

Identification of Mutations in the Coding Sequence of the Proto-oncogene *c-kit* in a Human Mast Cell Leukemia Cell Line Causing Ligand-independent Activation of *c-kit* Product

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Abstract

The *c-kit* proto-oncogene encodes a receptor tyrosine kinase. Binding of *c-kit* ligand, stem cell factor (SCF) to *c-kit* receptor (*c-kitR*) is known to activate *c-kitR* tyrosine kinase, thereby leading to autophosphorylation of *c-kitR* on tyrosine and to association of *c-kitR* with substrates such as phosphatidylinositol 3-kinase (PI3K). In a human mast cell leukemia cell line HMC-1, *c-kitR* was found to be constitutively phosphorylated on tyrosine, activated, and associated with PI3K without the addition of SCF. The expression of SCF mRNA transcript in HMC-1 cells was not detectable by means of PCR after reverse transcription (RT-PCR) analysis, suggesting that the constitutive activation of *c-kitR* was ligand independent. Sequencing of whole coding region of *c-kit* cDNA revealed that *c-kit* genes of HMC-1 cells were composed of a normal, wild-type allele and a mutant allele with two point mutations resulting in intracellular amino acid substitutions of Gly-560 for Val and Val-816 for Asp. Amino acid sequences in the regions of the two mutations are completely conserved in all of mouse, rat, and human *c-kit*. In order to determine the causal role of these mutations in the constitutive activation, murine *c-kit* mutants encoding Gly-559 and/or Val-814, corresponding to human Gly-560 and/or Val-816, were constructed by site-directed mutagenesis and expressed in a human embryonic kidney cell line, 293T cells. In the transfected cells, both *c-kitR* (Gly-559, Val-814) and *c-kitR* (Val-814) were abundantly phosphorylated on tyrosine and activated in immune complex kinase reaction in the absence of SCF, whereas tyrosine phosphorylation and activation of *c-kitR* (Gly-559) or wild-type *c-kitR* was modest or little, respectively. These results suggest that conversion of Asp-816 to Val in human *c-kitR* may be an activating mutation and responsible for the constitutive activation of *c-kitR* in HMC-1 cells. (*J. Clin. Invest.* 1993. 92:1736–1744.) Key words: proto-oncogene *c-kit* • point mutation • leukemia • tyrosine kinase • activation

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Introduction

The proto-oncogene *c-kit* was identified as the cellular homologue of the oncogene *v-kit* present in the genome of Hardy-Zuckerman 4-feline sarcoma virus that induces multicentric fibrosarcomas in the domestic cat (1). The proto-oncogene *c-kit* encodes a transmembrane tyrosine kinase receptor that is a member of receptors for PDGF and CSF-1 (2, 3). The characteristic features of the receptor tyrosine kinase subfamily are an extracellular domain made up of five immunoglobulin-like repeats, and an intracellular tyrosine kinase, which is split into two domains by a hydrophilic insert sequence (1–4). The ligand for *c-kit* product has recently been cloned and variously designated as stem cell factor (SCF),¹ mast cell growth factor, *kit* ligand, or *steel* factor (5–12).

The *c-kit* gene is allelic with the dominant *white spotting* (*W*) locus on mouse chromosome 5 (13, 14), whereas the gene encoding SCF resides at *steel* (*S1*) locus on mouse chromosome 10 (7, 9, 12). Mutations at either the *W/c-kit* locus or the *S1/SCF* locus affect various aspects of hematopoiesis and the proliferation and/or migration of primordial germ cells and melanoblasts during embryogenesis (4, 15–17). In hematopoiesis, both *W/c-kit* and *S1/SCF* mutations affect the hematopoietic stem cell compartment and give rise to hypoplastic anemia and a deletion of mast cells (4, 15–17). Recent experiments have shown that *c-kit* product is expressed on mast cells (4, 16, 18) and on a small, progenitor-cell fraction (~4%) of adult bone marrow cells (19–21), and that SCF acts synergistically with other hematopoietic growth factors to stimulate the growth of a variety of progenitor cells (5, 6, 22–24).

In addition to normal hematopoietic cells, we and others have investigated the expression and function of *c-kit* product in human leukemia cells (25–32). In human leukemia cell lines, the expression of *c-kit* product was detected in erythroleukemia cell lines (HEL, LAMA, and TF-1), megakaryocytic leukemia cell lines (M07E and MEG-01), and mast-cell leukemia cell line (HMC-1) (25, 27, 31, 32). Also, *c-kit* product was found to be expressed in most cases of human acute myeloblastic leukemia (AML) and in some cases of chronic myelogenous leukemia (CML) in blastic crisis (26, 28–31). Further, SCF induced rapid tyrosine phosphorylation and activation of *c-kit* product, leading to proliferation of M07E cells and blast cells in a substantial fraction of AML cases (27, 28). These results

1. Abbreviations used in this paper: AML, acute myeloblastic leukemia; CML, chronic myelogenous leukemia; HMC-1, human mast-cell leukemia cell line-1; PI3K, phosphatidylinositol 3-kinase; rh, recombinant human; RTK, receptor tyrosine kinase; *S1*, *steel* locus; SCF, stem cell factor; *W*, *white spotting* locus.

suggest that *c-kit* activation may be involved in some aspect of excessive proliferation and aberrant differentiation of human leukemia cells. However, structural abnormalities of *c-kit* gene contributing to constitutive, ligand-independent activation of the *c-kitR* kinase have not been reported.

In this study, we have investigated the state of tyrosine phosphorylation and activation of *c-kit* product in human leukemia cells, and found that, regardless of stimulation with SCF, *c-kit* product is constitutively phosphorylated on tyrosine residues in HMC-1. Neither SCF mRNA transcript nor biological activity of SCF were detectable from HMC-1 cells, suggesting that the activation of *c-kit* was not due to autocrine mechanism. The sequence of the *c-kit* gene revealed unique, as-yet-unidentified, two-point mutations in the cytoplasmic domain, resulting in amino acid substitutions of Gly for Val in codon 560 and Val for Asp in codon 816. Further, introduction of the mutated *c-kit* genes into cells of a human embryonic kidney cell line, 293T, resulted in the generation of the *c-kit*-encoded protein that was constitutively phosphorylated on tyrosine and activated without the addition of SCF. Thus, our data provide the first evidence that point mutation(s) in *c-kit* gene is capable of inducing constitutive activation of *c-kit* product, and raise the possibility that the activating mutation(s) described here may be involved in oncogenesis of some cell types, including mast cells and hematopoietic stem cells.

