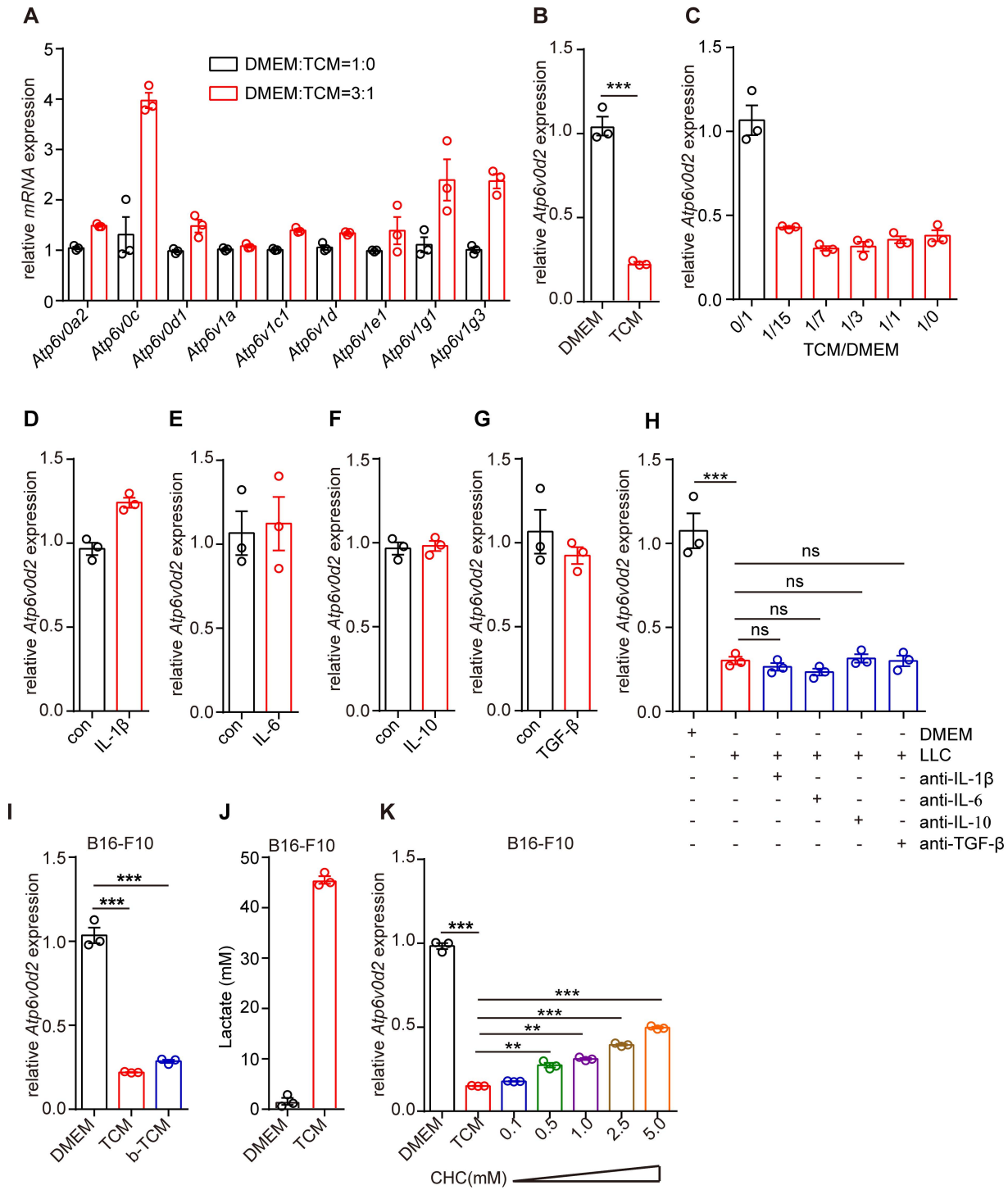
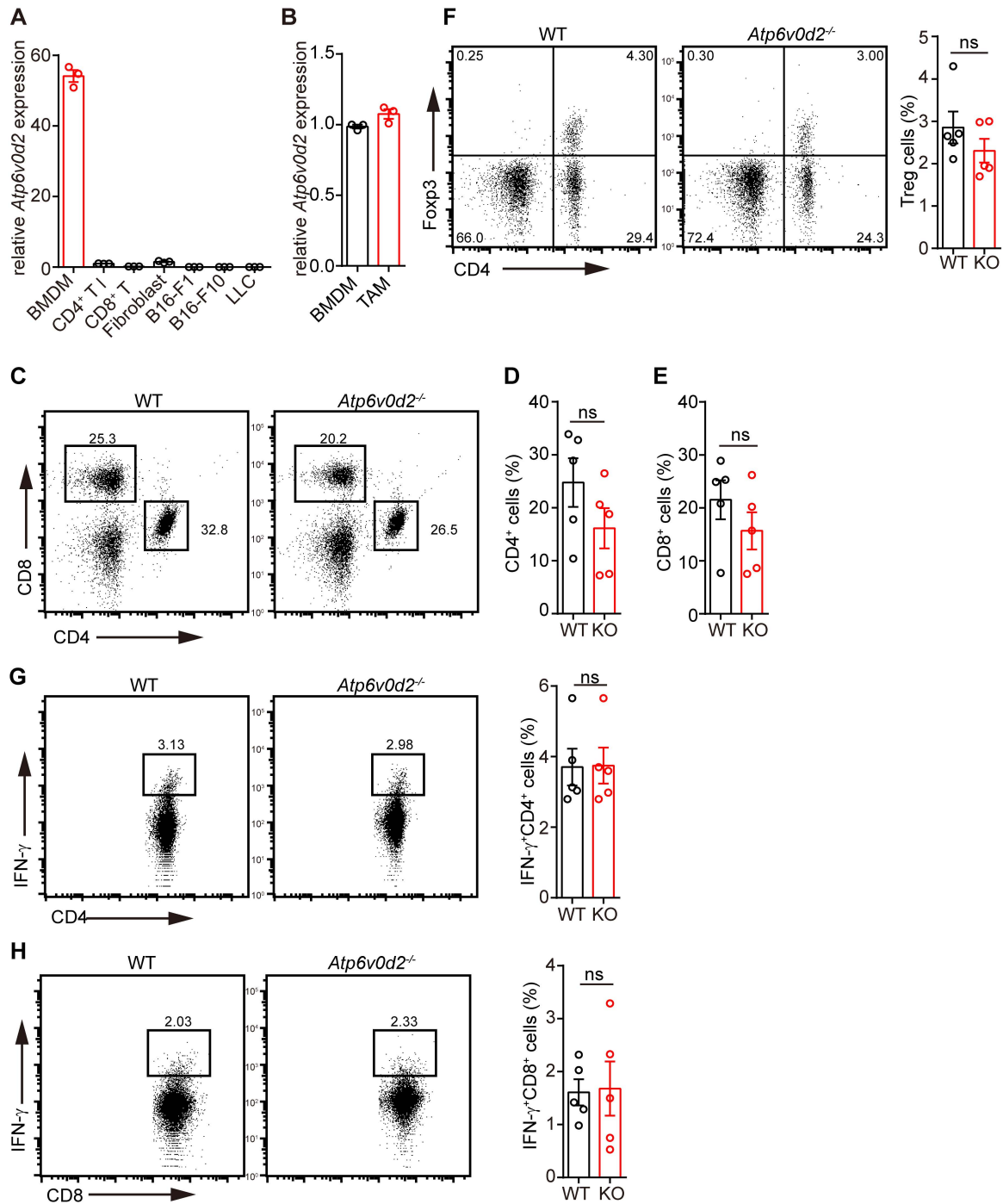


