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Research Article

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Ipodate (Oragrafin), 12 g orally in two subjects, caused much smaller changes in hepatic and serum radioactivity. However, ipodate also caused a doubling of the percent free T_4 in the serum as judged by equilibrium dialysis, and the ratio of hepatic radioactivity to free [125 I] T_4 in serum declined markedly after ipodate, similar to the fall in hepatic:serum 125 I ratios after tyropanoate. BSP, 20 mg/kg i.v. in three subjects, caused a smaller change; the decline in hepatic T_4 content [...]

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Inhibition of Hepatic Binding of Thyroxine by Cholecystographic Agents

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ABSTRACT Subsequent to studies indicating that cholecystographic agents and sulfobromophthalein (BSP) inhibit uptake of thyroxine (T_4) by rat liver slices, the effect of such compounds on hepatic storage of T_4 in man has been examined. After intravenous administration of [125 I] T_4 to five normal subjects, hepatic radioactivity, estimated by external gamma counting, rose to a peak in ~4 h and then declined in parallel with serum radioactivity. When a 6-g dose of the cholecystographic agent, tyropanoate (Bilopaque), was administered orally 3 d later, estimated hepatic extravascular radioactivity fell 50–60% within 4 h and then rose toward the pretyropanoate value. Concomitant with the fall in hepatic radioactivity, serum radioactivity rose 57–70%, as did stable T_4 levels in serum, suggesting that hormone discharged from the liver entered the serum. Both uptake of T_4 and discharge by tyropanoate were much less in two patients with liver disease.

Ipodate (Oragrafin), 12 g orally in two subjects, caused much smaller changes in hepatic and serum radioactivity. However, ipodate also caused a doubling of the percent free T_4 in the serum as judged by equilibrium dialysis, and the ratio of hepatic radioactivity to free [125 I] T_4 in serum declined markedly after ipodate, similar to the fall in hepatic:serum 125 I ratios after tyropanoate. BSP, 20 mg/kg i.v. in three subjects, caused a smaller change; the decline in hepatic T_4 content averaged 19%.

We conclude that these organic anions, tyropanoate, ipodate, and BSP, all can displace T_4 from the liver. The results imply a link between T_4 transport and organic anion transport, and indicate a mechanism for altering hepatic T_4 content in man that could be the site of physiologic regulation or of disease. A method is described whereby analysis of the change in hepatic and plasma radioactivity after tyropanoate could be employed to estimate hepatic T_4 content in diverse clinical circumstances.

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INTRODUCTION

It was once thought that iodine-containing radiographic contrast agents affect thyroid function chiefly by releasing inorganic iodide. However, two drugs employed for oral cholecystography, iopanoate (Telepaque, Winthrop Laboratories, N. Y.) and ipodate (Oragrafin, E. R. Squibb & Sons, Princeton, N. J.), are now known to cause a major decline in circulating levels of the potent thyroid hormone triiodothyronine (T_3)¹ by virtue of their ability to inhibit peripheral conversion of thyroxine (T_4) to T_3 (1, 2). Cholecystographic agents also inhibit conversion of T_4 to T_3 in vitro (3–5).

Yet another effect of iopanoate and ipodate on thyroid hormone economy has recently been demonstrated in our laboratory, namely, their ability to inhibit uptake of T_4 and T_3 by rat liver slices; similar effects were produced by sulfobromophthalein (BSP) and bilirubin (4). The findings suggested that the agents might be able to discharge thyroid hormones from the liver in vivo. To study this possibility in man, we chose the agent sodium tyropanoate (Bilopaque, Winthrop Laboratories) because Suzuki et al.,² in comparing the effect of various agents on hormone levels, found that this drug provoked an acute and significant increase in serum T_4 . Our results indicate that a major discharge of T_4 from the liver follows tyropanoate ingestion; ipodate and BSP cause a similar but less striking discharge.

METHODS

Subjects. Five healthy volunteers were studied: three white men and two white women (Table I). All had normal

¹ *Abbreviations used in this paper:* BSP, sulfobromophthalein; ISF, interstitial fluid, RIA, radioimmunoassay; T_3 , triiodothyronine; T_4 , thyroxine.

