (G) Fluorescence micrographs and analysis of collagen III⁺ and collagen IV⁺ region in naïve LNs and DLNs following transplantation of WT and ischemic organs. Scale bars 50μm (n=3). Percentage of area stained positive in fluorescence micrographs was assessed in 3-6 random microscopic fields for each mouse. **p* < 0.05; ***p* < 0.01 by Student's t test and 2-way ANOVA with Tukey's multiple-comparisons test.

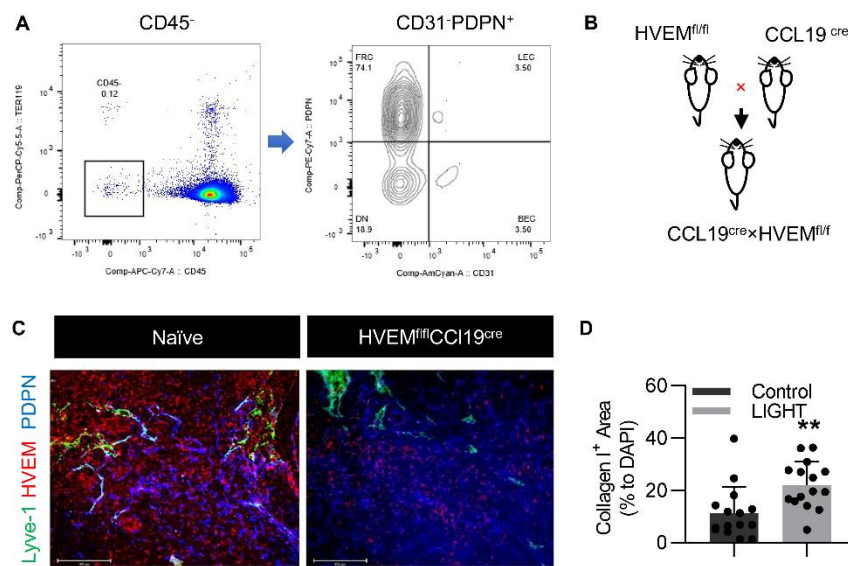


Supplementary Figure 2: Persistent accumulation of collagen 1 in DLNs following transplantation. (A) Fluorescence microscopic analysis shows marked increase of collagen 1 fibers (red) at 7, 42, and 100 days following skin and heart transplantation. Scale bar 100 μ m (n=3). (B) Schematic for the timelines of repetitive skin transplantation and heart transplantation models. (C) Fluorescence micrograph of collagen I⁺ (red) and Lyve-1⁺ (green) staining and semi-quantitative analysis of collagen 1⁺ region in DLN^{Rep} following repetitive BALB/c \rightarrow C57/BJ6 heart transplantation. Scale bar 1000 μ m (n=3). (D) Electron micrograph of one to two mast cells

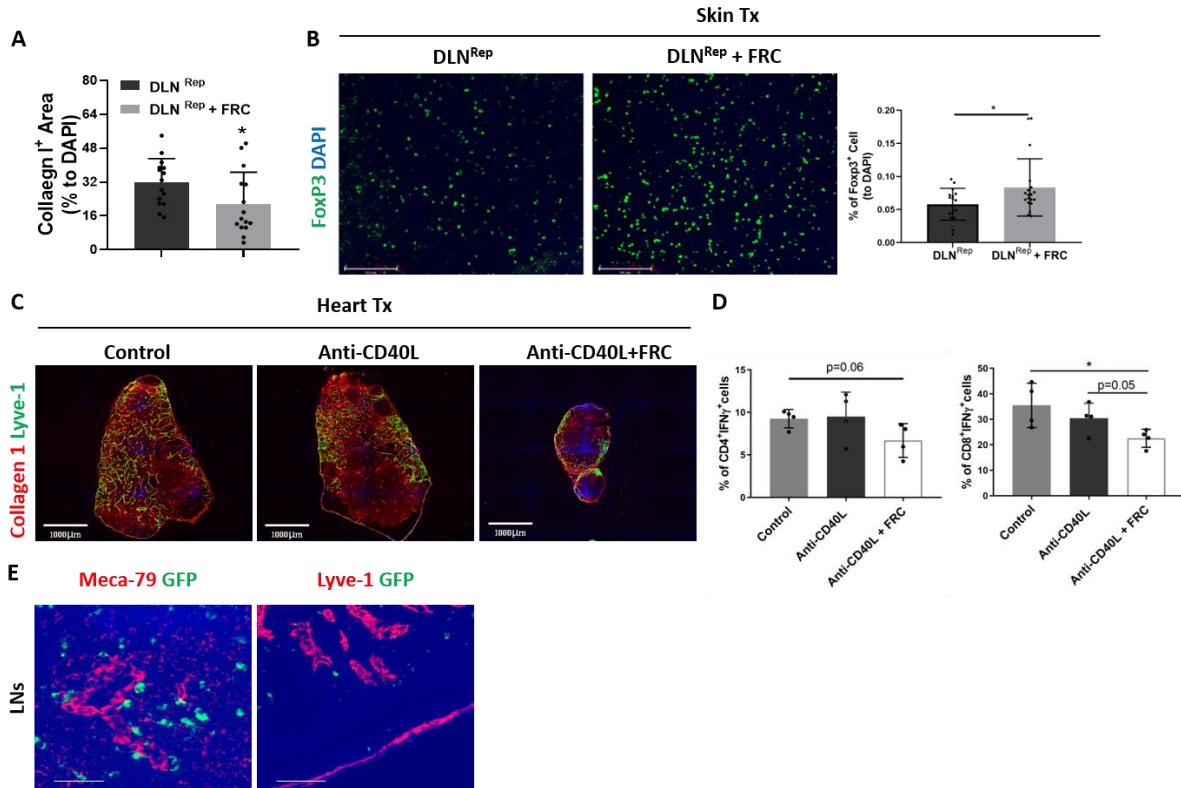
located near lymphatics, HEVs, and collagen 1 fibers (red arrow) in naïve LN and higher number in DLN^{Rep}, along with obliteration of HEV lumen, detachment of HEV, and higher collagen 1 fiber density. Scale bar 2 μ m in mast cell and HEV images, 8 μ m in collagen 1 fiber images. (E) Fluorescence micrographs of collagen I⁺ fibers (red) and B220⁺ B cells (green) in the spleens of mice that underwent repetitive transplantation. Scale bars 1000 μ m and 50 μ m (n=3). (F) Gene expression levels of collagen 1⁺ (red) in LNs of naïve mice and DLN^{Rep} of recipients of syngeneic and allogeneic skin transplantation. Scale bar 1500 μ m. (G) Assessment of α -SMA⁺ staining in naïve LNs and DLNs following repetitive skin transplantation. Scale bar 1000 μ m (n=4). Data in graphs are represented as means \pm SD. Percentage of area stained positive in fluorescence micrographs was assessed in 3-6 random microscopic fields for each mouse. * $p < 0.05$; ** $p < 0.01$ by Student's t test and 2-way ANOVA with Tukey's multiple-comparisons test.