Figure S1



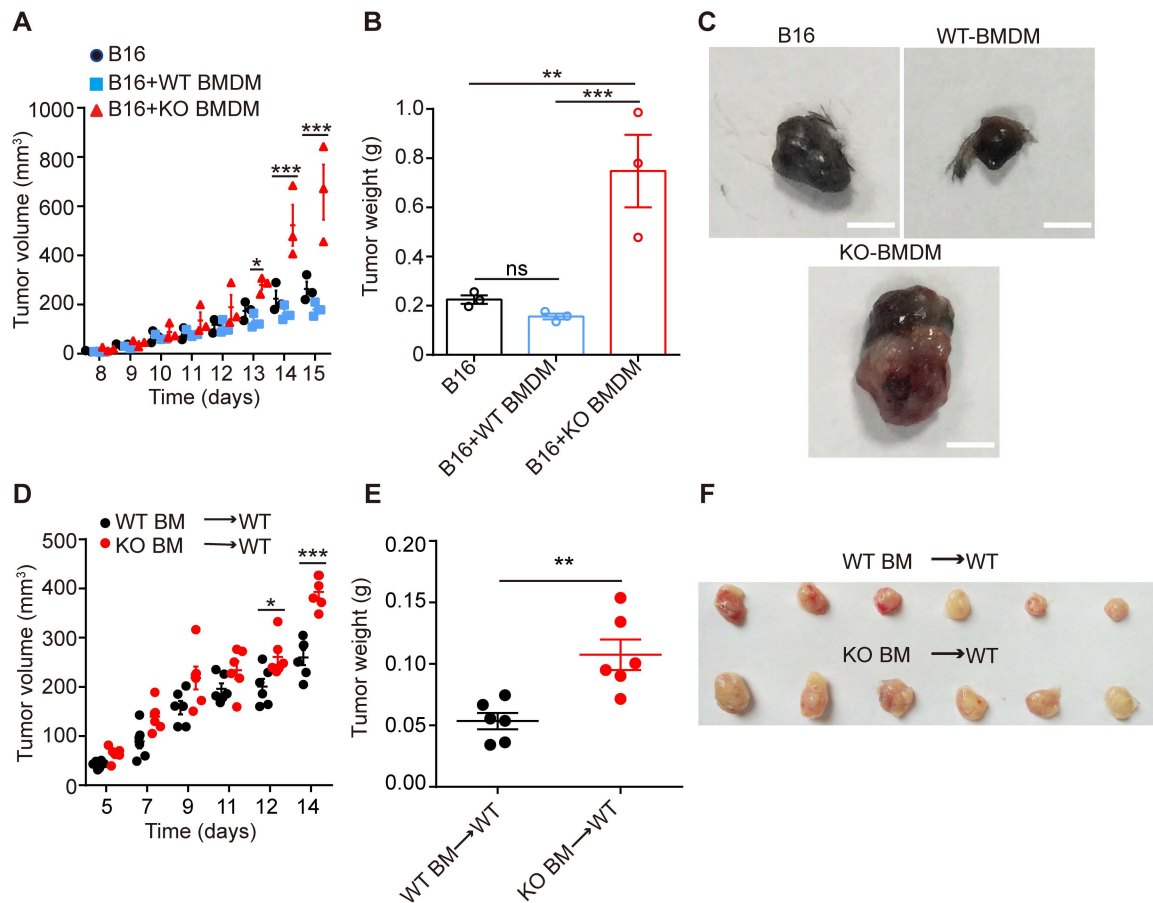
Supplementary Figure 1. TCM but not cytokines presented in tumors inhibits *Atp6v0d2* expression. (A) Expression of different V-ATPase subunits were determined by q-PCR in BMDMs cultured with complete DMEM or stimulated with DMEM:LLC-TCM (3:1) for 6h. (B-K, except J) *Atp6v0d2* mRNA expression was determined by q-PCR. (B and C) BMDMs stimulated with B16-F10-TCM (B) or a concentration gradient of B16-F10-TCM (C) for 6h. (D-G) Expression of *Atp6v0d2* mRNA was determined by q-PCR in BMDMs stimulated with IL-1 β (D), IL-6 (E), IL-10 (F) and TGF- β (G) for 6h. (H) BMDMs were stimulated with LLC-TCM or with the addition of different neutralization antibodies (10 ng/ml) for 6h. (I) BMDMs were stimulated with B16-F10-TCM or boiled B16-F10 TCM for 6h. (J) Lactate concentration in the B16-F10-TCM was determined. (K) BMDMs stimulated with B16-F10-TCM alone or with the addition of different doses of CHC for 6h. Data are representative of two independent experiments. Data are assessed by Student *t* test (A, B, D-G) or one-way ANOVA with Turkey's test (C, H, I, K). mean \pm SEM. ***p* < 0.01, ****p* < 0.001.