Methods

Reagents. Highly purified recombinant human (rh) GM-CSF (rhGM-CSF) was a gift from Drs. S. Clark and G. Wong (Genetic Institute, Cambridge, MA). rhSCF was generously provided by Dr. K. M. Zsebo (Amgen Inc., Thousand Oaks, CA). The anti-phosphotyrosine antibody, a murine mAb generated against phosphotyramine, was generously supplied by Dr. B. Druker (Dana-Farber Cancer Institute, Boston, MA). The specificity of the anti-phosphotyrosine mAb was described previously (33, 34). Rabbit anti-human *c-kit* serum was kindly provided by Dr. A. Ullrich (Max-Planck Institute für Biochemie, Martinsried, Germany). This antiserum was generated against a synthetic peptide corresponding to COOH-terminal 16 amino acid residues of *c-kitR* (2). A murine mAb (YB5.B8) that reacts with the extracellular domain of human *c-kit* protein was originally raised against human AML blast cells (21, 35). The rabbit antiserum against the whole murine *c-kit* protein (36) was generously donated by Dr. P. Besmer (Cornell University Graduate School of Medical Science, New York). A rat mAb (ACK-2) against the extracellular domain of murine *c-kit* product (19) and the full length of murine *c-kit* cDNA clone were generously donated by Dr. S-I. Nishikawa (Kumamoto University, Kumamoto, Japan). Rabbit anti-phosphatidylinositol 3-kinase (PI3K) serum was purchased from Upstate Biotechnology, Inc. (Lake Placid, NY); the anti-PI3K serum recognizes an 85-kD subunit of PI3K. Chemically defined serum-free medium (ASF-102) was purchased from Ajinomoto (Tokyo, Japan); it contains human transferrin, insulin, and BSA.

Cell lines. HMC-1, a human mast cell leukemia cell line, established from the peripheral blood from a patient with mast cell leukemia, was grown in Iscove's modified Dulbecco's medium supplemented with 10% horse serum (Flow Lab, North Ryde, Australia) as described previously (37). M07E, a human GM-CSF and IL-3 dependent cell line, was obtained from Dr. Steve Clark, Genetics Institute, and was originally established by Avanzi et al. from the peripheral blood from an infant with acute megakaryocytic leukemia (38). M07E cells were cultured in RPMI-1640 medium (Nacalai Tesque, Kyoto, Japan) supplemented with 10% FCS (Flow Lab) and 10 ng/ml rhGM-CSF. HEL, a human erythroleukemia cell line, and NTI-4, human fetus-derived fibroblast cell line, were obtained from the Japanese

Cancer Research Resources Bank, and this cell line were adapted to grow and maintained in RPMI 1640 supplemented with 10% FCS. A human embryonic kidney cell line, 293T was kindly provided by Dr. D. Baltimore (The Rockefeller University, New York), and was derived from human embryonic kidney cells transformed by DNA from human adenovirus type 5 (39). 293T cells were maintained in DMEM (Nacalai Tesque) supplemented with 10% FCS. In a preliminary study, 293T cells were found to be capable of expressing a greater amount of *c-kit* product than COS cells after transfection of *c-kit* gene.

Flow cytometry. Cells (5×10^5) were washed with PBS and resuspended in PBS containing 0.5% BSA (Sigma Chemical, St. Louis, MO), 0.1% NaN_3 , and human Ig (1 mg/ml) to block binding of antibodies to Fc receptors on the cells. Cells were incubated with YB5.B8 or mouse monoclonal IgG, control (Becton Dickinson and Co., Mountain View, CA) at 4°C for 30 min, then washed three times with the buffer. Cells were subsequently incubated with fluorescein-conjugated goat (Fab')₂ anti-mouse IgG antibody (Tago, Inc., Burlingame, CA) at 4°C for 30 min, and washed twice before analysis using a FACScan® flow cytometer (Becton Dickinson and Co.).

Immunoblotting. The procedures of cell lysis, gel electrophoresis, and immunoblotting were performed according to the methods described previously (27, 28, 33, 34). Briefly, exponentially growing cells were washed free of serum and growth factors and incubated in serum-free ASF-102 medium for 16 h at 37°C to factor-deprive the cells. The cells (10^7 cells suspended in 1 ml of ASF-102 medium) were then exposed to rhSCF (100 ng/ml) for 15 min at 37°C. After stimulation, cells were washed with cold PBS and lysed in NP-40 buffer (20 mM Tris-HCl pH 8.0, 137 mM NaCl, 10% glycerol, 1% NP-40) containing protease and phosphatase inhibitors at 4°C for 20 min. In some experiments, lysis buffer containing 1% digitonin (Wako Pure Chemical Industries, Ltd., Osaka, Japan) instead of 1% NP-40 was used to examine the association of *c-kit* product with PI3K. Insoluble material was removed by centrifugation at 10,000 g for 15 min at 4°C, and cell lysates were subjected to SDS-PAGE. Proteins were electrophoretically transferred from the gel onto a polyvinylidene difluoride membrane (Immobilon; Millipore Corp., Bedford, MA), and immunoblotting was performed with either anti-phosphotyrosine mAb or anti-*c-kit* serum.

For immunoprecipitation, the lysates from rhSCF-stimulated or unstimulated cells were precleared with protein A-Sepharose beads (Pharmacia AB, Uppsala, Sweden) for 2 h at 4°C. The precleared lysates were then incubated with a YB5.B8 mAb and protein A-Sepharose beads to collect the antigen-antibody complexes. The immunoprecipitates were washed five times with lysis buffer containing protease and phosphatase inhibitors, and subjected to SDS-PAGE and immunoblotting with either anti-phosphotyrosine mAb, anti-*c-kit* serum, or anti-PI3K serum.

Immune complex kinase assay. The cell lysates were prepared from rhSCF-unstimulated cells using NP-40 lysis buffer. The product of *c-kit* was immunoprecipitated with the YB5.B8 mAb and protein A-Sepharose beads, and the immune complexes were washed once with PBS, twice with 0.5 M LiCl and 50 mM Tris-HCl, pH 7.4, and once with kinase buffer (10 mM MnCl_2 , 20 mM Tris-HCl, pH 7.4) at 4°C. Then, the immune complexes were incubated in kinase buffer containing 10 $\mu\text{Ci/ml}$ [γ -³²P]ATP (DuPont/NEN Research Products, Boston, MA) for 20 min at 25°C, washed, and separated by SDS-PAGE. The gel was dried, and radioactive proteins were detected by autoradiography.

