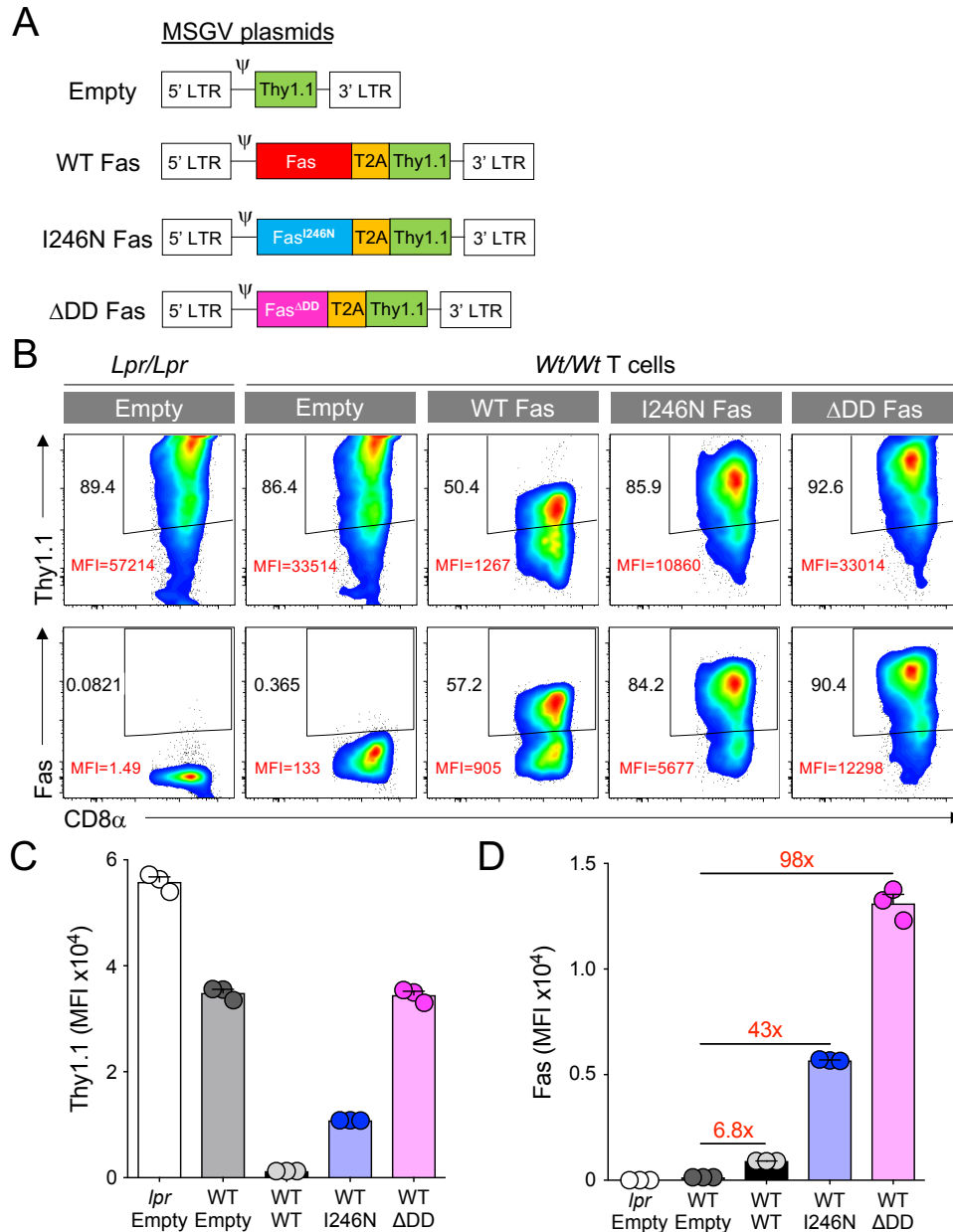


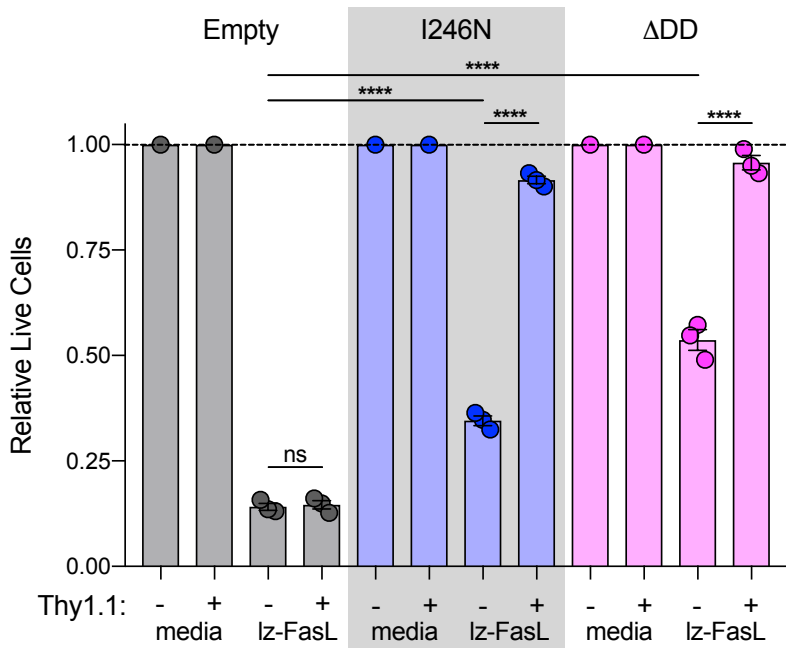
SUPPLEMENTARY TABLES AND LEGENDS:

Supplemental Figure 1:



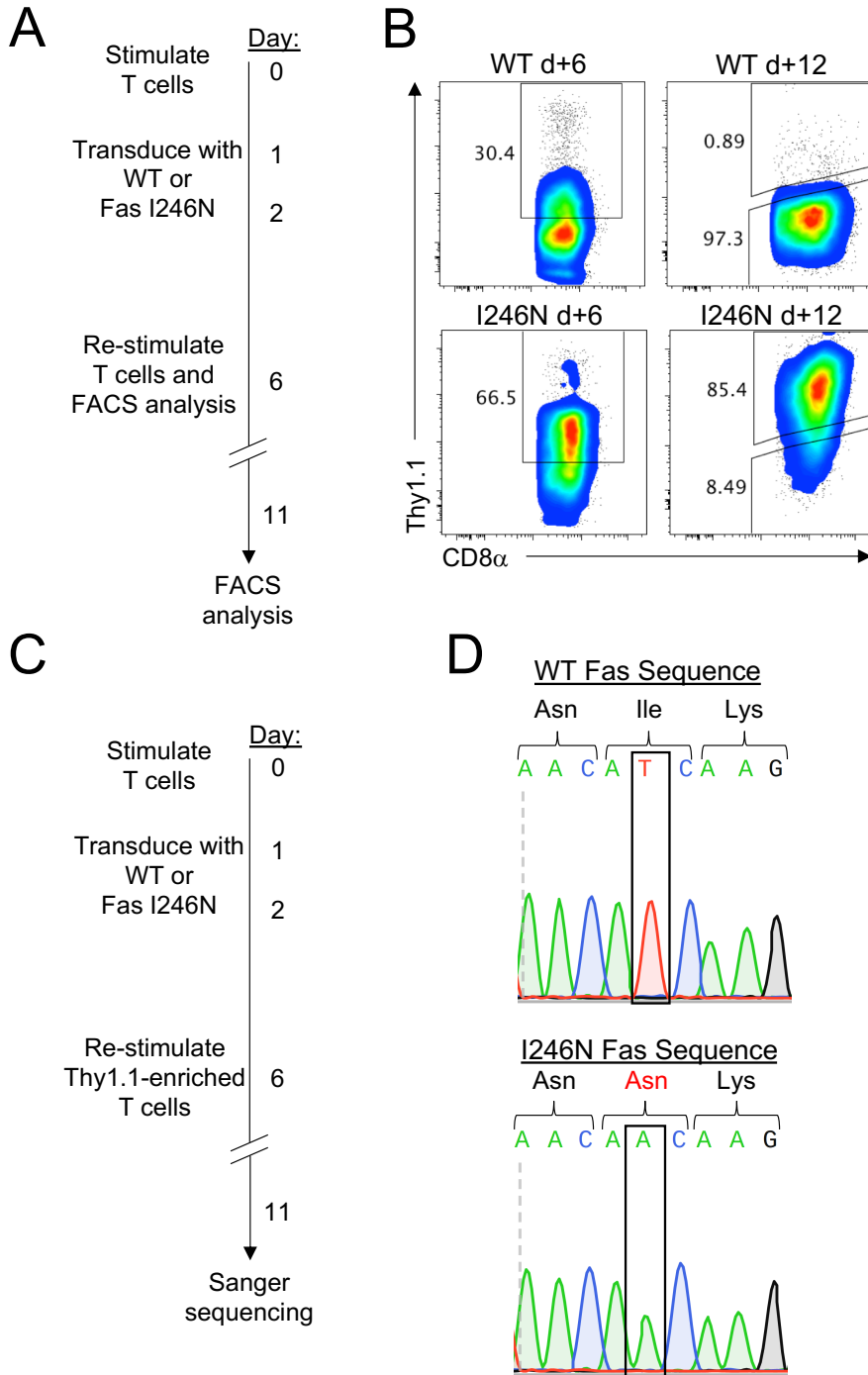
Supplemental Figure 1: Design and expression of retrovirally-encoded Fas DNR constructs and controls in mouse CD8⁺ T cells. (A) Schematic overview of the designs for retroviral constructs encoding murine wildtype (WT) Fas or mutant versions of Fas impaired in their ability to bind the intracellular adapter molecule Fas-associated via death domain. WT Fas, Fas^{I246N}, or Fas^{ΔDD} were cloned into an MSGV1 expression vector in front of a T2A cleavage site and the Thy1.1 reporter gene. An empty vector containing only the Thy1.1 reporter gene (Empty) was used as a negative control. (B) Representative FACS plots and summary bar graphs of (C) Thy1.1 and (D) Fas expression 4d following retroviral transduction of Fas-deficient B6-*lpr* or WT CD8⁺ T cells. The percentage of gated Thy1.1⁺ or Fas⁺ cells is shown in black, MFI of Thy1.1⁺ or Fas⁺ cells is shown in red (C). Fold change in Fas MFI over endogenous, WT Fas expression in WT, Empty-transduced cells is shown in red in (D). Data in (C) and (D) are displayed as mean ± SEM with *n*=3 per condition and is representative of 12 independent experiments.