Figure S2



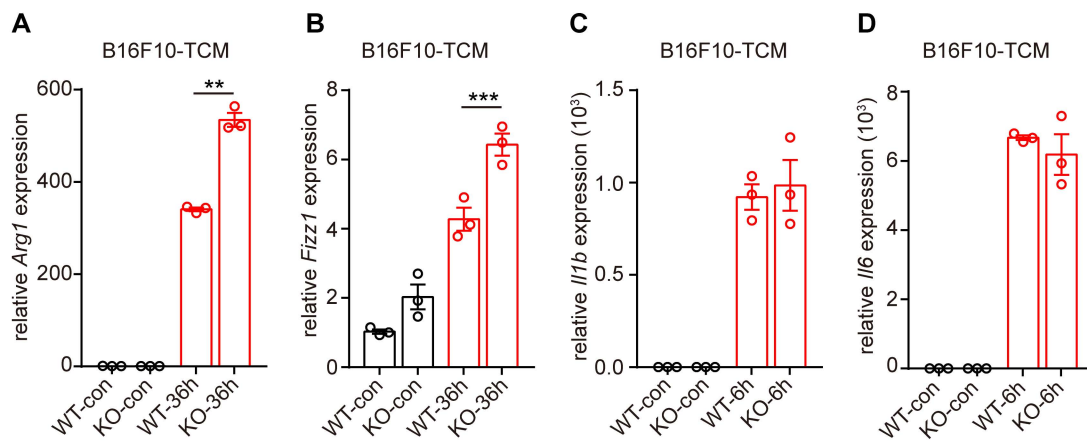
Supplementary Figure 2. Deficiency of *Atp6v0d2* does not affect T cell functions within tumor. (A) q-PCR analysis of *Atp6v0d2* expression in BMDMs, CD4⁺T cell, CD8⁺ T cell, fibroblast, B16-F1, B16-F10 and LLC cells. (B) The expression of *Atp6v0d2* in BMDMs and TAMs from tumor-bearing WT mice isolated with CD11b magnetic beads was analyzed by q-PCR. (C-H) WT and *Atp6v0d2*^{-/-} mice were injected s.c. with 5×10⁵ LLC cells. On day 15 post-inoculation, mice were sacrificed (n=5). (C-E) Representative flow cytometric plot showing T cells fractions in LLC tumor tissues (C), comparisons of the percentages of CD4⁺ (D) and CD8⁺ (E) in LLC tumors between WT and *Atp6v0d2*^{-/-} mice. (F-H) Representative dot plots showing percentages of Foxp3⁺CD4⁺ T cells (F), IFN- γ ⁺CD4⁺ T cells (G), IFN- γ ⁺CD8⁺ T cells (H) in the tumor tissues. Right, cumulative results for the staining at left. Data Student *t* test and are representative of two independent experiments.

