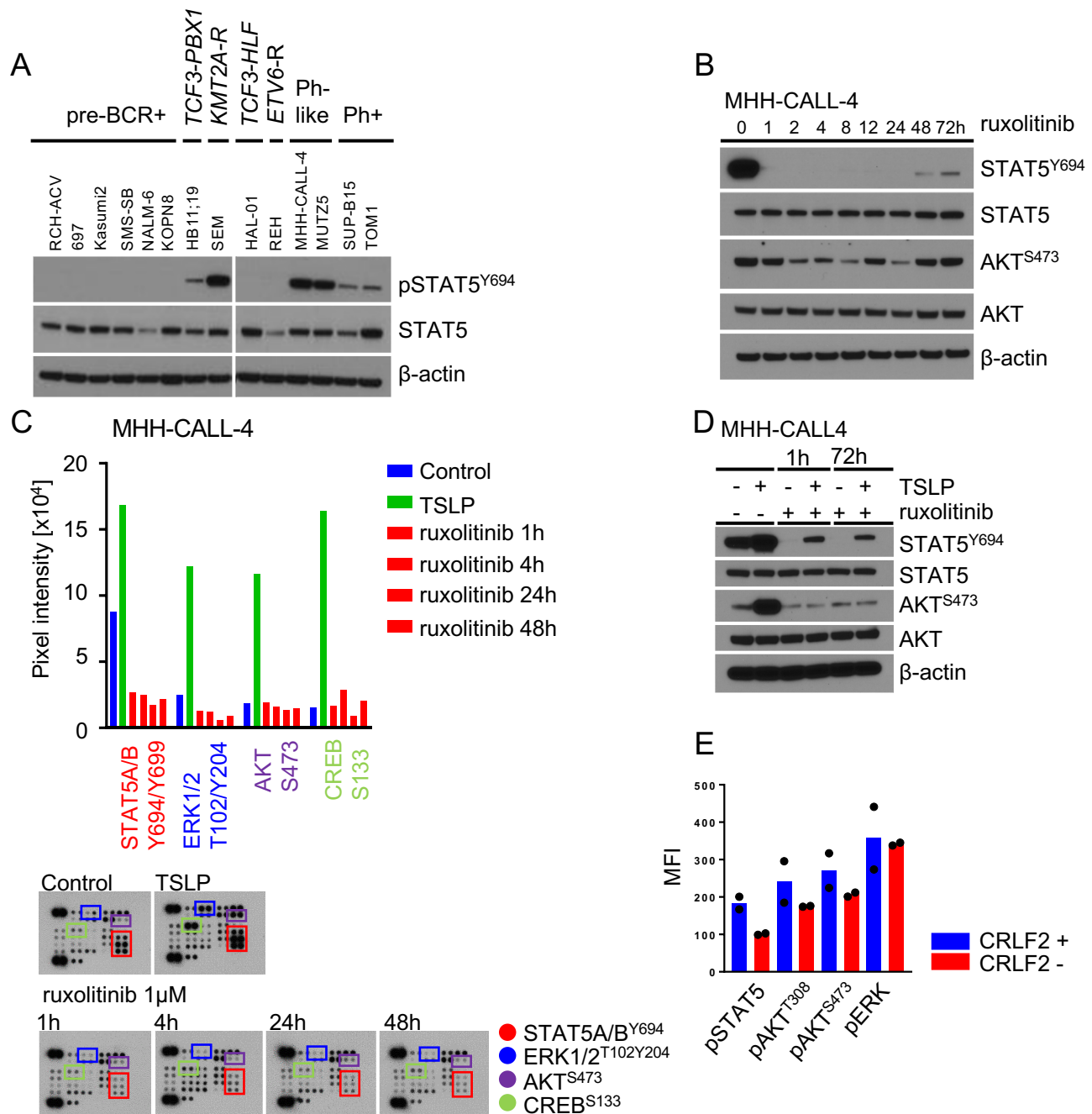
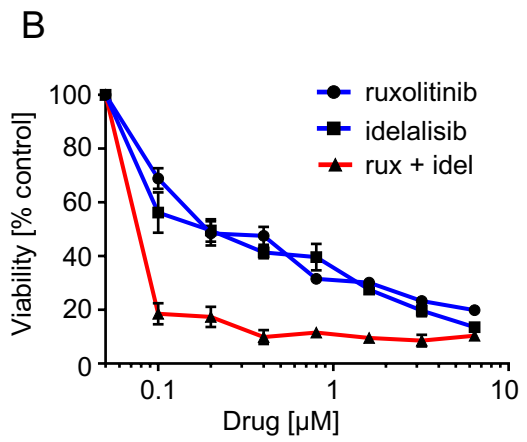
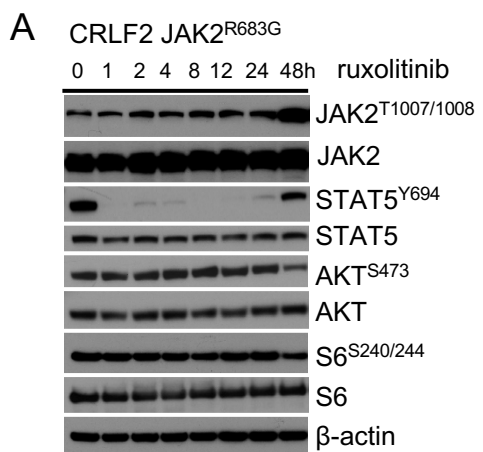


