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CNS-targeting pharmacological interventions for the metabolic syndrome

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The metabolic syndrome (MetS) encompasses medical conditions such as obesity, hyperglycemia, high blood pressure, and dyslipidemia that are major drivers for the ever-increasing prevalence of type 2 diabetes, cardiovascular diseases, and certain types of cancer. At the core of clinical strategies against the MetS is weight loss, induced by bariatric surgery, lifestyle changes based on calorie reduction and exercise, or pharmacology. This Review summarizes the past, current, and future efforts of targeting the MetS by pharmacological agents. Major emphasis is given to drugs that target the CNS as a key denominator for obesity and its comorbid sequelae.

Introduction

The metabolic syndrome (MetS) encompasses a cluster of pernicious metabolic diseases that include visceral obesity, dyslipidemia, hyperglycemia, and hypertension (1). It is considered to be a silent killer owing to increases in the risk of heart attacks and related cardiovascular maladies (2). Additional evidence suggests a role for the MetS in the etiology of certain types of cancer (3) and cognitive impairments, particularly Alzheimer's disease (4). Reducing body weight by 5%-10% substantially lowers all MetS components, and thereby the risk of fatal concomitant diseases (5). However, in most obese individuals, dieting and exercise fail to achieve persistent weight loss (6). These obese individuals could benefit from pharmacological interventions that decrease energy intake by enhancing satiety and reducing hunger and food cravings or increase energy expenditure and improve glycemic control (7).

Homeostatic and hedonic mechanisms underlying CNS-regulated metabolism

The CNS plays a pivotal role in regulating food intake and energy balance by adjusting daily energy requirements and sustaining bodily functions (8). The CNS receives satiation signals about energy input and availability from the gastrointestinal (GI) tract, as well as adiposity signals about energy storage from the white adipose tissue (WAT). These inputs are integrated in multiple centers within the CNS and incorporated into humoral and neuronal outputs to peripheral effector organs to tightly balance energy, glucose, and lipid metabolism (ref. 9 and Figure 1).

Homeostatic control centers in the hypothalamus and the brainstem are of particular importance for metabolic control.

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Both of these brain areas are in close proximity to circumventricular organs (e.g., the median eminence or area postrema) that contain "leaky," fenestrated capillaries to allow access of peripheral nutrients, metabolites, and hormones. The brainstem integrates short-term satiation signals from the GI tract either directly via the blood, or via input from vagal afferents that innervate the esophagus, stomach, and small intestine. The nerve endings respond to mechanical stimuli such as gastric dilatation, as well as to chemical satiety signals including the postprandially secreted GI hormones cholecystokinin (CCK) (10), glucagon-like peptide-1 (GLP-1) (11, 12), peptide YY (PYY) (13), and apolipoprotein A-IV (ApoAIV) (14). After binding to specific receptors on the vagal afferents, all of these signals converge in the nucleus of the solitary tract in the brainstem and are subsequently relayed to other brain areas to be finally incorporated into output signals to induce satiety.

The hypothalamus, particularly the arcuate nucleus (ARC), provides the pivotal sensing region for adiposity signals including leptin (15) and insulin (16), as well as for glucose. It also receives input from many other parts of the CNS, including the hindbrain. In the ARC, glucoregulatory and glucose-sensing neurons exist alongside two distinct and functionally antagonistic populations of neurons, each characterized by the expression of specific neuropeptides: the anorexigenic proopiomelanocortin-expressing (POMC-expressing) neurons, which are active during a positive energy balance, and orexigenic neurons, which coexpress agoutirelated peptide (AgRP) and neuropeptide Y (NPY) and are active during a negative energy balance (17, 18). Neurons within the ventromedial, dorsomedial, and lateral hypothalamus and the paraventricular nucleus play an equally important role in controlling energy and glucose homeostasis. Together, they form a hypothalamic network that integrates with multiple neurocircuits outside of the hypothalamus in order to govern food intake, energy expenditure, glucose metabolism, and insulin sensitivity (18).

Homeostatic signals can be overpowered by nonhomeostatic cues of high hedonic valence (19). For instance, food enriched with fat and sugar can serve as potent reward stimulus. Consequently, highly rewarding food can initiate eating even in the absence of an

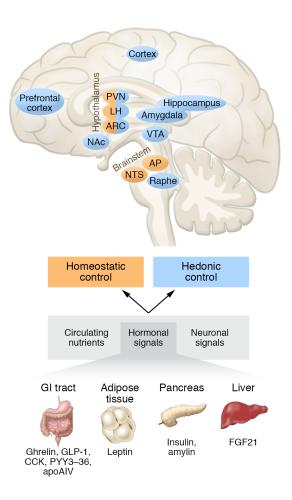


Figure 1. Homeostatic and hedonic control centers in the brain. Drugs targeting control of metabolism by the CNS act mainly via homeostatic and hedonic control centers that govern feeding behaviors, energy and glucose homeostasis, and body weight. The related brain areas are densely interconnected, and receive direct input from circulating nutrients such as glucose or fatty acids, peripheral neuronal networks, and hormonal satiation signals such as GLP-1 or amylin, or hormonal adiposity signals such as leptin. Within the homeostatic and hedonic control centers, the peripheral signals are integrated with sensory input, past experiences, and cues arising from the prevailing stress situation, emotional context, and mood. Ultimately, the signals converge in nuclei such as the hypothalamic paraventricular nucleus and lateral hypothalamus, and induce both adaptations to our ingestive behavior and brain stem-mediated changes to peripheral organ functions and our control of energy and glucose metabolism. AP, area postrema; ARC, arcuate nucleus; FGF21, fibroblast growth factor 21; GI tract, gastrointestinal tract; GIP, glucose-dependent insulinotropic polypeptide; GLP-1, glucagon-like peptide-1; LH, lateral hypothalamus; NAc, nucleus accumbens; NTS, nucleus of the solitary tract; PVN, paraventricular nucleus; PYY3-36, peptide YY 3-36; VTA, ventral tegmental area.

energetic requirement. Several brain regions and neurotransmitter systems, including dopamine, serotonin, endocannabinoids, and opioids, are involved in the rewarding effect of food (20–23). Also, homeostatic signals such as leptin (24), insulin (25), and ghrelin (26) affect the brain reward system.

Reward in the context of ingestive behavior is built upon two separable functional components: first, the hedonic "liking" of food, which is related to pleasure and palatability and primarily involves the opioid and cannabinoid systems in the nucleus accumbens, ventral pallidum, parabrachial nucleus, and nucleus of the solitary tract; and second, the "wanting" of food, which is related to appetite and the incentive motivation to eat and which is mainly related to the mesolimbic dopaminergic system with its projections from the ventral tegmental area to the nucleus accumbens and neural circuits involving the prefrontal cortex, amygdala, and hypothalamus (27–29).

Small-molecule CNS stimulants have been shown to tackle both components of the food reward system to ultimately suppress appetite. They have thus long been recognized as potential antiobesity drugs, and were the first drugs in use, as outlined below.

Principles and strategies in targeting the CNS-regulated metabolism

In the 1920s, at a time before it was recognized that obesity accounts for a growing prevalence of harmful chronic diseases (30), attitudes concerning body weight began to shift in favor of a slimmer and athletic appearance. The perceptual change boosted

the search for pharmacological strategies to facilitate weight loss. The first weight-lowering drugs were identified at a time when the mechanisms for food intake and weight control were largely unknown. Today we know that these appetite suppressants were mainly targeting monoamine neurotransmitter systems, which comprise a network of neurons within homeostatic and hedonic circuits of the brain that use monoamine neurotransmitters including the catecholamines dopamine and norepinephrine and the indolamine serotonin.

Amphetamines, the first monoamine-targeting weight loss drugs. The first monoamine neurotransmitter-based weight loss drug was introduced in the 1930s, when Smith, Kline & French Laboratories synthesized and commercialized the two optical enantiomers of amphetamine: dextroamphetamine and levoamphetamine. Benzedrine contained the racemic mixture of both isomers, while Dexedrine only included the more potent dextroamphetamine. Originally advertised as a treatment for narcolepsy or postencephalitic parkinsonism, the cognitive-enhancing properties of amphetamine were quickly recognized (31). The observation of its potent appetite-suppressing side effect caused an erratic increase in the use of amphetamines as weight loss therapy (31, 32). Therapies arose that combined amphetamine with barbiturates to counter adverse side effects, such as insomnia, restlessness, and increased blood pressure (Dexamyl). Clarkotabs added thermogenic thyroid hormone to enhance weight loss, along with phenobarbital, aloin, and atropine sulfate to reduce undesirable adverse effects. Furthermore, N-methyl-substituted amphet-

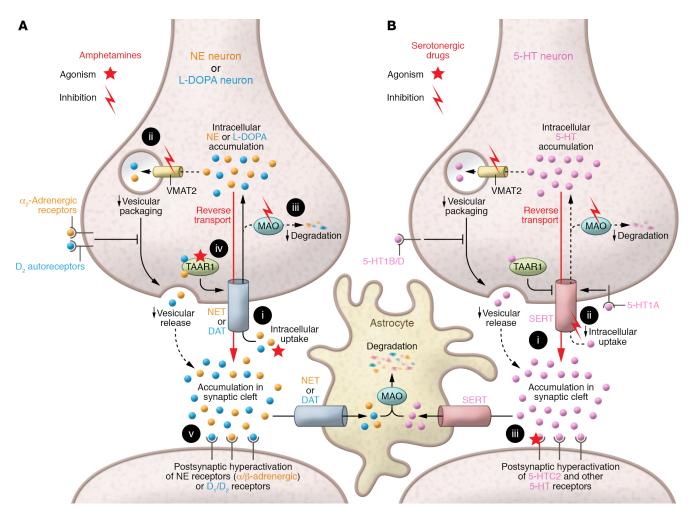


Figure 2. Central monoaminergic drug action. Pharmacological effects of amphetamines and their congeners are primarily mediated by increased synaptic release of monoamine neurotransmitters norepinephrine (NE), dopamine (DA), and, to a lesser extent, serotonin (5-HT). (A) (i) Amphetamines are competitive agonists for NET and DAT. (ii) Upon entering the presynaptic neuron, amphetamines bind to VMAT2, thereby inhibiting monoamine translocation from the cytosolic pool into storage vesicles. (iii) Amphetamines also weakly inhibit monoamine oxidase-mediated (MAO-mediated) monoamine breakdown, resulting in intracellular increase of monoamines. (iv) Amphetamines can further activate the intracellular trace amine-associated receptor 1 (TAAR1) to promote DA efflux. All processes contribute to reverse transport via NET or DAT, enhancing extracellular monoamine release. (v) Elevated monoamine release induces satiety and decreases feeding by activating postsynaptic α - and β -adrenergic (NE) and D1/D2 (DA) receptors. Increased DA signaling within the mesocorticolimbic system contributes to the addictive properties of amphetamines and their congeners. (B) Selective serotonergic drugs act either as (i) serotonin-releasing agents (SRAs), (ii) selective serotonin reuptake inhibitors (SSRIs), or (iii) selective 5-HT2C receptor agonists. SRAs (e.g., fenfluramine) increase synaptic 5-HT release, augmenting serotonergic function. Although SRAs' precise mechanisms remain unclear, they may be comparable to NE and DA releasers, i.e. reversing SERT- or VMAT2-mediated 5-HT transport. SSRIs (e.g., sibutramine) selectively bind SERT to inhibit 5-HT re-uptake. Postsynaptic 5-HT2C receptors appear to mediate the main effects of 5-HT on food intake and are the target of selective 5-HT2C receptor agonists such as lorcaserin. Presynaptic autoreceptor 5-HT1A and postsynaptic 5-HT1B, 5-HT2B, and 5-HT6 receptors may also contribute to the regulation of food intake by 5-HT. Monoaminergic drugs act at pre- and postsynaptic neurons, and they also interact with monoaminergic signaling on astrocytes. Astrocytic expression of NET, DAT, SERT, and metabolizing enzymes such as MAO can regulate monoamine levels in the synaptic cleft, neurotransmitter release from astrocytes and its transport into presynaptic neurons, and postsynaptic neuron activity.

amine (methamphetamine) derivatives, including Desoxyn and Methedrine, were hailed as weight loss drugs (33).