Figure S3



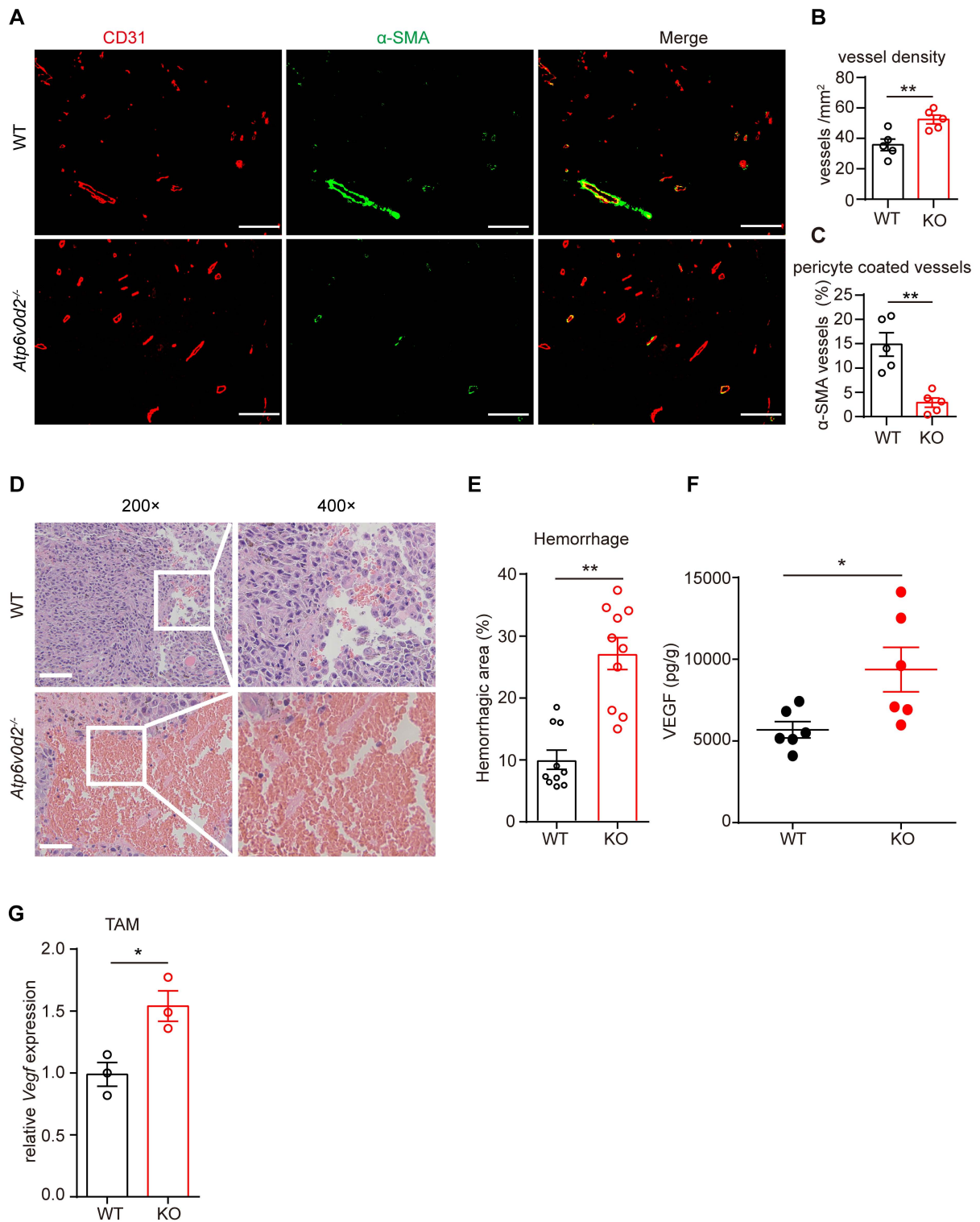
Supplementary Figure 3. *Atp6v0d2*-deficient BMDMs and bone marrows promote tumor growth. (A-C) *Atp6v0d2*^{-/-} mice were challenged s.c. with 5×10^5 B16-F10 cells alone, or plus with 2×10^5 either WT BMDMs or *Atp6v0d2*^{-/-} BMDMs (n=3). Tumor size was measured every 2 days (A). Tumor mass was determined at day 15 after inoculation (B). Representative images of tumor were presented (C). (D-F) WT mice were irradiated with 7Gy and reconstituted with either WT or *Atp6v0d2*^{-/-} BM (n=6). After 4 weeks, mice were injected s.c. with 5×10^5 LLC cells, tumor growth rate (D), tumor weight (E) and images of excised tumor (F). Data were assessed by one-way ANOVA with Turkey's test (A, B) or Student's t test (D, E). mean \pm SEM. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$. Scale bars: 5 mm