Oligonucleotide primers. PCR was performed by using two complementary oligonucleotide primers for amplifying human SCF cDNA: 5'-ATGAAGAAGACACAACCTTG-3' (184 to 203, sense) and 5'-AAGGCATCAATGGATCTATT-3' (615 to 634, antisense). For the amplification of human *c-kit* cDNA, the following seven oligonucleotide primers were used: primer 1, 5'-GATCCCATCGCAGCTACCGCG-3' (1 to 21, sense); primer 2, 5'-ATCTTCCCCATGATAAACAC-3' (964 to 983, sense); primer 3, 5'-AGGAGATAAATGGAAACAATT-ATGT-3' (1703 to 1727, sense); primer 4, 5'-TTGAGCATCTTTACAGCGACAGTCA-3' (1874 to 1898, antisense); primer 5, 5'-GCCACCTGGTCATTACAGAAT-3' (2013 to 2035, sense); primer 6, 5'-AAAATCCCATAGGACCAG-3' (2577 to 2594, antisense); primer 7,

5'-AGTGTGCTCAGAAAGACAGGATTGC-3' (3068 to 3092, antisense). The numbers in parentheses represent the nucleotide numbers on the complementary strands of each cDNA sequence.

cDNA synthesis and PCR amplification. Total cellular RNA was isolated with a guanidine isocyanate method in combination with a cesium chloride modification (40, 41). Genomic DNA was prepared from HMC-1 cells by the standard method. Total RNA was converted into cDNA using the primer specific for the subsequent PCR amplification. cDNA synthesized from 4 µg of total RNA was amplified in a 100 µl reaction mixture by a DNA thermal cycler (Perkin-Elmer Cetus Corp., Norwalk, CT) using *Taq* DNA polymerase in 25 cycles of 1-min denaturation at 94°C, 2-min annealing at 55°C, and 3-min synthesis at 72°C. To minimize possible artifacts that could be caused by priming with PCR, the number of PCR cycles was kept to a minimum.

Detection of human SCF mRNA transcript by means of PCR after reverse transcription (RT-PCR) analysis. The single-strand cDNA was synthesized from total mRNA of HMC-1 and NTI-4 cells with both the specific and random primers, and subjected to PCR to amplify the 451-bp DNA fragment between nucleotide 184 and 634 of human SCF cDNA. PCR products were examined on 1% agarose gel, transferred to nitrocellulose filter (Schleicher & Schuell, Dassel, Germany), and then hybridized with random ³²P-labeled probe.

Sequencing of *c-kit* cDNA. Sequencing of the whole coding region of *c-kit* cDNA was performed in HMC-1 cells by using four sets of primers: primers 1-4, 2-6, 3-6, and 5-7 as described above. The PCR products amplified by these primers were gel purified, sequentially treated with T4 polynucleotide kinase and the Klenow fragment of DNA polymerase I, and subcloned into the *EcoRV* site of Bluescript KS(-) (Stratagene Corp., La Jolla, CA). DNA sequences of four subclones obtained from each PCR product were determined and compared with normal sequence of human *c-kit* cDNA (2). The DNA sequencing reaction was performed by the method of Sanger et al. (42) using [³²P]α-deoxycytidine triphosphate (DuPont/NEN Research Products). In order to further confirm the mutations of *c-kit* mRNA, the sequencing in the regions of *c-kit* mutations was performed directly using DNA fragments amplified by independent RT-PCR.

Site-directed mutagenesis and transfection. To directly examine the causal role of the *c-kit* mutations in activation of *c-kit* tyrosine kinase activity, site-directed mutagenesis was performed using murine *c-kit* cDNA because of the following reasons. First, amino acid sequences in the regions of the two *c-kit* mutations are completely conserved between mouse and human. Second, human SCF is known to have little effect on cells expressing murine *c-kit* receptor (*c-kitR*) (6, 7), whereas murine SCF can exert its actions through interaction with both murine and human *c-kitRs* (6, 7, 27, 43). We therefore used a combination of murine *c-kit* cDNA and a human 293T cell line as a transfectant.

The gene encoding murine wild-type *c-kitR* was cloned into Bluescript I KS(-) and the orientation of the insert was determined by sequencing. Single-stranded uracil-containing DNA was prepared from *Escherichia coli* strain CJ236 (*dut⁻ ung⁻*) as described by Kunkel (44). To generate genes containing G-559 and V-814 mutations, two oligonucleotides of 5'-CAATGGAAGGTTGGCGAGGAGATAAAT-3' and 5'-GGGCTAGCCAGAGTCATCAGGAATGAT-3' were synthesized, annealed to the template and extended. The double-stranded mismatch containing mutant plasmids were transformed into *ung⁺ E. coli* strain, and the clones containing each mutation were then screened by sequencing. The *DraI*-*Bss*HII fragment with G-599 mutation and the *NheI*-*MroI* fragment with V-814 mutation were isolated and inserted into the wild type *c-kit* clone to construct mutants encoding G-559, V-814, or both of them. The full coding sequences of the wild-type and mutated *c-kit* cDNA were then released by *SmaI* and *HindIII* restriction digestion, isolated, and inserted into the expression vector pSV2. The inserts were resequenced to confirm the orientation and mutations within the expression vectors.

The expression vectors containing wild-type and mutated *c-kit* cDNA were transfected into 293T cells by the calcium phosphate method as described previously (45). The cells were cultured in DMEM containing 10% FCS and fed every 3 d with fresh medium. To

determine the expression of *c-kit* product in the transfected 293T cells, the transfectants were incubated with 100 µCi/ml [³⁵S]methionine (DuPont/NEN Research Products) for 6 h, and lysed with NP-40 lysis buffer. The labeled *c-kit* product was then immunoprecipitated with a rat anti-mouse *c-kit* mAb (ACK-2) and analyzed by SDS-PAGE and autoradiography. For the analysis of protein tyrosine phosphorylation in the transfectants, cell lysates were subjected to SDS-PAGE and immunoblotting with an anti-phosphotyrosine mAb. To examine the activation and tyrosine phosphorylation of *c-kit* product, *c-kit* product of each transfectant was immunoprecipitated with rabbit antiserum against murine *c-kit* protein from the lysates, and then subjected to *in vitro* immune complex kinase assay and immunoblotting with an anti-phosphotyrosine mAb as described above.