Supplemental Figure 2:



Supplemental Figure 2: Fas DNRs can protect non-transduced cells from FasL-mediated apoptosis. Summary bar graph showing the relative frequency of cell viability of non-transduced and transduced T cells after 20h following exposure to lz-FasL (100 ng mL⁻¹). Results shown after gating on live CD8α⁺ lymphocytes, and viability shown relative to the media for each transduction condition. Data shown is representative of 3 independently performed experiments and is displayed as mean ± SEM with *n* = 3 per condition. *****P*<0.0001, ns = not significant (one-way ANOVA, corrected with Tukey's multiple comparisons).

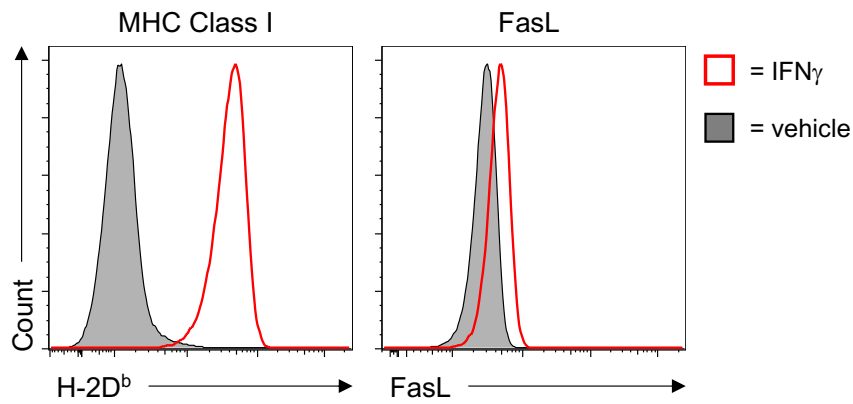
Supplemental Figure 3:



Supplemental Figure 3: Expression of FasI246N in T cells does not cause reversion to WT Fas. (A)

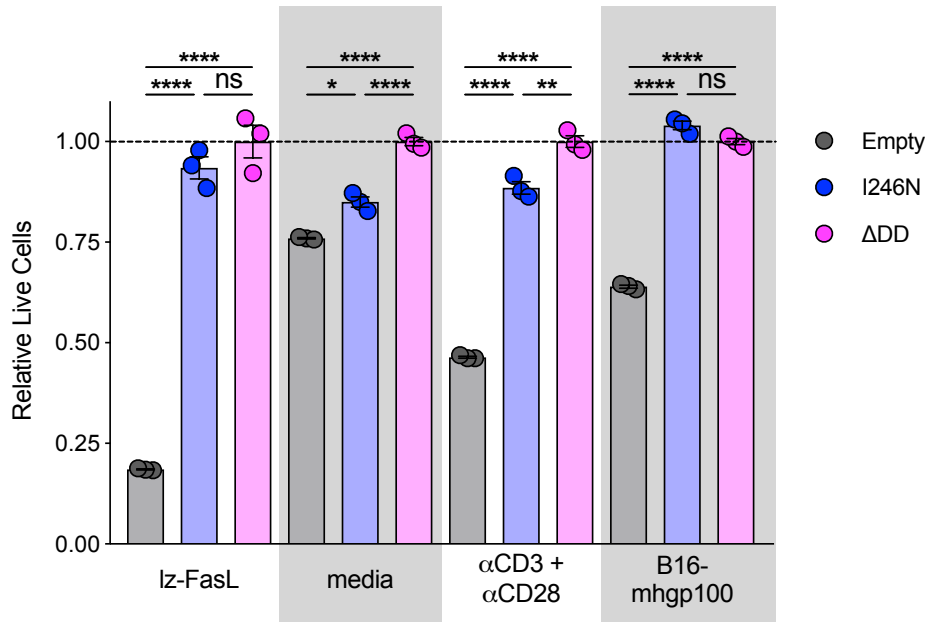
Experimental timeline for the stimulation, retroviral transduction, and analysis of WT CD8 α^+ T cells modified with Fas^{WT} or Fas^{I246N}. **(B)** Representative FACS plots of Thy1.1 expression at days 6 and 12 for Fas^{WT} or Fas^{I246N} transduced cells. **(C)** Experimental timeline for the stimulation, transduction, Thy1.1-enrichment, and sequencing of WT CD8 α^+ T cells modified with Fas^{WT} or Fas^{I246N}. **(D)** Representative sequencing data showing WT Fas maintains the A-T-C sequence encoding the isoleucine at amino acid position 246, whereas the Fas^{I246N} sequence is A-A-C, encoding an asparagine at amino acid position 246 in the introduced Fas DNR construct.