Figure S4



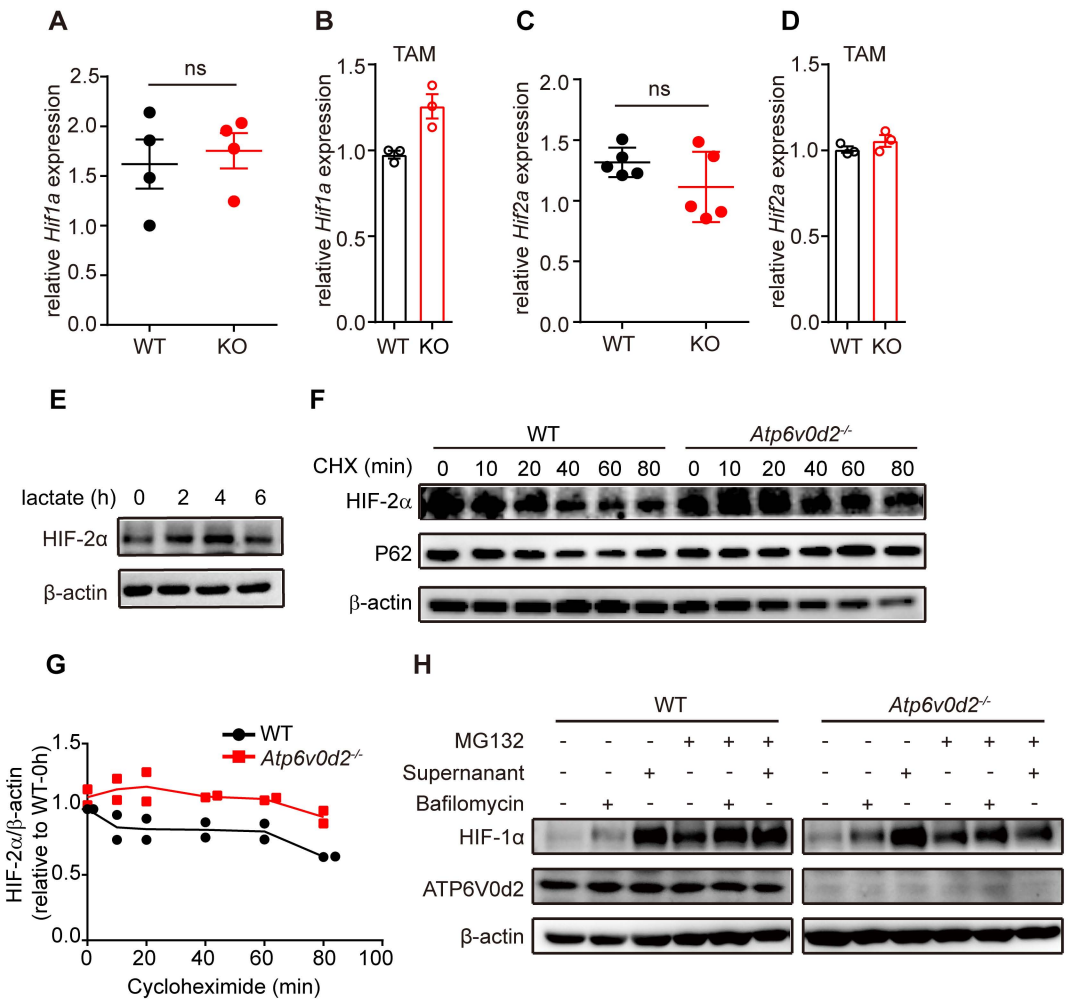
Supplementary Figure 4. Deletion of *Atp6v0d2* leads to enhanced protumoral-related gene expression. (A and B) Expression analysis by q-PCR of *Arg1* (A) and *Fizz1* (B) in WT and *Atp6v0d2*^{-/-} BMDMs stimulated with B16-F10-conditioned medium for 36h. (C and D) Expression of *Il1b* (C) and *Il6* (D) in WT and *Atp6v0d2*^{-/-} BMDMs stimulated with B16-F10-conditioned medium for 6h. Data are assessed by student *t* test and representative representative of three independent experiments. mean±SEM. ***p* < 0.01, ****p* < 0.001.