Results

Constitutive tyrosine phosphorylation of *c-kit* product in HMC-1 cells. The expression of *c-kit* product was first examined with an anti-*c-kit* mAb, YB5.B8, which recognizes the extracellular domain of human *c-kit* product (35). Flow cytometry analysis showed that *c-kit* product was expressed on HMC-1, M07E, and HEL cells at an almost equal level (Fig. 1 A). Despite the high level of *c-kit* expression on HMC-1 cells, proliferation of HMC-1 cells was not augmented by the treatment with rhSCF (data not shown), whereas rhSCF was reported to induce proliferation of M07E and HEL cells (27, 31).

To examine the impaired effect of rhSCF on proliferation of HMC-1 cells, we investigated the effect of rhSCF on tyrosine kinase activity in HMC-1, M07E, and HEL cells. The cells were removed from growth factors for 16 h and then stimulated with 100 ng/ml of rhSCF for 15 min at 37°C. Changes in tyrosine phosphorylation were detected by immunoblotting with a mAb specific for phosphotyrosine. After stimulation of M07E and HEL cells with rhSCF, increased phosphotyrosine was observed in proteins particularly at molecular mass of 145 kD (Fig. 1 B). In HMC-1 cells, by contrast, tyrosine phosphorylation of 145-kD protein was observed even in the absence of exogenous rhSCF, and rhSCF treatment resulted in only a modest increase in tyrosine phosphorylation of the protein (Fig. 1 B).

In order to determine if the 145 protein was *c-kit* product, *c-kit* product was immunoprecipitated and analyzed by immunoblotting with either anti-phosphotyrosine mAb or anti-*c-kit* serum. As shown in Fig. 1 C, tyrosine phosphorylation of *c-kit* product was detected in HMC-1 cells regardless of rhSCF stimulation, while it was considerably increased by stimulation with rhSCF in M07E and HEL cells.

Activation of *c-kit* product in HMC-1 cells. Ligand binding to *c-kit* product is known to activate the *c-kit* receptor tyrosine kinase, thereby leading to *c-kit*-tyrosyl autophosphorylation and the association of *c-kit* with its substrates such as phosphatidylinositol 3-kinase (PI3-kinase) (4, 46-48). To determine whether or not the constitutive tyrosine phosphorylation of *c-kit* product in HMC-1 cells was accompanied by activation of *c-kit* receptor tyrosine kinase, we examined the association of *c-kit* product with PI3K and the autokinase activity of *c-kit* products in the cells.

HMC-1, M07E, and HEL cells were factor starved and exposed to medium alone or rhSCF (100 ng/ml) for 15 min at 37°C, and then lysed with lysis buffer containing 1% digitonin. Before and after stimulation with rhSCF, *c-kit* products were immunoprecipitated with a YB5.B8 mAb from the cell lysates and divided into two aliquots, one of which was subjected to

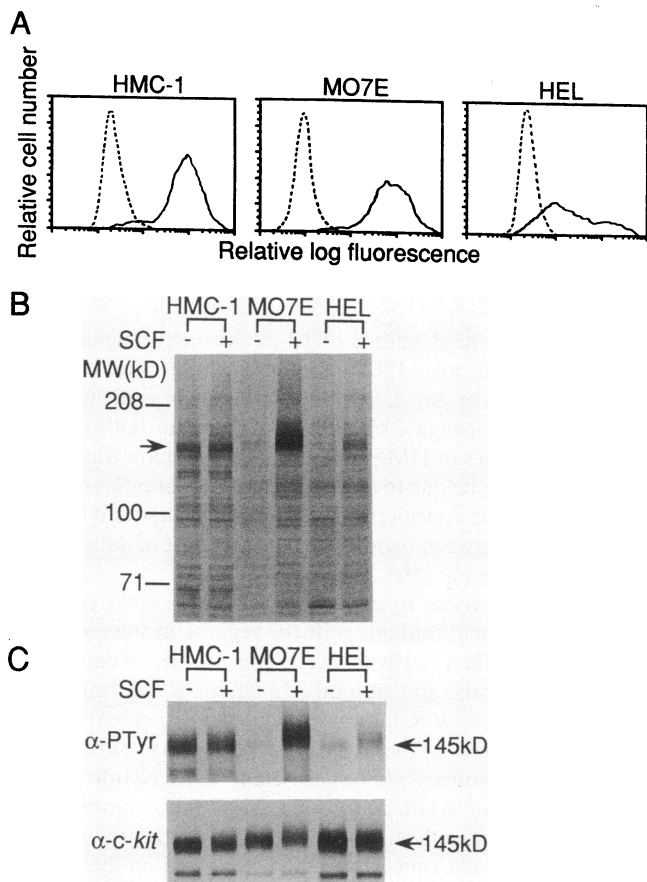


Figure 1. Expression of *c-kit* product on HMC-1, M07E, and HEL cells (A) and effects of rhSCF on tyrosine phosphorylation of cellular proteins (B) and *c-kit* product (C) in the cell lines. Flow cytometric analysis of the surface binding of a monoclonal anti-*c-kit* antibody (YB5.B8) to HMC-1, M07E, and HEL cells (A). Cells were incubated in either negative control antibody (---) or YB5.B8 (—), washed, incubated with fluorescein-conjugated goat (Fab')₂ anti-mouse IgG antibody, and analyzed on a FACScan®. For identification of phosphotyrosine-containing proteins, cell lysates were obtained before and after treatment with rhSCF (100 ng/ml) for 15 min at 37°C and subjected to immunoblot with anti-phosphotyrosine mAb (B). To examine changes in tyrosine phosphorylation of *c-kit* product, *c-kit* product was immunoprecipitated with YB5.B8 from cell lysates before and 15 min after rhSCF stimulation, and immunoblot was performed with either anti-phosphotyrosine mAb or anti-*c-kit* serum (C).

immunoblotting with anti-PI3K serum and another with anti-*c-kit* serum. Based on α -*c-kit* immunoblot data as shown in the lower portion of Fig. 2, approximately equivalent amounts of *c-kit* products were immunoprecipitated before and after stimulation with rhSCF in each cell line. By contrast, α -PI3K immunoblot as shown in the upper portion of Fig. 2 revealed that the 85-kD subunit of PI3K was co-immunoprecipitated with anti-*c-kit* antibody only after stimulation with rhSCF in two cell lines, M07E and HEL, whereas it was co-immunoprecipitated in HMC-1 cells even before stimulation with rhSCF.