Supplemental Figure 1



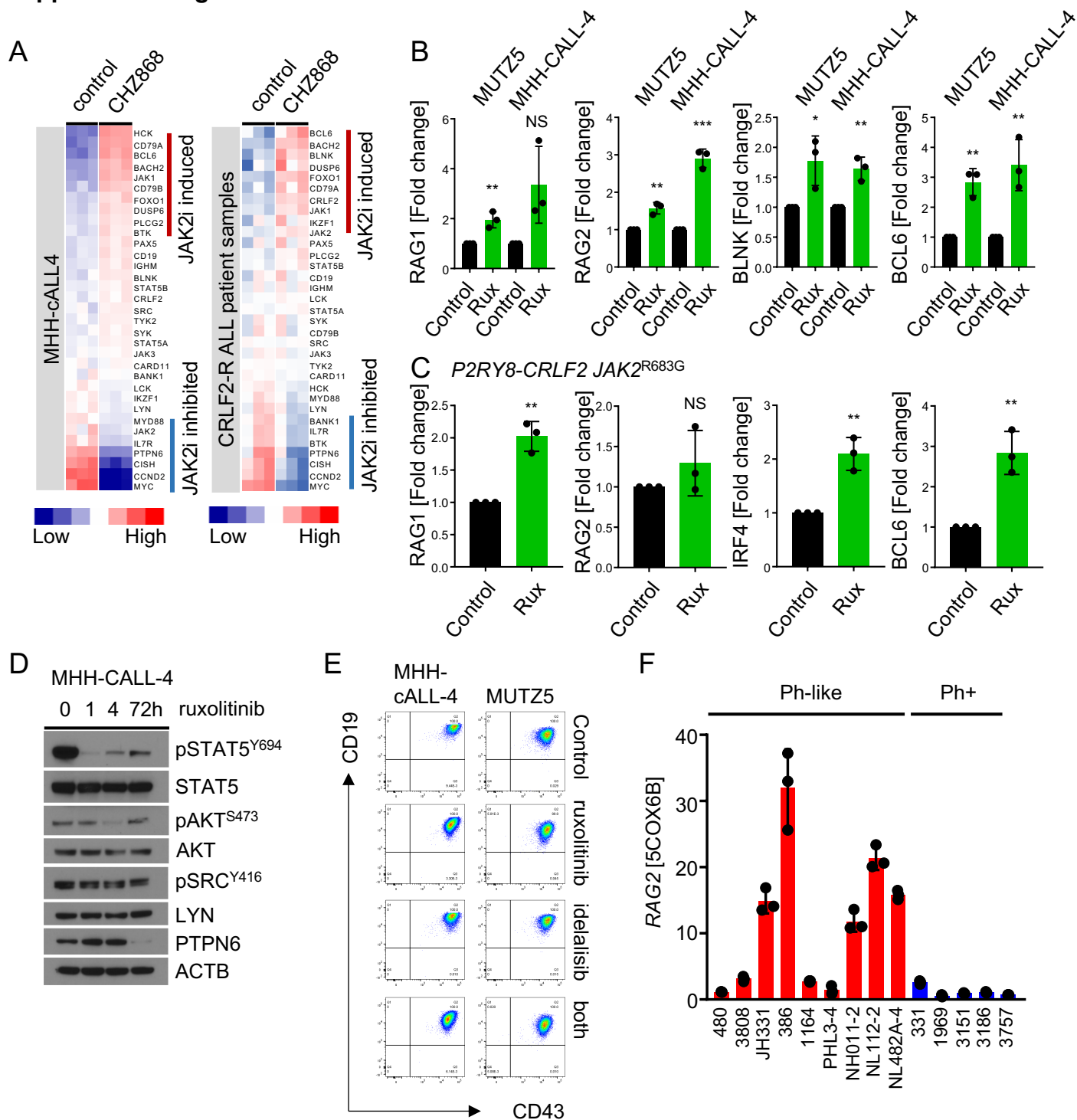
(A) Western blot analysis of pSTAT5 in multiple B-ALL cell lines using beta-actin as loading control. **(B)** Protein analysis of indicated targets in MHH-CALL-4 cells treated with 1 μ M ruxolitinib for the indicated timepoints. **(C)** Phosphoprotein analysis using the human phospho-kinase array from R&D Systems in MHH-CALL-4 cells treated with TSLP (25 nM x 30 min) or ruxolitinib (1 μ M x 1h). A scan of each blot for each condition is included on the bottom. The colored boxes indicate changes that were observed with TSLP stimulation (n=1). **(D)** MHH-CALL-4 cells were treated either with TSLP (25 nM x 30min) or ruxolitinib (1 μ M x 1h or 72h) alone or TSLP was added to the ruxolitinib treated samples (25nM; 30 min). Protein expression of pSTAT5^{Y694}, pAKT^{S473}, and the corresponding total protein controls were tested via Western blot using beta-actin as control. **(E)** MUTZ5 cells isolated from the spleen of terminally sick NSG mice (shown in Figure 2E) were tested for signaling activation via phosphoflow in CRLF2-null and CRLF2-expressing cells by gating on the corresponding cells (see Figure 2E). Only two of the three injected mice were available for this analysis.

Supplemental Figure 2



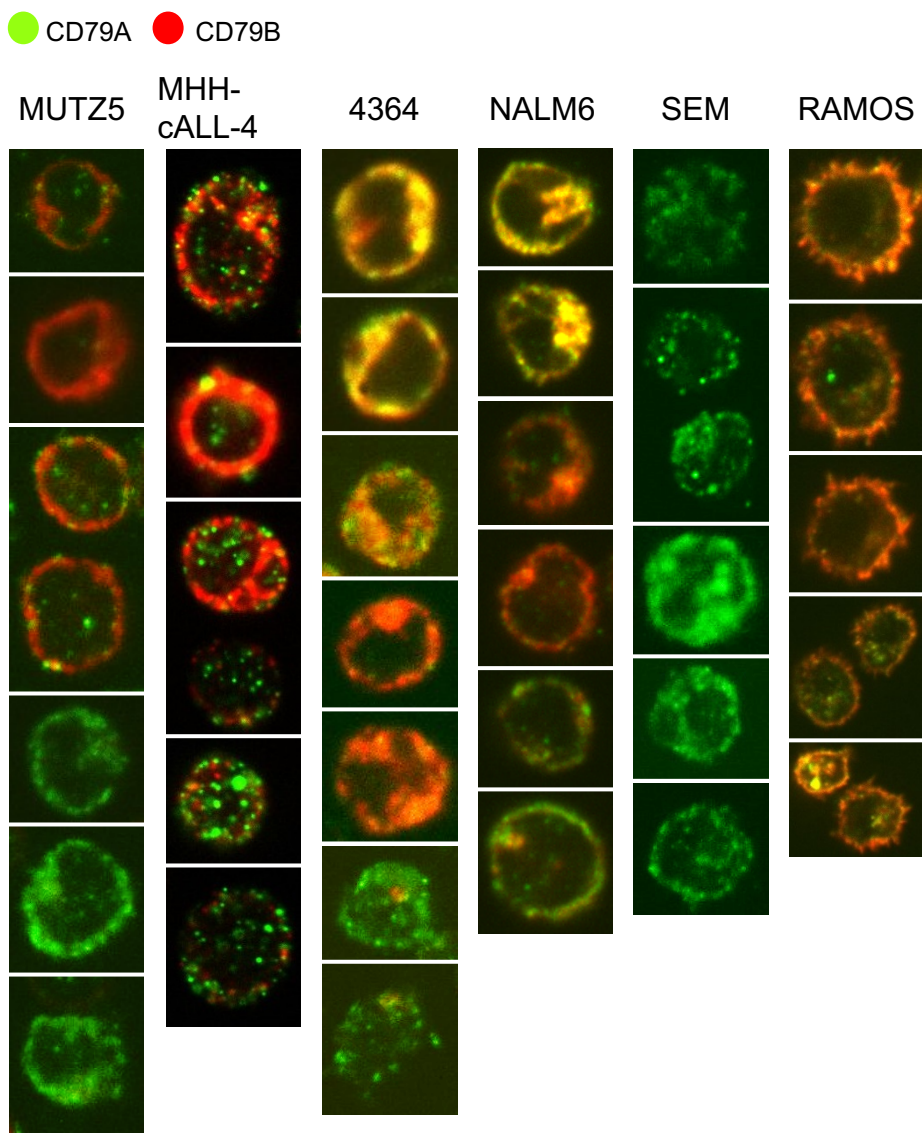
(A) Protein analysis of indicated targets in murine CRLF2/JAK2^{R683G} Ph-like ALL cells treated with 500 nM ruxolitinib for the indicated time points. β-actin was used as loading control. **(B)** Viability of mCRLF2-JAK2 cells treated with increasing concentrations of ruxolitinib and idelalisib (blue) or with 1 μM idelalisib and increasing concentrations of ruxolitinib (red) was assessed (n=3). Data are represented as individual values with mean ± SEM bars.