² The following work was published after manuscript was written: Suzuki, H., N. Kadana, K. Takeushi, and S. Nakagawa. Effects of three-day oral cholecystography on serum iodothyronines and TSH concentrations: comparison of the effects among some cholecystographic agents and the effects of iopanoic acid on the pituitary-thyroid axis. *Acta Endocrinol.* 92: 477–488.

TABLE I
Clinical Data of Subjects

Subject	Age	Sex	Height	Weight	T ₄ RIA	T ₃ RIA	T ₃ resin uptake	Thyrotropin	Comment
	<i>yr</i>		<i>cm</i>	<i>kg</i>	$\mu\text{g/dl}$	ng/dl	%	$\mu\text{U/ml}$	
1	43	M	183	68	6.0	106	42	6.0	Recurrent pancreatitis
2	57	M	169	65	5.9	107	44	3.0	Normal
3	27	M	166	59	9.5	104	37	3.2	Normal
4	57	F	152	62	6.8	102	40	1.9	Hashimoto's thyroiditis*
5	55	F	166	64	15.0	198	49	1.5	Hashimoto's thyroiditis*
6	41	M	185	82	7.6	63	49	3.0	Alcoholic hepatitis
7	58	M	175	68	3.4	50	63	3.5	Viral hepatitis
Normal range					4.1–11.3	70–200	33–45	<6.0	

* Both women were receiving 0.2 mg levothyroxine daily.

liver function as evidenced by normal serum tests of liver function (bilirubin, alkaline phosphatase, glutamic-oxaloacetic transaminase) and normal values for BSP excretion. All were clinically euthyroid, and four had normal values for serum T₄ by radioimmunoassay (RIA), T₃ resin uptake, and thyrotropin. The two women had euthyroid goiter, clinically diagnosed as Hashimoto's thyroiditis and treated with 0.2 mg levothyroxine daily; subject 5 had modestly elevated hormone levels. Two male in-patients with liver disease were also studied (Table I). During the period of study, serum bilirubin levels averaged 12.0 mg/dl in both subject 6 and 7, and respective values were 141 and 31 U/liter (normal, <25 U/liter) for serum glutamic-oxaloacetic transaminase and 3.6 and 2.6 g/dl for serum albumin. The study was approved by the Human Subjects Review and Radiation Safety Committees of the University of Washington; informed consent was obtained in writing from each subject.

Standard protocol. Each subject was first started on saturated solution of potassium iodide, 5 drops orally every 12 h, to block thyroidal uptake of labeled iodide. After intravenous injection of 50 μCi of ¹²⁵I-labeled T₄ (Industrial Nuclear Co., St. Louis, Mo.), serial counts were made over the hepatic area, employing a shielded sodium iodide crystal detector. Additionally, serum samples were obtained at the same times as the liver counts, and counted together with standards prepared from the injection solution in a well-type scintillation detector (Beckman Gamma 4000, Beckman Instruments, Inc., Fullerton, Calif.). All counting was performed with a relative error of <3%.

After ~72 h, long enough for hepatic and serum counts to reach a steady state of decline, a 6-g dose of sodium tyropanoate was administered orally; liver counting and blood sampling continued for another 24–48 h. The patient was fasting on the morning of drug administration to promote more rapid absorption of the gallbladder agent; the fast was ended 4 h after drug ingestion.

4–7 d after tyropanoate administration, when liver:serum ¹²⁵I ratios had stabilized near their original level, three subjects were given BSP intravenously, 5 mg/kg on one day and 20 mg/kg on the following day. Serial liver counts and blood specimens were obtained as in the standard protocol.

Two volunteers were retested with ipodate. The protocol was identical except that the dose of ipodate was 12 rather than 6 g. One subject was also retested using a 3-g and later a 9-g dose of tyropanoate.