We next examined the autokinase activity of *c-kit* products in immune complex kinase assay. Products of *c-kit* were immunoprecipitated from NP-40 lysates of HMC-1, M07E, and HEL cells before treatment with rhSCF, and assayed for autokinase activity. Tyrosine kinase activity of *c-kit* products was

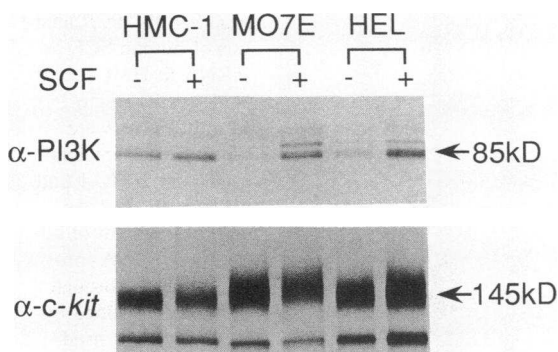


Figure 2. Association of *c-kit* product with PI3K in HMC-1, M07E, and HEL cells. Cells were treated with rhSCF (100 ng/ml) for 15 min at 37°C and lysed with 1% digitonin. *c-kit* product and *c-kit*-associated molecules were immunoprecipitated with YB5.B8 and protein A-Sepharose beads from the cell lysates before and after stimulation with rhSCF. The immunoprecipitates were divided into two aliquotes, separated by SDS-PAGE, and subjected to immunoblot analyses of PI3K (upper panel) and *c-kit* product (lower panel).

readily detectable in HMC-1 cells in the absence of SCF stimulation, whereas it was only faint in M07E or HEL cells (Fig. 3). In this experiment, almost equal amounts of *c-kit* products were immunoprecipitated in these samples (data not shown).

Absence of SCF mRNA transcripts in HMC-1 cells. One of the possible explanations for the results shown in Figs. 1–3 was that HMC-1 cells might produce SCF. The expression of SCF mRNA transcripts was examined by RT-PCR Southern blot analysis in HMC-1 cells and also in NTI-4 cells that were used as a positive control. As shown in Fig. 4, SCF mRNA transcript was not observed in HMC-1 cells, while a 451-bp fragment of SCF mRNA was easily detected in NTI-4 cells. Further, the culture supernatant of HMC-1 did not have any activities in stimulating proliferation or inducing tyrosine phosphorylation of *c-kit* product in M07E cells (data not shown). Thus, the constitutive tyrosine phosphorylation and activation of *c-kit* product in HMC-1 cells were not attributed to autocrine mechanism.

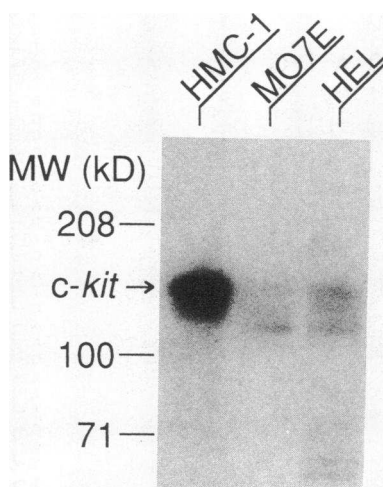


Figure 3. Immune complex kinase assay for *c-kit* tyrosine kinase activity in HMC-1, M07E, and HEL cells. *c-kit* product was immunoprecipitated with YB5.B8 from NP-40 lysates before stimulation with rhSCF, and the immunoprecipitates were incubated in kinase buffer containing 10 μ Ci/ml [γ -³²P]ATP for 20 min at 25°C, washed, and separated by SDS-PAGE. Incorporation of ³²P into *c-kit* product was visualized by autoradiography. In

this study, the immunoprecipitates from HMC-1, M07E, and HEL cells contained approximately equivalent levels of *c-kit* products.

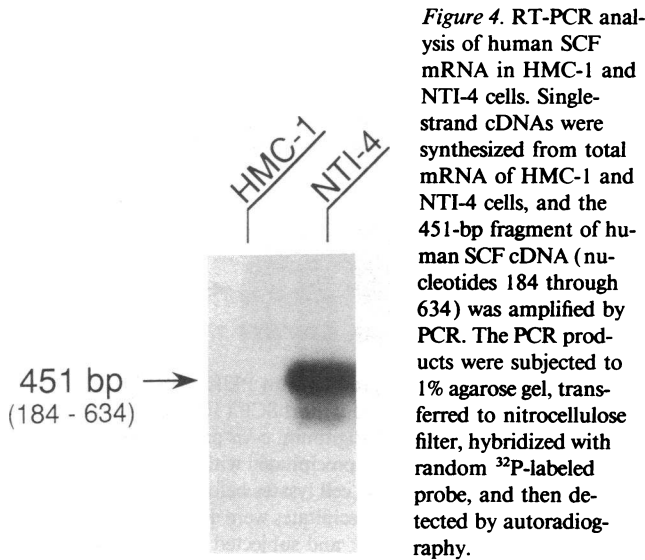


Figure 4. RT-PCR analysis of human SCF mRNA in HMC-1 and NTI-4 cells. Single-strand cDNAs were synthesized from total mRNA of HMC-1 and NTI-4 cells, and the 451-bp fragment of human SCF cDNA (nucleotides 184 through 634) was amplified by PCR. The PCR products were subjected to 1% agarose gel, transferred to nitrocellulose filter, hybridized with random ³²P-labeled probe, and then detected by autoradiography.

Mutations in *c-kit* cDNA. An alternate mechanism which might explain the constitutive tyrosine phosphorylation and activation of *c-kit* products in HMC-1 cells would be due to mutations in the *c-kit* proto-oncogene. Southern blot analyses of genomic DNA from HMC-1 using *EcoRI*, *HindIII*, and *BamHI* showed neither rearrangement nor amplification in *c-kit* gene as compared with normal human placenta (data not shown). However, sequencing of *c-kit* cDNA demonstrated that *c-kit* cDNA of HMC-1 cells carries heterozygous transitions in codon 560 (GTT → GGT, nucleotides 1699 through 1701 in the region between the membrane and the ATP-binding domains) and codon 816 (GAC → GTC, nucleotides 2467 through 2469 in the phosphotransferase domain), which result in the amino acid substitutions of Gly-560 for Val and Val-816

for Asp, respectively. These two point mutations were observed in cDNA clones generated from three independent PCR products, and also confirmed by direct sequencing of PCR products amplified independently (Fig. 5). Further, sequencing of cDNA fragment (nucleotides 964 through 2594) covering the regions of both mutations was performed in four subclones. This analysis showed that two subclones had both mutations, but the remaining two subclones had neither mutation. These results indicate that *c-kit* genes of HMC-1 cells are composed of a normal wild-type allele and a mutant allele with two point mutations.