Supplemental Figure 4:



Supplemental Figure 4: IFN γ upregulates FasL on surface of B16 tumor cells. B16 cells were treated with vehicle (PBS) or IFN γ (100 ng mL⁻¹) for 24 hours, then analyzed for surface expression of MHC Class I (H-2D^b; left panel) or FasL (right panel) by flow cytometry.

Supplemental Figure 5:



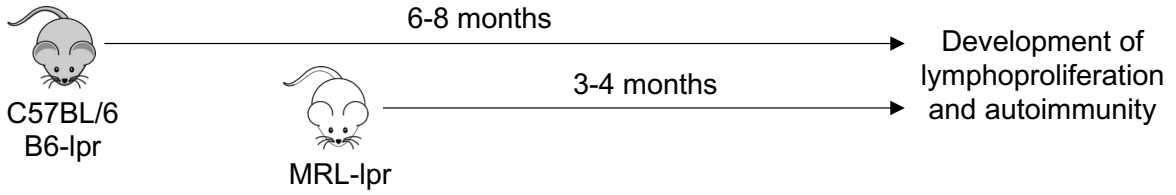
Supplemental Figure 5: T cells engineered with Fas DNRs prevent apoptosis from various stimuli.

Summary bar graph showing the relative frequency of cell viability of transduced T cells after 20h following exposure to lz-FasL (100 ng mL⁻¹). Results are shown after gating on Thy1.1⁺ cells, and viability is shown relative to Fas^{ΔDD}. Data shown is representative of 10 independently performed experiments and is displayed as mean ± SEM with *n* = 3 per condition. **P*<0.05 ***P*<0.01

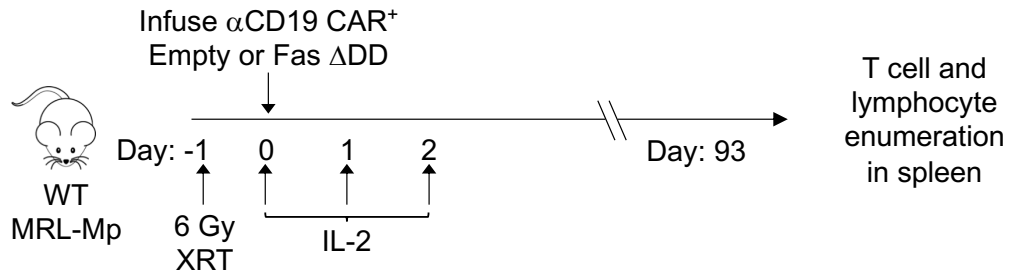
*****P*<0.0001, ns = not significant (one-way ANOVA, corrected with Tukey's multiple comparisons).

Supplemental Figure 6:

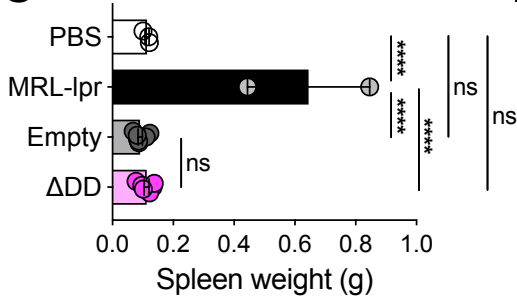
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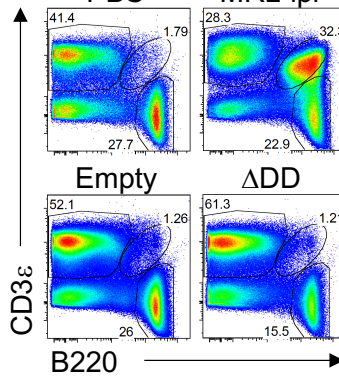
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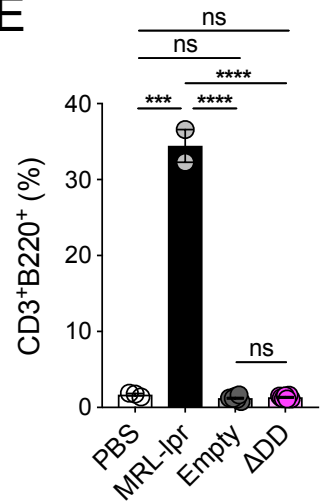
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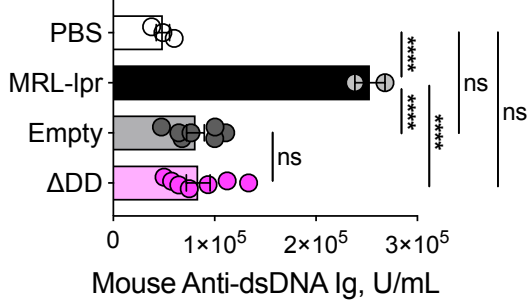
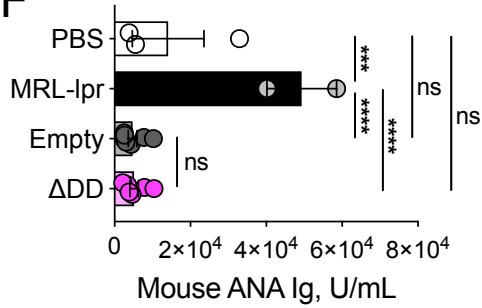
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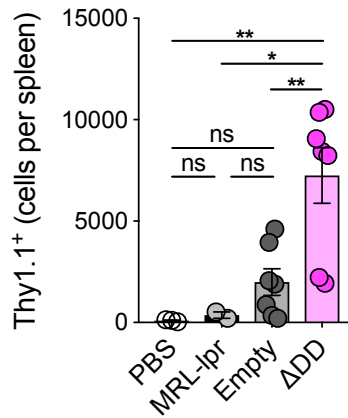
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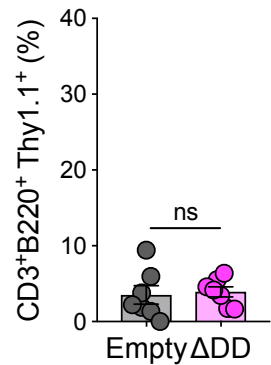
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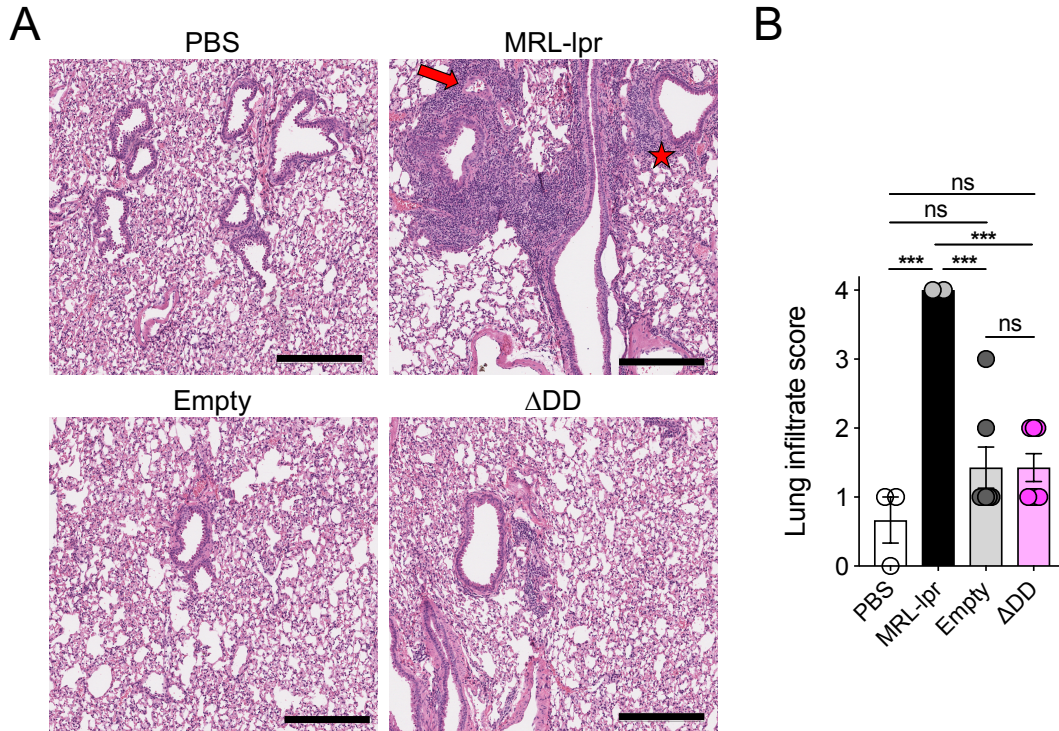


H



Supplemental Figure 6. Fas DNR expression does not induce lymphoproliferation in the ALPS-susceptible MRL strain. (A) Schematic comparing the onset of lymphoproliferation in C57BL/6 B6-lpr mice at 6-9 months (top) to the MRL-lpr strain at 3-4 months. (B) Experimental design to analyze long-term persistence of WT anti-CD19 CAR expressing CD8 α ⁺ T cells modified with Fas^{ADD} or empty vector control in WT MRL-Mp mice. A total of 3x10⁶ of anti-CD19 CAR⁺ CD8 α ⁺ T cells were infused i.v. into sublethally irradiated (6 Gy XRT) mice. Recipient mice received IL-2 by daily i.p. injection for 3d and the spleens were harvested for analysis after 93d. (C) Summary numbers of spleen weight in recipient mice, compared to age-matched wild type mice and Fas-deficient B6-lpr mice (negative and positive controls, respectively). (D, E) Representative FACS plots (D) and (E) summary bar graph of the frequency of CD3⁺B220⁺ double negative lymphocytes in the spleens of recipient and control mice. (F) Summary bar graphs of levels of anti-nuclear antibody (ANA) Ig (top) and anti-dsDNA Ig (bottom) as measured by ELISA. (G, H) Summary bar graphs demonstrating the persistence (G) and surface phenotype (H) of transferred Thy1.1⁺ T cells modified with Fas^{ADD} DNR or an empty vector control. *n*=2-7 mice per cohort. *****P*<0.0001, ****P*<0.001, ***P*<0.01, **P*<0.05, ns = not significant (one-way ANOVA, corrected with Tukey's multiple comparisons).