The weight-lowering effect of amphetamine was mainly assigned to a decrease in food intake. When humans were given amphetamine or placebo and required to maintain constant eating, the weight-lowering effect was eradicated (34). Later studies in rodents demonstrated that intraperitoneally injected amphetamine is less effective in suppressing appetite in rats with lateral hypothalamic lesions (35). Moreover, direct hypothalamic injections of amphetamine decreased food intake, and amphetamine

action on the lateral hypothalamus was inhibited by local administration of dopaminergic and β -adrenergic antagonists, and by inhibitors of catecholamine synthesis (36). Amphetamine-induced anorexia was linked to a decreased hypothalamic expression of orexigenic NPY (37, 38). Amphetamine therapy was further shown to increase the expression of cocaine- and amphetamine-regulated transcript (CART) (39), a neuropeptide secreted by anorexigenic POMC neurons that decreases food intake (40).

Over time, the widespread consumption of amphetamines displayed a dark side. Multiple users experienced addictive behav-

Table 1. Withdrawn	monoaminergic	antichesity drugs
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Drug	Mode of action	First approval	Withdrawn from market	Reason for suspension
Phenylproanolamine (Accutrim and generic)	Nonselective adrenergic receptor agonist and norepinephrine reuptake inhibitor (162, 163)	1910 as nasal anticongestive (US), 1976 as weight- lowering drug (US)	2000	Case reports of intracranial hemorrhage and stroke in young women due to an unresolved mechanism (164)
Phenmetrazine (Preludin)	Norepinephrine/dopamine-releasing agent (165)	1954 (EU)	1965	Psychoactive effects including euphoria, delusions, and paranoia (166)
Aminorex (Menocil)	Serotonin-releasing agent and uptake inhibitor (167)	1965 (EU)	1968	Pulmonary hypertension and related death cases (168)
Fenfluramine (Pondimin)	Serotonin-releasing agent by binding to the serotonin transporter (165, 169)	1964 (France), 1973 (US)	1997	Valvular heart disease and pulmonary hypertension (52), likely as a result of 5-HT ₂₈ receptor activation, expressed on cardiac valvular interstitial cells (170)
Dexfenfluramine (Redux)	Serotonin-releasing agent and reuptake inhibitor (169)	1996	1997	Valvular heart disease and pulmonary hypertension (52)
Phentermine/Fenfluramine	Phentermine: releasing agent of norepinephrine and dopamine (171); fenfluramine: see above	1953 (phentermine), 1973 (fenfluramine), off-label combination	1997	See fenfluramine above
Sibutramine (Meridia)	Combined norepinephrine and serotonin reuptake inhibitor (172)	1997	2010	Excess of nonfatal cardiovascular events in the SCOUT trial (53)

iors that went beyond a mere habituation to the effects of amphetamines. This addictive behavior was later assigned to the competitive binding of amphetamine to the norepinephrine transporter (NET) and the dopamine transporter (DAT) (41), which inhibited the reuptake of endogenous norepinephrine and dopamine into the presynaptic neurons. Amphetamines were further shown to promote the reverse transport (efflux) of both monoamines, and to slow catecholamine catabolism by inhibiting monoamine oxidase (ref. 42 and Figure 2). In consequence, amphetamines induced an amplification of the mesolimbic dopaminergic signal transmission in the striatum that profoundly escalated their rewarding and addicting properties (43).

Past failures and evolution of monoaminergic drugs. The abusive potential of amphetamines prompted the pharmaceutical industry to develop structural derivatives with the goal of decreasing the dopaminergic effect and the risk of habituation (31). Several amphetamine congeners were developed and put into clinical use, some of them with catastrophic results. Aminorex, phenylpropanolamine, and phenmetrazine have been withdrawn from the market because of severe adverse effects (Table 1). At present, four amphetamine congeners - phendimetrazine, diethylpropion, phentermine, and benzphetamine — are approved for the treatment of obesity (Table 2). However, the safety concerns regarding their addictive potential were never fully reconciled. In 1977, all approved amphetamine-derived drugs were restricted to shortterm use and were categorized as controlled substances by the US Drug Enforcement Administration (DEA), indicating their respective likelihood for physical addiction and mental dependence. In their capacity as CNS stimulants, their typical unwanted effects include, besides insomnia and nervousness, an increased heart rate. This particular side effect renders them counterindicated for patients with existing cardiovascular problems, including uncontrolled hypertension. Nevertheless, amphetamine congeners, and phentermine in particular, rank as some of the most prescribed antiobesity medications in the United States, either as monotherapy or as combination treatment with the anticonvulsant topiramate (Table 2). In 2017, bupropion, which chemically resembles the amphetamine derivative diethylpropion, was approved for weight loss in combination with the μ/κ -opioid receptor antagonist naltrexone (ref. 44, Table 2, and Figure 3). Phase III clinical trials are currently investigating the weight-lowering effects of bupropion in combination with the anticonvulsant zonisamide (ref. 45 and Table 2).

To overcome some of the challenges associated with amphetamines mainly acting on dopaminergic and noradrenergic circuitry, novel classes of monoaminergic drugs were developed with a preference for targeting the serotonin system. Serotonin (5-HT) acts as a hormone and a neurotransmitter that regulates a variety of physiological processes in the CNS and in peripheral organs. Serotonin cannot cross the blood-brain barrier, which explains why the peripheral and the central serotonergic systems are functionally separated. In the CNS, serotonin is synthesized and released by serotonergic neurons, which are organized into nine nuclei (B1-B9) and located in the midbrain and hindbrain areas. The most substantial portion of total brain serotonin is synthesized in the dorsal raphe (B7) of the brain stem, which has projections to hypothalamic nuclei and other feeding-related forebrain areas (46). Serotonin acts as a key anorexigenic signal mainly via two distinct types of serotonin receptor-expressing neurons. First, the activation of serotonin 2C receptors (5-HT₂₀Rs) on POMC neurons (47, 48) leads to an increased release of α -melanocyte-stimulating hormone (α-MSH) and subsequent stimulation of the melanocortin-3 and -4 receptor system (MC3/4Rs) (49). Second, the stimulation of 5-HT, Rs on NPY and AgRP or exigenic neurons blocks the release of NPY and AgRP and abolishes the inhibitory effect of GABA on POMC neurons (50). In addition, 5-HT R antagonists can potently reduce food intake and body weight gain in rodents, but the underlying mechanisms remain to be determined (51). Overall, the sero-

Table 2. Monoaminergic antiobesity drugs				
Drug, first approval, DEA schedule	CNS-based mode of action	Dosage	Placebo-subtracted weight loss (PSWL)	Other effects
Phendimetrazine (Bontril and generics), 1959, III	Norepinephrine- and dopamine-releasing agent. After oral administration, ~30% of phendimetrazine is converted to the active metabolite phenmetrazine; based on its prodrug character and its slower release, phendimetrazine has a milder onset and likely less abusive potential than the N-demethylated metabolite phenmetrazine (165, 173)	p.o. 35 mg (2–3 times daily) or 105 mg (once daily); short- term treatment	7% weight loss relative to the baseline body weight after up to 32 wk of treatment (174)	CNS stimulant, which raises the heart rate and blood pressure; not recommended for MetS patients with a history of heart disease, pulmonary hypertension, or high blood pressure
Diethylpropion (Tenuate and generics), 1959, IV	Norepinephrine- and dopamine-releasing agent. The keto substitution at the β-carbon of the phenylamine backbone leads to a strong reduction of its dopaminergic action (165); an additional peripheral thermogenic effect that may contribute to weight loss was demonstrated in rats (175)	p.o. 25 mg (thrice daily) or 75 mg sustained release (once daily); short- term treatment	3.0 kg PSWL in studies ranging from 6 to 52 wk (176)	CNS stimulant, which raises the heart rate and blood pressure; not recommended for MetS patients with a history of heart disease, pulmonary hypertension, or high blood pressure
Phentermine hydrochloride (Adipex-P and generic), 1959, IV	Norepinephrine- and dopamine-releasing agent. As α -methylated amphetamine, it potently stimulates norepinephrine release (IC ₅₀ = 39.4 nM), with less effect on the release of dopamine (IC ₅₀ = 262 nM) (177)	p.o. (1–3 times daily), 15–37.5 mg; short- term treatment	Average PSWL of 3.5 kg in a 28-wk study (178) and in studies ranging from 2 to 24 wk (176)	CNS stimulant, which raises the heart rate and blood pressure; not recommended for MetS patients with a history of heart disease, pulmonary hypertension, or high blood pressure
Benzphetamine (Regimex , Didrex) , 1960 , III	Norepinephrine- and dopamine-releasing agent	p.o. (twice daily), 25–50 mg; short- term treatment	PSWL of 3.3 kg in studies with an average duration of 8.9 wk (176)	CNS stimulant, which raises the heart rate and blood pressure; not recommended for MetS patients with a history of heart disease, pulmonary hypertension, or high blood pressure
Phentermine/ topiramate ER (Qsymia), 2012, IV	Synergistic action of the norepinephrine- and dopamine-releasing agent phentermine (see above) and the anticonvulsant topiramate. The weight-lowering and insulin-sensitizing mechanism of topiramate is uncertain; it may promote satiety and appetite suppression due to effects on neurotransmitters, neurotransmission, or inhibition of carbonic anhydrase (179, 180)	p.o. (once daily), 3.75 mg/23 mg, 7.5 mg/46 mg, 15 mg/92 mg; escalating dose regimen depending on individual response	EQUIP (52 wk, BMI ≥ 35): mean PSWL: 4.1 kg at 3.75 mg/23 mg, 10.7 kg at 15 mg/92 mg (181); CONQUER (56 wk, BMI ≥ 27 ± clinical comorbidity): mean PSWL: 6.7 kg at 7.5 mg/46 mg, 8.8 kg at 15 mg/92 mg (182)	Significant improvements in cardiometabolic risk factors and glycemic control (183)
Lorcaserin (Belviq), 2012, IV	Selective 5-HT _{xc} agonist that is thought to decrease food intake through the hypothalamic POMC-melanocortin axis without additional effects on energy expenditure (55)	p.o. (twice daily), 10 mg	BLOOM (52 wk, BMI ≥ 30 or BMI ≥ 27 ± clinical comorbidity); mean PSWL: 3.6 kg (184)	Improvement in multiple cardiovascular risk factors, including lipids, blood pressure, blood glucose, and renal function; decreases risk for incident diabetes, induces remission of hyperglycemia, and reduces the risk of microvascular complications (56)
Naltrexone/bupropion (Contrave), 2017, not scheduled	Synergistic action of the dopamine and norepinephrine reuptake inhibitor (bupropion) and the μ/κ -opioid receptor antagonist (naltrexone). By blocking the opioid receptor, naltrexone prevents autoinhibition of β -endorphin on POMC neuron activity	p.o. (twice daily), 8.0 mg/90 mg; long-term treatment	COR-I (52 wk, BMI ≥ 30 or BMI ≥ 27 ± clinical comorbidity); mean PSWL: 3.5 kg at 16 mg/360 mg ER, 4.7 kg at 32 mg/360 mg ER (44)	Weight loss associated with improvements in glycemic control and select cardiovascular risk factors in T2D patients (185)
Tesofensine (NS2330), in phase III trials, not scheduled	Serotonin-noradrenaline-dopamine reuptake inhibitor (186)	To be determined	Phase II (24 wk, BMI ≥ 30); mean PSWL: 4.5 kg at 0.25 mg, 9.1 kg at 0.5 mg, 10.6 kg at 1.0 mg (187)	Improvements in serum insulin, HbA1c, triglycerides, and cholesterol, but also an increase in blood pressure at the highest dose (187)
Bupropion/zonisamide (Empatic) ER, in phase III trials, not scheduled	Synergistic action of the norepinephrine- and dopamine-releasing agent bupropion with the anticonvulsant zonisamine. The weight-lowering effect of zonisamine is unknown and may include glutaminergic and GABAergic neurotransmission	To be determined	Phase II (24 wk, BMI \geq 30 or BMI \geq 27 \pm clinical comorbidity); patients treated with a 360 mg/360 mg combination lost 9.9% of their baseline body weight (45)	Improvements in cardiometabolic risk factors such as serum triglycerides, fasting insulin, and blood pressure