Supplemental Figure 3



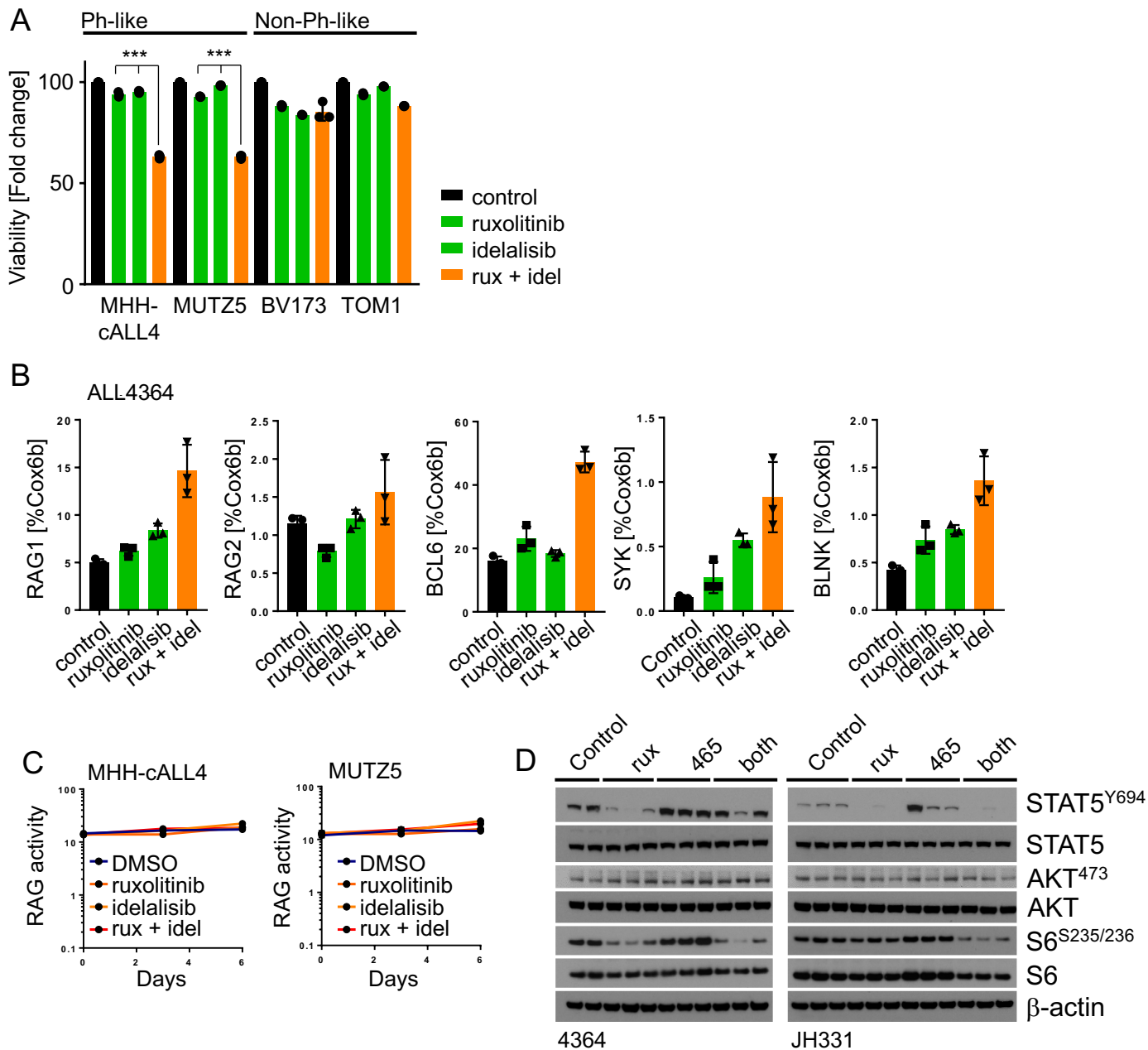
(A) Supervised meta-analysis of gene expression data (GSE61696). RT-PCR validation of gene expression data for the indicated targets in **(B)** MUTZ5 and MHH-CALL-4 and **(C)** Ph-like ALL4364 PDX model treated in vitro with 1 μ M ruxolitinib for 12h (n=3 independent experiments/biologic replicates each with 3 technical replicates). **(D)** MHH-CALL4 cells were treated with 1 μ M ruxolitinib for indicated timepoints. Protein levels for the indicated targets were determined via Western blotting. **(E)** Flow cytometric analysis of CD19 and CD43 expression in MHH-CALL-4 cells treated with ruxolitinib (1 μ M), idelalisib (1 μ M) or both (n=3). **(F)** *RAG2* expression levels were tested in the indicated PDX samples (Ph-like red; Ph+ blue) via RT-PCR using *COX6B* as control (n=3). Data are represented as individual values with mean \pm SEM bars.

Supplemental Figure 4



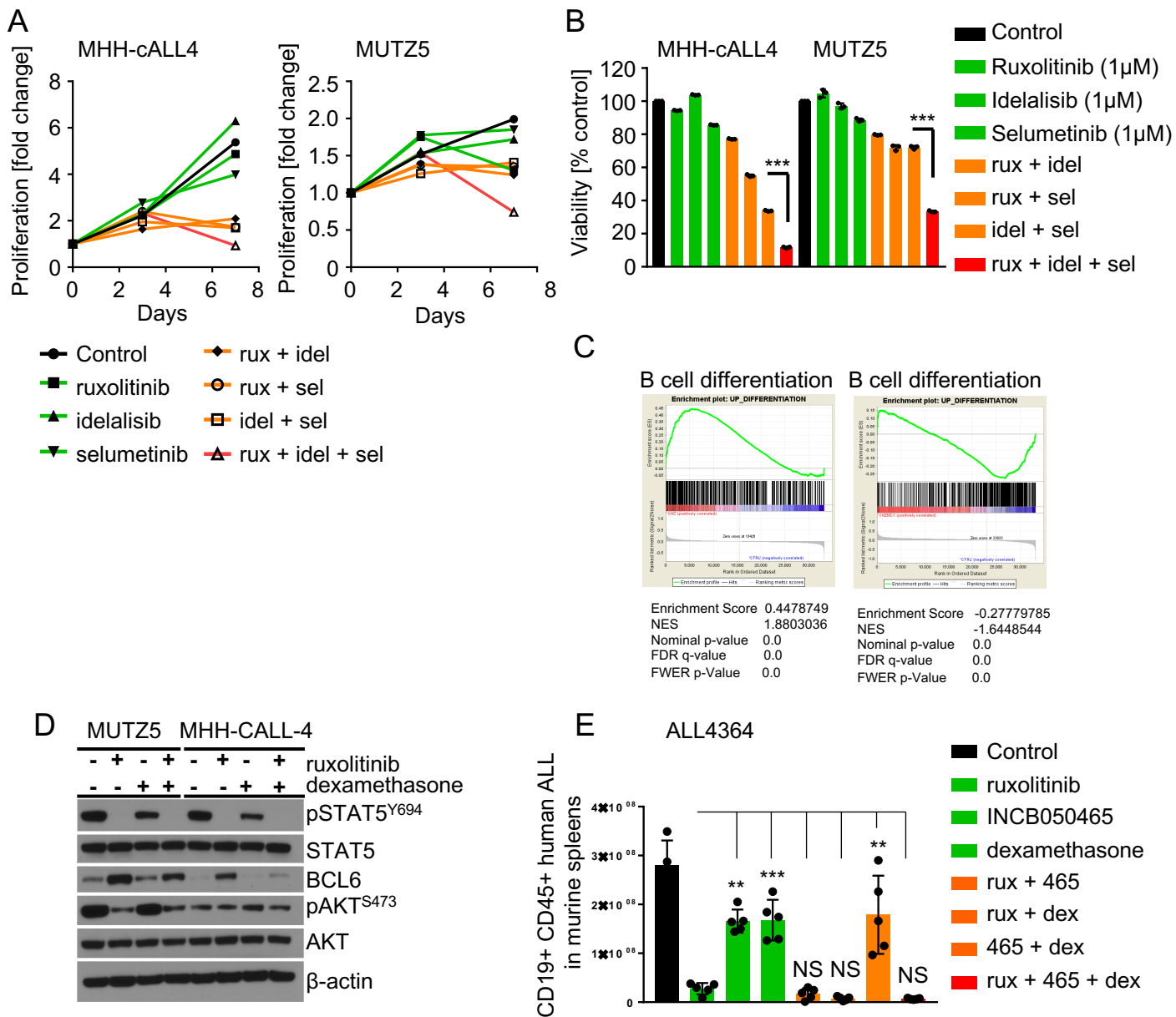
Immunofluorescence microscopy for CD79A (green) and CD79B (red) in Ph-like ALL cell lines (MUTZ5, MHH-CALL-4) and one Ph-like ALL PDX sample (ALL4364). The CD79B-negative *KMT2A-R* B-ALL cell line SEM was used as negative control, and the CD79A+/CD79B+ lymphoma cell line Ramos was used as positive control.

