Suppression of normal human erythropoiesis by gamma interferon in vitro. Role of monocytes and T lymphocytes.

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Research Article

Interferons (IFN) have been shown to suppress the proliferation of human erythroid progenitors (erythroid burst-forming units [BFU-E] and colony-forming units [CFU-E]) in vitro. To examine the mechanism(s) underlying this inhibitory activity, the effect of different doses (50-10,000 U) of a highly purified preparation of recombinant DNA produced human gamma-IFN on erythroid colony formation by normal human bone marrow BFU-E and CFU-E in the presence and absence of monocytes and/or T lymphocytes was studied. The addition of gamma-IFN to whole marrow caused suppression of BFU-E (6-65%) and CFU-E (31-79%) in a dose-dependent fashion. This inhibition occurred both with the direct addition of gamma-IFN to the culture plates as well as by the preincubation of marrow cells with gamma-IFN followed by the washing of the cells; at the highest concentration of gamma-IFN (10,000 U), near-maximal inhibition of colony formation occurred with as little as 15 min of preexposure (BFU-E, 50%; CFU-E, 81%). Removal of monocytes and/or T lymphocytes before the addition of gamma-IFN significantly reduced the inhibitory effects of this lymphokine (BFU-E, -1 to 38%; CFU-E, -8 to 67%). Co-culture of purified autologous monocytes or T cells preexposed to gamma-IFN with monocyte and T cell-depleted marrow cells resulted in highly significant inhibition of erythroid colony formation even when these treated cells comprised less than 1% of the total nucleated cell populations in […]

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Suppression of Normal Human Erythropoiesis by Gamma Interferon In Vitro

Role of Monocytes and T Lymphocytes

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Abstract

Interferons (IFN) have been shown to suppress the proliferation of human erythroid progenitors (erythroid burst-forming units [BFU-E] and colony-forming units [CFU-E]) in vitro. To examine the mechanism(s) underlying this inhibitory activity, the effect of different doses (50–10,000 U) of a highly purified preparation of recombinant DNA produced human γ-IFN on erythroid colony formation by normal human bone marrow BFU-E and CFU-E in the presence and absence of monocytes and/or T lymphocytes was studied. The addition of γ-IFN to whole marrow caused suppression of BFU-E (6–65%) and CFU-E (31–79%) in a dose-dependent fashion. This inhibition occurred both with the direct addition of γ-IFN to the culture plates as well as by the preincubation of marrow cells with γ-IFN followed by the washing of the cells; at the highest concentration of γ-IFN (10,000 U), near-maximal inhibition of colony formation occurred with as little as 15 min of preexposure (BFU-E, 50%; CFU-E, 81%). Removal of monocytes and/or T lymphocytes before the addition of γ-IFN significantly reduced the inhibitory effects of this lymphokine (BFU-E, −1 to 38%; CFU-E, −8 to 67%). Co-culture of purified autologous monocytes or T cells preexposed to γ-IFN with monocyte and T cell-depleted marrow cells resulted in highly significant inhibition of erythroid colony formation even when these treated cells comprised <1% of the total nucleated cell populations in culture. The inhibitory action of γ-IFN was not prevented or reversed by erythropoietin. These results demonstrate that the inhibitory effects of γ-IFN on erythropoiesis are mediated to a significant degree through accessory cell populations, and suggest that γ-IFN may represent a useful tool in the study of the role of immunocompetent cells in the regulation of erythropoiesis in vitro.

Introduction

Interferons (IFN)[1] are biologically active molecules that are produced by nucleated cells in response to viral infections as well as to a number of other stimuli (1). As a class of molecules, IFN are capable of a diverse range of functions both in vivo and in vitro. Although initially recognized and defined by their antiviral properties (2–4), it has become obvious that this group of glycoproteins may play a significant role in cellular differentiation (5), proliferation (6, 7), and immunoregulation (8, 9).