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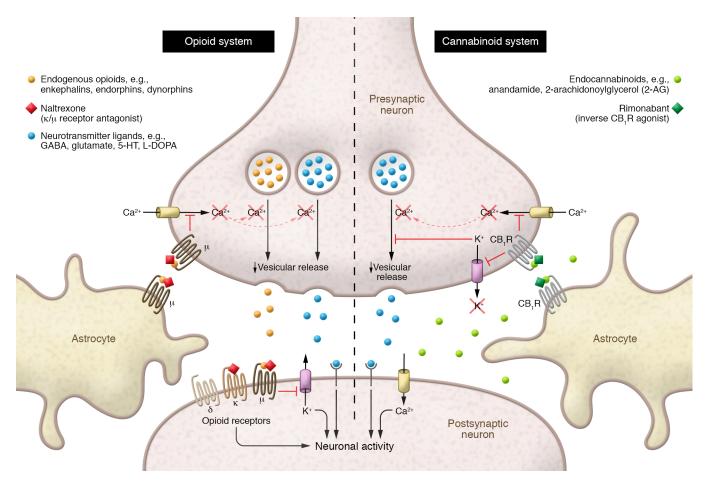


Figure 3. Drugs targeting the opioid and cannabinoid system. Multiple homeostatic and hedonic control centers of food intake express δ -, κ -, and/or μ -opioid receptors as well as cannabinoid receptor type 1. Endogenous opioids such as enkephalins, endorphins, or dynorphins are important in our response to and moderation of pain and pleasure, and influence both homeostatic and hedonic aspects of eating behavior. Similar actions on food intake are reported for endocannabinoids such as anandamide or 2-arachidonoylglcerol. Accordingly, both systems have been at the focus of the development of antiobesity drugs based on receptor antagonists. To date, only the μ/κ -opioid receptor antagonist naltrexone and the type 1 cannabinoid receptor (CB,R) antagonist rimonabant have gained market access as weight loss drugs, but psychiatric liabilities led to withdrawal of rimonabant. On presynaptic neurons, both drugs act via inhibition of presynaptic intracellular calcium influx and/or potassium efflux, which ultimately blocks calcium-dependent neurotransmitter vesicle release. Postsynaptically, the antagonist naltrexone inhibits μ - and to a lesser extent κ -opioid signaling to decrease neuronal activity. Rimonabant and naltrexone may further activate astrocyte cannabinoid and opioid signaling to modulate both presynaptic and postsynaptic neuronal processes.

tonin system continues to be a viable target for weight control and has led to the development of three classes of serotonergic drugs: serotonin-releasing agents, serotonin reuptake inhibitors, and selective 5-HT₂₀R agonists (Figure 2).

In the 1990s, fenfluramine, a first-generation serotonin-releasing agent, was combined with the sympathomimetic drug phentermine to create the weight-lowering drug Fen-Phen. This combination drug gained great popularity until an increasing number of valvular heart disease and pulmonary hypertension cases were associated with its use (52), ultimately causing the suspension of the combination drug as well as fenfluramine and its derivative dexfenfluramine (Table 1). Similarly, sibutramine, a selective reuptake inhibitor for serotonin and norepinephrine, was withdrawn owing to severe cardiovascular side effects (53) (Table 1). In 1995, the finding that 5-HT_{2c}Rs act as key regulators of satiety in rodents stirred the development of a third generation of serotonergic drugs (54). Lorcaserin is a selective 5-HT_{2c}R agonist that is thought to decrease food intake through the hypo-

thalamic POMC-melanocortin axis (55) in the absence of serotonergic adverse events (Table 2). Importantly, lorcaserin treatment results in the reduction of multiple cardiovascular risk factors and leads to an improved glycemic control, which renders it applicable for the treatment of the MetS (56). The combined serotonin-noradrenaline-dopamine reuptake inhibitor tesofensine (NS2330) is currently being investigated in phase II clinical trials and may have weight-lowering properties beyond those of existing monoaminergic weight loss medications (Table 2).

Drugs targeting the endocannabinoid system. In the late 1980s, the discovery of type 1 and type 2 cannabinoid receptors (CB₁R and CB₂R) and their endogenous ligands, the endocannabinoids, prompted the development of synthetic receptor agonists and antagonists in order to study the physiological function of the endocannabinoid system (ECS). Major attention has been paid to CB₁R, which is the more abundant CBR in the CNS, particularly the hippocampus, basal ganglia, and hypothalamus (57). CB₁R has also been identified in the GI tract, adipose tissue, skeletal mus-

cle, and cardiovascular system. One of the first described CB₁R inverse agonists (functional antagonist) was SR141716A (rimonabant) (ref. 58 and Figure 3). Chemically, rimonabant is a pyrazole and piperidine derivative, which upon daily i.p. (intraperitoneal) injection caused a profound reduction in body weight and food intake in lean rats (59). This finding was in line with the hypophagic and lean phenotype of mice lacking CB₁R (60). The weight-lowering effect of chronic rimonabant administration was further confirmed in diet-induced obese (DIO) mice (61) and in hyperphagic Lep^{ob} mice (62). Peripheral CB₁R antagonism was shown to contribute to the weight-lowering effect by enhancing lipolysis in adipocytes (63). The finding of reduced drug-seeking behavior in rimonabant-treated rats (64), and of an attenuated reward behavior in the CB₁R-KO mouse (65), provided strong evidence for the involvement of the ECS in motivation and hedonic behaviors.

Clinical trials confirmed the weight loss efficacy of rimonabant (20 mg) by showing a placebo-subtracted weight loss of 2.6 to 6.3 kg (66, 67). In addition, rimonabant caused a significant improvement in cardiovascular risk factors associated with the MetS (66, 67). In 2009, only three years after rimonabant was introduced to the European market, it was withdrawn based on novel data that linked it with depression and an increased risk for suicide (68). Accumulating evidence suggests that the mood-changing effects were caused by rimonabant's inverse agonism, which rendered CB₁R in the amygdala and the ventral tegmental area constitutively active (69).

Recently, neutral CB₁R antagonists were developed. They lack the inverse agonist properties of rimonabant and the mood-changing effects, but continue to reduce weight gain and food intake (69). Whether such neutral CB₁R antagonists can represent a novel and safer alternative for the treatment of the MetS remains to be determined. Currently, a novel neutral peripheral cannabinoid antagonist (AM6545) with limited CNS penetration is under investigation (70).

Weight loss drugs that mimic WAT adiposity signals. The increasing understanding of the physiology of food intake and energy balance, and the pathophysiology of its dysregulation, resulted in the development of drugs that interfere with neuropeptide hormone signaling pathways, such as leptin-melanocortin signaling. The adipokine leptin is secreted in direct proportion to fat mass. As an adiposity signal it targets hypothalamic leptin receptors (LepRs) and their downstream JAK2/STAT3, MAPK, and PI3K signaling to decrease food intake and increase energy expenditure in lean individuals. Its main action is driven by LepR-positive AgRP (71, 72) and POMC (73, 74) neurons in the ARC. These first-order neurons sense leptin levels and numerous other hormonal and nutritional cues, and orchestrate the activation of melanocortin-3 and -4 receptor-positive (MC3/4R-positive) neurons in the paraventricular nucleus via direct synaptic innervation or via the concomitant release of the neuropeptide MC3/4R agonist AgRP or the MC3/4R antagonist α -MSH, a cleavage product of POMC (75). The finetuning of melanocortin tone by competing neuropeptides ultimately governs ingestive behaviors and behaviors beyond feeding (76-78) as well as non-CNS processes such as thermogenesis and WAT browning (79) or bone metabolism (80).

Subjects with loss-of-function mutations in leptin, LepR, or downstream signaling components such as POMC or MC4R suffer from severe forms of morbid obesity and comorbid sequelae (81). Treatment with recombinant leptin can fully normalize body weight in leptin-deficient patients, but has no beneficial effects in patients with mutations in LepR or its downstream signaling. Currently, only one recombinant leptin analog, metreleptin (Myalepta), is approved for patients with leptin deficiency. The search for downstream mediators of leptin deficiency resulted in the discovery of the orexigenic hypothalamic peptide melanin-concentrating hormone (MCH) (82). Pharmacological blockade of MCH receptor 1 (MCHRI) emerged as promising drug target for the treatment of obesity. However, years of efforts failed to validate the MCHR1 antagonist concept in phase I clinical trials (83).

While monogenetic forms of obesity may often involve mutations in leptin melanocortin signaling, they remain rare and insignificant for the overall majority of obese individuals. These individuals have high leptin levels but exhibit leptin resistance, i.e., a relative inability of endogenous leptin or exogenous recombinant leptin to decrease food intake and body weight. Molecular underpinnings for the insensitivity toward leptin action are not entirely understood and need further investigation. Impaired leptin transport, LepR trafficking, and leptin feedback signaling have been discussed (84), but more recent reports found little evidence for perturbed transport or signaling (85) and suggest fully intact CNS leptin action even in a state of diet-induced obesity (86).

Although leptin resistance remains an enigma, recent results have nonetheless encouraged reconsideration of therapeutic antiobesity strategies built on leptin sensitization. Increasing evidence has demonstrated that leptin sensitivity can be restored by pharmacologically induced weight loss (87-90). Notably, calorie restriction alone was not sufficient to restore leptin sensitivity (89). Pramlintide (Symlin), a synthetic analog of pancreatic amylin, sensitizes mice to the effects of leptin (90). Currently, pramlintide is clinically approved as adjunct therapy to mealtime insulin for the control of blood sugar. The combination of pramlintide with metreleptin resulted in a mean weight loss of 12.7% (90), and future weight loss therapies based on amylinomimetics or combinatorial therapies (e.g., with leptin) appear plausible. In addition, inhibition of the protein tyrosine phosphatase PTP1B, a negative regulator of the leptin and insulin signaling pathway, by trodusquemine (MSI-1436) and related analogs was shown to elicit weight loss and leptin resensitization (91, 92).