Figure S5



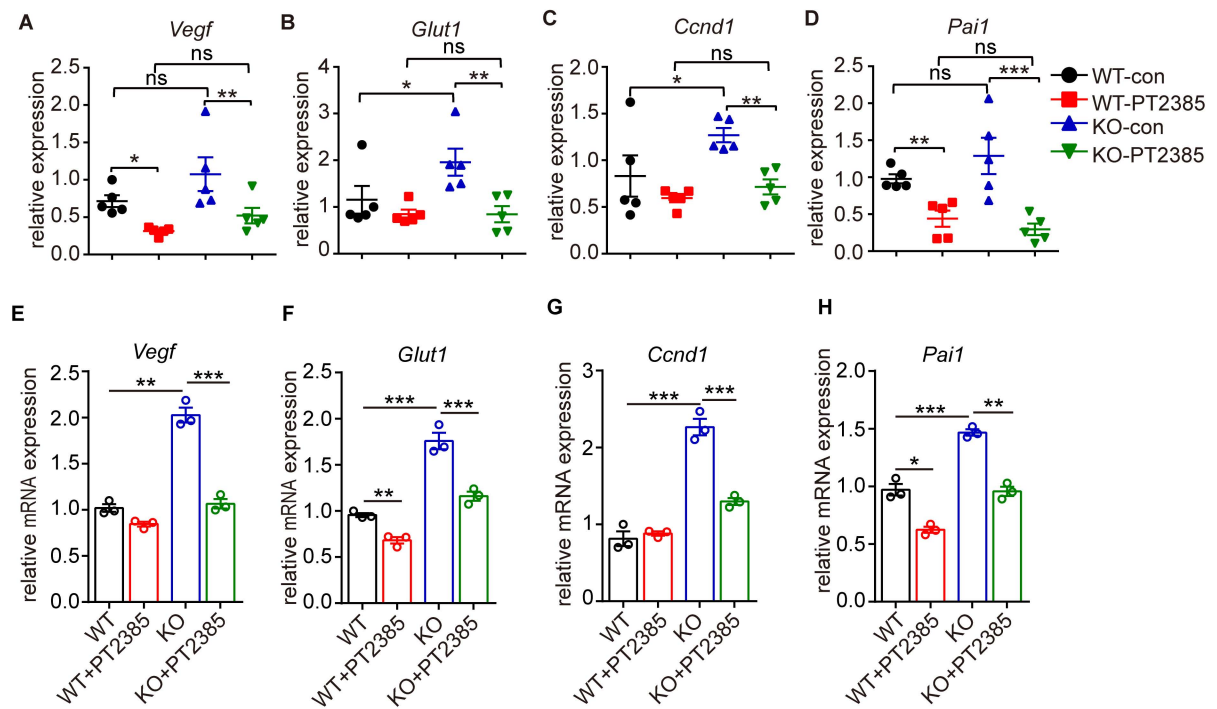
Supplementary Figure 5. *Atp6v0d2*-deficient mice have enhanced angiogenesis. (A-G) WT and *Atp6v0d2*^{-/-} mice were injected s.c. with 2×10^5 B16-F10 cells. On day 15 post-inoculation, mice were sacrificed. (A-C) Double immunostaining for CD31 (red) and α -SMA (green) vessels in tumor tissues isolated from WT and *Atp6v0d2*^{-/-} mice (A), quantifications of the percentage CD31⁺ vessels (B), and α -SMA⁺CD31⁺ vessels (C) (n=5). Bars in all tissue section panels were 100 μ m. (D and E) H&E staining (200 \times , 400 \times) showing hemorrhaging in B16 tumors tissues isolated from WT and *Atp6v0d2*^{-/-} mice (D). Comparison of hemorrhagic area (% of tumor area) was shown in (E). (F) The levels of VEGF were determined in tumor tissues from WT and *Atp6v0d2*^{-/-} mice by ELISA. (G) The mRNA of *Vegf* was determined in TAMs isolated from WT and *Atp6v0d2*^{-/-} tumor bearing mice. Data are assessed by student *t* test and representative of two independent experiments. mean \pm SEM. **p* < 0.05, ***p* < 0.01.