IFN have been shown to suppress the proliferation and/or differentiation of normal human bone marrow pluripotent (CFU-GEMM) (10), granulocyte-macrophage (granulocyte-macrophage colony-forming units [CFU-GM]) (11), and erythroid (7, 12) progenitors (erythroid burst-forming units [BFU-E] and colony-forming units [CFU-E]) in vitro. In vitro suppression of murine bone marrow megakaryocyte colony-forming cells has also been reported (13). More recently, the possibility that IFN may play a role in the pathogenesis of aplastic anemia has been suggested (14, 15). In support of this hypothesis, it has been demonstrated that (a) IFN was a mediator of the in vitro suppression of hematopoiesis by cells from patients with aplastic anemia (15), and (b) IFN was present in higher than normal concentrations in bone marrow of seven of seven aplastic anemia patients (16). Moreover, the development of red cell aplasia, responsive to immunotherapy, in a patient with well-differentiated lymphocytic lymphoma after treatment with IFN has been documented (17). The mechanism(s) underlying the suppressive effect of IFN on hematopoiesis is not known. In this regard, it is generally accepted that immunocompetent cells, principally T cells and monocyte-macrophages (MO) play an important regulatory role in hematopoietic progenitor cell activity (18–21). Furthermore, increasing evidence points to IFN as a potent modulator of a variety of cell-mediated immune responses (22, 23); IFN have been shown to augment the cytotoxic activities of MO and natural killer cells (8, 24). The present study was designed to examine the role of T cells and MO in the mediation of the suppressive effect of a highly purified preparation of recombinant DNA-derived human γ-IFN on normal human bone marrow BFU-E and CFU-E in vitro. The results demonstrate that (a) γ-IFN is a potent suppressor of human erythropoiesis in vitro, (b) this inhibitory activity, which is not prevented/reversed by erythropoietin (Ep), is exercised more at the level of CFU-E than BFU-E, and more importantly, (c) T cells and MO play an important part in the mediation of this effect.

Methods

Cells and cell separation procedures. All bone marrow aspirations were performed at the posterior iliac crest. Heparinized (preservative-free) bone marrow and peripheral blood were obtained from normal healthy adult donors subsequent to obtaining informed consent. Bone marrow cells (BMC) and blood mononuclear cells were isolated by Ficoll-

1. *Abbreviations used in this paper:* BFU-E, erythroid burst-forming unit; BMC, bone marrow cell; BPA, burst-promoting activity; CFU-E, colony-forming unit; Ep, erythropoietin; FCS, fetal calf serum; IFN, interferon; IMDM, Iscove's modified Dulbecco's medium; MO, monocyte-macrophage; SRBC, sheep erythrocyte.

Hypaque density-gradient centrifugation (1.077 sp gr). When required, BMC were depleted of MO by adherence to plastic surfaces. 15 ml of BMC (5×10^6 cells/ml), suspended in Iscove's modified Dulbecco's medium (IMDM) with 20% fetal calf serum (FCS) was placed in 75-cm^2 plastic tissue culture flasks and allowed to adhere for 2 h at 37°C in a humidified atmosphere of 5% CO_2 in air. Nonadherent cells (BMC−MO) were collected by gentle agitation of the flasks and slow removal of the supernatant. The mixture was spun at 1,400 rpm for 10 min, washed once in IMDM and resuspended in IMDM at desired concentrations. Adherent cells for use in co-culture studies were removed by either gentle scraping with a rubber policeman or by exposure to EDTA (2% in phosphate-buffered saline) for 15 min followed by washing three times and resuspension in IMDM at desired concentrations. Removal of T lymphocytes from BMC−MO (BMC−MO−T) was achieved by double rosetting with sheep erythrocytes (SRBC); the second rosetting was performed overnight at 37°C, 5% CO_2 in humidified air. These procedures resulted in complete removal of MO and T cells as determined by the use of monoclonal antibodies LeuM3 (Becton, Dickinson Monoclonal Center, Inc., Mountain View, CA), MO2 (Coulter Immunology, Hialeah, FL), and OKT3 (Ortho Diagnostic Systems, Inc., Raritan, NJ) (25−28). Autologous T cells for co-culture experiments were obtained from PBMC−MO by single SRBC-rosetting and lysis of SRBC. Isolated T cells were on the average 95−97% SRBC$^+$ and OKT3$^+$. Isolated MO populations were evaluated for purity by a nonspecific esterase staining reaction (29), which revealed 90−99% nonspecific esterase-positive cells. Each of these separation procedures also resulted in a significant loss of total marrow nucleated cells. The percentages of total marrow nucleated cells recovered were: BM−MO, 49.9±5.0%; BM−T, 28.7±3.6%; BM−MO−T, 23.4±3.2%.

Clonal assays. Erythroid progenitors (BFU-E, CFU-E) were assayed in a plasma clot culture system as described (21). Bone marrow cells (BMC, 3−4 × 10^5 cells/1.1 ml; BMC−MO, 4 × 10^5 cells/1.1 ml; BMC−T, 2 × 10^5 cells/1.1 ml; BMC−MO−T, 2 × 10^5 cells/1.1 ml) were cultured in quadruplicate wells (0.1 ml/well) with Ep (0.5 IU/1.1 ml, unless otherwise specified) in the presence or absence of different concentrations of γ-IFN and/or autologous MO/T. The plates were incubated at 37°C in a humidified atmosphere of 5% CO_2 in air for 7 d (CFU-E) and 15 d (BFU-E). The clots were then removed, placed on glass slides, fixed with gluteraldehyde and stained with benzidine. Ep (CAT-1, 1972 IU/mg protein) used in these studies was generously supplied by the Blood Resources Division of the National Heart, Lung and Blood Institute of the National Institutes of Health.