Screenings for novel leptin-sensitizing molecules using the bioinformatical Connectivity Map (CMAP) tool led to the identification of the plant constituents celastrol and withaferin A, which increase leptin sensitivity and reduce body weight of obese mice (93, 94). The leptin-sensitizing properties of celastrol were later confirmed (95) and attributed to the hypothalamic inhibition of the protein tyrosine phosphatases PTP1B and TCPTP (96) and to an upregulation of the hypothalamic interleukin-1 receptor 1 (IL1R1) (97).

Restoring leptin sensitivity constitutes a challenge in the field of obesity and offers the unprecedented opportunity to develop an efficient weight loss and weight maintenance therapy. However, clinical data on these novel small-molecule sensitizing drugs are not yet available. They may further be complemented by additional drugs that elicit weight-lowering actions via the leptin-melanocortin system. These drugs include a new generation of small-molecule MC4R agonists such as setmelanotide (RM-493), which has

Table 3. Incretin mimetics for the treatment of obesity and type 2 diabetes

Drug	First approval	Chemical specifications	Dosage	Major efficacy results in phase III clinical trials
Exenatide (Byetta)	2005	Synthetic analog of exendin-4 with 53% homology to native GLP-1; increased half-life of 2.4 hours due to resistance to DPP-4–mediated degradation and an enhanced stability of the secondary structure (188)	s.c. (twice daily), 5 μg or 10 μg	DURATION-1 (30-week trial in inadequately controlled T2D patients, 10 μg twice daily) (189); HbA1c reduction from baseline: –1.5%; patients achieving HbA1C > 7.0: 61%; mean weight loss from baseline: –3.6%
Lixisenatide (Adlyxin)	2013 (EU)	Synthetic analog of exendin-4 with a C-terminal deletion of a proline residue and the addition of 6 lysine residues, leading to a half-life of 3 hours (190)	s.c. (once daily), 10 μg or 20 μg	GetGoal-X (24-week trial in inadequately controlled T2D patients, 20 μg once daily) (191); HbA1c reduction from baseline: –0.8%; patients achieving HbA1C > 7.0: 48.5%; mean weight loss from baseline: –2.8%
Exenatide ER (Bydureon)	2012	Exenatide (formulation in microsphere permits a prolonged absorption of exenatide from the subcutaneous depot allowing once-weekly dosing) (192)	s.c. (once weekly), 2.0 mg	DURATION-1 (30-week trial in inadequately controlled T2D patients, 2 mg once weekly) (189); HbA1c reduction from baseline: -1.9%; patients achieving HbA1C > 7.0: 77%; weight loss from baseline: -3.7%
Albiglutide (Tanzeum)	2014–2018 (discontinued for economic reasons)	Genetic fusion of a DPP-4—resistant GLP-1 dimer to human albumin, leading to a reduced renal clearance and an increased half-life of 5—8 days (193)	s.c. (once weekly), 30 mg or 50 mg	HARMONY-7 (32-week trial in inadequately controlled T2D patients, 50 mg once weekly) (194); HbA1c reduction from baseline: –0.78%; mean weight loss from baseline: –0.64 kg
Dulaglutide (Trulicity)	2014	Fusion of a DPP-4–resistant GLP-1 dimer to a human IgG4-Fc heavy chain by a small peptide linker, leading to a reduced renal clearance and an increased half-life of 4 days (195)	s.c. (once weekly), 0.75 mg or 1.5 mg	AWARD-6 (26-week trial in inadequately controlled T2D patients, 1.5 mg once weekly) (196); HbA1c reduction from baseline: –1.42%; patients achieving HbA1C > 7.0: 68%; weight loss from baseline: –2.9 kg
Liraglutide (Victoza or Saxenda)	2009	GLP-1 analog with 97% sequence homology to human GLP-1 with only 2 amino acid changes and the addition of a palmitic acid through a γ-glutamyl spacer; this lipid anchor causes strong albumin binding leading to a reduced renal clearance and a prolongation of the half-life to 13 hours (197)	s.c. (once daily), 0.6 mg, 1.2 mg, 1.8 mg, and 3.0 mg	LEAD-6 (26-week trial in inadequately controlled T2D patients, 1.8 mg once daily) (198); HbA1c reduction from baseline: -1.12%; patients achieving HbA1C > 7.0: 54%; weight loss from baseline: -3.24 kg
Semaglutide (Ozempic)	2016	GLP-1 analog with 94% sequence homology to human GLP-1; it resembles liraglutide with an additional Aib8 to prevent DPP-4—mediated cleavage and a C18-based fatty acid chain connected to Lys26 through a miniPEG space leading to a half-life of 160 hours (199)	s.c. (once weekly); escalating dose up to 1.0 mg	SUSTAIN-7 (40-week trial in inadequately controlled T2D patients, 1 mg once weekly) (200); HbA1c reduction from baseline: –1.8%; patients achieving HbA1C > 7.0: 79%; weight loss from baseline: –6.5 kg

recently been successfully used to treat patients with LepR deficiency (98) or with mutations in POMC (98, 99). Earlier small-molecule MC4R agonists had shown limited weight-lowering efficacy and/or severe cardiovascular liabilities, i.e., increases in blood pressure or heart rate (100, 101). Nonetheless, efforts continue to search for safe yet efficacious MC4R agonists, but their full potential as antiobesity drugs in obese patients remains underexplored.

Weight loss drugs that mimic GI satiety signals. Bariatric surgery is an effective albeit highly invasive option for obese subjects to achieve and sustain long-term weight loss and reductions in all MetS-related symptoms. The finding that bariatric surgery leads to profound changes in the secretion of gut hormones that have effects on food intake and glycemic control provided guidance to the search for new drugs that harness the CNS response to multiple satiety signals from the GI tract.

CCK mainly targets type 1 CCK receptors (CCK1Rs) on vagal afferent neurons to regulate satiety by terminating meals (102). Accordingly, CCK1R agonists were considered as promising antiobesity drugs. However, to date, their therapeutic utility has been limited by compensatory increases in meal frequency (103), by the development of drug tolerance in response to prolonged drug application (104), and by limited weight loss efficacy in phase II

clinical trials (105). Additional efforts have been directed toward exploring antiobesity effects of gut-derived PYY3-36. However, discrepant results in rodents (106, 107) and high levels of nausea in humans (108) impeded further clinical developments. PYY3-36 has high affinity for the NPY receptor Y2, which is one of several NPY receptors that play important roles in the regulation of food intake. Major ongoing efforts have been directed toward finding centrally acting agonists or antagonists against Y1, Y2, Y4, or Y5 receptors, but progress to date has been limited (109).

Extensive efforts were directed toward the generation of drugs that mimic the actions of the incretin GLP-1 (Table 3). In the periphery, GLP-1 receptor (GLP-1R) agonists exhibit properties ranging from glucose-dependent stimulation of insulin secretion (110, 111), suppression of glucagon secretion (112), and preservation of β cell mass (113), to the reduction of hepatic glucose output (114), which together leads to improvements in glycemic control. GLP-1R signaling in the brain is crucially involved in the anorectic and weight-lowering effects of GLP-1 (115), which are in part mediated via direct activation of hypothalamic POMC/ CART neurons in the ARC (116) or GLP-1R-positive neurons in the nucleus of the solitary tract of the hindbrain (117). Moreover, there is evidence that the inhibitory effect of GLP-1R agonists on food

intake goes beyond satiation and includes effects on food reward and motivation (118).

Native GLP-1 has a half-life of 2-3 minutes due to rapid degradation by dipeptidyl peptidase-4 (DPP-4), and several GLP-1R agonists have been developed to provide prolonged bioavailability. Depending on their half-life, they can be categorized either as short- or long-acting compounds (Table 3). The short-acting compounds include a synthetic version of exendin-4, exenatide (Byetta), and lixisenatide (Adlyxin). The long-acting compounds include albiglutide (Tanzeum), dulaglutide (Trulicity), exenatide long-acting release (Bydureon), liraglutide (1.8 mg Victoza or 3.0 mg Saxenda), and semaglutide (Ozempic). Differences in the bioavailability of these compounds lead to important differences in their biological actions. Short-acting GLP-1R agonists are applied before a meal and cause a profound deceleration of gastric emptying and a reduction in postprandial glycemia (119, 120). In contrast, long-acting GLP-1R agonists exert stronger effects on fasting glucose levels by causing prolonged stimulation of insulin secretion, but the effects on gastric emptying are subject to rapid tachyphylaxis (121). Consequently, short-acting GLP-1R agonists could be more suitable for the treatment of patients suffering primarily from postprandial hyperglycemia, whereas long-acting GLP-1R agonists would be more suitable for patients with predominant fasting hyperglycemia (122).

Head-to-head comparisons of incretin mimetics so far rendered liraglutide as the most effective antiglycemic GLP-1R agonist (123). The weight-lowering effect of GLP-1R agonists are dose-dependent and are most pronounced for high-dose liraglutide (3 mg) or semaglutide treatment. The latter caused a placebosubtracted body weight loss of up to 16% in obese patients after 52 weeks of treatment (124), which for the first time comes close to the weight loss achieved by bariatric surgery. Remarkably, an alternative formulation of semaglutide is currently being evaluated as a precedent-setting peptide-based antiobesity/antidiabetes drug that is given by oral administration (125).

The most common adverse effects seen with all GLP-1 therapies include nausea, vomiting, and injection-site reactions. Importantly, GLP-1R agonists do not seem to negatively affect cardiovascular risk in type 2 diabetes (T2D) patients. Novel findings even suggest a cardioprotective action of GLP-1R agonists (126, 127), which may render them as the treatment of choice for MetS patients with cardiovascular symptoms.

A new generation of combinatorial peptide drugs. Structural similarity between GLP-1, glucagon, and the incretin glucose-dependent insulinotropic polypeptide (GIP) and their low-potency cross-reactivity at their respective receptors facilitated integration of each activity into sequence-intermixed unimolecular hybrids. GLP-1 has now been successfully combined with glucagon (128, 129) or GIP into unimolecular dual or tri-agonists (130, 131) in order to achieve synergistic reductions of adiposity and hyperglycemia.

The first GLP-1-based multi-agonist was GLP-1 combined with glucagon action. Apart from its hyperglycemic effect, glucagon is a potent anorectic hormone. It mediates its weight-lowering effect mainly by acting on the CNS as a satiety signal to reduce food intake and by increasing energy expenditure and thermogenesis (132). Accordingly, it was hypothesized that glucagon would increase the weight-lowering effect of GLP-1, while the insulino-

tropic actions of GLP-1 would counter the hyperglycemic liability of glucagon. A large variety of GLP-1/glucagon receptor coagonists have been developed and advanced to clinical evaluation (133). Two of them, SAR425899 and MEDIO382, were recently shown to induce clinically meaningful reductions in blood glucose and body weight in obese T2D patients (134, 135).