Supplemental Figure 5



(A) Viability analysis via flow cytometry in 2 Ph-like and 2 Ph+ ALL cell lines after 3 days of treatment with ruxolitinib, idelalisib, or a combination of both (each 1 μ M, n=3 independent experiments). **(B)** Cells from the Ph-like *CRLF2-R* PDX ALL4364 model were treated for 4h with 1 μ M ruxolitinib and 2 μ M idelalisib or both drugs. RT-PCR was performed, and expression levels of the indicated targets was tested in comparison to *COX6B* as control (n=3 technical repeats). **(C)** MUTZ5 and MHH-CALL-4 cells were lentivirally transduced with a RAG enzyme activity reporter. Both cell lines were treated with ruxolitinib and idelalisib for 6 days with a concentration of 1 μ M for each drug. **(D)** Protein analysis of pSTAT5 and pS6 of isolated ALL PDX cells from the in vivo experiment described in Figure 4 D and E. Three mice were used for each treatment group. Data for (A) are represented as individual values with mean \pm SEM bars, *p<0.05, **p<0.01, ***p<0.001, ****p<0.0001 by unpaired t-test.

Supplemental Figure 6



(A) MHH-CALL-4 and MUTZ5 cells were treated with the indicated kinase inhibitors, and cell proliferation was monitored over time using an automated cell counter (7 days, n=3). (B) 7AAD/ANNEXIN V flow cytometry analysis of MHH-CALL-4 and MUTZ5 cells treated with indicated kinase inhibitors (1 µM each; 3 days, n=3). (C) Gene set enrichment analysis (GSEA) plots demonstrate i) enrichment of B cell differentiation gene sets in CRLF2-rearranged B-ALL PDXs cells treated with CHZ868 compared to vehicle-treated controls (left panel) or ii) cotreatment with dexamethasone (right panel) for treatment details see⁴⁴. Statistics are shown. (D) MHH-CALL-4 and MUTZ5 cells were treated with ruxolitinib (1µM) or dexamethasone (10 nM) for 4h and subjected to a protein analysis of the indicated target genes. (E) Mice were injected via tail vein with Ph-like ALL PDX ALL4364 cells. Once 5% blast were detected in the blood, therapy of the indicated drug combination was initiated (see Material and Methods for dosing). Mice were treated for 4 weeks and sacrificed 12h after the treatment has stopped and spleen and blood cells were analysis via flow cytometry for CD19+/CD45+ ALL cells. Data are represented as individual values with mean ± SEM bars, *p<0.05, **p<0.01, ***p<0.001, ****p<0.0001 by unpaired t-test (B) or ANOVA with Dunnett post-test for multiple comparisons (E).

Supplemental Table 1

Patient	USI	Age (years)	WBC Count (10e3 cells/uL)	Sex	Race/Ethnicity	Genomic Lesion(s)	Cytogenetics	Notes	Induction Chemotherapy Regimen or Trial	TKI	HSCT
Ph-like B-ALL (n=14)											
1	UP_386	64	28.6	F	Caucasian	<i>P2RY8-CRLF2, JAK2 R683G</i>	46,XX		hyperCVAD	no	unknown
2	UP_480	49	2.9	M	Caucasian	<i>IGH-CRLF2</i>	46,XY		E2993	no	yes
3	UP_810	62	9.7	M	Caucasian	* <i>CRLF2+</i>	46,XY		hyperCVAD	no	no
4	UP_1164	42	19.3	M	Caucasian	<i>P2RY8-CRLF2, JAK2 K682delinsGG E</i>	n/a		hyperCVAD	no	yes
5	UP_2142	29	136.2	M	Caucasian	<i>IGH-CRLF2</i>	46,XY,del(9)(p21p21)[6]/46,X,Y[24]	Positive FISH for deletion of 9p21 (<i>CDKN2A</i> and <i>CDKN2B</i>); nuc ish(9p11-q11.1x2),(CDKN2Ax1)[130/200]	C10403	no	yes
6	UP_2549	28	2.2	M	Caucasian	<i>IGH-CRLF2, JAK2 R938Q</i>	46,XY		C10403	no	no
7	UP_3211	63	14.5	M	Caucasian	<i>IGH-CRLF2, JAK2 F694L</i>	n/a		hyperCVAD	no	no
8	UP_3808	43	42.1	M	Caucasian	<i>IGH-CRLF2, JAK2 R683G</i>	46,XY		hyperCVAD	no	unknown
9	UP_4986	37	193.5	M	Hispanic/Latino	<i>IGH-CRLF2, JAK2 L977L and C616_C618delinsGV</i>	46,XY		E1910/arm A	no	no
10	UP_4988	60	60.5	F	Hispanic/Latino	<i>IGH-CRLF2, JAK2 F694S</i>	46,XX,del(7)(p11.2)[7]/46,XX[13]	<i>KRAS G12R & NOTCH1 VUS</i>	E1910/arm A	no	yes
11	UP_5049	41	46	F	Caucasian	Ph-like kinase fusions not detected	45,XX,add(3)(q?24),-7[6]/46,XX[4]	<i>FLT3</i> internal tandem duplication	yes	no	yes
12	UP_5318	29	23.2	M	Caucasian	truncated <i>EPOR, TBL1XR1-PIK3CA</i>	46,XY,t(2;19)(p21;p13.1)[2]/46,XY[18]	Negative for <i>ABL1</i> and <i>JAK2</i> fusion genes on panel.	C10403	no	no
13	UP_6005	75	24.2	M	Hispanic/Latino	<i>IGH-CRLF2, KRAS G12D</i>	48,XY,+8,+21[12]/46,XY[2]	<i>BCORL1 P562Rfs*43 and FLT3 N676K</i> pathogenic mutations on CPD	ECOG 1910	no	no
14	UP_6096	76	48.2	F	Caucasian	<i>TP53 R280S</i>	35-38,XX,-3,-7,-9,dic(10;18)(q26;p11.2),der(12)t(9;12)(q13;p13),-13,-15,-16,-17,add(17)(p11.2),-19,-20,+1-2mar[c p11]/46,XX[11]	<i>BCOR D1712N (VUS) and TPMT A154T and Y240C</i>	weekly vincristine, IT cytarabine	no	no