Preincubation studies. The effect of γ-IFN on erythroid colony formation was examined in the following way. (*a*) Varying doses of γ-IFN (50−20,000 U/1.1 ml) were added to the different cell preparations at the start of incubation and the mixture was cultured for 7 and 15 d as described. (*b*) The exact numbers of bone marrow target cells to be cultured were preincubated with either control medium (IMDM, 10% FCS) or medium containing different doses of γ-IFN (800−20,000 U/ml) at 37°C in a water bath with gentle shaking for 15−60 min, washed one to four times (IMDM, 10% FCS, 1,400 rpm for 10 min), resuspended in complete culture medium and cultured as above. (*c*) Autologous T cells or MO were preincubated with either medium alone (IMDM, 10% FCS) or medium containing varying doses of γ-IFN (800−20,000 U/ml) at 37°C in a water bath with gentle shaking for 15−60 min, washed one to four times (IMDM, 10% FCS, 1,400 rpm for 10 min), resuspended in IMDM at desired concentrations and co-cultured with the different bone marrow cell preparations for 7 and 15 d. The numbers of MO or T cells used in co-culture studies comprised 0.24−8% of the total nucleated cell populations in culture.

Human interferon. The two highly purified recombinant DNA-derived preparations of human γ-IFN (Biogen, Inc., Cambridge, MA) used in these studies (93−97% pure) had the following characteristics: activity, 11.3×10^7 U/ml (National Institutes of Health reference units); sp act, 7×10^7 U/mg; pyrogens, 1.3 ng/ml; penicillin, ≤7.8 ng/ml; and *Escherichia coli* protein, 60 ng/ml. All dilutions were carried out in IMDM.

Results

The addition of γ-IFN to BMC resulted in significant inhibition of erythroid colony formation in a dose-dependent fashion (Fig. 1). This was evident not only by a significant decrease in the numbers of colonies formed, but also by the reduced size of individual colonies. Although the degree of inhibition was dependent on the dose of γ-IFN used, generally two plateaus were noted. The level of suppression was unchanged with doses between 50 and 800 U/1.1 ml, peaked, and plateaued again at 1,600−10,000 U/1.1 ml range (Fig. 1). Overall, the effect was more pronounced on CFU-E than BFU-E at lower doses of γ-IFN. However, a similar degree of suppression was seen when higher doses were employed (Fig. 1). The inhibitory effect of γ-IFN was completely abolished by the use of a monospecific anti-γ-IFN antibody (data not shown). In the studies reported here, except those described in Table I, the cultures were established in the presence of 0.5 IU of Ep/1.1 ml. This dose of Ep is used because in our hands, near-maximal stimulation of CFU-E-derived colony formation occurs at this dose and the effect on BFU-E falls within the linear portion of the dose-response curve; we frequently observe a plateau for BFU-E with doses of Ep ≥ 1 IU/1.1 ml. In addition, the suppressive effect of γ-IFN on both CFU-E and BFU-E is not affected by the dose of Ep employed (Table I).

Results presented in Table I demonstrate that the inhibition of erythroid colony formation by the direct addition of γ-IFN to the culture was independent of the dose of Ep used; concentrations of Ep between 0.5 and 5 IU/1.1 ml failed to prevent the suppression of CFU-E and/or BFU-E activity by γ-IFN. This was true both when the different Ep doses were added to culture at the start of incubation (Table I), and when BMC were preincubated with up to 10 IU of EP for 1 h before the addition of γ-IFN (data not shown). In these studies, cells were exposed to Ep (0.5−10 IU/ml) for 1 h, washed, and replated in the presence of 0.5−10 IU of Ep/1.1 ml.

The effect of different concentrations of γ-IFN on erythroid colony formation by BMC−MO (0% LeuM3$^+$, 0% MO2$^+$, 0.5% nonspecific esterase$^+$, 3−6% OKT3$^+$, 3−6% SRBC$^+$) is shown in Fig. 2. In these, as well as studies presented in Figs. 3 and 4, no attempt was made to present the results in terms of colonies produced by a given number of non-MO and/or T cells. As a result, the numbers for both classes of progenitors
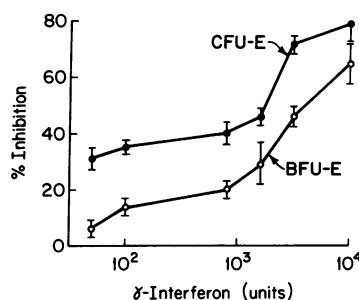

Figure 1. Effect of γ-IFN on erythroid colony formation by intact normal human bone marrow cells in vitro. Each data point on this linear-log plot represents mean±1 SEM of results from eight separate studies each involving a different normal marrow donor. Percentage of inhibition was calculated by reference to values obtained in control cultures: CFU-E, 305±25; BFU-E, 201±39 colonies/10^5 cells. When compared to control cultures, the effect on CFU-E was statistically significant (*P* < 0.05) beginning at γ-IFN dose of 50 U/ml; statistically significant (*P* < 0.05) inhibition of BFU-E was first detected at the 3,200 U/ml dose.