Like GLP-1, the incretin GIP is secreted from the gut in response to nutrient ingestion and promotes insulin secretion in a glucose-dependent manner. While insulinotropic effects of GIP are well defined, controversy exists regarding its weight-lowering potential. Surprisingly, the pharmacological targeting of the GIP receptor (GIPR) by agonists (130, 136-138) as well as by antagonists (139, 140) led to body weight loss in obese rodents. Notably, a recent study aimed at disentangling these contradictory observations by comparing the in vivo potency of several structurally diverse GIPR agonists with a potent long-acting antagonist (138). This study confirmed weight loss in DIO mice only for selective GIPR agonists, but not for the GIPR antagonist. A combination of GLP-1R and GIPR agonism may thus have superior effects on glucose tolerance and body weight loss. Indeed, several studies on GLP-1R/GIPR dual agonists favor beneficial effects of GIP activation in glycemic control in preclinical (130) and clinical trials (141, 142). Tirzepatide (LY3298176), a once-weekly GLP-1/GIP coagonist, was recently shown to be superior to the GLP-1R agonist dulaglutide in terms of body weight loss and improved glycated hemoglobin (HbA1c) in obese human subjects with T2D (142). Whether GIP-based coagonists can provide greater maximal clinical efficacy and fewer side effects compared with the current best-in-class GLP-1R mono-agonist, semaglutide, will require the development of additional coagonist variants and a thorough clinical evaluation.

Based on the promising clinical trials using GLP-1/GIP and GLP-1/glucagon dual agonists, it was predicted that tri-agonist molecules with agonism at all three receptors would provide superior metabolic improvements. Indeed, in DIO mice and obese monkeys, the reduction of body weight by a GLP-1/GIP/glucagon tri-agonist was greater than that by the same dose of a GLP-1/GIP dual agonist (131). The potential benefits of GLP-1/GIP/glucagon tri-agonism for the management of obese individuals with T2D are currently being investigated in clinical trials (133).

The above-described hybrid GLP-1-based multi-agonists are limited to structurally similar molecules. In addition to this approach, fusion peptides have been generated in which structurally diverse hormones or oligonucleotides can be connected to GLP-1 via a chemical linker. GLP-1 fusion molecules with other peptide hormones including gastrin, amylin, and CCK have been generated and shown to achieve enhanced metabolic efficacy (143-145). Finally, there are recently reported successes in developing hybrid drugs that use GLP-1 as a hormonally active peptide for the cell typespecific delivery of chemically conjugated nuclear receptor agonists (146, 147) and antisense oligonucleotides (148). For instance, GLP-1R targeting has been leveraged to deliver estrogen to metabolically relevant tissues, where it enhanced the body weight-lowering, insulinotropic, and islet-preserving effects of estrogen through complementary pharmacology. Importantly, endocrine toxicities in non-GLP-1R-expressing organs were absent, which highlights the cell type-specific delivery (146, 149). In preclinical mouse models, the combination of GLP-1 with the glucocorticoid receptor agonist dexamethasone synergistically drove weight loss, likely mediated by a concomitant decrease in hypothalamic inflammation and GLP-1R-dependent activation of anorexigenic neurons (147). Currently, hybrid drugs are still in preclinical testing, and their clinical safety and efficacy remain to be determined.

Outlook and perspective

Weight reduction plays a fundamental role in managing the MetS. With emerging knowledge about neuronal pathways and peripheral feedback mechanisms controlling hunger and appetite, CNS-targeted weight loss pharmacology continues to evolve toward safer and more efficacious strategies. Currently, targeting strategies are mostly directed toward neuronal networks involved in the regulation of systemic metabolism. Built on the recent observation that systemic metabolism is also functionally controlled by non-neuronal cells in the CNS, including astrocytes, microglia, and tanycytes (150), future targeting strategies may require a wider focus and extraordinary approaches. However, at present it remains largely elusive whether and how disrupted non-neuronal glial networks are functionally involved in the development of the MetS.

Novel therapies may be built on the hormonal signals and CNS pathways discussed above, but they may also use entirely different concepts and strategies. For instance, the past decades saw the discovery of multiple new, hitherto unknown peripheral factors such as meteorin (151), meteorin-like (152), adipsin (153), irisin (154), or GDF15 (155), which have all been linked to energy and glucose homeostasis. These novel factors may hold great promise as backbones for future therapies against the MetS. GDF15 appears to be at center stage in this competitive search for new antiobesity drugs, and has recently been reported as a potent anorexigen that exerts its weight-lowering action via the receptor GDNF family receptor α-like (GFRAL) (156-158). The large family of fibroblast growth factors (FGFs) has gained similar attention in the search for antiobesity and antidiabetes drugs. Secreted by multiple tissues, FGF21 has been shown to exert weight loss and other multisystemic metabolic benefits in rodent models, and several FGF21 mimetics and receptor antagonists have hence entered the clinical testing phase (159). A single dose of FGF1 injected into the hypothalamus was further shown to induce a sustained and full remission of diabetic hyperglycemia in rodents (160, 161), which highlights the potential of FGF-based drugs in the fight against the MetS.

Overall, it is becoming increasingly clear that the complex and individual manifestation of the MetS requires pursuit of tailored therapies that ensure improved efficacy and safety in specific patient cohorts. Such novel therapies further require pioneering new pharmacological concepts and drugs that help close the current therapeutic gap and the relative lack of CNS-driven antiobesity drugs. Lastly, novel therapeutic concepts will greatly benefit from the increasing availability of large data sets and the development of advanced algorithms that facilitate an earlier and individualized patient diagnosis to enrich the prediction of individual risks for the development of comorbidities.

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- Alberti KG, Zimmet P, Shaw J, IDF Epidemiology Task Force Consensus Group. The metabolic syndrome — a new worldwide definition. *Lancet*. 2005;366(9491):1059–1062.
- Alberti KG, et al. Harmonizing the metabolic syndrome: a joint interim statement of the International Diabetes Federation Task Force on Epidemiology and Prevention; National Heart, Lung, and Blood Institute; American Heart Association; World Heart Federation; International Atherosclerosis Society; and International Association for the Study of Obesity. Circulation. 2009;120(16):1640-1645.
- Pothiwala P, Jain SK, Yaturu S. Metabolic syndrome and cancer. Metab Syndr Relat Disord. 2009;7(4):279–288.
- Vanhanen M, et al. Association of metabolic syndrome with Alzheimer disease: a populationbased study. Neurology. 2006;67(5):843–847.
- Magkos F, et al. Effects of moderate and subsequent progressive weight loss on metabolic function and adipose tissue biology in humans with

- obesity. Cell Metab. 2016;23(4):591-601.
- Look AHEAD Research Group. Eight-year weight losses with an intensive lifestyle intervention: the look AHEAD study. Obesity (Silver Spring). 2014;22(1):5–13.
- Gadde KM, Martin CK, Berthoud HR, Heymsfield SB. Obesity: pathophysiology and management. J Am Coll Cardiol. 2018;71(1):69–84.
- Crowley VE, Yeo GS, O'Rahilly S. Obesity therapy: altering the energy intake-andexpenditure balance sheet. *Nat Rev Drug Discov*. 2002;1(4):276–286.
- Woods SC, May-Zhang AA, Begg DP. How and why do gastrointestinal peptides influence food intake? *Physiol Behav.* 2018;193(pt B):218-222.
- Gibbs J, Young RC, Smith GP. Cholecystokinin decreases food intake in rats. J Comp Physiol Psychol. 1973;84(3):488-495.
- Turton MD, et al. A role for glucagon-like peptide-1 in the central regulation of feeding. *Nature*. 1996;379(6560):69-72.
- 12. Tang-Christensen M, et al. Central administra-

- tion of GLP-1-(7-36) amide inhibits food and water intake in rats. *Am J Physiol*. 1996;271(4 pt 2):R848-R856.
- Batterham RL, et al. Gut hormone PYY(3-36) physiologically inhibits food intake. *Nature*. 2002;418(6898):650-654.
- Tso P, Liu M, Kalogeris TJ, Thomson AB. The role of apolipoprotein A-IV in the regulation of food intake. *Annu Rev Nutr.* 2001;21:231–254.
- Zhang Y, Proenca R, Maffei M, Barone M, Leopold L, Friedman JM. Positional cloning of the mouse obese gene and its human homologue. *Nature*. 1994;372(6505):425-432.
- Woods SC, Lotter EC, McKay LD, Porte D. Chronic intracerebroventricular infusion of insulin reduces food intake and body weight of baboons. *Nature*. 1979;282(5738):503–505.
- 17. Horvath TL, Naftolin F, Kalra SP, Leranth C. Neuropeptide-Y innervation of beta-endorphincontaining cells in the rat mediobasal hypothalamus: a light and electron microscopic double immunostaining analysis. Endocrinology.

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- 1992;131(5):2461-2467.
- 18. Waterson MJ, Horvath TL. Neuronal regulation of energy homeostasis: beyond the hypothalamus and feeding. Cell Metab. 2015;22(6):962-970.
- 19. Berthoud HR, Münzberg H, Morrison CD. Blaming the brain for obesity: integration of hedonic and homeostatic mechanisms. Gastroenterology. 2017;152(7):1728-1738.
- 20. Berridge KC, Robinson TE. What is the role of dopamine in reward: hedonic impact, reward learning, or incentive salience? Brain Res Brain Res Rev. 1998;28(3):309-369.
- 21. Leibowitz SF, Alexander JT. Hypothalamic serotonin in control of eating behavior, meal size, and body weight. Biol Psychiatry. 1998;44(9):851-864.
- 22. Cota D, Tschöp MH, Horvath TL, Levine AS. Cannabinoids, opioids and eating behavior: the molecular face of hedonism? Brain Res Rev. 2006;51(1):85-107.
- 23. Nogueiras R, Romero-Picó A, Vazquez MJ, Novelle MG, López M, Diéguez C. The opioid system and food intake: homeostatic and hedonic mechanisms. Obes Facts. 2012;5(2):196-207.
- 24. Baicy K, et al. Leptin replacement alters brain response to food cues in genetically leptin-deficient adults. Proc Natl Acad Sci USA. 2007;104(46):18276-18279.
- 25. Figlewicz DP, Bennett JL, Naleid AM, Davis C, Grimm JW. Intraventricular insulin and leptin decrease sucrose self-administration in rats. Physiol Behav. 2006;89(4):611-616.
- 26. Malik S, McGlone F, Bedrossian D, Dagher A. Ghrelin modulates brain activity in areas that control appetitive behavior. Cell Metab. 2008;7(5):400-409.
- 27. Berridge KC. 'Liking' and 'wanting' food rewards: brain substrates and roles in eating disorders. Physiol Behav. 2009;97(5):537-550.
- 28. Cannon CM, Palmiter RD. Reward without dopamine. J Neurosci. 2003;23(34):10827-10831.
- 29. Peciña S, Cagniard B, Berridge KC, Aldridge JW, Zhuang X. Hyperdopaminergic mutant mice have higher "wanting" but not "liking" for sweet rewards. J Neurosci. 2003;23(28):9395-9402.
- 30. Van Itallie TB. Health implications of overweight and obesity in the United States. Ann Intern Med. 1985;103(6 pt 2):983-988.
- 31. Rasmussen N. America's first amphetamine epidemic 1929-1971: a quantitative and qualitative retrospective with implications for the present. Am J Public Health. 2008;98(6):974-985.
- 32. Nathanson MH. The central action of β-aminopropyl-benzene (Benzedrine). JAMA. 1937;108:528-531.
- 33. Nichols DE. Medical Chemistry and Structure-Activity Relationships. In: Cho AK, Segal DS, eds. Amphetamine and its Anlogs: Psychopharmacology, Toxicology, and Abuse. San Diego, California, USA: Academic Press: 1994:3-42.
- 34. Harris SC, Ivy AC, Searle LM. The mechanism of amphetamine-induced loss of weight; a consideration of the theory of hunger and appetite. J Am Med Assoc. 1947;134(17):1468-1475.
- 35. Carlisle HJ. Differential effects of amphetamine on food and water intake in rats with lateral hypothalamic lesions. J Comp Physiol Psychol. 1964:58:47-54.
- 36. Leibowitz SF. Catecholaminergic mechanisms