Supplemental Table 1

Patient	USI	Age (years)	WBC count (10e3 cells/uL)	Sex	Race/Ethnicity	Genomic lesion(s)	Cytogenetics	Notes	Induction Chemotherapy Regimen or Trial	TKI	HSCT
Ph+ B-ALL (n=33)											
15	UP_408	21.9	31.3	M	Caucasian	BCR-ABL1 p190	46,XY[11]	BCR-ABL1+ by RT-PCR	E2993	no	unknown
16	UP_1120	n/a	161	M	n/a	BCR-ABL1 p190	n/a		unknown	unknown	unknown
17	UP_1182	61	35.1	F	Caucasian	BCR-ABL1 p190	46,XX,t(9;22)(q34;q11.2)[9]		hyperCVAD	yes - imatinib	unknown
18	UP_1441	37	35.1	F	Hispanic/Latino	BCR-ABL1 p190	n/a		hyperCVAD	yes - imatinib	yes
19	UP_1866	64	19.4	M	n/a	BCR-ABL1 p190	53-55,XY,+X,+der(1)t(1;7)(p13;q11.2),+2,+6,-7,t(9;22)(q34;q11.2),+10,+11,+14,+18,+21,+der(22)t(9;22),+mar[cp16]/46,XY[4]		hyperCVAD	yes - imatinib	no
20	UP_1969	52	17.7	F	Caucasian	BCR-ABL1 p210	46,XX,t(9;22)(q34;q11.2)[10]/45,XX,-7,t(9;22)(q34;q11.2)[3]/46,XX[12]		hyperCVAD	yes - imatinib	no
21	UP_2270	54	12.6	M	Caucasian	BCR-ABL1 p210	46,XY,t(9;22)(q34;q11.2)[10]/46,XY[15]		hyperCVAD	yes - dasatinib	no
22	UP_2802	78	6.8	F	Caucasian	BCR-ABL1 p190	46,XX,t(9;22)(q34;q11.2)[6]/46,XX[2]		methotrexate & cytarabine	yes - dasatinib	no
23	UP_3151	65	58.3	F	Black/African American	BCR-ABL1 p190	n/a		hyperCVAD	yes - dasatinib	no
24	UP_3186	53	75.4	F	Caucasian	BCR-ABL1 p210	46,XX		hyperCVAD	yes - dasatinib	yes
25	UP_3285	88	56	M	Caucasian	BCR-ABL1 p190	46,XY,der(9)del(9)(p22)t(9;22)(q34;q11.2),der(22)t(9;22)[10]/46,XY,-8,der(9)t(8;9)(q12;p22)t(9;22)(q34;q11.2)x2,der(22)t(9;22)[8]		prednisone	yes - dasatinib	no
26	UP_3304	42	18.6	F	Caucasian	BCR-ABL1 p210	46,XX		hyperCVAD	yes - dasatinib	no
27	UP_3360	40	22.7	F	Caucasian	BCR-ABL1 p190	46,XX		S0805	yes - dasatinib	yes
28	UP_3434	59	30.9	M	Caucasian	BCR-ABL1 p190	46,XY		S0805	yes - dasatinib	yes
29	UP_3529	79	126.1	F	Caucasian	BCR-ABL1 p210	46,XX,t(9;22)(q34;q11.2)[9]		prednisone, cytarabine, methotrexate	yes - dasatinib	unknown
30	UP_3757	45	45.3	M	Caucasian	BCR-ABL1 p210	46,XY,der(2)t(1;2)(q12;q37),t(7;9;22)(q11.2;q34;q11.2)[7]		S0805	yes - dasatinib	yes
31	UP_4260	53	38.6	M	Black/African American	BCR-ABL1 p210	46,XY,t(9;22)(q34;q11.2)[2]/44,XY,der(4)t(9;22)(q34;q11.2)t(4;9)(p15.2;q21),-7,-9,der(22)t(9;22)[14]/46,XY[4]		hyperCVAD	yes - dasatinib	no
32	UP_5204	74.3	73.5	M	Caucasian	BCR-ABL1 p210	46,XY,t(9;22)(q34;q11.2)[5]/50,idem,+Y,t(7;15)(p13;q15),+8,+10,+21[6]		prednisone	yes - dasatinib	no

* HSCT = hematopoietic stem cell transplantation, TKI = tyrosine kinase inhibitor, USI = unique specimen identifier, WBC = white blood cell

Supplemental Table 2

Cell line	Genetic rearrangement	JAK2 mutation	Malignancy type	LDA positive	Source
MUTZ5	<i>IGH-CRLF2</i>	R683G	B-ALL	yes	DSMZ
MHH-CALL-4	<i>IGH-CRLF2</i>	I682F	B-ALL	yes	DSMZ
BV173	<i>BCR-ABL1</i>		B-ALL		DSMZ
TOM-1	<i>BCR-ABL1</i>		B-ALL		DSMZ
SUP-B15	<i>BCR-ABL1</i>		B-ALL		DSMZ
HAL-01	<i>TCF3-HLF</i>		B-ALL		DSMZ
RCH-ACV	<i>TCF3-PBX1</i>		B-ALL		DSMZ
697	<i>TCF3-PBX1</i>		B-ALL		DSMZ
Kasumi-2	<i>TCF3-PBX1</i>		B-ALL		DSMZ
HAL-01	<i>TCF3-HLF</i>		B-ALL		DSMZ
NALM-6	<i>ETV6-PDGFRB</i>		B-ALL		DSMZ
KOPN-8	<i>KMT2A-MLL1</i>		B-ALL		DSMZ
HB11;19	<i>KMT2A-MLL1</i>		B-ALL		Gift from Dr Patrick Brown (Johns Hopkins University; Baltimore, Maryland)
SEM	<i>KMT2A-AFF1</i>		B-ALL		DSMZ
REH	<i>ETV6-RUNX1</i>		B-ALL		DSMZ
SMS-SB	<i>PDGFRB+</i>		B-ALL		Gift from Dr Markus Muschen (City of Hope; Duarte, California)
Ramos	<i>MYC-IGH</i>		Burkitt lymphoma		DSMZ