In addition, the deletions of 12-nucleotide sequence (nucleotides 1550 through 1561), the extracellular 4-amino acid sequence (Gly-Asn-Asn-Lys), and 3-nucleotide sequence (nucleotides 2163 through 2165) were observed in both normal and mutant alleles of HMC-1. The former deletion was previously reported to be due to alternative splicing of mRNA (49); the latter was also considered as the result of alternative splicing of mRNA between exon 14 and 15, in light of data on the human *c-kit* genome (50).

Mutation(s) leads to activation of *c-kit* receptor tyrosine kinase. Amino acid sequences in the regions of the two *c-kit* mutations identified in HMC-1 cells are completely conserved in all of mice, rats, and humans. In order to determine the direct role of the *c-kit* mutations in ligand-independent activation of *c-kit* products, the expression vectors containing normal or mutated murine *c-kit* genes were transfected into cells of a human embryonic kidney cell line, 293T. This combination could minimize the effect of SCF, which might be produced by the transfected cells, on *c-kit* product expressed on 293T cells (see Methods).

We used site-directed mutagenesis to construct mutant murine *c-kitR* containing substitutions of Gly for Val at codon 559 and/or Val for Asp at codon 814, which corresponded to Gly-560 and/or Val-816 of mutant human *c-kitR*, respec-

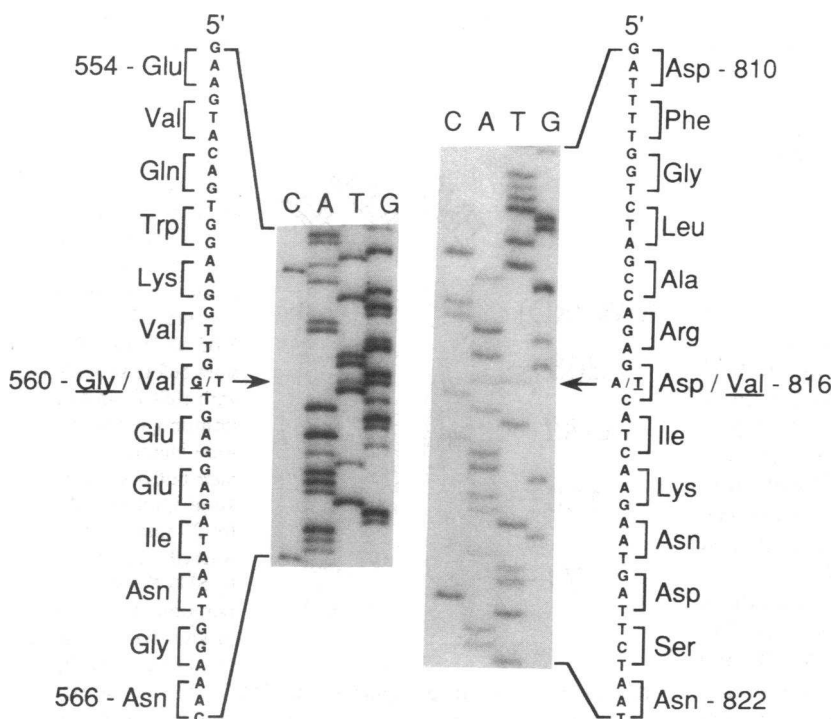


Figure 5. Nucleotide sequence including the mutation sites of *c-kit* cDNA obtained from HMC-1 cells. Shown are results from direct sequencing of *c-kit* DNA fragments from amino acid residues 554 to 566 and those 810 to 822. HMC-1 cells were found to carry heterologous mutations of *c-kit* gene at codons 560 and 816. These two mutations were observed in cDNA clones from three independent PCR products, and sequencing of cDNA fragments (nucleotides 964 through 2594) in four clones revealed that the *c-kit* gene of HMC-1 is composed of a normal and a mutant allele. The arrows indicate the mutation sites, and the altered nucleotides and amino acids are underlined.

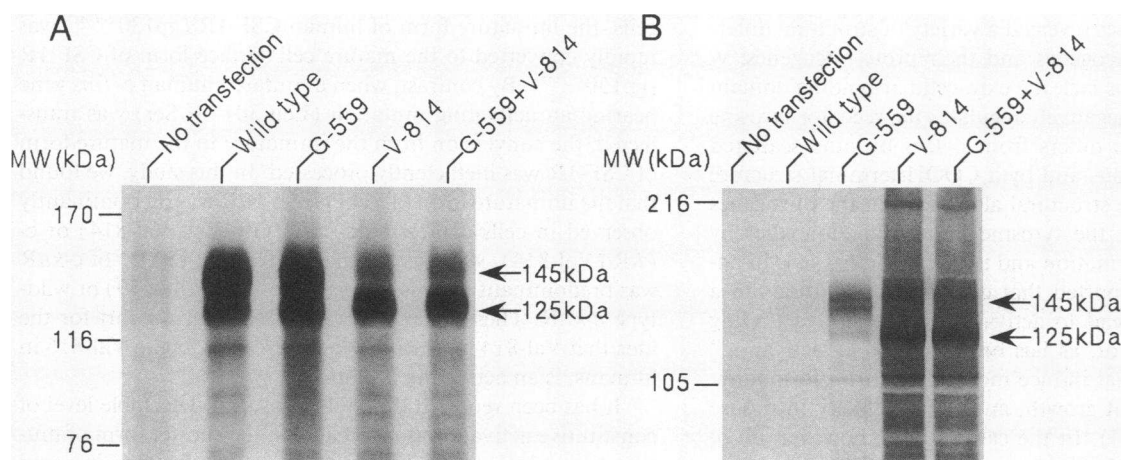


Figure 6. Expression of *c-kit* product (A) and identification of phosphotyrosine-containing proteins (B) in 293T cells transfected with wild-type and mutated murine *c-kit* genes. The cells were transfected with wild-type or mutated murine *c-kit* genes encoding wild-type *c-kitR* (wild type), *c-kitR* (Gly-559; G-559), *c-kitR* (Val-814; V-814), or *c-kitR* (Gly-559, Val-814; G-559, V814). Also, 293T cells that did not receive transfection were used as a negative control (no transfection). (A) The indicated cells were labeled for 6 h with 100 $\mu\text{Ci/ml}$ [^{35}S]methionine and lysed. *c-kit* product was then immunoprecipitated and analyzed by SDS-PAGE and autoradiography. (B) Lysates of the indicated cells were normalized for protein content, and were subjected to SDS-PAGE and immunoblotting with an anti-phosphotyrosine mAb. The mobilities of the mature (145-kD) and immature (125-kD) forms of *c-kit* products are indicated at right.