- of the lateral hypothalamus: their role in the mediation of amphetamine anorexia. Brain Res. 1975;98(3):529-545.
- 37. Kuo DY. Further evidence for the mediation of both subtypes of dopamine D1/D2 receptors and cerebral neuropeptide Y (NPY) in amphetamine-induced appetite suppression. Behav Brain Res. 2003;147(1-2):149-155.
- 38. Kotz CM, Briggs JE, Grace MK, Levine AS, Billington CJ. Divergence of the feeding and thermogenic pathways influenced by NPY in the hypothalamic PVN of the rat. Am J Physiol. 1998;275(2):R471-R477.
- 39. Douglass J, McKinzie AA, Couceyro P. PCR differential display identifies a rat brain mRNA that is transcriptionally regulated by cocaine and amphetamine. J Neurosci. 1995;15(3 pt 2):2471-2481.
- 40. Kristensen P, et al. Hypothalamic CART is a new anorectic peptide regulated by leptin. Nature. 1998;393(6680):72-76.
- 41. Burn JH, Rand MJ. The action of sympathomimetic amines in animals treated with reserpine. J Physiol (Lond). 1958;144(2):314-336.
- 42. Miller HH, Shore PA, Clarke DE. In vivo monoamine oxidase inhibition by d-amphetamine. Biochem Pharmacol. 1980;29(10):1347-1354.
- 43. Bunney BS, Aghajanian GK. d-Amphetamineinduced depression of central dopamine neurons: evidence for mediation by both autoreceptors and a striato-nigral feedback pathway. Naunyn Schmiedebergs Arch Pharmacol. 1978;304(3):255-261.
- 44. Greenway FL, et al. Effect of naltrexone plus bupropion on weight loss in overweight and obese adults (COR-I): a multicentre, randomised, double-blind, placebo-controlled, phase 3 trial. Lancet. 2010;376(9741):595-605.
- 45. Valentino MA, Lin JE, Waldman SA. Central and peripheral molecular targets for antiobesity pharmacotherapy. Clin Pharmacol Ther. 2010;87(6):652-662.
- 46. Lam DD, Garfield AS, Marston OJ, Shaw J, Heisler LK. Brain serotonin system in the coordination of food intake and body weight. Pharmacol Biochem Behav. 2010;97(1):84-91.
- 47. Xu Y, et al. 5-HT2CRs expressed by pro-opiomelanocortin neurons regulate energy homeostasis. Neuron. 2008;60(4):582-589.
- 48. Berglund ED, et al. Serotonin 2C receptors in pro-opiomelanocortin neurons regulate energy and glucose homeostasis. J Clin Invest. 2013;123(12):5061-5070.
- 49. Lam DD, et al. Serotonin 5-HT2C receptor agonist promotes hypophagia via downstream activation of melanocortin 4 receptors. Endocrinology. 2008;149(3):1323-1328.
- 50. Heisler LK, et al. Serotonin reciprocally regulates melanocortin neurons to modulate food intake. Neuron, 2006;51(2):239-249.
- 51. Halford JC, Harrold JA, Boyland EJ, Lawton CL, Blundell JE. Serotonergic drugs: effects on appetite expression and use for the treatment of obesity. Drugs. 2007;67(1):27-55.
- 52. Connolly HM, et al. Valvular heart disease associated with fenfluramine-phentermine. N Engl J Med. 1997;337(9):581-588.
- 53. James WP, et al. Effect of sibutramine on cardiovascular outcomes in overweight and obese subjects. N Engl J Med. 2010;363(10):905-917.

- 54. Tecott LH, et al. Eating disorder and epilepsy in mice lacking 5-HT2c serotonin receptors. Nature. 1995;374(6522):542-546.
- 55. Martin CK, et al. Lorcaserin, a 5-HT(2C) receptor agonist, reduces body weight by decreasing energy intake without influencing energy expenditure. J Clin Endocrinol Metab. 2011;96(3):837-845.
- 56. Bohula EA, et al. Effect of lorcaserin on prevention and remission of type 2 diabetes in overweight and obese patients (CAMELLIA-TIMI 61): a randomised, placebo-controlled trial. Lancet. 2018:392(10161):2269-2279.
- 57. Herkenham M, et al. Cannabinoid receptor localization in brain. Proc Natl Acad Sci USA. 1990;87(5):1932-1936.
- 58. Rinaldi-Carmona M, et al. SR141716A, a potent and selective antagonist of the brain cannabinoid receptor. FEBS Lett. 1994;350(2-3):240-244.
- 59. Colombo G, Agabio R, Diaz G, Lobina C, Reali R, Gessa GL. Appetite suppression and weight loss after the cannabinoid antagonist SR 141716. Life Sci. 1998;63(8):PL113-PL117.
- 60. Cota D, et al. The endogenous cannabinoid system affects energy balance via central orexigenic drive and peripheral lipogenesis. J Clin Invest. 2003;112(3):423-431.
- 61. Ravinet Trillou C, et al. Anti-obesity effect of SR141716, a CB1 receptor antagonist, in dietinduced obese mice. Am J Physiol Regul Integr Comp Physiol. 2003;284(2):R345-R353.
- 62. Di Marzo V, et al. Leptin-regulated endocannabinoids are involved in maintaining food intake. Nature. 2001;410(6830):822-825.
- 63. Jbilo O, et al. The CB1 receptor antagonist rimonabant reverses the diet-induced obesity phenotype through the regulation of lipolysis and energy balance. FASEB J. 2005;19(11):1567-1569.
- 64. De Vries TJ, Homberg JR, Binnekade R, Raasø H, Schoffelmeer ANM. Cannabinoid modulation of the reinforcing and motivational properties of heroin and heroin-associated cues in rats. Psychopharmacology (Berl). 2003;168(1-2):164-169.
- 65. Thanos PK, Dimitrakakis ES, Rice O, Gifford A, Volkow ND. Ethanol self-administration and ethanol conditioned place preference are reduced in mice lacking cannabinoid CB1 receptors. Behav Brain Res. 2005;164(2):206-213.
- 66. Pi-Sunyer FX, Aronne LJ, Heshmati HM, Devin J, Rosenstock J, RIO-North America Study Group. Effect of rimonabant, a cannabinoid-1 receptor blocker, on weight and cardiometabolic risk factors in overweight or obese patients: RIO-North America: a randomized controlled trial. JAMA. 2006:295(7):761-775.
- 67. Van Gaal LF, Rissanen AM, Scheen AJ, Ziegler O, Rössner S, RIO-Europe Study Group. Effects of the cannabinoid-1 receptor blocker rimonabant on weight reduction and cardiovascular risk factors in overweight patients: 1-year experience from the RIO-Europe study. Lancet. 2005;365(9468):1389-1397.
- 68. Christensen R, Kristensen PK, Bartels EM, Bliddal H, Astrup A. Efficacy and safety of the weight-loss drug rimonabant: a meta-analysis of randomised trials. Lancet. 2007;370(9600):1706-1713.
- 69. Meye FJ, Trezza V, Vanderschuren LJ, Ramakers GM, Adan RA. Neutral antagonism at the canna-

- binoid 1 receptor: a safer treatment for obesity. *Mol Psychiatry*. 2013;18(12):1294–1301.
- Randall PA, et al. The novel cannabinoid CB1 antagonist AM6545 suppresses food intake and food-reinforced behavior. *Pharmacol Biochem* Behav. 2010;97(1):179–184.
- Ebihara K, et al. Involvement of agouti-related protein, an endogenous antagonist of hypothalamic melanocortin receptor, in leptin action. *Diabetes*. 1999;48(10):2028–2033.
- Xu J, et al. Genetic identification of leptin neural circuits in energy and glucose homeostases. *Nature*. 2018;556(7702):505–509.
- Cheung CC, Clifton DK, Steiner RA. Proopiomelanocortin neurons are direct targets for leptin in the hypothalamus. *Endocrinology*. 1997;138(10):4489-4492.
- Schwartz MW, et al. Leptin increases hypothalamic pro-opiomelanocortin mRNA expression in the rostral arcuate nucleus. *Diabetes*. 1997;46(12):2119–2123.
- Kühnen P, Krude H, Biebermann H. Melanocortin-4 receptor signalling: importance for weight regulation and obesity treatment. *Trends Mol Med*. 2019;25(2):136–148.
- Dietrich MO, Zimmer MR, Bober J, Horvath TL. Hypothalamic Agrp neurons drive stereotypic behaviors beyond feeding. *Cell*. 2015;160(6):1222–1232.
- Dietrich MO, et al. AgRP neurons regulate development of dopamine neuronal plasticity and nonfood-associated behaviors. *Nat Neurosci*. 2012;15(8):1108-1110.
- Zimmer MR, Schmitz AE, Dietrich MO. Activation of Agrp neurons modulates memory-related cognitive processes in mice. *Pharmacol Res.* 2019;141:303–309.
- Ruan HB, et al. O-GlcNAc transferase enables AgRP neurons to suppress browning of white fat. Cell. 2014;159(2):306–317.
- 80. Kim JG, et al. AgRP neurons regulate bone mass. *Cell Rep.* 2015;13(1):8-14.
- Farooqi IS, O'Rahilly S. Mutations in ligands and receptors of the leptin-melanocortin pathway that lead to obesity. *Nat Clin Pract Endocrinol Metab*. 2008;4(10):569–577.
- Qu D, et al. A role for melanin-concentrating hormone in the central regulation of feeding behaviour. *Nature*. 1996;380(6571):243–247.
- Johansson A. Evolution of physicochemical properties of melanin concentrating hormone receptor 1 (MCHr1) antagonists. *Bioorg Med Chem Lett*. 2016;26(19):4559–4564.
- Quarta C, Sánchez-Garrido MA, Tschöp MH, Clemmensen C. Renaissance of leptin for obesity therapy. *Diabetologia*. 2016;59(5):920–927.
- Harrison L, et al. Fluorescent blood-brain barrier tracing shows intact leptin transport in obese mice. *Int J Obes (Lond)*. 2019;43(6):1305–1318.
- Ottaway N, et al. Diet-induced obese mice retain endogenous leptin action. *Cell Metab*. 2015;21(6):877-882.
- 87. Kim YW, et al. Metformin restores leptin sensitivity in high-fat-fed obese rats with leptin resistance. *Diabetes*. 2006;55(3):716-724.
- 88. Clemmensen C, et al. GLP-1/glucagon coagonism restores leptin responsiveness in obese mice chronically maintained on an obesogenic