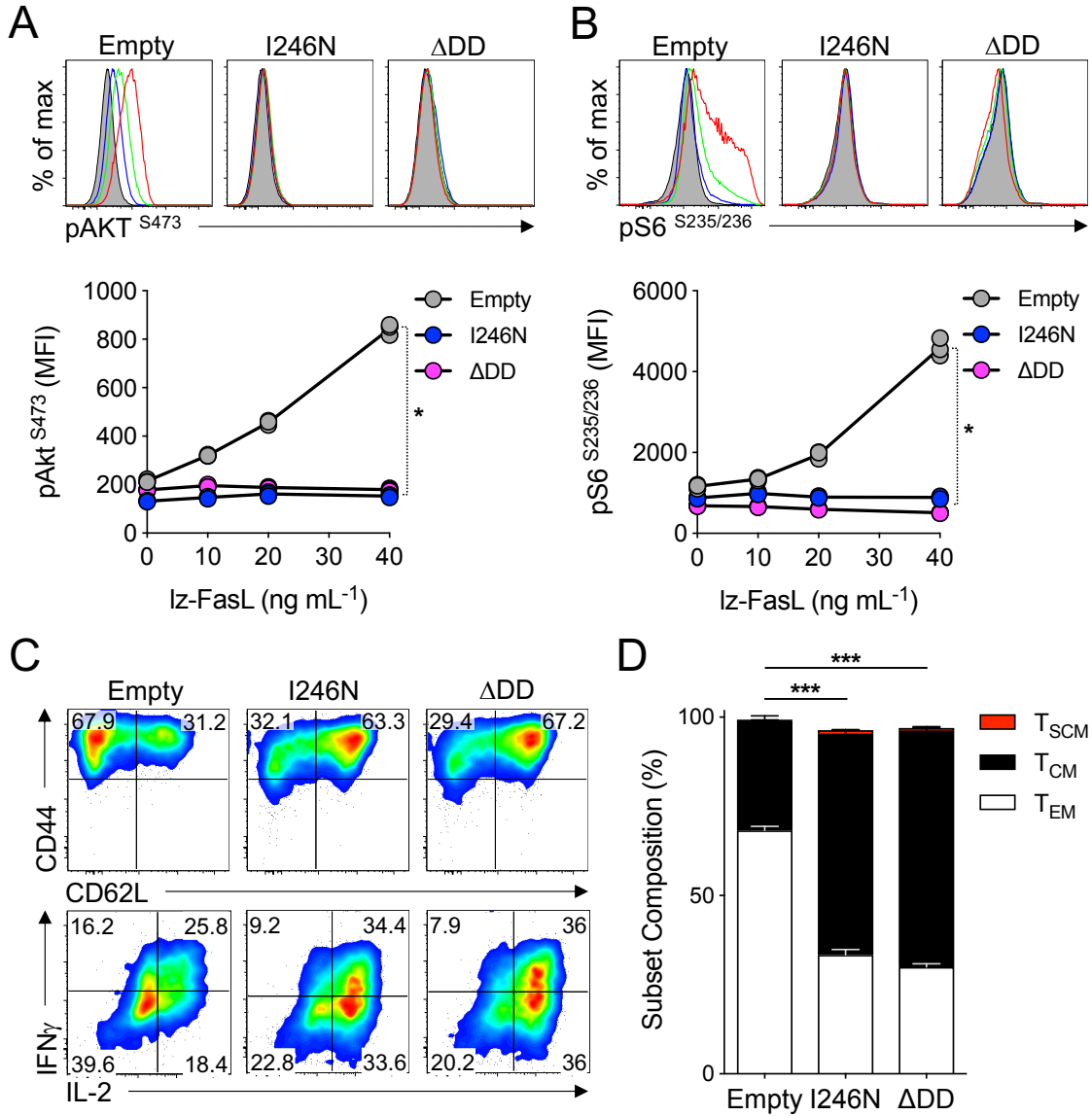
Supplemental Figure 7:



Supplemental Figure 7: Adoptively transferred T cell modified with Fas DNR do not induce an inflammatory infiltrate in the lungs of ALPS-susceptible MRL host mice. (A) Representative H&E stained micrographs and (B) summary graph demonstrating the intensity of inflammatory mononuclear cell infiltrates in the lungs of treated mice. The red arrow and red star point to areas of dense peri-vascular and peri-bronchiolar mononuclear inflammatory infiltrates, respectively. Scale bar = 300 μ m. All images were scored in a blinded fashion by an interpreting pathologist. *** P <0.001, ns = not significant (one-way ANOVA, corrected with Tukey's multiple comparisons).

