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Research Article

Once infected by obligate intracellular pathogens, monocytes/macrophages release cytokines that activate natural killer (NK) cells. NK cells in turn produce and secrete monocyte/macrophage activating factors such as interferongamma (IFN-gamma), which are important in the early control of these infections. Here we demonstrate that human NK cells are potent producers of another monocyte/macrophage-activating factor, macrophage inflammatory protein-1 alpha (MIP-1 alpha). Fresh NK cells produce negligible amounts of MIP-1 alpha after stimulation with the monocyte-derived cytokines IL-12, TNF-alpha, IL-1 beta, or IL-10, while stimulation with IL-15 alone results in modest MIP-1 alpha production. Abundant NK cell production MIP-1 alpha is seen after costimulation with IL-12 and IL-15, and is dose-dependent. Combinations of IL-12, with TNF-alpha, IL-1 beta, or IL-10 are substantially less effective inducers of MIP-1 alpha production by NK cells. NK cell MIP-1 alpha mRNA transcripts were detectable within 1 h after costimulation with IL-12 plus IL-15 and steadily increased over 24 h, with a concomitant increase in protein production detectable at 12 h. Resting NK cells constitutively express mRNA transcript for a MIP-1 alpha receptor, and costimulation with IL-12 and IL-15 upregulates its level of expression. Equilibrium binding studies with radioiodinated MIP-1 alpha were consistent with the induction of a single class of high affinity MIP-1 alpha receptors on NK cells costimulated with IL-12 and IL-15. Addition of exogenous MIP-1 [...]

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Human Natural Killer Cells Produce Abundant Macrophage Inflammatory Protein-1 α in Response to Monocyte-derived Cytokines

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Abstract

Once infected by obligate intracellular pathogens, monocytes/macrophages release cytokines that activate natural killer (NK) cells. NK cells in turn produce and secrete monocyte/macrophage activating factors such as interferon-gamma (IFN- γ), which are important in the early control of these infections. Here we demonstrate that human NK cells are potent producers of another monocyte/macrophage-activating factor, macrophage inflammatory protein-1 α (MIP-1 α). Fresh NK cells produce negligible amounts of MIP-1 α after stimulation with the monocyte-derived cytokines IL-12, TNF- α , IL-1 β , or IL-10, while stimulation with IL-15 alone results in modest MIP-1 α production. Abundant NK cell production of MIP-1 α is seen after costimulation with IL-12 and IL-15, and is dose-dependent. Combinations of IL-12 with TNF- α , IL-1 β , or IL-10 are substantially less effective inducers of MIP-1 α production by NK cells. NK cell MIP-1 α mRNA transcripts were detectable within 1 h after costimulation with IL-12 plus IL-15 and steadily increased over 24 h, with a concomitant increase in protein production detectable at 12 h. Resting NK cells constitutively express mRNA transcript for a MIP-1 α receptor, and costimulation with IL-12 and IL-15 upregulates its level of expression. Equilibrium binding studies with radioiodinated MIP-1 α were consistent with the induction of a single class of high affinity MIP-1 α receptors on NK cells costimulated with IL-12 and IL-15. Addition of exogenous MIP-1 α to resting NK cells did not enhance cytokine production, but did increase NK cytotoxic activity. The requirement for IL-15 as a critical cofactor for NK cell production of MIP-1 α suggests a potentially unique role for this monocyte-derived cytokine in combination with IL-12. As MIP-1 α is known to potentiate the actions of IFN- γ on monocytes and to suppress human immunodeficiency virus replication, the NK cell's production of MIP-1 α may be important during the innate immune response to infection. (*J. Clin. Invest.* 1996; 97: 2722–2727.) Key words: chemokine • innate immunity • IL-15 • IL-12 • monokine

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Introduction

Natural killer (NK)¹ cells are innate immune effector cells thought to be important during the early stages of infection. Monocytes infected with obligate intracellular pathogens such as *Listeria monocytogenes*, *Toxoplasma gondii*, and *Leishmania major* release cytokines which in turn activate NK cells to secrete interferon gamma (IFN- γ), tumor necrosis factor-alpha (TNF- α), and granulocyte-macrophage colony stimulating factor (GM-CSF) in a positive feedback loop. Indeed, a number of experimental immunodeficient animal models have clearly demonstrated the importance of NK cell IFN- γ production in preventing overwhelming infection from several obligate intracellular pathogens (reviewed in references 1 and 2). Additionally, NK cell production of both IFN- γ and TNF- α appears to play a role in the pathogenesis of septic shock (3, 4). The number of NK-derived monocyte-activating factors that may contribute to protection from microbial invasion and/or septic shock is currently unknown.