tively. When wild-type or mutated *c-kit* genes were transfected in 293T cells, the *c-kit* gene products were synthesized in the cells as 145- and 125-kD proteins (Fig. 6 A), which were the mature and immature forms of *c-kit* products, respectively (51). Although each transfectant expressed approximately equal amounts of *c-kit* products, a major *c-kit* product that incorporated [^{35}S]methionine was the immature form in the cells expressing *c-kitR* (Gly-559, Val-814) or *c-kitR* (Val-814), whereas it was the mature form in the cells expressing *c-kitR* (Gly-559) or wild-type *c-kitR* (Fig. 6 A).

Before and after transfection of *c-kit* genes, changes in tyrosine phosphorylation of proteins were detected by immunoblotting with anti-phosphotyrosine mAb (Fig. 6 B). As compared with nontransfected 293T cells, little or no increase in tyrosine phosphorylation was observed in the cells expressing wild-type *c-kitR*. However, a number of proteins, including 125- and 145-kD proteins, were heavily phosphorylated on tyrosine in the cells expressing *c-kitR* (Val-814) or *c-kitR* (Gly-559, Val-814). Also, a modest degree of tyrosine phosphorylation of 125- and 145-kD proteins was noted in the cells expressing *c-kitR* (Gly-559).

In order to further examine the tyrosine phosphorylation and activation of *c-kit* products in the transfected cells, *c-kit* products were immunoprecipitated and assayed by immunoblotting with an anti-phosphotyrosine mAb and also for auto-kinase activity. In the absence of SCF, both *c-kitR* (Val-814) and *c-kitR* (Gly-559, Val-814) were found to be abundantly phosphorylated on tyrosine, and *c-kitR* (Val-814) was also phosphorylated on tyrosine, albeit to a much lesser degree, whereas wild-type *c-kitR* was not (Fig. 7 A). Further, in accord with the findings on the immunoblotting analysis, immune complex kinase assay showed a striking autokinase activity in both *c-kitR* (Val-814) and *c-kitR* (Gly-559, Val-814), a weak activity in *c-kitR* (Gly-559), and little or no activity in wild-type *c-kitR* in the absence of SCF (Fig. 7 B).

Discussion

Many receptor tyrosine kinases were first identified as retroviral oncogenes such as *v-erbB*, *v-fms*, and *v-kit* that encode oncogenic forms of epidermal growth factor R (EGFR), CSF-1R, and *c-kitR*, respectively (1, 46, 52). Comparative nucleo-

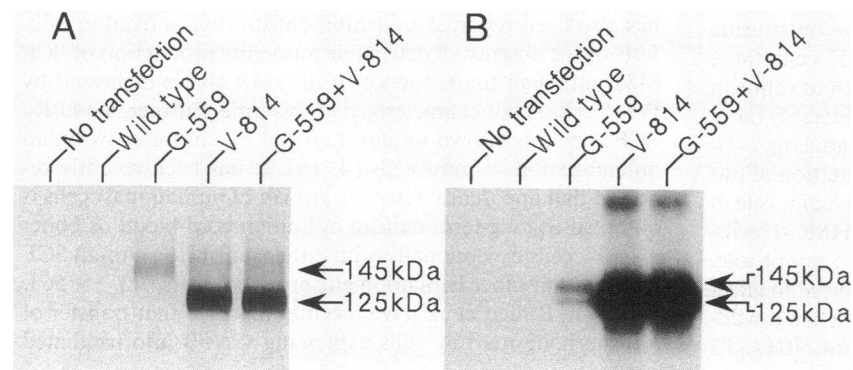


Figure 7. Tyrosine phosphorylation (A) and activation (B) of wild-type and mutated murine *c-kitR* expressed in 293T cells in the absence of exogenous SCF. (A) *c-kit* product was immunoprecipitated from the lysates of the indicated cells, and subjected to SDS-PAGE and immunoblotting with an anti-phosphotyrosine mAb. (B) Immune complex kinase assay for *c-kit* tyrosine kinase activity. *c-kit* product was immunoprecipitated from lysates from the indicated cells, and the immunoprecipitates were incubated in kinase buffer containing 10 $\mu\text{Ci/ml}$ [$\gamma\text{-}^{32}\text{P}$]ATP for 20 min at 25°C, washed, and separated by SDS-PAGE. Incorporation of ^{32}P into *c-kit* product was visualized by autoradiography. The mobilities of the mature (145-kD) and immature (125-kD) forms of *c-kit* products are indicated at right.

tide sequencing analyses revealed a variety of structural differences between the oncogenes and their proto-oncogenes: *v-erbB* and *v-kit* proteins lack the extracellular binding domain of the receptors that negatively regulates the receptor tyrosine kinase activity; *v-fms* differs from *c-fms* by nine scattered amino acid substitutions and by a COOH-terminal structure (3, 46, 53, 54). These structural alterations in the oncogenes activate constitutively the tyrosine kinase function, thereby leading to cell transformation and tumorigenicity (46). In addition, it has been reported that even point mutations in a proto-oncogene can lead to activation of the corresponding tyrosine kinase receptor, as has been shown by "activating" mutations of *c-fms* that induce morphologic transformation, anchorage-independent growth, and tumorigenicity in mouse NIH 3T3 cells (53, 54). In the case of *c-kit*, however, all of mutations so far described in mice, rats, and humans are reported to affect either the structure of *c-kit* or *c-kit* expression, resulting in a decrease in *c-kit* tyrosine kinase activity (4).

In this study, we have shown that *c-kit* product is constitutively phosphorylated on tyrosine and activated in HMC-1 cells even in the absence of SCF-stimulation. Further, *c-kit* product was found to be associated with PI3K without the exogenous SCF. The ligand-induced association of *c-kit* product with PI3K is reported to depend on autophosphorylation of *c-kit* product on tyrosine residues where the association occurs (48). Therefore, tyrosine residues that are constitutively phosphorylated in HMC-1 cells may be indistinguishable from those phosphorylated by ligand stimulation. However, neither mRNA transcript nor biological activity of SCF were detectable from HMC-1 cells. These results suggest that *c-kit* product in HMC-1 cells is constitutively activated in a ligand-independent manner.