ALL PDX USI	Genetic rearrangement	JAK2 mutation	Other genetic alterations	LDA positive	Diagnosis/ Relapse	Source
UP_ALL2142	<i>IGH-CRLF2</i>			yes	diagnosis	Penn SCXC
ALL121	<i>IGH-CRLF2</i>	R683G		yes	relapse	CHOP CCCR
JH331	<i>IGH-CRLF2</i>	R683G		yes	diagnosis	COG
ALL2128	<i>IGH-CRLF2</i>			no	diagnosis	CHOP CCCR
UP_ALL588	<i>P2YR8-CRLF2</i>			yes	relapse	Penn SCXC
ALL4364	<i>P2YR8-CRLF2</i>	R683G		yes	relapse	CHOP CCCR
ALL185GD	<i>P2RY8-CRLF2</i>		<i>PAX5-AUTS2, CDKN2A/B del</i>	yes	diagnosis	COG
NL482A	<i>BCR-JAK2</i>			yes	diagnosis	COG
NL112	<i>STRN3-JAK2</i>			yes	diagnosis	COG
PAXDBJ	<i>GOLAG5-JAK2</i>			yes	diagnosis	CHOP CCCR
UP_ALL240	<i>IGH-EPOR</i>			yes	diagnosis	Penn SCXC
NL432	<i>EBF1-PDGFRB</i>			yes	diagnosis	COG
NH011	<i>NUP214-ABL1</i>		<i>IKZF1, CDKN2A, PAX5 dels</i>	yes	diagnosis	COG
ALL-NT	<i>RCSD1-ABL1</i>			yes	relapse	CHOP CCCR
PHL3	<i>ETV6-ABL1</i>			yes	diagnosis	COG
TVA1	<i>ETV6-ABL1</i>			yes	diagnosis	Gift from Drs Vo and Fruman (University of California, Irvine)
UP_ALL3529	<i>BCR-ABL1</i>			(yes; Ph+)	diagnosis	Penn SCXC
UP_ALL5204	<i>BCR-ABL1</i>			(yes; Ph+)	diagnosis	Penn SCXC
ALL1807 (STL-CHPB1719)	<i>TCF3-HLF</i>		<i>IKZF1 del</i>	no	relapse	CHOP CCCR

B-ALL patient specimen (adult)	Genetic rearrangement	JAK2 mutation	LDA positive	Diagnosis/ Relapse	Source
240	<i>IGH-EPOR</i>		yes	diagnosis	Penn SCXC
480	<i>IGH-CRLF2</i>		yes	diagnosis	Penn SCXC
1003	<i>IGH-CRLF2</i>		yes	relapse	Penn SCXC
2142	<i>IGH-CRLF2</i>		yes	diagnosis	Penn SCXC
3808	<i>IGH-CRLF2</i>	R938Q	yes	diagnosis	Penn SCXC
4988	<i>IGH-CRLF2</i>	F694S	yes	relapse	Penn SCXC
386	<i>P2RY8-CRLF2</i>	R683G	yes	diagnosis	Penn SCXC
331	<i>BCR-ABL1</i>			relapse	Penn SCXC
1969	<i>BCR-ABL1</i>			diagnosis	Penn SCXC
3186	<i>BCR-ABL1</i>			diagnosis	Penn SCXC
3151	<i>BCR-ABL1</i>			diagnosis	Penn SCXC
3757	<i>BCR-ABL1</i>			diagnosis	Penn SCXC
4835	<i>BCR-ABL1</i>			diagnosis	Penn SCXC
3958	trisomy 13, del(17p)			diagnosis	Penn SCXC

* CHOP CCCR = Children's Hospital of Philadelphia Center for Childhood Cancer Research (Philadelphia, Pennsylvania), COG = Children's Oncology Group (Monrovia, California), DSMZ = Deutsche Sammlung von Mikroorganismen und Zellkulturen (<https://www.dsmz.de/>); Leibniz Institute; Braunschweig, Germany), Penn SCXC = University of Pennsylvania Stem Cell and Xenograft Core (Philadelphia, Pennsylvania)

Supplemental Table 3

Drug Name	Target	Vendor	Product ID#
ruxolitinib (formerly INCB18424)	JAK1/2	Incyte Corporation	
ruxolitinib	JAK1/2	SelleckChem	S1378
idelalisib (formerly CAL-101)	PI3K δ	SelleckChem	S2226
parsaclisib (INCB050465)	PI3K δ	Incyte Corporation	
dasatinib	SRC/ABL/BTK	LC Laboratories	D-3307
selumetinib	MEK	LC Laboratories	S-4490
dexamethasone	glucocorticoid receptor	Mylan	67457-422-54

Supplemental Table 4

Antibodies used for Western blotting			
Antigen	Clone	Manufacturer	ID #
pJAK2-Tyr1007/1008		Cell Signaling	3771S
JAK2	D2E12	Cell Signaling	3230S
pSTAT5-Tyr694	D47E7	Cell Signaling	4322S
STAT5	D2O6Y	Cell Signaling	94205S
STAT5	C-17	Santa Cruz	SC-835-G
pAKT-Ser473	D9E	Cell Signaling	4060L
AKT	C67E7	Cell Signaling	4691L
pERK-Thr202/Tyr204	D13.14.4E	Cell Signaling	4370S
ERK	3A7	Cell Signaling	9107S
pS6-Ser240/244	D68F8	Cell Signaling	5364L
S6	5G10	Cell Signaling	2217L
pSRC-Tyr416		Cell Signaling	2101S
LYN	C13F9	Cell Signaling	2796S
PTPN6	C14H6	Cell Signaling	3759S
CD79A		Cell Signaling	3351S
CD79B	D7V2F	Cell Signaling	96024S
CD19	D4V4B	Cell Signaling	90176S
pBTK-Tyr223	D9T6H	Cell Signaling	87141S
BTK	D6T2C	Cell Signaling	56044S
BCL6	D65C10	Cell Signaling	5650S
beta-actin	AC-15	Sigma-Aldrich	A5441
pFOXO1-T24/FOXO3-T32		Cell Signaling	9464S
FOXO1	c29h4	Cell Signaling	2880S
Immunohistochemistry			
Antigen	Clone	Manufacturer	ID #
CD79A	HM47/A9	ThermoFisher	MA-513212
CD79B	EPR6861	Abcam	ab134147
Flow cytometry			
Antigen (mouse)	Clone	Manufacturer	ID #
CD19-APC-Cy7	1D3	BD	557655
TSLPR (CRLF2)-PE	eBio1A6	ThermoFisher	12-5499-42
Antigen (human)	Clone	Manufacturer	ID #
CD10-PE-Cy7	eBioCB-CALLA	ThermoFisher	25-0106-42
CD19-APC-Cy7	SJ25-C1	ThermoFisher	A15429
CD19-BV786	SJ25C1	BD	563325
CD19-PE	H1B19	ThermoFisher	12-0199-42
CD43-PE	CD43-10G7	Biolegend	343204
CD45-APC	HI30	ThermoFisher	MHCD4505
CD45-APC	2D1	ThermoFisher	17-9459-42
CD79A-Alexa488	ZL7-4	BioRad	MCA1298A488
CD79B-PE-Cy5	CB3-1	BD	551063
TSLPR (CRLF2)-PE	eBio1A6	ThermoFisher	12-5499-42
TSLPR (CRLF2)-PerCP	1A6	ThermoFisher	46-5499-41
pAKT ^{T308} -Alexa 488	D25E6	Cell Signaling	43506S
pAKT ^{S473} -Alexa 647	D9E	Cell Signaling	4075S
pERK ^{T202/Y204} -Alexa 647	D13.14.4E	Cell Signaling	4284S