Macrophage inflammatory protein-1 α (MIP-1 α) is a member of the chemokine superfamily of low molecular mass secretory proteins with phagocyte-stimulating and proinflammatory properties. MIP-1 α has been shown to potentiate monocyte/macrophage activation induced by IFN- γ (5) and most recently, to suppress replication by the human immunodeficiency virus (HIV) (6). Here we provide the first evidence that human NK cells produce MIP-1 α after costimulation with a specific combination of monocyte-derived cytokines. Further, we characterize MIP-1 α production, receptor expression and binding, as well as the consequences of its binding on human NK cell function.

Methods

Isolation of CD3⁻CD56⁺ human cells and cell lines. Human NK cells were isolated from fresh blood to $\geq 97\%$ purity using immunomagnetic bead depletion followed by sorting for CD56⁺ NK cells or CD56^{bright} NK cells on a FACstar Plus[®] (Becton Dickinson, San Jose, CA) as previously described (7). The COLO 205 cell line was obtained from the ATCC (Rockville, MD) and the NK-92 cell line was a generous gift of Dr. H.-G. Klingemann (8).

Cytokines, NK cytokine production, and monoclonal antibodies (mAbs). Recombinant human (rh) IL-15 was provided by Immunex Corp. (Seattle, WA). rhIL-12 (sp act 4.5×10^6 U/mg) was provided by Genetics Institute (Cambridge, MA). rhIL-10 (sp act 11.0×10^6 IRU/mg) was provided by Schering-Plough (Kenilworth, NJ). rhTGF- β_2 (TGF- β) was a generous gift of Dr. Thomas B. Tomasi

1. *Abbreviations used in this paper:* MIP, macrophage inflammatory protein; NK, natural killer.

(Roswell Park Cancer Institute, Buffalo, NY). rhTNF- α was provided by Asahi Brewing Co. (Tokyo, Japan, sp act 2×10^3 U/mg) and rhIL-1 β was purchased from Pharmacia Biotech, Inc. (Piscataway, NJ). rhMIP-1 α was purchased from Biosource International (Carmalito, CA). rhIL-2 (sp act 1.53×10^7 IU/mg) was provided by Hoffmann-La Roche (Nutley, NJ). All cytokines used in this study were diluted to working concentrations in 0.1% human albumin (Armor Pharmaceuticals, Kankakee, IL) in RPMI 1640 (Sigma Diagnostics, St. Louis, MO). Anti-CD25 (anti-IL-2R α) and non-reactive isotype control mAbs were purchased from Biosource International.

For all assays of NK cell cytokine production, 2×10^4 purified ($\geq 97\%$) fresh human NK cells were plated in 96-round bottom wells containing 250 μ l of RPMI 1640 medium (Sigma Chemical Co., St. Louis, MO) supplemented with 10% human AB serum (C-Six Diagnostics, Mequon, WI), antibiotics, and various amounts of rh cytokines as indicated. Cultures were incubated for 72 h at 37°C unless otherwise indicated, at which time 200 μ l of supernatant was removed for measurement of MIP-1 α production without disturbing the cell pellet. Measurement of human MIP-1 α production was performed by ELISA (sensitivity: 2.0 pg/ml, R & D Systems, Minneapolis, MN) following the manufacturer's instructions.

Analysis of RNA. 2.0×10^7 NK cells ($\geq 85\%$ CD3 $^-$ CD56 $^+$) were isolated from fresh leukopacs using immunomagnetic bead depletion and either placed immediately in RNazol (Biotex Laboratories, Friendswood, TX), or incubated with IL-12 (10 U/ml) and/or IL-15

(10 ng/ml) at 37°C for indicated times, followed by lysis in RNazol. Total RNA was extracted according to the manufacturer's recommendation, and Northern blot analysis was performed as previously described (9). The GOS19.1 cDNA (a generous gift of Dr. D.R. Forsdyke, Queens University, Hamilton, Ontario), is the full length 1 kb cDNA encoding the human MIP-1 α protein (10). The C-C CKR-1 cDNA (a generous gift of Dr. T. Schall, DNAX Research Institute, Palo Alto, CA) is a 1 kb cDNA encoding a known protein receptor for MIP-1 α (11). Both probes were labeled with the Prime It $^{\text{®}}$ II random primer labeling kit (Stratagene Inc., LaJolla, CA) as per the manufacturer's instructions, prior to hybridization.