Table I. Effect of Increasing Concentrations of Ep on the Inhibitory Action of γ-IFN on Erythroid Colony Formation by Normal Human BMC in Vitro*

Additions to cultures	BFU-E	CFU-E	BFU-E	CFU-E
	Colonies/10⁵ cells‡	*Colonies/10⁵ cells‡*	*% inhibition*	*% inhibition*
None	0	0	—	—
0.5 IU Ep§	76±10	186±11	—	—
0.5 IU Ep§ + 3,200 U IFN	36±4	28±3	53	85
0.5 IU Ep§ + 10,000 U IFN	26±5	35±6	66	81
1.0 IU Ep	108±18	264±29	—	—
1.0 IU Ep + 3,200 U IFN	50±7	54±9	54	80
1.0 IU Ep + 10,000 U IFN	38±5	35±6	65	87
2.0 IU Ep	139±14	225±23	—	—
2.0 IU Ep + 3,200 U IFN	83±4	71±16	40	68
2.0 IU Ep + 10,000 U IFN	67±13	79±24	52	65
5.0 IU Ep	160±6	263±17	—	—
5.0 IU Ep + 3,200 U IFN	62±5	48±21	61	62
5.0 IU Ep + 10,000 U IFN	56±4	28±6	65	89

* Ep and γ-IFN were added at the start of culture. See text for details. ‡ Each value represents mean±1 SEM of results from two separate studies each involving a different normal donor. § Ep (CAT-1) used in these studies was provided by the Blood Resources Division, National Heart, Lung and Blood Institute, National Institutes of Health; 0.5 IU Ep/1.1 ml of culture was used.

are higher than those given in Fig. 1 and Table I, which simply reflects the concentration effects of the depletion procedures employed. It is interesting to note, however, that the relative increase in BFU-E density when both T cells and MO were removed was not as high as when the marrow was depleted of T cells alone (BMC−T, 650±89 vs. BMC−MO−T, 375±65 BFU-E/10⁵ cells). Similarly, removal of MO alone did not result in a significant increase in detectable BFU-E density (BMC, 201±39 vs. BMC−MO, 200±35 BFU-E/10⁵ cells). We interpret these results to indicate that adherent cells play a significant helper function in bone marrow BFU-E development in vitro. We did not feel it necessary, however, to perform these studies in the presence of added burst-promoting activity (BPA) because γ-IFN-mediated inhibition of erythropoiesis occurred even when intact BMC were used. Moreover, γ-IFN

was previously shown to inhibit erythroid colony development in cultures supplemented with exogenous BPA (12).

The results presented in Fig. 2 show that only modest inhibition of BFU-E occurred when γ-IFN was added directly to BMC−MO. Moreover, the suppressive effect of γ-IFN on the more mature CFU-E, although significant, was decreased when compared to BMC (4–8% LeuM3⁺, 2–5% MO2⁺, 4–11% OKT3⁺, 4–11% SRBC⁺). When γ-IFN was added directly to bone marrow cells depleted of T cells (BMC−T) (0% OKT3⁺, 0% SRBC⁺, 2–5% LeuM3⁺, 2–8% MO2⁺), again less inhibition of either CFU-E or BFU-E occurred over the dose range of 50–3,200 U (Fig. 3) when compared to γ-IFN effect on whole marrow. When γ-IFN was added to bone marrow cells depleted of both T cells as well as monocytes (0% OKT3⁺, 0% SRBC⁺,
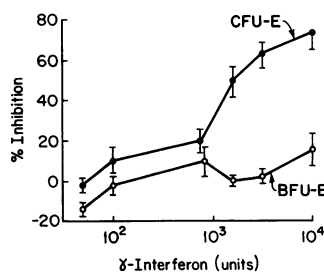

Figure 2. Effect of γ-IFN on erythroid colony formation by monocyte-depleted normal human bone marrow cells in vitro. Each data point on this linear-log plot represents results from six separate studies each involving a different normal marrow donor. Percentage of inhibition was calculated by reference to values obtained in control cultures: CFU-E, 271±33; BFU-E, 200±35 colonies/10⁵ cells. When compared to control cultures, the effect on CFU-E became statistically significant at γ-IFN dose of 1,600 U/ml (P < 0.05); the effect on BFU-E did not reach statistically significant levels at any dose of γ-IFN.
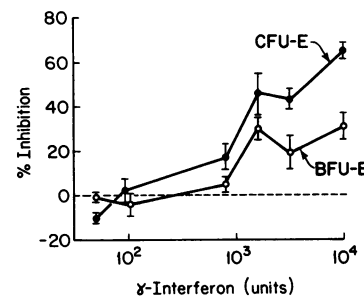

