

Microfilament-dependent activation of $\text{Na}^+/\text{K}^+/\text{2Cl}^-$ cotransport by cAMP in intestinal epithelial monolayers

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Correction

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Figure 3 has been revised to look as follows:

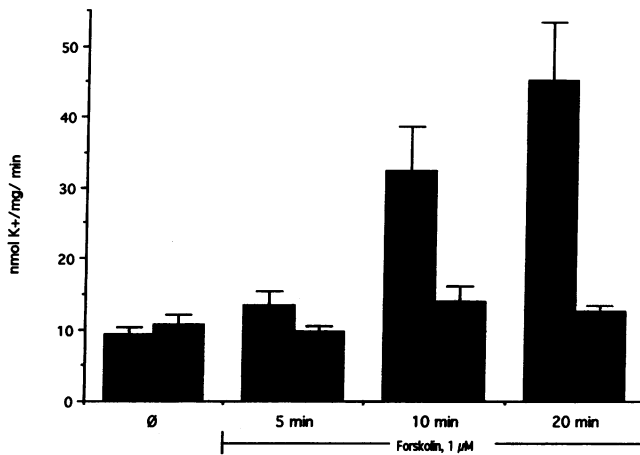


Figure 3. cAMP activates Na⁺/K⁺/2Cl⁻ cotransport in T84 cells. Forskolin (1 μM) increases Na⁺/K⁺/2Cl⁻ cotransporter activity (solid bars), measured by 1-min bumetanide-sensitive ⁸⁶Rb uptake using semiconfluent monolayers grown on 35-mm dishes 5 min ($n = 57$), 10 min ($n = 14$), or 20 min ($n = 7$) after addition of agonist compared to controls ($n = 35$) (by ANOVA $F = 16.49$, $P < 0.001$). In contrast, the bumetanide-insensitive component (hatched bars) shows no significant increase with forskolin stimulation. The vertical axis expresses ⁸⁶Rb as corrected for the specific activity of K⁺, for which it acts as a tracer, calculated as in (28). ■, Sensitive; ▨, insensitive.

Figure 4 has been revised to look as follows:

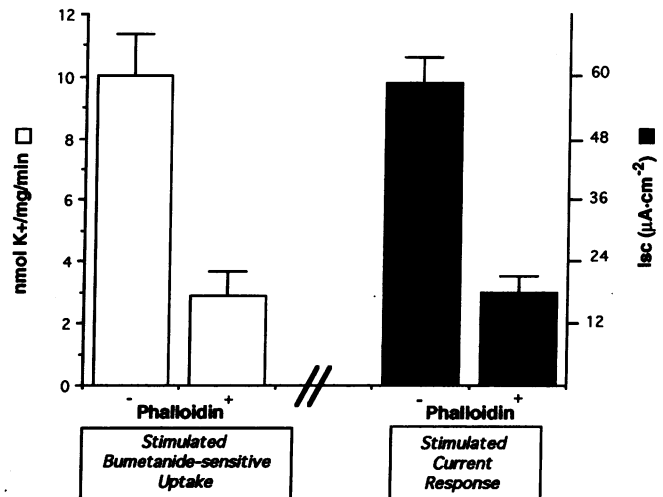


Figure 4. Phalloidin-loading inhibits cAMP-stimulation of Na⁺/K⁺/2Cl⁻ cotransporter activity in T84 cells. The left hand side of the figure (white bars) indicates that Na⁺/K⁺/2Cl⁻ cotransporter activity as measured by bumetanide-sensitive ⁸⁶Rb uptake under forskolin-stimulated conditions is inhibited under phalloidin-loaded conditions compared to controls ($n = 12$ for each, $P < 0.001$); the percent reduction in bumetanide-sensitive ⁸⁶Rb uptake is comparable to the inhibition of the forskolin-stimulated peak I_{sc} (black bars) under these conditions, shown in the right side of the figure, data taken from Fig. 1. The relative reduction of ⁸⁶Rb uptake was similar between cells grown on 35-mm dishes or permeable supports.