^{125}I -MIP-1 α binding and unlabeled ligand competition. NK cells ($\geq 85\%$ CD3 $^-$ CD56 $^+$) were isolated from fresh leukopacs using immunomagnetic bead depletion, and incubated in medium alone or medium with IL-15 (10 ng/ml) and IL-12 (10 U/ml) for 8 h, followed by overnight shipment at room temperature under otherwise identical culture conditions. Upon receipt, NK cells were first checked for viability by trypan blue ($\geq 90\%$), and tested for specific binding to radiolabeled ^{125}I -MIP-1 α as described (12). Briefly, the NK cells were washed twice with PBS (pH 7.4) and resuspended at 2.0×10^6 cells/ml in binding buffer consisting of Iscove's modified Dulbecco's medium (IMDM) containing 25 mmol/liter HEPES and 4 mg/ml BSA (pH 7.4). The cells were incubated for 1 h at 23°C with varying concentrations of ^{125}I -MIP-1 α , ranging from 0.01 to 2 nmol/liter, in the presence or absence of a 100-fold excess of unlabeled MIP-1 α in a final volume of

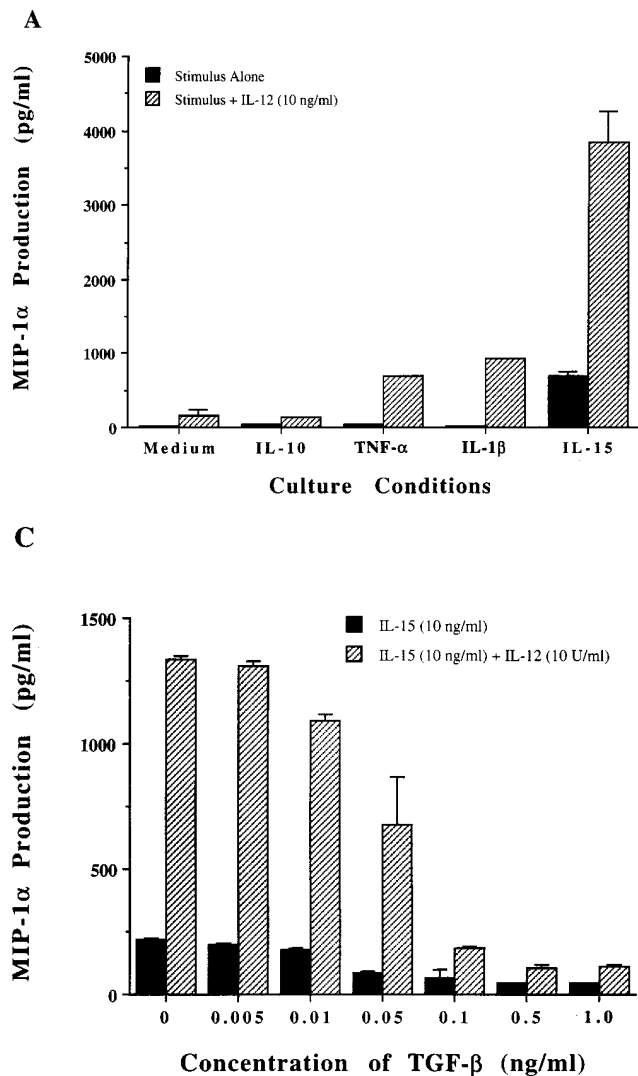


Figure 1. NK cell production of MIP-1 α in response to monocyte-derived cytokines. Human NK cells ($\geq 97\%$) were plated at 2×10^4 cells per well in medium containing various rh cytokines. After 72 h of culture, supernatants were harvested and analyzed for MIP-1 α protein by ELISA. Values represent mean \pm SE and are representative of three independent experiments. (A) MIP-1 α production by CD56 $^+$ cells cultured with IL-10 (50 ng/ml); TNF- α (2000 U/ml); IL-1 β (100 ng/ml); IL-15 (100 ng/ml) in the presence or absence of IL-12 (10 U/ml). (B) NK cell MIP-1 α production in response to increasing concentrations of IL-15, in the presence and absence of IL-12. (C) The suppressive effect of TGF- β on MIP-1 α production by CD56 $^+$ NK cells is dose dependent. Increasing concentrations of TGF- β were added to cultures containing 10 ng/ml IL-15, with or without 10 U/ml IL-12.