Supplemental Table 5

MUTZ5						
Sequence variant(s)						
Gene	Genomic Position (hg19)	Reference	Nucleotide	Amino Acid	VAF	Comment
<i>IKZF1</i>	chr7:50444470-50444471	NM_006060.5	c.400_401insGGG CTCCG	p.Val134Glyfs* 62	0.58	
<i>JAK2</i>	chr9:5078360-5078360	NM_004972.3	c.2047A>G	p.Arg683Gly	0.41	COSM29300
<i>CDC25C</i>	chr5:137621491-137621491	NM_001790.4	c.1312C>G	p.Gln438Glu	0.41	
<i>NSD1</i>	chr5:176637415-176637415	NM_022455.4	c.2015C>T	p.Thr672Ile	0.49	
<i>SF1</i>	chr11:64534522-64534522	NM_004630.3	c.1432A>T	p.Met478Leu	0.53	
<i>SH2B3</i>	chr12:111885984-111885984	NM_005475.2	c.1606G>A	p.Ala536Thr	0.42	rs140649197

Copy Number Variant(s)					
Chromosome	Band	Abnormality	Gene	Comments	
X	partial Xp	loss	<i>KDM6A</i>	Homozygous deletion	
2	partial 2p	loss	<i>MSH6</i>		
4	partial 4p	loss	<i>LEF1</i>		
5	partial 5p	loss	<i>EBF1</i>		
7	partial 7p	loss	<i>IKZF1</i>		
9	partial 9p	loss	<i>CDKN2A/B, PAX5</i>	Homozygous deletion of <i>CDKN2A/2B</i>	
10	partial 10p	loss	<i>PTEN</i>		
10	partial 12p	loss	<i>ETV6</i>	Homozygous deletion	

MHH-CALL-4						
Sequence variant(s)						
Gene	Genomic Position (hg19)	Reference	Nucleotide	Amino Acid	VAF	Comment
<i>JAK2</i>	chr9:5078357-5078357	NM_004972.3	c.2044A>T	p.Ile682Phe	0.48	COSM303887
<i>ETV6</i>	chr12:12038883- 12038896	NM_001987.4	c.1177_1189del	p.Lys393Profs*8	0.83	
<i>KRAS</i>	chr12:25398251- 25398251	NM_033360.3	c.68T>G	p.Leu23Arg	0.44	rs730880472;C OSM303853
<i>KMT2D</i>	chr12:49426916- 49426917	NM_003482.3	c.11571_11572insT	p.Gln3858Serfs*154	0.1	
<i>USH2A</i>	chr1:216052344- 216052344	NM_206933.2	c.8320G>A	p.Ala2774Thr	0.46	rs111033533;C OSM4783112
<i>ABL1</i>	chr9:133760372- 133760372	NM_005157.5	c.2695C>T	p.Pro899Ser	0.45	rs767171554

Copy Number Variant(s)					
Chromosome	Band	Abnormality	Gene	Comments	
7	partial 7p	loss	<i>IKZF1</i>		
9	partial 9p	loss	<i>CDKN2A/2B, PAX5</i>	Homozygous loss of <i>CDKN2A/2B</i>	
12	partial 12p	loss	<i>ETV6</i>		

Supplemental Table 6

Viral construct	Insert	Notes	Vector	Ligation method
<i>CRLF2</i> retro myc IRES mcherry	PCR product, Genescript OHu23392D myc tag (NotI, AgeI cloning sites, for C-terminal myc epitope tag)		lentiCRISPRv2 blast Addgene 398239 ¹	Gibson (NEB HiFi) oligo ligation/T4 ligase (invitrogen)
<i>PAX-JAK2</i>	PCR of N-term PAX5, Genescript OHu14569D	Junction in PMID: 2289784 I	IRES mCherry Addgene 80139 ² retro myc IRES mCherry	Gibson (NEB HiFi)
<i>EBF-PDGFRB</i>	PCR of C-term JAK2 (Addgene 23915) PCR of N-term EBF1, Genescript OHu20245D	Junction in PMID: 2289784 I	retro myc IRES mCherry	Gibson (NEB HiFi)
<i>JAK2 R683G</i>	PCR of C-term PDGFRB, Genescript OHu25829D PCR product, Addgene JAK2 (23915) ³	I	retro myc IRES mCherry	Gibson (NEB HiFi)

Viral construct	Notes	Source
MSCV-RFP	Control vector for GFPi	Kind gift from Jocelyne Demengeot ⁴
MSCV-GFPi	RAG activity reporter	Kind gift from Jocelyne Demengeot ⁴
Luciferase-T2A-GFP		

References for constructs:

1. Stringer, B.W., et al. A reference collection of patient-derived cell line and xenograft models of proneural, classical and mesenchymal glioblastoma. *Scientific reports* 9, 4902 (2019).
2. Dick, H.M., Wright, P., Chapman, C.M., Zacharias, F.J. & Nicholls, J.T. Adverse reactions to practolol: some observations on the possible relevance to immune mechanisms. *Allergy* 33, 71-75 (1978).
3. Wang, S., et al. JAK2-binding long noncoding RNA promotes breast cancer brain metastasis. *The Journal of clinical investigation* 127, 4498-4515 (2017).
4. Trancoso I, Bonnet M, Gardner R, Carneiro J, Barreto VM, Demengeot J, and Sarmiento LM. A Novel Quantitative Fluorescent Reporter Assay for RAG Targets and RAG Activity. *Frontiers in immunology*. 2013;4(110).

Supplemental Table 7

Quantitative RT-PCR primers	Sequence
<i>BCL6</i>	F_TGAGAAGCCCTATCCCTGTG R_TGTGACGGAAATGCAGGTTA
<i>BLNK</i>	F_CCAGCTGGCCTTCAGAGAAA R_ATGAAAAGCTGGGTAGGGGC
<i>RAG1</i>	F_ATAGAAGAAAGCAACACAAAAGC R_ATACTGAGTTCAATCCCTGAAGA
<i>RAG2</i>	F_ATAGCAAGAGCTCTACACACTCC R_AAAAATCAGATCAGAAATCCTCA
<i>SYK</i>	F_CACAAAGCAAAGGCAGTCCC R_AGGGGGAAAGTAAGCCAAGC
<i>COX6B</i>	F_AACTACAAGACCGCCCCTTT R_GCAGCCAGTTCAGATCTTCC
sgRNAs purchased from IDT	Sequence
<i>CD79B</i>	CD79B.1.AA_AACACCTCGGAGGTCTACCA
<i>CRLF2</i>	CRLF2.1.AC ACCTGCAACGTCACCATAGA