Figure 3. Effect of γ-IFN on erythroid colony formation by T cell-depleted normal human bone marrow cells in vitro. Each data point on this linear-log plot represents mean±1 SEM of results from three separate studies each involving a different normal marrow donor. Percentage of inhibition was calculated by reference to values obtained in control cultures: CFU-E, 821±79; BFU-E, 650±89 colonies/10⁵ cells. When compared to control cultures, the effect of CFU-E was statistically significant (P < 0.05) beginning at γ-IFN dose of 1,600 U/ml; the effect on BFU-E was not statistically significant.
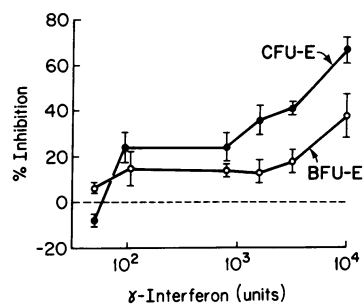
Figure 4. Effect of γ-IFN on erythroid colony formation by T cell and monocyte-depleted normal human bone marrow cells in vitro. Each data point on this linear-log plot represents results from four separate studies each involving a different normal marrow donor. Percentage of inhibition was calculated by reference to values obtained in control cultures: CFU-E, 613±76; BFU-E, 375±65 colonies/10^5 cells. When compared to control cultures, statistically significant (P < 0.05) inhibition of CFU-E was seen starting with 1,600 U/ml of γ-IFN; the effect on BFU-E was not statistically significant.

Table III. Effect of IFN on Erythroid Colony Formation by Normal Human BMC Preexposed to IFN*

Culture conditions	CFU-E/10^5 cells‡	% inhibition
BM + Ep	293±60	—
BM + Ep + 10,000 U IFN§	61±11	79
BM (preincubated) + Ep‖	262±8	—
BM (preincubated with 10,00 U IFN) + Ep	73±8	72
BM (preincubated with 10,000 U IFN) + Ep + 800 U IFN	41±3	84
BM (preincubated with 10,000 U IFN) + Ep + 1600 U IFN	44±4	83

* BMC were incubated with IFN or medium alone for 1 h, washed, and cultured in plasma clot in the presence of 0.5 IU Ep with and without IFN.
‡ Each value represents mean±1 SEM of results from two separate studies each involving a separate normal donor.
§ γ-IFN.
‖ Erythropoietin.

0% LeuM3⁺, 0% MO2⁺), only modest inhibition of BFU-E was seen (31% with 10,000 U of γ-IFN) and the inhibitory effect on CFU-E was also moderated (Fig. 4). To determine whether the suppression of erythropoiesis by γ-IFN in vitro requires the continuous presence of the lymphokine, we performed preincubation studies where BMC were exposed to varying doses of γ-IFN (1,600, 3,200, or 10,000 U) for 15, 30, or 60 min, washed, and then cultured in plasma clots in the presence of Ep alone. The results of these studies are shown in Table II and indicate that the γ-IFN-mediated suppression of CFU-E and BFU-E is both dose and time dependent. Thus at a dose of 10,000 U of γ-IFN, maximal inhibition of CFU-E and BFU-E occurred with as little as 15 min of preincubation. As seen in Table III, preincubation with 10,000 U of γ-IFN for 15 min, washing the cells, followed by the direct addition to culture of up to 1,600 U of γ-IFN resulted in no further inhibition of erythroid colony formation. These results suggest that the inhibitory effect of γ-IFN on CFU-E and BFU-E can result from very brief exposure to γ-IFN, that γ-IFN mediated suppression of erythroid colony formation is both dose and time dependent, and that at the doses utilized, near-maximal inhibitory effect can be achieved by preincubation.

To further delineate the role of immunocompetent cells in the mediation of γ-IFN effect on CFU-E and BFU-E, autologous T cells and MO (1–16 × 10^3 cells/1.1 ml) were preincubated with either control medium or γ-IFN (3,200–20,000 U), washed, and co-cultured with BMC−MO−T in the presence of Ep alone. Results presented in Tables IV and V demonstrate that the addition of as few as 1,000 γ-IFN treated T cells (Table IV) or MO (Table V) (comprising 0.24% of total nucleated cells present in culture) to autologous BMC−MO−T resulted in near-maximal inhibition of CFU-E-derived colony formation. The effect on BFU-E-derived colony formation, although significant, was less pronounced.