Figure S6



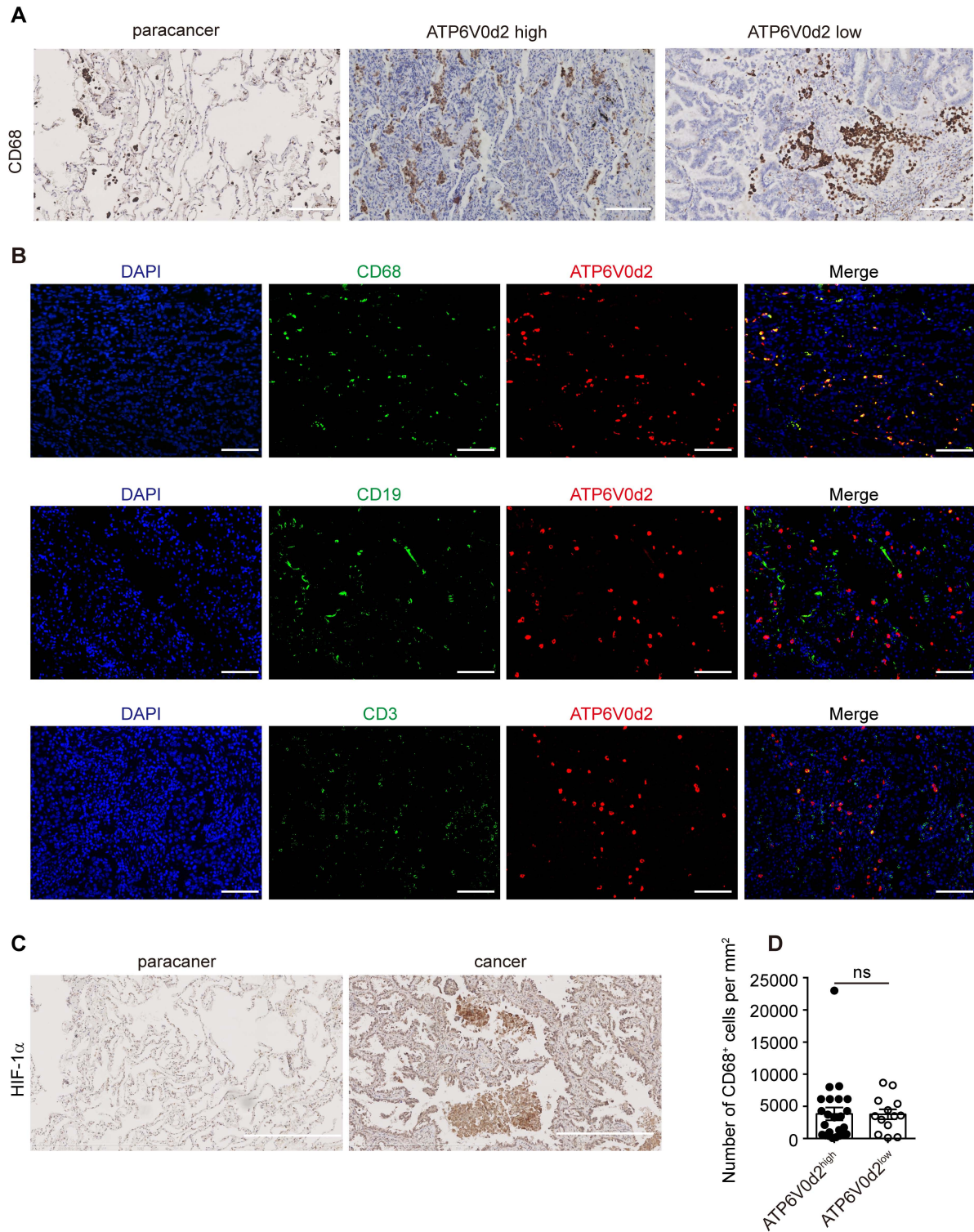
Supplementary Figure 6. ATP6V0d2 does not regulate HIF-1α stability. (A and B) *Hif1a* mRNA expression in LLC-tumor tissues (A) and TAMs (B) from WT and *Atp6v0d2*^{-/-} mice. (C and D) *Hif2a* mRNA expression in LLC-tumor tissues (C) and TAMs (D) from WT and *Atp6v0d2*^{-/-} mice. (E) Immunoblotting of HIF-2α on WT and *Atp6v0d2*^{-/-} BMDMs that were stimulated with lactate (40mM) for the indicated times. (F and G) Immunoblotting of HIF-2α and P62 expression in WT and *Atp6v0d2*^{-/-} BMDMs incubated with cycloheximide (100μg/ml) for indicated times (F) and band intensities were plotted (G). (H) Immunoblotting of HIF-1α expression in WT and *Atp6v0d2*^{-/-} BMDMs stimulated with Bafilomycin (100nM), LLC-TCM, MG132 (20μM), MG132 together with Bafilomycin, or MG132 in addition to LLC-TCM for 6h. Data are assessed by Student *t* test and representative of two (A-D, F-G) or three (E, H) independent experiments.

Figure S7



Supplementary Figure 7. PT2385 inhibits HIF-2 α target genes in macrophages. (A-D) q-PCR analysis of HIF-2 α target genes *Vegf* (A), *Glut1* (B), *Ccnd1* (C), *Pai1* (D) in the tumor tissues of the WT and *Atp6v0d2*^{-/-} mice treated with vehicle or PT2385. (E-H) q-PCR of HIF-2 α target genes *Vegf* (E), *Glut1* (F), *Ccnd1* (G) and *Pai1* (H) in WT and *Atp6v0d2*^{-/-} BMDMs stimulated with LLC-TCM for 6h with or without PT2385 (10mM). PT2385 was added 18h prior to the TCM stimulation. Experiments were repeated for two times. Data are assessed by two-way ANOVA with Turkey's test and presented as Mean \pm SEM, * p < 0.05, ** p < 0.01, *** p < 0.001.

Figure S8



Supplementary Figure 8. ATP6V0d2 is specifically expressed in macrophages in human primary lung cancers. (A) Immunohistochemistry(IHC) analysis of CD68 in the lung sections of adjacent non-tumorous lungs or tumor tissues isolated from two lung adenocarcinoma patients with differential ATP6V0d2 expression. **(B)** Immunofluorescence staining of human primary lung adenocarcinoma tissues with DAPI (blue), anti-CD68 (green), anti-CD20 (green), anti-CD3 (green), and anti-ATP6V0d2 (red) (n=3). **(C)** A representative image of IHC staining the HIF-1 α in the lung sections of adjacent non-tumorous or tumor tissues of human adenocarcinoma patients. **(D)** Quantification of numbers of CD68⁺ TAMs in 5 areas of each patient were determined manually in two groups of lung adenocarcinoma patients with high ATP6V0d2 expression (n=24, IRS>3) and low ATP6V0d2 expression (n=13, IRS<3). Scale bars, (A and C) 200 μ m; B 100 μ m.