Supplemental Table 8

Gene expression signature									
Up-differentiation									
BCL6	PAFAH1B3	PHKA2	CMTM7	PGLS	WDR90	CLK1	TRAPPC1	ECH1	ABI1
FCRLA	BLNK	HDAC5	PSRC1	PNRC2	SFSWAP	GABARAP	DESI1	CENPN	WDR81
CD72	DDIT4	B3GNT8	DYNLT1	APOBEC1	PCYT1A	RNF167	UTRN	CD24	ZFP36L1
TMPRSS3	CCNG2	DNTT	RHOQ	ZBTB20	ZCCHC6	RMND5B	FOXP1	RAB37	PPP2R5A
LAPTM5	TIFA	TMEM71	KLHL24	CSRP2	CARHSP1	INPL1	SYAP1	REEP5	PRR14
UBLCP1	AP003419.11	CXCR4	IRF8	GPR155	CCNB2	KIF23	RDH11	ERO1B	FAM107B
NATD1	ARNTL	SORD	PLEKHO1	B3GNT2	MANBA	GIT2	CD53	LYRM2	PRKCG
SPP1	WBP1	LGR5	CDC42EP3	SRGN	RFX5	TGFBR2	SKAP2	HIVEP1	PLEKHO2
YPEL5	IL2RA	PIK3AP1	ITM2B	MTURN	PIGH	ZFP36L2	DDIT3	LAMP2	CARNS1
ADSSL1	ZFPM1	IGHV1-69	TESK2	WDFY1	CHKB	TNFRSF13C	RSPH9	AMDHD1	KDM3B
RGS2	RPL3L	EQTN	EBF1	RDH12	RAMP1	PTPRC	AC027682.1	TPP1	ZC3H7A
IL12A	SMARCA4	MTSS1	ATP6AP1	STX5	CENPF	TCTA	EHD2	KIF22	NR3C1
PECAM1	CTD-2545M3.6	NUSAP1	ARHGAP17	RELB	CENPE	ASPM	FBXO11	UCP2	TTC28
NTMT1	HERPUD1	MYLIP	SLC15A2	POLM	PCMTD2	ATP6V1D	IQSEC1	STK17B	TRIM6-TRIM34
PIPOX	HIST3H2A	MXD3	TSPYL4	SYK	CIC	SLA	PXK	BIRC3	TRIP4
RAB43	CDKN1A	IL7R	SERP1	OGT	MPP1	NT5C2	HEXB	GTPBP2	STK11IP
MYL4	BLK	SPATA6	TSPAN17	MYB	PIK3CG	CDCA3	ATXN7L1	RELL1	ZDHHC14
SBK1	TXNIP	RSPH1	COX2A2L	TCIRG1	DNASE1	FCHO1	TMA16	MKI67	GPSM3
PFN2	CPEB4	KLF13	ELK3	PLGRKT	CCPG1	IFFO1	MDM1	CRLF2	ZBTB18
BACH2	UNC93B1	IRF4	LGALS9	SKIL	NDUFAF3	ZEB2	TRIM59	IIFT22	CCNI
MXD4	PHKA1	ATP2A3	PNRC1	TRIOBP	VAMP8	UNC119B	LMO2	NCAPD2	ATG12
EGFL6	PLEKHG2	CDC25B	ZFAND3	PRKCB	MYO18A	TOB1	NRM	ITGAM	IRF2
MBD4	PIAS3	TBX6	PINK1	LITAF	TUBA1A	GF11B	TRAFD1	CTNNA1	IFNA7
SELL	SIT1	NFKBIE	DDX54	RDM1	HINT3	TRAM1	CCDC88A	PCBD1	IFNA7
YPEL3	GFRA1	EMP1	PITPNM1	ARHGAP45	DUSP2	FOXO1	GRINA	CDC37L1	PHF1
CXXC5	CPT1A	ABC4	ITPR2	PQLC3	INPP5K	AC013264.1	IFT122	PIAS1	PNPLA8
SLC23A1	H2AFV	PLEKHA2	TCF19	CRLF1	FRYL	SPEF1	SIRT7	AGFG2	ZRSR1
PREP	BTG2	IFT80	TSPO	PARP4	SAT1	HSPA2	CEP89	ADCY7	REEP2
FCGR2C	HBP1	ARHGAP18	MGST1	KMT2A	DUSP6	SDC4	HIPK3	TNFRSF13B	PPP1R18
EPS8	MYADM	ETS2	LIMD2	FMNL3	COTL1	CD37	PLAT	SLC7A7	STK10
CHST15	FAM63A	METTL7A	TCN2	ARPC5L	DUSP10	ZNF821	RHOH	RSRP1	SLC27A4
AP1S2	GFRA2	AKAP12	NEDD9	BAZ2B	TRIM16	RAB33B	FAM13B	BIRC2	TMEM229B
TP53INP1	ELOF1	BLVRB	IGKV1-6	TCF4	NNT	CEACAM1	XRCC6	CD79A	ARNT
ZAP70	SMIM14	CNP	NFAT5	P2RX3	CBFA2T3	CDK5RAP2	WSB2	CORO1B	EPC1
BTG1	GDI1	DNAH8	AKR1B10	PSAP	FAM64A	ATL2	MKRN1	RACGAP1	SRPK2
SEC63	ATP8A1	OTUB2	TAX1BP3	ABCD1	RCHY1	RB1	SERPINI1	SLAMF1	SLC30A4
RABGAP1L	TRIM11	GAS7	WDPCP	KANSL1L	MARCKS	NDC80	LASP1	RP11-286N22.8	CST5
PTGR1	IGKV4-1	SLC12A6	HIST1H1C	GADD45A	BTRC	TK2	OSBPL9	IQGAP1	STAT2
GLCC1	PSEN2	CAST	CEACAM3	NEIL3	IP6K1	ATP1B3	CHFR	DGKZ	RBM4B
EMP3	TLE6	CTSE	MAP1LC3B2	LUC7L2	RCSD1	CD38	GPM6A	SH3YL1	GENPA
MSH5	THBD	CD22	MYO7A	GSN	POC1B-GALNT4	STX7	SMIM3	SOX4	TM6SF1
DOK3	ID2	MTOR	KMT2E	PLD4	SNAP29	TGFB1	GP2	RRM2	CHCHD5
NFKBIA	RAG2	GPAM	CDKN2C	PDE4B	LGALS1	CCM2	GLRX	FGD6	CCDC28A
PHLDA1	TP53INP2	ABCA1	MAP7	ICAM2	NIPBL	MAST2	STK3	UBE2H	RP11-192H23.4
CERK	CDKN1B	CTDSP2	TRIB2	TMEM9B	ST8SIA4	SNAPC3	RNF103	RRM2B	SYVN1
CD93	CD79B	ACP5	CKLF	SLAMF6	AAK1	IL16	CNOT6L	CD19	ANKRD13A
RAG1	SLAMF7	CIT	HES6	LHPP	CNN3	VRK3	ZNF207	CMC2	C12orf57
CDKN2D	PBXIP1	EHBP1L1	RP11-307N16.6	MAP3K1	SESN3	TSPAN13	MEF2D	SPC25	CAT
HSPA1A	GRN	EIF2AK3	EDIL3	CASP6	SELP	HIVEP2	NSMF	ELK4	LTK
SLC12A3	TNFRSF19	PLXDC1	VPREB3	SH3KBP1	RB1CC1	SP4	BCL7A	CYBA	INPP5D