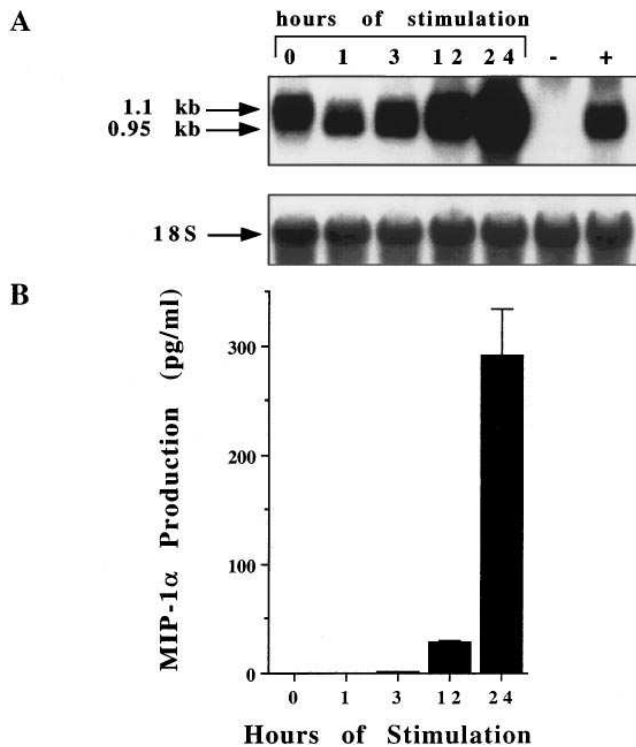


Figure 2. Kinetics of MIP-1 α gene expression in human NK cells after costimulation with IL-12 and IL-15. Freshly isolated ($\geq 85\%$) NK cells were cultured in the presence of IL-12 and IL-15 for the indicated lengths of time. Cells were processed for total RNA and supernatants were collected for protein measurements. (A) Representative Northern analysis for the induction of MIP-1 α gene expression over time. Resting NK cells expressed a 1.1-kb transcript previously only seen in cycloheximide-treated T cells, while the 0.95-kb transcript typically identified by the GOS19.1 probe is absent. After costimulation with IL-12 (10 U/ml) and IL-15 (10 ng/ml), the 0.95-kb transcript appears at 1 h and steadily increases, while the 1.1-kb transcript is absent at 1 h and then increases thereafter. (-) RNA from the COLO 205 cell line; (+) RNA from the NK-92 cell line stimulated for 3 h with IL-12 and IL-15. Autoradiographs were exposed to blots for 20 h. The same blot was reprobbed for 18 S rRNA (18 S) to confirm equal loading of each lane. (B) MIP-1 α protein production of NK cells over time. 2×10^4 NK cells isolated from the same source as those in A were plated into a 96-well plate with IL-12 and IL-15. Their supernatants were assayed for MIP-1 α production by ELISA at the indicated times.

400 μ l. After incubation, 0.75 ml of an ice-cold mixture of 75% FBS in binding buffer was added to the cells, the tubes were vortexed, and the cells were pelleted by centrifugation. The supernatant fluids were aspirated and the radioactivity contained in the cell pellets was measured in a gamma counter. All binding experiments were performed in duplicate. Specific binding was defined as the amount of binding competed by a 100-fold excess of unlabeled MIP-1 α . Equilibrium binding data were analyzed according to the method of Scatchard (13) and by weighted nonlinear least-squares curve fitting as described by Munson and Rodbard using the LIGAND program (14). Objective statistical criteria (F test, extra sum squares principle) were used to evaluate the goodness of fit and for discriminating between receptor binding models.

NK cell cytotoxicity assay. Purified ($\geq 97\%$) NK cells were cultured overnight in V-bottom 96-well plates with indicated concentrations of MIP-1 α . After overnight incubation at 37°C, ^{51}Cr -labeled NK-resistant COLO 205 target cells were added to each well to give effector:

target of 2.5. After a 4-h incubation, supernatants were harvested and specific cell lysis was calculated as previously described (15).

Results

Costimulation of NK cells with specific monocyte-derived factors induces MIP-1 α production by NK cells. In an effort to determine which, if any, monocyte-derived cytokines would stimulate the production of MIP-1 α by NK cells, we treated purified ($\geq 97\%$) CD56 $^+$ NK cells with several rh cytokines known to modulate the inflammatory response. Fig. 1 shows that culture of human NK cells in the presence of IL-10, IL-1 β , IL-12, or TNF- α did not stimulate significant production of MIP-1 α , compared with that seen with medium alone. Culture of NK cells with 100 ng/ml IL-15 alone resulted in modest production of MIP-1 α (500 pg/ml), comparable to the costimulation of NK cells with IL-12 and IL-1 β , or IL-12 and TNF- α (Fig. 1 A). However the costimulation of NK cells with IL-12 and IL-15 had a clear synergistic effect. Under these conditions MIP-1 α concentrations were 3851 ± 414 pg/ml. In the presence of 10 U/ml IL-12, a clear dose-response curve with IL-15 was noted. Significant MIP-1 α production first occurred at IL-15 concentrations of 10 ng/ml and continued to increase through concentrations of 1×10^3 ng/ml (Fig. 1 B). In contrast, the culture of NK cells with IL-15, IL-12, and TGF- β resulted in a clear dose-dependent inhibition of MIP-1 α production which was nearly complete at 0.5 ng/ml TGF- β (Fig. 1 C). IL-10 had no inhibitory effect, and actually showed a modest, dose dependent, additive effect on NK cell MIP-1 α production when combined with IL-15 (not shown).