Supplementary Figure 3: Senescent FRCs promote fibrosis. Analysis of the percentage of (A) β -gal⁺ cells and (B) p21⁺ cells in naïve LNs and DLN^{Rep} (n=4). Analysis of (C) surface area percentage occupied by β -gal⁺ cells, and (D) α -SMA⁺, collagen 1⁺, and p21⁺ cells among FRCs isolated from naïve LNs and DLN^{REP} (n=3). (E) IF staining shows that the expression levels of p16, p21, collagen 1, and α -SMA by cultured FRCs increase after etoposide treatment. Evaluation of collagen 1⁺ region in DLNs of (F) CCL19^{cre}×iDTR and (G) CCL19^{cre}×p16^{fl/fl} allogeneic skin transplant recipients (n=4). Data in graphs are represented as means \pm SD. Percentage of area stained positive in fluorescence micrographs was assessed in 3-6 random microscopic fields for each mouse. **p* < 0.05; ***p* < 0.01 by Student's t test.



Supplementary Figure 4: HVEM expression in FRCs of C57BL/6J and CCL19^{cre}×HVEM^{fl/fl} mice. (A) Gating strategy for flow cytometric analysis of FRCs. (B) Schematic for generation of CCL19^{cre} × HVEM^{fl/fl} mice. (C) IF staining shows HVEM (red) expression by PDPN (blue)⁺ LYVE-1(green)⁻ FRCs. (D) Semi-quantitative analysis of collagen I⁺ fibers in naïve (control) and LIGHT-treated FRCs (n=4). Data in graphs are represented as means \pm SD. **p* < 0.05; ***p* < 0.01 by Student's t test.



Supplementary Figure 5: FRC treatment improves immunosuppression post-transplantation. (A) Assessment of collagen I⁺ staining in the DLNs of untreated mice (DLN^{Rep}) and mice treated with FRCs (DLN^{Rep} + FRC) (n=4). (B) IF staining of Foxp3⁺ Tregs in DLN^{Rep} from mice treated with and without FRCs. (C) Co-staining of LYVE-1 (green) and collagen 1 (red) in DLNs at 7 days after heart transplantation. Scale bar 1000 μ m. (D) Percentages of CD4⁺IFN- γ ⁺ cells and CD8⁺IFN γ ⁺ cells in splenocytes as measured by flow cytometry (each dot represents one biological replicate). (E) Fluorescence micrographs of DLNs of C57BL/6J mice harvested 24 hours following injection of CMFDA-FRCs. GFP signal was detected along with Meca-79⁺ HEVs (Red) and Lyve-1⁺ lymphatic vessels (Red). Data in graphs are represented as means \pm SD. Percentage of area stained positive in fluorescence micrographs was assessed in 3-6 random microscopic fields for each mouse. **p* < 0.05; ***p* < 0.01 by Student's t test and 2-way ANOVA with Tukey's multiple-comparisons test.