Supplemental Figure 8:

Iz-FasL (ng/mL): ■ = 0 □ = 10 □ = 20 □ = 40



Supplemental Figure 8: Fas DNRs prevent Iz-FasL-induced Akt activation and T cell differentiation. (A, B) Representative FACS histograms (top) and summary plot (bottom) of the dose-response relationship between Iz-FasL exposure and (A) phospho- (p) Akt^{S473} and (B) pS6^{S235/236} in CD8 α^+ T cells transduced with Fas^{I246N}, Fas ^{Δ DD}, or empty vector control. Results shown 6d after activation, retroviral transduction, and expansion in the continuous presence of indicated concentrations of Iz-FasL. (C) Representative FACS plots of T cell differentiation (top) and intracellular IFN γ /IL-2 production (bottom) 11d after CD8 α^+ T cells were transduced Fas^{I246N}, Fas ^{Δ DD}, or empty vector control in the absence of exogenous FasL. Intracellular cytokine staining measured after ~5h incubation with PMA/ionomycin in brefeldin A and monensin. (D) Memory T cell subset composition of CD8 α^+ T cells 11d after activation, transduction, and expansion in culture. Graphs displayed as mean \pm SEM with $n=3$ per condition and is representative of 3 (A, B) and 5 (C, D) independent experiments. * $P<0.05$, (Wilcoxon rank sum test).

GTEX Tissue Type	TCGA Cancer Subtype	TCGA Abbr
Adrenal Gland	Adrenocortical Cancer	ACC
Bladder	Bladder Urothelial Carcinoma	BLCA
Whole Blood	Acute Lymphoblastic Leukemia	ALL
Brain - Amygdala Brain - Anterior Cingulate Cortex (Ba24) Brain - Caudate (Basal Ganglia) Brain - Cerebellar Hemisphere Brain - Cerebellum Brain - Cortex Brain - Frontal Cortex (Ba9) Brain - Hippocampus Brain - Hypothalamus Brain - Nucleus Accumbens (Basal Ganglia) Brain - Putamen (Basal Ganglia) Brain - Spinal Cord (Cervical C-1) Brain - Substantia Nigra	Brain Lower Grade Glioma	LGG
Brain - Amygdala Brain - Anterior Cingulate Cortex (Ba24) Brain - Caudate (Basal Ganglia) Brain - Cerebellar Hemisphere Brain - Cerebellum Brain - Cortex Brain - Frontal Cortex (Ba9) Brain - Hippocampus Brain - Hypothalamus Brain - Nucleus Accumbens (Basal Ganglia) Brain - Putamen (Basal Ganglia) Brain - Spinal Cord (Cervical C-1) Brain - Substantia Nigra	Glioblastoma Multiforme	GBM
Breast - Mammary Tissue	Breast Invasive Carcinoma	BRCA
Cervix - Ectocervix Cervix - Endocervix	Cervical & Endocervical Cancer	CESC
Kidney - Cortex	Kidney Clear Cell Carcinoma	KIRC
Kidney - Cortex	Kidney Renal Papillary Cell Carcinoma	KIRP
Colon - Sigmoid Colon - Transverse	Colon Adenocarcinoma	COAD
Colon - Sigmoid	Rectum Adenocarcinoma	READ
Esophagus - Gastroesophageal Junction Esophagus - Mucosa	Esophageal Carcinoma	ESCA
Liver	Liver Hepatocellular Carcinoma	LIHC
Lung	Lung Adenocarcinoma	LUAD
Lung	Lung Squamous Cell Carcinoma	LUSC
Spleen	Diffuse Large B-Cell Lymphoma	DLBC
Adipose - Subcutaneous Artery - Tibial Nerve - Tibial Muscle - Skeletal	Sarcoma	SARC
Fallopian Tube Ovary	Ovarian Serous Cystadenocarcinoma	OV
Prostate	Prostate Adenocarcinoma	PRAD
Pancreas	Pancreatic Adenocarcinoma	PAAD
Skin - Not Sun Exposed (Suprapubic) Skin - Sun Exposed (Lower Leg)	Skin Cutaneous Melanoma	SKCM
Stomach	Stomach Adenocarcinoma	STAD
Testis	Testicular Germ Cell Tumor	TGCT
Thyroid	Thyroid Carcinoma	THCA
Uterus	Uterine Carcinosarcoma	UCS
Uterus	Uterine Corpus Endometrioid Carcinoma	UCEC

Supplemental Table 1: Summary of TCGA cancer subtypes and matched tissues of origin from the GTEx database. A list of the normal tissues of origin and corresponding cancer subtypes ($n=26$) were obtained from the GTEx and TCGA datasets, respectively.