Kinetics of MIP-1 α gene expression. We assayed NK cell production of MIP-1 α mRNA transcript and protein over time. Results are shown in Fig. 2, A and B. Interestingly, resting NK cells express a 1.1-kb transcript previously only seen in cycloheximide-treated T cells after PHA activation (16). The 0.95-kb transcript normally identified with the full length GOS19.1 probe was absent in resting NK cells, but appeared after a 1 h costimulation with IL-12 and IL-15 (Fig. 2 A). Expression of this smaller transcript increased over the 24 h of the assay. The 1.1-kb transcript initially diminished over the first hour of costimulation and then increased in parallel with the 0.95-kb species. MIP-1 α protein was first detected in the supernatant of an identical culture at 12 h after costimulation and increased to approximately 300 pg/ml after 24 h (Fig. 2 B). Production continued to increase for several days afterward into the ng/ml range (not shown). Importantly, the kinetics of MIP-1 α production are nearly identical to the kinetics of NK cell IFN- γ production following the same costimulation (17).

NK cell expression of C-C CKR-1 mRNA and ^{125}I -MIP-1 α binding affinity. We performed a Northern analysis on total RNA obtained from fresh NK cells and NK cells cultured for 24 h in IL-15, or IL-15 and IL-12, to assess for the MIP-1 α receptor gene product. Fig. 3 A shows that resting and IL-15-activated NK cells express similar amounts of C-C CKR-1 transcript. Costimulation with IL-15 and IL-12 resulted in a substantial increase in gene expression. These results were confirmed by the reverse transcription polymerase chain reaction (data not shown). Receptor binding studies were also performed on untreated NK cells and those costimulated with IL-12 and IL-15. Only low levels of specific binding of MIP-1 α could be detected on untreated NK cells (data not shown). Treatment of NK cells with a combination of IL-12 and IL-15