Discussion

The results presented here demonstrate that human γ-IFN is a potent inhibitor of human erythropoiesis in vitro. Thus the

Table II. Effect of Preexposure of Normal Human BMC to γ-IFN on Erythroid Colony Formation in Response to Ep In Vitro*

	IFN					
	1,600 U		3,200 U		10,000 U	
Time	BFU-E	CFU-E	BFU-E	CFU-E	BFU-E	CFU-E
min	*% inhibition‡*	*% inhibition‡*	*% inhibition‡*	*% inhibition‡*	*% inhibition‡*	*% inhibition‡*
15	26±8	31±11	38±14	49±15	50±10	81±10
30	29±12	39±15	59±28	82±18	65±8	68±7
60	42±12	46±9	64±19	76±21	55±4	68±8

* Cells were incubated with the indicated doses of γ-IFN for 15–60 min, washed, and cultured in the presence of 0.5 IU Ep (see text for details.) ‡ Calculated from mean colony counts obtained in three separate studies each involving a different normal marrow donor. Mean control (incubated without IFN) colony counts used to calculate the % inhibitions (mean±1 SEM) were: BFU-E, 140±15 colonies/10^5 cells and CFU-E, 231±26 colonies/10^5 cells.

Table IV. Effect of Autologous T Cells Preincubated
with γ-IFN on Erythroid Colony Formation by
T- and Monocyte-depleted Normal Human BMC in Vitro

Additions to cultures	BFU-E	CFU-E
	Colonies/10^5 cells*	Colonies/10^5 cells*
None	0	0
Ep‡	292±14	600±55
Ep‡ + (10^3 T + medium)§	299±40	389±104
Ep‡ + (10^3 T + 3,200 U IFN)	298±45 (0%)ǁ	96±22 (75%)
Ep‡ + (10^3 T + 10,000 U IFN)	144±20 (52%)	22±5 (95%)
Ep‡ + (10^3 T + 20,000 U IFN)	176±22 (41%)	16±5 (96%)
Ep‡ + (8 × 10^3 T + medium)	260±16	513±77
Ep‡ + (8 × 10^3 T + 3,200 U IFN)	250±30 (4%)	136±11 (74%)
Ep‡ + (8 × 10^3 T + 10,000 U IFN)	206±48 (21%)	86±6 (84%)
Ep‡ + (8 × 10^3 T + 20,000 U IFN)	185±23 (29%)	88±11 (83%)
Ep‡ + (16 × 10^3 T + medium)	321±39	559±39
Ep‡ + (16 × 10^3 T + 3,200 U IFN)	378±23 (−18%)	347±49 (38%)
Ep‡ + (16 × 10^3 T + 10,000 U IFN)	142±33 (56%)	99±22 (83%)
Ep‡ + (16 × 10^3 T + 20,000 U IFN)	144±23 (56%)	100±11 (82%)

* Mean±1 SEM of results from quadruplicate cultures.
‡ See legend to Table I; 0.5 IU Ep/1.1 ml was used.
§ T cells were incubated with medium or γ-IFN for 1 h, washed, and co-cultured with marrow cells.
ǁ Values in parentheses indicate % inhibition by direct comparison to medium controls for each T cell concentration used.

Table V. Effect of Autologous Monocytes Preincubated
with γ-IFN on Erythroid Colony Formation by T- and
Monocyte-depleted Normal Human BMC in Vitro

Additions to cultures	BFU-E	CFU-E
	Colonies/10^5 cells*	Colonies/10^5 cells*
None	0	0
Ep‡	292±14	600±55
Ep‡ + (10^3 monocytes + medium)§	307±34	493±66
Ep‡ + (10^3 monocytes + 3,200 U IFN)	353±46 (−15%)ǁ	277±66 (44%)
Ep‡ + (10^3 monocytes + 10,000 U IFN)	202±27 (34%)	97±17 (80%)
Ep‡ + (10^3 monocytes + 20,000 U IFN)	165±11 (46%)	
Ep‡ + (8 × 10^3 monocytes + medium)	320±22	504±72
Ep‡ + (8 × 10^3 monocytes + 3,200 U IFN)	377±25 (−17%)	301±39 (40%)
Ep‡ + (8 × 10^3 monocytes + 10,000 U IFN)	238±45 (26%)	106±6 (79%)
Ep‡ + (8 × 10^3 monocytes + 20,000 U IFN)	172±15 (46%)	112±16 (78%)
Ep‡ + (16 × 10^3 monocytes + medium)	311±22	514±33
Ep‡ + (16 × 10^3 monocytes + 3,200 U IFN)	327±45 (−5%)	326±31 (37%)
Ep‡ + (16 × 10^3 monocytes + 10,000 U IFN)	160±10 (49%)	105±11 (80%)
Ep‡ + (16 × 10^3 monocytes + 20,000 U IFN)	121±6 (61%)	126±23 (75%)