- diet. Diabetes. 2014;63(4):1422-1427.
- 89. Müller TD, et al. Restoration of leptin responsiveness in diet-induced obese mice using an optimized leptin analog in combination with exendin-4 or FGF21. J Pept Sci. 2012;18(6):383–393.
- 90. Roth JD, et al. Leptin responsiveness restored by amylin agonism in diet-induced obesity: evidence from nonclinical and clinical studies. *Proc Natl Acad Sci USA*. 2008;105(20):7257–7262.
- Lantz KA, et al. Inhibition of PTP1B by trodusquemine (MSI-1436) causes fat-specific weight loss in diet-induced obese mice. *Obesity* (Silver Spring). 2010;18(8):1516–1523.
- Krishnan N, Konidaris KF, Gasser G, Tonks NK.
 A potent, selective, and orally bioavailable inhibitor of the protein-tyrosine phosphatase PTP1B improves insulin and leptin signaling in animal models. *J Biol Chem.* 2018;293(5):1517–1525.
- 93. Liu J, Lee J, Salazar Hernandez MA, Mazitschek R, Ozcan U. Treatment of obesity with celastrol. *Cell*. 2015;161(5):999–1011.
- Lee J, et al. Withaferin A is a leptin sensitizer with strong antidiabetic properties in mice. *Nat Med*. 2016;22(9):1023–1032.
- 95. Pfuhlmann K, et al. Celastrol-induced weight loss is driven by hypophagia and independent from UCP1. *Diabetes*. 2018;67(11):2456–2465.
- Kyriakou E, et al. Celastrol promotes weight loss in diet-induced obesity by inhibiting the protein tyrosine phosphatases PTP1B and TCPTP in the hypothalamus. J Med Chem. 2018;61(24):11144–11157.
- Feng X, et al. IL1R1 is required for celastrol's leptin-sensitization and antiobesity effects. Nat Med. 2019;25(4):575–582.
- Clément K, et al. MC4R agonism promotes durable weight loss in patients with leptin receptor deficiency. Nat Med. 2018;24(5):551–555.
- Kühnen P, et al. Proopiomelanocortin deficiency treated with a melanocortin-4 receptor agonist. N Engl J Med. 2016;375(3):240-246.
- 100.Greenfield JR, et al. Modulation of blood pressure by central melanocortinergic pathways. N Engl J Med. 2009;360(1):44–52.
- 101. Krishna R, et al. Potent and selective agonism of the melanocortin receptor 4 with MK-0493 does not induce weight loss in obese human subjects: energy intake predicts lack of weight loss efficacy. Clin Pharmacol Ther. 2009;86(6):659-666.
- 102. Lorenz DN, Goldman SA. Vagal mediation of the cholecystokinin satiety effect in rats. *Physiol Behav*. 1982;29(4):599–604.
- 103. West DB, Fey D, Woods SC. Cholecystokinin persistently suppresses meal size but not food intake in free-feeding rats. Am J Physiol. 1984; 246(5 pt 2):R776-R787.
- 104. Crawley JN, Beinfeld MC. Rapid development of tolerance to the behavioural actions of cholecystokinin. *Nature*. 1983;302(5910):703-706.
- 105. Jordan J, et al. Stimulation of cholecystokinin-A receptors with GI181771X does not cause weight loss in overweight or obese patients. Clin Pharmacol Ther. 2008;83(2):281–287.
- 106.Batterham RL, et al. Inhibition of food intake in obese subjects by peptide YY3-36. N Engl J Med. 2003;349(10):941-948.
- 107. Tschöp M, et al. Physiology: does gut hormone PYY3-36 decrease food intake in rodents? *Nature*. 2004;430(6996):1 p following 165; discussion 2

- p following 165.
- 108.le Roux CW, Borg CM, Murphy KG, Vincent RP, Ghatei MA, Bloom SR. Supraphysiological doses of intravenous PYY3-36 cause nausea, but no additional reduction in food intake. Ann Clin Biochem. 2008;45(pt 1):93-95.
- 109. Yulyaningsih E, Zhang L, Herzog H, Sainsbury A. NPY receptors as potential targets for anti-obesity drug development. Br J Pharmacol. 2011;163(6):1170-1202.
- 110. Kreymann B, Williams G, Ghatei MA, Bloom SR. Glucagon-like peptide-17-36: a physiological incretin in man. *Lancet*. 1987;2(8571):1300-1304.
- 111. Mojsov S, Weir GC, Habener JF. Insulinotropin: glucagon-like peptide I (7-37) co-encoded in the glucagon gene is a potent stimulator of insulin release in the perfused rat pancreas. *J Clin Invest*. 1987;79(2):616–619.
- 112. de Heer J, Rasmussen C, Coy DH, Holst JJ. Glucagon-like peptide-1, but not glucose-dependent insulinotropic peptide, inhibits glucagon secretion via somatostatin (receptor subtype 2) in the perfused rat pancreas. *Diabetologia*. 2008;51(12):2263-2270.
- 113. Li Y, Hansotia T, Yusta B, Ris F, Halban PA, Drucker DJ. Glucagon-like peptide-1 receptor signaling modulates beta cell apoptosis. *J Biol Chem*. 2003;278(1):471–478.
- 114. Lee YS, et al. Glucagon-like peptide-1 gene therapy in obese diabetic mice results in long-term cure of diabetes by improving insulin sensitivity and reducing hepatic gluconeogenesis. *Diabetes*. 2007;56(6):1671-1679.
- 115. Patel S, et al. GDF15 provides an endocrine signal of nutritional stress in mice and humans. *Cell Metab*. 2019;29(3):707-718.e8.
- 116. Secher A, et al. The arcuate nucleus mediates GLP-1 receptor agonist liraglutide-dependent weight loss. J Clin Invest. 2014;124(10):4473-4488.
- 117. Hayes MR, et al. Intracellular signals mediating the food intake-suppressive effects of hindbrain glucagon-like peptide-1 receptor activation. *Cell Metab.* 2011;13(3):320–330.
- 118. Hayes MR, Schmidt HD. GLP-1 influences food and drug reward. Curr Opin Behav Sci. 2016;9:66-70.
- 119. Linnebjerg H, et al. Effect of exenatide on gastric emptying and relationship to postprandial glycemia in type 2 diabetes. *Regul Pept*. 2008;151(1-3):123-129.
- 120. Lorenz M, et al. Effects of lixisenatide once daily on gastric emptying in type 2 diabetes — relationship to postprandial glycemia. *Regul Pept*. 2013;185:1–8.
- 121. Nauck MA, Kemmeries G, Holst JJ, Meier JJ.
 Rapid tachyphylaxis of the glucagon-like peptide
 1-induced deceleration of gastric emptying in
 humans. *Diabetes*. 2011;60(5):1561–1565.
- 122. Abd El Aziz MS, Kahle M, Meier JJ, Nauck MA. A meta-analysis comparing clinical effects of shortor long-acting GLP-1 receptor agonists versus insulin treatment from head-to-head studies in type 2 diabetic patients. *Diabetes Obes Metab*. 2017;19(2):216–227.
- 123. Trujillo JM, Nuffer W, Ellis SL. GLP-1 receptor agonists: a review of head-to-head clinical studies. *Ther Adv Endocrinol Metab*. 2015;6(1):19–28. 124.O'Neil PM, et al. Efficacy and safety of sema-

The Journal of Clinical Investigation

- glutide compared with liraglutide and placebo for weight loss in patients with obesity: a randomised, double-blind, placebo and active controlled, dose-ranging, phase 2 trial. *Lancet*. 2018;392(10148):637–649.
- 125. Davies M, Pieber TR, Hartoft-Nielsen ML, Hansen OKH, Jabbour S, Rosenstock J. Effect of oral semaglutide compared with placebo and subcutaneous semaglutide on glycemic control in patients with type 2 diabetes: a randomized clinical trial. *JAMA*. 2017;318(15):1460–1470.
- 126. Bethel MA, et al. Cardiovascular outcomes with glucagon-like peptide-1 receptor agonists in patients with type 2 diabetes: a meta-analysis. Lancet Diabetes Endocrinol. 2018;6(2):105–113.
- 127. Drucker DJ. The ascending GLP-1 road from clinical safety to reduction of cardiovascular complications. *Diabetes*. 2018;67(9):1710-1719.
- 128. Pocai A, et al. Glucagon-like peptide 1/glucagon receptor dual agonism reverses obesity in mice. *Diabetes*. 2009;58(10):2258–2266.
- 129. Day JW, et al. A new glucagon and GLP-1 coagonist eliminates obesity in rodents. Nat Chem Biol. 2009;5(10):749-757.
- 130. Finan B, et al. Unimolecular dual incretins maximize metabolic benefits in rodents, monkeys, and humans. Sci Transl Med. 2013;5(209):209ra151.
- 131. Finan B, et al. A rationally designed monomeric peptide triagonist corrects obesity and diabetes in rodents. Nat Med. 2015;21(1):27-36.
- 132. Habegger KM, Heppner KM, Geary N, Bartness TJ, DiMarchi R, Tschöp MH. The metabolic actions of glucagon revisited. *Nat Rev Endocrinol*. 2010;6(12):689-697.
- 133. Brandt SJ, Götz A, Tschöp MH, Müller TD. Gut hormone polyagonists for the treatment of type 2 diabetes. *Peptides*. 2018;100:190-201.
- 134. Ambery P, et al. MEDIO382, a GLP-1 and glucagon receptor dual agonist, in obese or overweight patients with type 2 diabetes: a randomised, controlled, double-blind, ascending dose and phase 2a study. *Lancet*. 2018;391(10140):2607–2618.
- 135. Tillner J, et al. A novel dual glucagon-like peptide and glucagon receptor agonist SAR425899: results of randomized, placebo-controlled firstin-human and first-in-patient trials. *Diabetes Obes Metab*. 2019;21(1):120–128.
- 136. Tatarkiewicz K, et al. A novel long-acting glucose-dependent insulinotropic peptide analogue: enhanced efficacy in normal and diabetic rodents. *Diabetes Obes Metab.* 2014;16(1):75–85.
- 137. Nørregaard PK, et al. A novel GIP analogue, ZP4165, enhances glucagon-like peptide-1induced body weight loss and improves glycaemic control in rodents. *Diabetes Obes Metab*. 2018:20(1):60-68.
- 138. Mroz PA, et al. Optimized GIP analogs promote body weight lowering in mice through GIPR agonism not antagonism. *Mol Metab*. 2019;20:51–62.
- 139. Pathak V, Gault VA, Flatt PR, Irwin N. Antagonism of gastric inhibitory polypeptide (GIP) by palmitoylation of GIP analogues with N- and C-terminal modifications improves obesity and metabolic control in high fat fed mice. Mol Cell Endocrinol. 2015;401:120–129.
- 140. Killion EA, et al. Anti-obesity effects of GIPR antagonists alone and in combination with GLP-