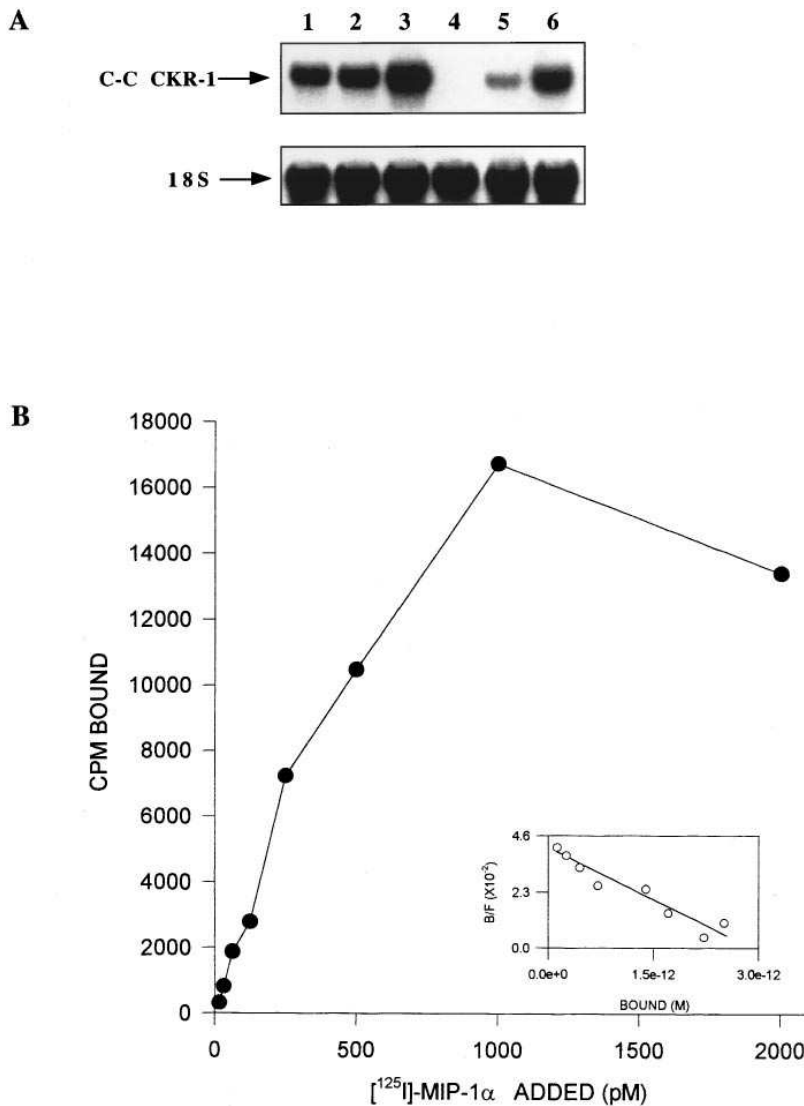


Figure 3. Characterization of MIP-1 α receptor expression on human NK cells. (A) Human NK cells ($\geq 85\%$) were cultured at 5×10^5 cells/ml for 24 h under one of the following conditions: medium alone, medium and IL-15 (10 ng/ml), or medium, IL-15 (10 ng/ml) and IL-12 (10 U/ml). Total cellular RNA was isolated, prepared for Northern analysis and probed for C-C CKR-1 message. Lane 1, resting NK cells; lane 2, IL-15; lane 3, IL-15 plus IL-12; lane 4, COLO 205 (negative control); lane 5, LPS-activated monocytes (positive control); and lane 6, resting monocytes (positive control). The same blot was reprobbed for 18 S rRNA to confirm equal loading of lanes. Autoradiographs were exposed for 20 h. (B), MIP-1 α receptor protein expression. Equilibrium binding curve and Scatchard plot (inset) are shown for NK cells following a 24 h incubation in medium containing IL-12 (10 U/ml) and IL-15 (10 ng/ml). CD56⁺ NK cells were incubated with varying concentrations of ¹²⁵I-MIP-1 α with or without a 100-fold excess of unlabeled MIP-1 α as described in Methods. The binding curves are plotted as CPMs bound versus total concentration of ¹²⁵I-MIP-1 α added and illustrate the difference between total binding and nonspecific binding. The Scatchard analysis is plotted as the ratio of bound (B) to free (F) ¹²⁵I-MIP-1 α versus the concentration of bound ¹²⁵I-MIP-1 α . Results are representative of two independent experiments.

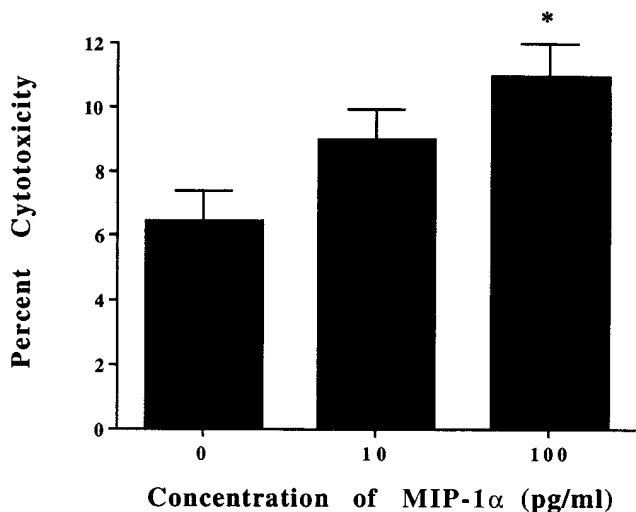


Figure 4. Exogenous MIP-1 α enhances NK cell cytotoxicity. NK cells ($\geq 97\%$) were first incubated overnight in medium containing varying concentrations of MIP-1 α . Standard 4 h cytotoxicity assays were

resulted in the induction of a single class of high affinity receptors for MIP-1 α with a K_d of 69–95 pM and an expression density of 125–173 sites per cell (Fig. 3 B).

Functional response of human NK cells to MIP-1 α receptor activation. Despite the fact that NK cells produced MIP-1 α and expressed receptors for MIP-1 α , the chemokine itself had no effect on augmenting NK cell production of IFN- γ , TNF- α , or GM-CSF (not shown). However, low concentrations of rhMIP-1 α did enhance NK cell cytotoxic activity against NK-resistant tumor target cells in the absence of other cytokines (Fig. 4), and in the presence of IL-15 (not shown).

Induction of MIP-1 α production by IL-15 versus IL-2. IL-15 activates human NK cells through the intermediate affinity IL-2 receptor (IL-2R $\beta\gamma$), but does not utilize the alpha (α) subunit of the high affinity IL-2 receptor (IL-2R $\alpha\beta\gamma$). IL-2 can activate NK cells through IL-2R $\beta\gamma$ and IL-2R $\alpha\beta\gamma$ (15). The CD56^{bright} NK cell subset constitutively expresses both the in-

then conducted using the NK-resistant COLO 205 cell line as targets. Results represent the mean percent cytotoxicity for indicated conditions of triplicate wells. The results are representative of three independent experiments. * $P \leq 0.025$.