* Mean±1 SEM of results from quadruplicate cultures.
‡ See legend to Table I; 0.5 IU Ep/1.1 ml was used.
§ Monocytes were incubated with medium or γ-IFN for 1 h, washed, and co-cultured with marrow cells.
ǁ Values in parentheses indicate % inhibition by direct comparison to medium controls for each monocyte concentration used.

addition of γ-IFN to normal human BMC resulted in highly significant suppression of erythroid progenitor activity in vitro. This activity, which was completely prevented by anti-γ-IFN, was not mediated through the alteration of the activity and/or target-recognition ability of Ep, inasmuch as the addition of large concentrations of Ep to the cultures or preexposure of marrow cells to Ep failed to prevent/reverse the γ-IFN-mediated suppression of CFU-E and BFU-E. This is in accord with results reported by other investigators demonstrating the Ep-independent nature of interferon effect on erythropoiesis in vitro (30, 31). It is of interest to note, however, that increasing the concentration of colony-stimulating factor has been reported to completely overcome the inhibitory effects of interferons on CFU-GM in the murine system (32).

These results also demonstrate that the inhibitory action of γ-IFN on erythropoiesis is mediated, at least in part, through MO and T cells; removal of these cells before the addition of γ-IFN significantly reduced the suppressive effect of γ-IFN. In

this regard, γ-IFN appears to exert a differential effect on CFU-E and BFU-E. When utilizing BMC, CFU-E exhibited a greater sensitivity to γ-IFN, especially at lower doses, than BFU-E. This was also true when BMC−MO was cultured with γ-IFN; monocyte depletion resulted in the ablation of γ-IFN-mediated suppression of BFU-E but not CFU-E. On the other hand, removal of T cells and/or MO and T cells before the addition of γ-IFN caused a significant decrease in both BFU-E and CFU-E suppression. It should be pointed out that the observed decreased sensitivity of erythroid progenitors to γ-IFN in the absence of MO and/or T cells was not due to individual marrow donor differences; on several occasions, intact and depleted marrows from the same donor were used. In all cases, significantly reduced inhibition was seen when MO and/or T cells were removed.

The finding that the removal of these immunocompetent cells causes a significant reduction in the γ-IFN-mediated suppression of human erythropoiesis in vitro suggests that the

primary effect of γ-IFN may not be against BFU-E and CFU-E directly but rather through changes in the activity of accessory cell populations. However, a direct effect of γ-IFN on erythroid-committed progenitors or inhibition through activation of other non-monocyte, non-T accessory cells can not be ruled out. The bisigmoid nature of the CFU-E and BFU-E inhibition curves shown in Fig. 1, and the near-total elimination of the second (i.e., high dose) phase of the inhibitory effect by the removal of adherent cells (Fig. 2) suggest that γ-IFN more likely acts on two cell populations (erythroid progenitors and accessory cells). The findings reported here also show that, although the addition of either activated MO or activated T cells alone to BMC or BMC−MO−T can suppress erythropoiesis, removal of only one of these cell types markedly decreased γ-IFN-induced suppression of erythroid colony formation in vitro. This apparent paradox can best be explained by the possibility of involvement of a MO−T cell cooperation in the mediation of γ-IFN effect on erythropoiesis in vitro. In this regard, we have found that even when BMC rigorously depleted of MO and T cells were cultured, significant numbers of MO and T cells were detected by day 7 of incubation. It is possible that these newly formed cells participate in the mediation of the overall effect of γ-IFN on erythropoiesis in vitro.

It is now well established that the proliferation and differentiation of erythroid progenitors in vitro is subject to regulatory influences of a number of accessory cell populations (18–21). Normal T cells and MO through production/release of soluble products, stimulate the growth of BFU-E and CFU-E in vitro (18, 19). By contrast, T cells from some patients with blood dyscrasias, e.g., aplastic anemia (33), and MO from some patients with disseminated fungal infections (21) have been found to suppress the growth of autologous and heterologous BFU-E and CFU-E in vitro. Moreover, activation of normal T cells by exposure to mitogenic factors (e.g., phytohemagglutinin, PHA) can render these cells inhibitory to hematopoietic progenitors (34). It is possible that the inhibitory effect of γ-IFN reported here is mediated through a similar "activation" of T cells and/or MO. When purified populations of donor T cells or MO were exposed to γ-IFN for 60 min, washed, and co-cultured with autologous BMC, significant inhibition of erythroid colony formation occurred.