Sequencing analyses have revealed that *c-kit* genes of HMC-1 cells are composed of a normal, wild-type allele and a mutant allele with two point mutations resulting in amino acid substitutions of Gly-560 for Val and Val-816 for Asp in the intracellular domain. In order to analyze the effect of these previously unidentified substitutions on *c-kit* kinase function, mutated murine *c-kit* genes encoding murine *c-kitR* (Gly-559 and/or Val-814), which were constructed by site-directed mutagenesis to correspond to human *c-kitR* (Gly-560 and/or Val-816), were transfected in a human embryonic kidney cell line, 293T cells. In the transfected cells, wild-type *c-kitR* was neither phosphorylated on tyrosine *in vivo* nor activated in immune complex kinase reaction in the absence of SCF. By contrast, both *c-kitR* (Gly-559, Val-814) and *c-kitR* (Val-814) were strikingly phosphorylated on tyrosine and activated in the absence of SCF. Further, *c-kitR* (Gly-559) was also phosphorylated on tyrosine and activated, albeit to a much lesser degree. Given the fact that human SCF is hardly active on murine *c-kitR* (6, 7) even if it might be produced from 293T cells, these results indicate that conversion of aspartic acid-814 to valine in murine *c-kitR* is an activating mutation and considerably more efficient than that of valine-559 to glycine in inducing *c-kit* kinase activity. It is therefore suggested that a mutation of human *c-kit* gene at codon 816 (Asp → Val) plays a major role in the constitutive activation of *c-kit* product in HMC-1 cells, while a mutation at codon 560 (Val → Gly) has a minor role.

The activating mutation of *c-fms* gene is reported to affect intracellular transport of human CSF-1R (54). When a wild-type human *c-fms* gene was transfected into murine NIH/3T3

cells, the immature form of human CSF-1R (gp130^{*c-fms*}) was rapidly converted to the mature cell surface form of CSF-1R (gp150^{*c-fms*}). By contrast, when a mutated human *c-fms* gene bearing an activating mutation (Leu-301 → Ser) was transfected, the conversion from the immature to the mature form of CSF-1R was inefficiently processed. In this study, we found that the immature form (125 kD) of *c-kitR* was predominantly observed in cells expressing *c-kitR* (Gly-559, Val-814) or *c-kitR* (Val-814), whereas the mature form (145 kD) of *c-kitR* was predominant in cells expressing *c-kitR* (Gly-559) or wild-type *c-kitR*. These results provide additional support for the idea that Val-814 in murine *c-kit*, corresponding to Val-816 in humans, is an activating mutation.

It has been reported that only a low but detectable level of constitutive activation is noticeable in the shorter form of murine *c-kit* product, which lacks the extracellular 4 amino acid sequence (Gly-Asn-Asn-Lys) due to the alternative splicing of mRNA (49). This shorter form of *c-kit* is found in the half of normal murine or rat *c-kit* cDNA clones (40, 49). Also in HMC-1 cells, we found that the half of *c-kit* cDNA clones lacks the same four-amino acid sequence in both normal and mutated *c-kit* genes. In order to compare *c-kitR* (Gly-559 and/or Val-814) with the shorter form of wild-type *c-kitR* and to determine the direct role of the *c-kit* mutations in the constitutive activation, we used the shorter form of wild-type murine *c-kit* cDNA as a template for the site-directed mutagenesis in this study. In the transfected 293T cells, tyrosine phosphorylation and activation were barely detectable in the shorter form of *c-kitR* which was used as a normal control, whereas they were readily detectable in *c-kitR* (Gly-559 and/or Val-814). These results indicate that *c-kit* mutations identified in this study are considerably more active than the shorter form of *c-kit*, and that the shorter form is unlikely to contribute to the constitutive activation of *c-kit* product in HMC-1 cells.

Although the mutations described here are found to lead to the constitutive activation of *c-kit* tyrosine kinase activity, the role of the *c-kit* activating mutations in leukemogenesis remains to be elucidated. In a previous study, we showed that *c-kit* product was phosphorylated on tyrosine in freshly prepared leukemic cells of some AML cases even in the absence of SCF stimulation (28). However, occurrences of tyrosine phosphorylation of *c-kit* product observed in the AML cases were not so striking as that in HMC-1 cells (28). Therefore, an activating mutation at residue 816 (Asp → Val) may not occur frequently in *de novo* AML, while it is still possible that certain mutations of *c-kitR* other than *c-kitR* (Val-816) may contribute to the modest degree of *c-kit* activation in AML cells. It is interesting to note that HMC-1 is a human mast-cell leukemia cell line (37), and that a mouse mastocytoma cell line, P-815, has also been reported to exhibit constitutive activation of *c-kitR* in the absence of detectable autocrine production of SCF (48), although the sequence of the *c-kit* alleles expressed by P-815 cells is not characterized at this time. Further, a *c-kitR*/SCF system is known to play a crucial role in the growth and differentiation of mast cells (4, 16). It has been recently reported that apparently selective growth of human mast cells is observed in long-term culture of human cord-blood or bone-marrow cells exceptionally with either murine or human SCF but not with other hematopoietic growth factors (43, 55, 56). Also, von Ruden et al. have recently reported that transfer of murine bone marrow cells expressing *v-erbB* into irradiated

mice results in the development of lethal systemic mast cell leukemia, and suggested that *v-erbB* may use parts of a signal transduction pathway normally coupled to *c-kitR* (57). These findings raise the possibility that activating mutations of *c-kit* described here may be involved in neoplastic transformation of mast cells, although it is as yet unclear whether these mutations by themselves can lead to factor-independent growth and tumorigenicity. It would be of considerable interest to examine whether the transfection of nontumorigenic or factor-dependent cell lines with the mutant forms of *c-kit* genes results in the production of a tumorigenic or factor-independent phenotype.

In addition to hematological malignancies, expression of *c-kit* proto-oncogene has been found in various tumors such as lung cancer and germ cell tumors (58, 59). Further, the amino acid sequence in a region of the kinase domain containing Asp-816 of human *c-kit* product is widely conserved in many other RTKs such as receptors for PDGF, CSF-1, and insulin (60). Therefore, it is possible that the unique activating mutation in codon 816 of human *c-kitR*, or a similar mutation in other RTKs, may participate in the development of a variety of malignancies. Further studies on *c-kit* mutants will be necessary to understand the mechanisms by which growth factor receptors influence oncogenesis and normal growth control.

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