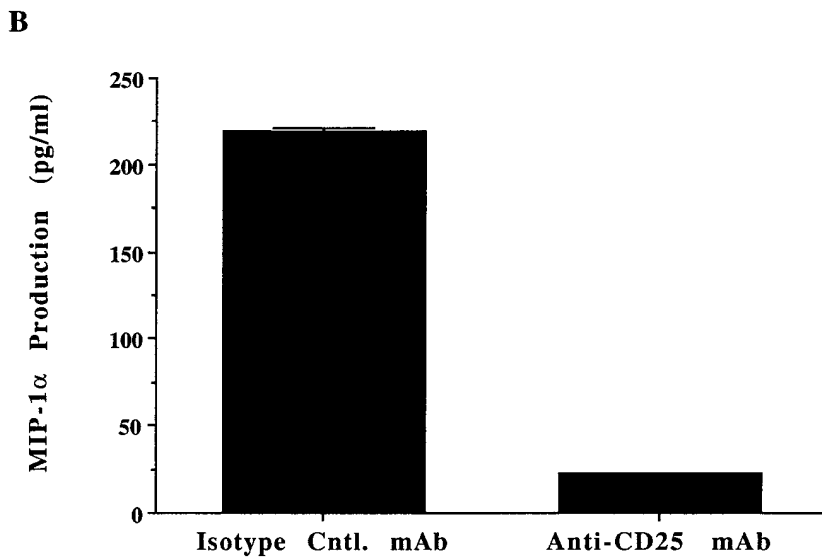
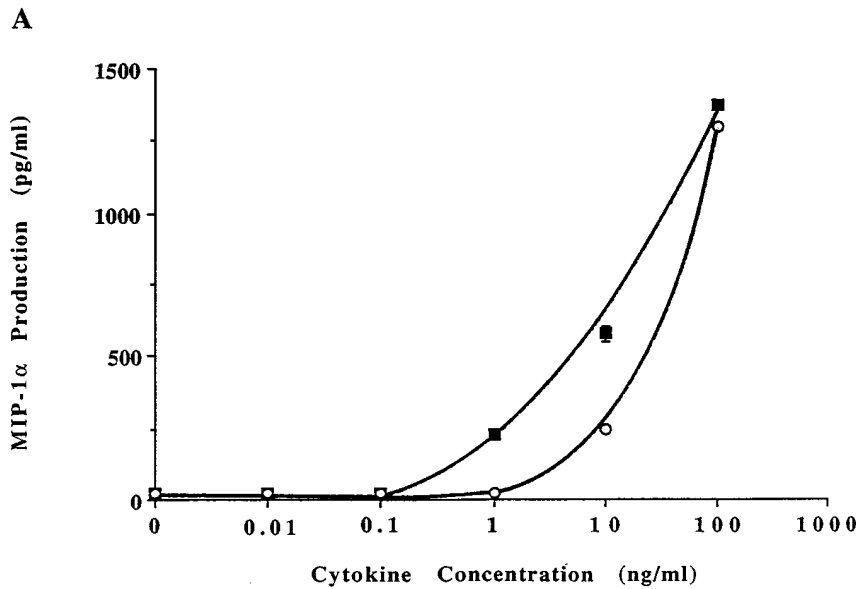


Figure 5. CD56^{bright} NK cell production of MIP-1 α in response to IL-2 versus IL-15. (A) Human CD56^{bright} NK cells ($\geq 97\%$) were plated at 2×10^4 cells per well in medium containing increasing concentrations of either IL-2 (■) or IL-15 (○). After 72 h of culture, supernatants were harvested and analyzed for MIP-1 α protein by ELISA. Values represent mean \pm SE of triplicate wells and are representative of three independent experiments. (B) Human CD56^{bright} NK cells ($\geq 97\%$) were plated at 2×10^4 cells per well in medium containing 1 ng/ml IL-2 with an isotype control mAb or 1 ng/ml IL-2 with an anti-CD25 (anti-IL-2R α) mAb. After 72 h of culture, supernatants were harvested and assayed for MIP-1 α protein by ELISA. Values represent mean \pm SE of triplicate wells and are representative of two independent experiments.