Supplemental Table 1

Oligonucleotides		
m. <i>Atp6v0d2</i> qPCR Forward primer TGCGGCAGGCTCTATCCAGAGG	This paper	N/A
m. <i>Atp6v0d2</i> qPCR Reverse primer CCACTGCCACCGACAGCGTC	This paper	N/A
m. <i>Atp6v0a2</i> qPCR Forward primer GTGGAGTTTGAGCCCACGTA	This paper	N/A
m. <i>Atp6v0a2</i> qPCR Reverse primer AGGCCAGAAACGAATCCCAG	This paper	N/A
m. <i>Atp6v0c</i> qPCR Forward primer TTGGCTAGAACTTCTTTCAC	This paper	N/A
m. <i>Atp6v0c</i> qPCR Reverse primer CACGGAGATGAGGAAGAAGC	This paper	N/A
m. <i>Atp6v0d1</i> qPCR Forward primer CTGTGCCGAAAGAGTTGA	This paper	N/A
m. <i>Atp6v0d1</i> qPCR Reverse primer AGAGCAGGACCTTGATGAG	This paper	N/A
m. <i>Atp6v1a</i> qPCR Forward primer GCCCACACTGTAGAACTTGAGAT	This paper	N/A
m. <i>Atp6v1a</i> qPCR Reverse primer TGCACTTAGGACCAGGCACA	This paper	N/A
m. <i>Atp6v1c1</i> qPCR Forward primer ATCCAGTTTAGCCAGTTC	This paper	N/A
m. <i>Atp6v1c1</i> qPCR Reverse primer CAATAATCTTGCCGTCTC	This paper	N/A
m. <i>Atp6v1d</i> qPCR Forward primer GCTTCCCTGCAGACTTCCTT	This paper	N/A
m. <i>Atp6v1d</i> qPCR Reverse primer TAGGCAAGGGTGCGTTCAAT	This paper	N/A
m. <i>Atp6v1e1</i> qPCR Forward primer TGGTAACGGGTCGTATCT	This paper	N/A
m. <i>Atp6v1e1</i> qPCR Reverse primer ATCAAGCAAGGCTCAAAG	This paper	N/A
m. <i>Atp6v1g1</i> qPCR Forward primer GCGGTTCTTTGACGGTGG	This paper	N/A
m. <i>Atp6v1g1</i> qPCR Reverse primer ACGGCCTAAGTCAACTGCTC	This paper	N/A
m. <i>Atp6v1g3</i> qPCR Forward primer AGAGCCACCTCTCGGATGAA	This paper	N/A
m. <i>Atp6v1g3</i> qPCR Reverse primer CGCACACCATGCTCAATAGC	This paper	N/A
m. <i>Vegf</i> qPCR Forward primer GCCGAAGCTCTCCACGATTT	This paper	N/A

m. <i>Vegf</i> qPCR Reverse primer ATTGGGGGAGCATTGGGTT	This paper	N/A
m. <i>Glut-1</i> qPCR Forward primer CAATGGCGGCGGTCTATAA	This paper	N/A
m. <i>Glut-1</i> qPCR Reverse primer CCCTGACGCACTTAAGACCC	This paper	N/A
m. <i>Pai-1</i> qPCR Forward primer GTGATAGATCTGGCCTGAAGCA	This paper	N/A
m. <i>Pai-1</i> qPCR Reverse primer TCTGAATGTATTTGGGTGACTCTGT	This paper	N/A
m. <i>Ccnd-1</i> qPCR Forward primer CCGGCCTCTGGCTAAACAAG	This paper	N/A
m. <i>Ccnd-1</i> qPCR Reverse primer CGCAGGCTTGACTCCAGAAG	This paper	N/A
m. <i>IL-1β</i> qPCR Forward primer ACCTTCCAGGATGAGGACATGA	This paper	N/A
m. <i>IL-1β</i> qPCR Reverse primer CTAATGGGAACGTCACACACCA	This paper	N/A
m. <i>IL-6</i> qPCR Forward primer GTTCTCTGGGAAATCGTGA	This paper	N/A
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m. <i>Hif-1α</i> qPCR Forward primer TCCAAGGAGCCTTAACCTGTC	This paper	N/A
m. <i>Hif-1α</i> qPCR Reverse primer GCGTATGTCAGAAAGTTGGC	This paper	N/A
m. <i>Hif-2α</i> qPCR Forward primer CACTGGCCCATGTCTTCCAT	This paper	N/A
m. <i>Hif-2α</i> qPCR Reverse primer GTGTTGGATCTGCCTCCCAT	This paper	N/A
m. <i>Arg-1</i> qPCR Forward primer CGTGGGTCCAACACCCTTAT	This paper	N/A
m. <i>Arg-1</i> qPCR Reverse primer TGGTCTGCTATTTGCCAGGA	This paper	N/A
m. <i>Fizz-1</i> qPCR Forward primer CAAGACTATGAACAGATGGGCCT	This paper	N/A
m. <i>Fizz-1</i> qPCR Reverse primer AGGAGATTGATGGGAGAGGACA	This paper	N/A
m. <i>Mrc-1</i> qPCR Forward primer ATGGATTGCCCTGAACAGCA	This paper	N/A
m. <i>Mrc-1</i> qPCR Reverse primer TGCTGACTTAAGCTTCGGCT	This paper	N/A
m. <i>β-actin</i> qPCR Forward primer TCTGAATGTATTTGGGTGACTCTGT	This paper	N/A

m. <i>β-actin</i> qPCR Reverse primer AACGCAGCTCAGTAACAGTCC	This paper	N/A
TFEB/Atp6v0d2 chip forward primer AAACTGGGCATGACCTTG	This paper	N/A
TFEB/Atp6v0d2 chip reverse primer GCAGCTCAGAAGGCACA	This paper	N/A