- 1R agonists in preclinical models. *Sci Transl Med*. 2018;10(472):eaat3392.
- 141. Frias JP, et al. The sustained effects of a dual GIP/GLP-1 receptor agonist, NNC0090-2746, in patients with type 2 diabetes. *Cell Metab*. 2017;26(2):343-352.e2.
- 142. Frias JP, et al. Efficacy and safety of LY3298176, a novel dual GIP and GLP-1 receptor agonist, in patients with type 2 diabetes: a randomised, placebo-controlled and active comparator-controlled phase 2 trial. *Lancet*. 2018;392(10160):2180-2193.
- 143. Hornigold DC, et al. A GLP-1:CCK fusion peptide harnesses the synergistic effects on metabolism of CCK-1 and GLP-1 receptor agonism in mice. *Appetite*. 2018;127:334–340.
- 144.Skarbaliene J, et al. The anti-diabetic effects of GLP-1-gastrin dual agonist ZP3022 in ZDF rats. Peptides. 2015;69:47–55.
- 145. Trevaskis JL, et al. Improved glucose control and reduced body weight in rodents with dual mechanism of action peptide hybrids. PLoS One. 2013;8(10):e78154.
- 146. Finan B, et al. Targeted estrogen delivery reverses the metabolic syndrome. *Nat Med*. 2012;18(12):1847–1856.
- 147. Quarta C, et al. Molecular integration of incretin and glucocorticoid action reverses immunometabolic dysfunction and obesity. *Cell Metab*. 2017;26(4):620-632.e6.
- 148.Ämmälä C, et al. Targeted delivery of antisense oligonucleotides to pancreatic β-cells. Sci Adv. 2018;4(10):eaat3386.
- 149. Schwenk RW, et al. GLP-1-oestrogen attenuates hyperphagia and protects from β cell failure in diabetes-prone New Zealand obese (NZO) mice. *Diabetologia*. 2015;58(3):604–614.
- 150. García-Cáceres C, et al. Role of astrocytes, microglia, and tanycytes in brain control of systemic metabolism. *Nat Neurosci*. 2019;22(1):7–14.
- 151. Nishino J, Yamashita K, Hashiguchi H, Fujii H, Shimazaki T, Hamada H. Meteorin: a secreted protein that regulates glial cell differentiation and promotes axonal extension. *EMBO J.* 2004;23(9):1998–2008.
- 152. Rao RR, et al. Meteorin-like is a hormone that regulates immune-adipose interactions to increase beige fat thermogenesis. *Cell*. 2014;157(6):1279–1291.
- 153. Lo JC, et al. Adipsin is an adipokine that improves β cell function in diabetes. *Cell*. 2014;158(1):41–53.
- 154. Boström P, et al. A PGC1-α-dependent myokine that drives brown-fat-like development of white fat and thermogenesis. *Nature*. 2012;481(7382):463-468.
- 155. Bootcov MR, et al. MIC-1, a novel macrophage inhibitory cytokine, is a divergent member of the TGF-β superfamily. *Proc Natl Acad Sci U S A*. 1997;94(21):11514–11519.
- 156. Emmerson PJ, et al. The metabolic effects of GDF15 are mediated by the orphan receptor GFRAL. Nat Med. 2017;23(10):1215–1219.
- 157. Yang L, et al. GFRAL is the receptor for GDF15 and is required for the anti-obesity effects of the ligand. *Nat Med.* 2017;23(10):1158–1166.
- 158. Mullican SE, et al. GFRAL is the receptor for GDF15 and the ligand promotes weight loss in mice and nonhuman primates. *Nat Med*.

- 2017;23(10):1150-1157.
- 159. Sonoda J, Chen MZ, Baruch A. FGF21-receptor agonists: an emerging therapeutic class for obesity-related diseases. Horm Mol Biol Clin Investig. 2017;30(2):20170002.
- 160.Scarlett JM, et al. Central injection of fibroblast growth factor 1 induces sustained remission of diabetic hyperglycemia in rodents. *Nat Med*. 2016;22(7):800–806.
- 161. Brown JM, et al. The hypothalamic arcuate nucleus-median eminence is a target for sustained diabetes remission induced by fibroblast growth factor 1. *Diabetes*. 2019;68(5):1054-1061.
- 162. Davies BT, Wellman PJ, Morien A. An assessment of the involvement of paraventricular hypothalamic α 2-adrenoceptors in phenylpropanolamine anorexia. *Physiol Behav.* 1993;54(1):121–128.
- 163. Forman HP, Levin S, Stewart B, Patel M, Feinstein S. Cerebral vasculitis and hemorrhage in an adolescent taking diet pills containing phenylpropanolamine: case report and review of literature. *Pediatrics*. 1989;83(5):737–741.
- 164. Kernan WN, et al. Phenylpropanolamine and the risk of hemorrhagic stroke. N Engl J Med. 2000;343(25):1826–1832.
- 165. Rothman RB, Baumann MH. Therapeutic potential of monoamine transporter substrates. Curr Top Med Chem. 2006;6(17):1845–1859.
- 166. Evans J. Psychosis and addiction to phenmetrazine (preludin). *Lancet*. 1959;2(7095):152–155.
- 167. Fishman AP. Aminorex to fen/phen: an epidemic foretold. *Circulation*. 1999;99(1):156–161.
- 168. Gurtner HP. Aminorex and pulmonary hypertension. A review. *Cor Vasa*. 1985;27(2-3):160-171.
- 169. Rowland NE, Carlton J. Neurobiology of an anorectic drug: fenfluramine. *Prog Neurobiol*. 1986;27(1):13–62.
- 170. Rothman RB, et al. Evidence for possible involvement of 5-HT(2B) receptors in the cardiac valvulopathy associated with fenfluramine and other serotonergic medications. *Circulation*. 2000;102(23):2836–2841.
- 171. Balcioglu A, Wurtman RJ. Effects of phentermine on striatal dopamine and serotonin release in conscious rats: in vivo microdialysis study. *Int J Obes Relat Metab Disord*. 1998;22(4):325–328.
- 172. Barkeling B, Elfhag K, Rooth P, Rössner S. Short-term effects of sibutramine (Reductil) on appetite and eating behaviour and the long-term therapeutic outcome. *Int J Obes Relat Metab Disord*. 2003;27(6):693–700.
- 173. Rothman RB, et al. Interaction of the anorectic medication, phendimetrazine, and its metabolites with monoamine transporters in rat brain. *Eur J Pharmacol*. 2002;447(1):51–57.
- 174. Le Riche WH, Van Belle G. Study of phendimetrazine bitartrate as an appetite suppressant in relation to dosage, weight loss and side effects. Can Med Assoc J. 1962;87:29–31.
- 175. Lang SS, Danforth E, Lien EL. Anorectic drugs which stimulate thermogenesis. *Life Sci*. 1983;33(13):1269–1275.
- 176. Haddock CK, Poston WS, Dill PL, Foreyt JP, Ericsson M. Pharmacotherapy for obesity: a quantitative analysis of four decades of published randomized clinical trials. *Int J Obes Relat Metab Disord*. 2002;26(2):262–273.
- 177. Rothman RB, et al. Amphetamine-type central

The Journal of Clinical Investigation

REVIEW SERIES: MECHANISMS UNDERLYING THE METABOLIC SYNDROME

- nervous system stimulants release norepinephrine more potently than they release dopamine and serotonin. *Synapse*. 2001;39(1):32–41.
- 178. Aronne LJ, Wadden TA, Peterson C, Winslow D, Odeh S, Gadde KM. Evaluation of phentermine and topiramate versus phentermine/topiramate extended-release in obese adults. Obesity (Silver Spring). 2013;21(11):2163–2171.
- 179. Coomans CP, et al. The insulin sensitizing effect of topiramate involves KATP channel activation in the central nervous system. *Br J Pharmacol*. 2013;170(4):908–918.
- 180. Picard F, Deshaies Y, Lalonde J, Samson P, Richard D. Topiramate reduces energy and fat gains in lean (Fa/?) and obese (fa/fa) Zucker rats. Obes Res. 2000;8(9):656-663.
- 181. Allison DB, et al. Controlled-release phentermine/topiramate in severely obese adults: a randomized controlled trial (EQUIP). Obesity (Silver Spring). 2012;20(2):330–342.
- 182. Gadde KM, et al. Effects of low-dose, controlled-release, phentermine plus topiramate combination on weight and associated comorbidities in overweight and obese adults (CON-QUER): a randomised, placebo-controlled, phase 3 trial. *Lancet*. 2011;377(9774):1341-1352.
- 183. Garvey WT, et al. Two-year sustained weight loss and metabolic benefits with controlled-release phentermine/topiramate in obese and overweight adults (SEQUEL): a randomized, placebocontrolled, phase 3 extension study. Am J Clin Nutr. 2012;95(2):297-308.
- 184.Smith SR, et al. Multicenter, placebo-controlled trial of lorcaserin for weight management. *N Engl J Med*. 2010;363(3):245–256.
- 185. Hollander P, et al. Effects of naltrexone sustained-release/bupropion sustained-release

- combination therapy on body weight and glycemic parameters in overweight and obese patients with type 2 diabetes. *Diabetes Care*. 2013;36(12):4022-4029.
- 186. Hauser RA, Salin L, Juhel N, Konyago VL. Randomized trial of the triple monoamine reuptake inhibitor NS 2330 (tesofensine) in early Parkinson's disease. Mov Disord. 2007;22(3):359–365.
- 187. Astrup A, Madsbad S, Breum L, Jensen TJ, Kroustrup JP, Larsen TM. Effect of tesofensine on bodyweight loss, body composition, and quality of life in obese patients: a randomised, double-blind, placebo-controlled trial. *Lancet*. 2008;372(9653):1906–1913.
- 188. Neidigh JW, Fesinmeyer RM, Prickett KS, Andersen NH. Exendin-4 and glucagon-like-peptide-1: NMR structural comparisons in the solution and micelle-associated states. *Biochemistry*. 2001;40(44):13188-13200.
- 189. Drucker DJ, et al. Exenatide once weekly versus twice daily for the treatment of type 2 diabetes: a randomised, open-label, non-inferiority study. *Lancet*. 2008;372(9645):1240–1250.
- 190.McCarty D, Coleman M, Boland CL. Lixisenatide: a new daily GLP-1 agonist for type 2 diabetes management. *Ann Pharmacother*. 2017;51(5):401-409.
- 191. Rosenstock J, et al. Efficacy and safety of lixisenatide once daily versus exenatide twice daily in type 2 diabetes inadequately controlled on metformin: a 24-week, randomized, open-label, active-controlled study (GetGoal-X). Diabetes Care. 2013;36(10):2945–2951.
- 192. Fineman M, et al. Pharmacokinetics and pharmacodynamics of exenatide extended-release after single and multiple dosing. *Clin Pharmacokinet*. 2011;50(1):65-74.

- 193. Bush MA, et al. Safety, tolerability, pharmacodynamics and pharmacokinetics of albiglutide, a long-acting glucagon-like peptide-1 mimetic, in healthy subjects. *Diabetes Obes Metab*. 2009;11(5):498-505.
- 194. Pratley RE, et al. Once-weekly albiglutide versus once-daily liraglutide in patients with type 2 diabetes inadequately controlled on oral drugs (HARMONY 7): a randomised, open-label, multicentre, non-inferiority phase 3 study. *Lancet Diabetes Endocrinol*. 2014;2(4):289–297.
- 195. Barrington P, et al. A 5-week study of the pharmacokinetics and pharmacodynamics of LY2189265, a novel, long-acting glucagon-like peptide-1 analogue, in patients with type 2 diabetes. *Diabetes Obes Metab*. 2011;13(5):426-433.
- 196. Dungan KM, et al. Once-weekly dulaglutide versus once-daily liraglutide in metformin-treated patients with type 2 diabetes (AWARD-6): a randomised, open-label, phase 3, non-inferiority trial. *Lancet*. 2014;384(9951):1349-1357.
- 197. Agersø H, Jensen LB, Elbrønd B, Rolan P, Zdravkovic M. The pharmacokinetics, pharmacodynamics, safety and tolerability of NN2211, a new long-acting GLP-1 derivative, in healthy men. *Diabetologia*. 2002;45(2):195–202.
- 198. Buse JB, et al. Liraglutide once a day versus exenatide twice a day for type 2 diabetes: a 26-week randomised, parallel-group, multinational, open-label trial (LEAD-6). *Lancet*. 2009;374(9683):39-47.
- 199.Lau J, et al. Discovery of the once-weekly glucagon-like peptide-1 (GLP-1) analogue semaglutide. J Med Chem. 2015;58(18):7370-7380.
- 200. Pratley RE, et al. Semaglutide versus dulaglutide once weekly in patients with type 2 diabetes (SUSTAIN 7): a randomised, open-label, phase 3b trial. Lancet Diabetes Endocrinol. 2018;6(4):275–286.