intermediate affinity IL-2R $\beta\gamma$ and the high affinity IL-2R $\alpha\beta\gamma$ (18–20). We therefore compared the effects of monocyte-derived IL-15 versus T cell-derived IL-2 on MIP-1 α production in the CD56^{bright} NK cell subset. Each cytokine induced MIP-1 α production in a dose dependent fashion (Fig. 5A). However, IL-2 was active at 10-fold lower concentrations than IL-15, presumably because IL-2R $\alpha\beta\gamma$ can selectively bind IL-2 with high affinity. This was confirmed by activating CD56^{bright} NK cells with 1 ng/ml IL-2 in the absence or presence of anti-CD25 mAb, which selectively blocks IL-2 binding to the high affinity IL-2R $\alpha\beta\gamma$ (18). CD56^{bright} NK cell MIP-1 α production at 1 ng/ml IL-2 was completely abrogated in the presence of anti-CD25 mAb (Fig. 5B). At higher concentrations of IL-2 and IL-15 that activate the intermediate affinity IL-2R $\beta\gamma$ (i.e., 100 ng/ml), each cytokine induced equivalent amounts of MIP-1 α production (Fig. 5A).

Discussion

After infection with obligate intracellular pathogens and gram negative organisms, monocytes are known to elaborate a num-

ber of cytokines, such as IL-12, IL-1 β , TNF- α , and IL-15, each of which can activate NK cells (15, 17, 21). NK cells, in turn, produce IFN- γ which is critical for elimination of these pathogens by monocytes (2). In this study, we examine NK cells for the production of additional factors which may be involved in this positive feedback loop between innate immune effector cells. We provide the first evidence that NK cells produce the chemokine MIP-1 α , and do so in response to monocyte/macrophage-derived factors. Unfractionated MIP-1, containing both the α and β fractions, has previously been shown to potentiate IFN- γ -inducible secretion of inflammatory cytokines by monocytes, such as IL-1 β and TNF- α (5). Recently, MIP-1 α has been shown to contribute to the suppression of the level of HIV replication in vitro (6). Thus, NK-produced MIP-1 α may assist in the control of both obligate intracellular and HIV infections in monocytes, via a paracrine positive feedback loop. Likewise, NK cell production of MIP-1 α may also contribute to septic shock, as has been proposed for IFN- γ and TNF- α (3, 4).

Significant MIP-1 α production by NK cells requires specific costimulation with IL-12 and IL-15, while combinations of IL-12 and IL-10, IL-1 β , or TNF- α were not effective. This is in

distinct contrast to NK cell IFN- γ production which can be induced by IL-12 in combination with any of these other monocyte-derived cytokines (15, 21, 22). NK cell MIP-1 α production therefore appears to define the first unique role for monocyte-derived IL-15 as a critical cofactor in NK activation. This may have relevance *in vivo*, as we have recently demonstrated that LPS-activated human monocytes secrete IL-15 protein over a similar time course as IL-12 (17). We also show TGF- β to be a potent inhibitor of NK cell MIP-1 α production induced by co-stimulation with IL-12 and IL-15. This may also have relevance *in vivo*, as HIV⁺ individuals have an increased plasma concentration of active TGF- β (23). IL-2 is a T cell-derived cytokine that is produced after antigen activation (24). Its ability to induce modest MIP-1 α production via the high affinity IL-2R $\alpha\beta\gamma$ in CD56^{bright} NK cells suggests a potential role for MIP-1 α during the antigen specific immune response.

NK cells upregulate a high affinity MIP-1 α receptor in response to costimulation by IL-12 and IL-15, as demonstrated by Northern blot and protein binding analyses. This observation supports the work of Maghazachi et al, showing that MIP-1 α is a chemoattractant for NK cells (25). Hence one role of NK cell MIP-1 α receptor expression may be to direct NK cells toward other important sources of MIP-1 α production, such as LPS-activated monocytes (26). This physical migration would assist in establishing the effective paracrine positive feedback loops required to control infection (1, 2, 15, 21). Our data suggest that MIP-1 α receptor expression may also serve to enhance NK cell killing of pathogens.

In summary, we provide the first evidence that human NK cells produce the chemokine MIP-1 α in response to co-stimulation by a specific combination of monocyte-derived cytokines. NK cell MIP-1 α and MIP-1 α receptor expression are likely to participate in the innate immune response after monocyte activation, via paracrine and autocrine positive feedback loops. The contribution of NK cell MIP-1 α to control of infection, inflammation and septic shock will likely be elucidated in studies that involve mice lacking the MIP-1 α gene product (27). The ability to selectively expand the NK cell population in HIV⁺ patients (as a potential source of MIP-1 α) without increasing HIV mRNA load (28) may provide an additional therapeutic modality to aid in slowing the clinical progression of AIDS.

Acknowledgments

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