Gene	<i>r</i>	Gene	<i>r</i>	Gene	<i>r</i>	Gene	<i>r</i>	Gene	<i>r</i>
SLA2	0.8580	CCL5	0.6757	NCKAP1L	0.6194	ZAP70	0.5731	PTPRCAP	0.5407
CD8A	0.8539	ARHGAP9	0.6747	LILRB1	0.6157	NCR1	0.5723	CD96	0.5387
CCR5	0.8510	KLRD1	0.6742	SIT1	0.6144	MS4A6A	0.5721	STAT1	0.5375
CD2	0.8440	SLFN12L	0.6739	GNGT2	0.6142	LCK	0.5720	FCER1G	0.5374
NKG7	0.8394	ARHGAP30	0.6597	C1QA	0.6137	ARHGAP15	0.5680	CST7	0.5370
GZMA	0.8383	ZNF683	0.6593	TNFAIP8L2	0.6107	CD86	0.5667	IGFLR1	0.5366
KLRK1	0.8337	IL10RA	0.6583	APOL3	0.6092	LAIR1	0.5656	TRAF3IP3	0.5364
CRTAM	0.8205	IL18BP	0.6576	FCGR1B	0.6073	GBP1	0.5605	HLA-DMA	0.5362
CXCR6	0.8097	TRAT1	0.6512	TLR8	0.6069	CD200R1	0.5605	CYBB	0.5360
SIRPG	0.8039	ABCD2	0.6506	DOCK2	0.6052	CD4	0.5594	LAT	0.5342
IFNG	0.8036	GPR65	0.6502	IKZF1	0.6031	GBP2	0.5590	TNFRSF1B	0.5342
UBASH3A	0.8005	SASH3	0.6493	LTA	0.6025	ABI3	0.5580	ITGB2	0.5334
EOMES	0.8000	CD6	0.6478	ARHGAP25	0.6021	HLA-E	0.5577	CD3G	0.5329
PRF1	0.7911	SLAMF7	0.6440	HLA-DPB1	0.6015	PLEK	0.5570	TBC1D10C	0.5304
CD247	0.7882	CYTH4	0.6436	TTC24	0.6014	LAPTM5	0.5557	TRIM22	0.5290
PYHIN1	0.7822	FAM78A	0.6416	C1QC	0.5971	SAMHD1	0.5553	JAK3	0.5289
CD3E	0.7818	PTPRC	0.6414	GIMAP2	0.5962	ZNF80	0.5547	CIITA	0.5288
CD3D	0.7780	XCL2	0.6397	BTN3A2	0.5936	CORO1A	0.5542	B2M	0.5280
LAG3	0.7708	RASAL3	0.6395	WIPF1	0.5896	IL16	0.5531	GAB3	0.5265
GZMH	0.7693	CD74	0.6386	TIFAB	0.5893	CLEC2D	0.5506	SIGLEC10	0.5264
CCL4	0.7582	IRF1	0.6385	GIMAP4	0.5892	C5orf56	0.5503	VAV1	0.5263
CXCR3	0.7568	SEPT1	0.6366	TNFRSF9	0.5889	GBP5	0.5494	TAP1	0.5254
GPR174	0.7559	ITK	0.6359	APOBEC3H	0.5887	LILRB2	0.5467	CXorf21	0.5239
GZMK	0.7358	CD53	0.6358	FCGR3A	0.5885	CARD16	0.5464	CD160	0.5232
TIGIT	0.7333	BIN2	0.6340	IL18RAP	0.5873	HLA-DQA1	0.5463	NCF1	0.5203
ITGAL	0.7237	GRAP2	0.6332	CCR2	0.5869	P2RY13	0.5462	GIMAP5	0.5200
IL12RB1	0.7188	AC008964.1	0.6305	CD48	0.5851	HLA-DOA	0.5458	PTPN7	0.5199
LCP2	0.7133	MYO1F	0.6296	CD72	0.5851	FMNL1	0.5452	FERMT3	0.5190
PDCD1	0.7126	IL21R	0.6285	LAP3	0.5846	SCIMP	0.5446	LST1	0.5189
SAMD3	0.7113	BTN3A3	0.6265	APOBEC3D	0.5836	C1orf162	0.5446	ITGAE	0.5188
FAM26F	0.7050	ICOS	0.6242	SELPLG	0.5808	IGSF6	0.5438	IL2RB	0.5180
SNX20	0.6990	PVRIG	0.6240	DOK2	0.5803	PSMB9	0.5435	SAMSN1	0.5160
CTSW	0.6979	SLAMF8	0.6235	AD000671.6	0.5802	EVI2A	0.5434	BTN2A2	0.5152
FCRL6	0.6945	C1QB	0.6235	AIF1	0.5797	HLA-DRB1	0.5421	GMFG	0.5139
PSTPIP1	0.6945	HLA-DPA1	0.6229	SLA	0.5780	FYB	0.5420	GIMAP7	0.5119
HCST	0.6833	HLA-DRA	0.6224	APOBEC3G	0.5778	PARVG	0.5419	APOL6	0.5112
SLAMF6	0.6826	TBX21	0.6219	GBP4	0.5776	P2RY10	0.5414	NLRC5	0.5099
SPN	0.6803	BTN3A1	0.6217	ACAP1	0.5762	LILRB4	0.5412	LY9	0.5092
CXCL9	0.6795	FCGR1A	0.6216	SP140	0.5752	WAS	0.5410	GPR31	0.5087
KLRC4-KLRK1	0.6773	KLRC4	0.6200	EVI2B	0.5744	C15orf53	0.5408	AKNA	0.5087

Supplemental Table 2: A pan-cancer analysis of the top 200 genes positively correlated with *FASLG* across the TCGA dataset. Gene names and Pearson's correlation coefficient (*r*) to *FASLG* are listed.