The mechanism(s) by which "activated" T cells or MO affect erythropoiesis is not known. Activation of T cells with phytohemagglutinin renders these cells inhibitory to erythropoiesis in vitro (34). A recent study suggests that the suppressive activity of lectin-treated T cells may be mediated through the production of γ-IFN by these cells (15). Activated T cells were found to produce an inhibitory supernatant with significant interferon activity (15). The suppressive activity of this supernatant was completely abrogated by treatment with antibody directed against γ-IFN (15). It is of interest to note, however, that similarly activated T cells have been documented to produce factors capable of promoting the growth and differentiation of hematopoietic precursors in vitro (18, 35). The studies of Zoumbos et al. (15) showed that the production of interferon by activated T cells peaked at days 3–5 of incubation and by day 7 were reduced to levels similar to those present in unstimulated cultures. This is in contrast to the finding that day 7 supernatants contain significant hematopoietic promoting factors (35). It appears, therefore, that the same stimulus can

instruct these cells to assume both inhibitory and stimulatory functions with regard to hematopoiesis.

It is possible that the inhibitory effect of γ-IFN-treated cells on erythroid progenitors is mediated via soluble factors. However, in preliminary studies, we found that media conditioned by these cells for 1–3 d were devoid of inhibitory activity. Thus it is unlikely that the inhibitory effects of γ-IFN-treated cells are mediated through the production of interferons. Moreover, the effect can not be attributed to the carry-over by the treated cells of amounts of interferon sufficient to significantly inhibit erythropoiesis in vitro; we observed similar degree of inhibitory activity regardless of whether the dose of cells used in co-culture studies were obtained from small or very large pools of cells treated with the same dose of γ-IFN. In addition, repeated washing of the treated cells before co-culture with autologous BMC failed to alter their inhibitory effects on erythropoiesis. An alternative possibility is that the observed inhibition is not mediated by a secretory product, but rather is the end result of specific cellular activation and an antiproliferative effect mediated via direct cell-to-cell contact.

It is now well established that the interferon-induced antiproliferative activity in other systems is mediated via cell-to-cell contact (36). The transfer of interferon-induced antiviral activity by a variety of cell types was not mediated by soluble factors released by these cells (37). The effect appears to require some type of as yet undefined attachment between the effector cell and the target cell (36); gentle agitation of a mixture of IFN-treated L cells and Daudi cells prevented the transfer of the antiproliferative activity of the former cells (36).

IFNs are increasingly being considered as therapeutic agents for solid tumors and a variety of hematologic malignancies (38). The studies described here provide in vitro evidence that the antiproliferative effects of IFN may not be limited to intended abnormal cells but may extend to normal hematopoietic progenitors as well. Indeed, hematologic toxicity has been observed with interferon therapy, neutropenia being a commonly used reason for discontinuation of treatment (39, 40). It has been suggested that increased production of endogenous interferons may represent a pathogenic mechanism for the development of pancytopenia in some patients with aplastic anemia (14–16). A number of studies have demonstrated the presence in some patients with aplastic anemia of T lymphocytes capable of inhibiting the orderly proliferation and/or differentiation activities of autologous and heterologous hematopoietic progenitors in vitro (17, 33). In view of the findings presented here, the demonstration of increased levels of IFN in bone marrow of a majority of patients with aplastic anemia (15, 16), and the observation that the addition of anti-interferon antibody to bone marrow of aplastic anemia patients in vitro results in marked increase in hematopoietic colony formation (15, 16), the possibility is raised that the presence of T cells with hematopoietic suppressor activity in these patients may be associated with IFN. Direct evidence in support of such an association has recently been described in a patient with diffuse well-differentiated lymphocytic lymphoma undergoing IFN therapy (17). Administration of IFN to this patient resulted in a severe pancytopenia characterized by reduced marrow CFU-E activity and increased numbers of marrow suppressor T cells (OKT8+ cells). Immunosuppressive therapy resulted in marked improvement associated with a

reduction in the numbers of OKT8$^+$ T cells (17). Whether IFN therapy was responsible for the increase in suppressor T cells is not known.

Our results also show that significant suppression of erythropoiesis occurred in the presence of relatively larger concentrations of γ-IFN than has been previously reported (12). It is likely that differences in experimental procedure and design played a significant role in bringing about these differences. In this regard, Broxmeyer et al. (12) employed an external source of BPA in their cultures that may have recruited a more immature population of BFU-E than those observed in our studies. Such cells may exhibit a different pattern of response to γ-IFN. It is also possible that the erythroid progenitors may exhibit a greater sensitivity to γ-IFN when cultured in methylcellulose (12) than plasma clot. Finally, the differences may be due to the different preparations of γ-IFN used. The natural γ-IFN used by Broxmeyer et al. (12) may be intrinsically more effective than the recombinant γ-IFN used in these studies.

Acknowledgments

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