External counting over the liver. Subjects were studied at two hospitals; at one the detector has a 2-in. sodium iodide crystal and at the other a 3-in. crystal. Gamma energies

from 20 to 65 keV were counted and were corrected for room background. Using a point source of ¹²⁵I in a water bath, the half-thickness value for both probes was 0.75 cm, i.e., the count rate decreased 50% for each 0.75-cm distance in water from the crystal. Because of this limited range, heavy collimation to block out radioactivity from distant perihepatic tissues seemed unnecessary, and most subjects were studied without a collimator, applying the detector directly to the chest wall. The limited range did mean, however, that chest wall tissues could make a significant contribution to net count rates. Assuming a chest wall thickness of 2.5 cm, and an "infinite" thickness of liver beyond, chest wall tissues would be counted at about eight times the efficiency of the liver and could contribute 10% to net counts even though the relative isotope concentrations in liver and chest wall were 50:1. Although the major conclusions of the present study are not compromised by this factor, a correction for chest wall activity would be necessary for precise quantitation of hepatic ¹²⁵I content. The chest wall factor would be less, employing ¹³¹I-labeled hormones that have a more energetic gamma emission; [¹³¹I]T₄ was not readily available to us.

Special analyses on serum. Two additional procedures were used to determine the nature of the increased serum radioactivity produced by tyropanoate. First, a serum sample obtained from one subject at the peak of the tyropanoate effect was subjected to column chromatography on Sephadex G-25, using a technique previously described (6). Second, serum samples obtained from subject 5 just before tyropanoate and also at the peak of the tyropanoate effect were subjected to precipitation with trichloroacetic acid (TCA), using a modification of the technique of Lepp et al. (7). After initial counting, 0.125 ml serum was mixed with 1.75 ml distilled H₂O; 0.5 ml of 40% TCA was then added. After mixing, the tube was centrifuged for 20 min at 2,000 rpm, the supernate decanted, and the tube recounted.

Serum samples from the ipodate studies were also subjected to equilibrium dialysis to determine the free T₄ fraction, using the technique of Sterling and Brenner (8).³

Analysis of data. Data analysis was similar to that of Cavalieri and Searle (9). L(t) is the count rate over the liver at time (t) after T₄ injection, normalized to a value of 100 at t = 0. Similarly, S(t) is the count rate per unit volume of

³ We are grateful to Dr. Margaret Kenny of the Department of Laboratory Medicine, University of Washington, for performing these determinations.

serum or plasma, normalized to 100 at $t = 0$. Count rates at $t = 0$ were estimated by extrapolation of the early portions of the hepatic uptake and serum disappearance curves, respectively. It is assumed that $L(0)$ is the result of radioactivity in the hepatic vascular space seen by the probe, and that the contribution of hepatic intravascular counts to total liver counts at any later time t can be estimated as $L(0) \times S(t)/S(0)$, or, because $L(0)$ and $S(0)$ are both normalized to 100, simply as $S(t)$. Therefore, a ratio $L(t):S(t) > 1.0$ implies entry of tracer into the hepatic extravascular space, and the difference, $L(t) - S(t)$, should be directly proportional to the hepatic extravascular radioactivity. Also, the percent change in hepatic extravascular T_4 content between any two times, t_1 and t_2 , can be expressed as

$$\Delta(L - S) = \frac{L(t_1) - S(t_1) - (L[t_2] - S[t_2])}{L(t_1) - S(t_1)} \times 100. \quad (1)$$

RESULTS

Effects of tyropanoate. Fig. 1 shows the results of the complete study in subject 3. There was a rapid initial uptake of hormone by the liver, reaching a maximum ~ 4 h after injection. Hepatic and serum ^{125}I then gradually declined over the next 72 h, approximately in parallel. When tyropanoate was given, a prompt fall in hepatic radioactivity ensued, reaching a nadir ~ 4 h later. There was a marked rise in the serum radioactivity at the same time that hepatic radioactivity fell, thus providing strong evidence that tyropanoate causes a discharge into the circulation of thyroxine stored within the liver. Over the next 48 h, both liver and serum radioactivity returned toward values that would have been predicted had the drug not been administered, suggesting that the effect is reversible. This is consistent with tyropanoate's relatively brief biologic half-life, being rapidly excreted into bile and then removed in the feces (10).

The results in each of the five normal subjects were similar to those depicted in Fig. 1. As shown in Fig. 2, liver:serum ^{125}I ratios rose to a plateau; individual plateau values varied from 10.4 to 15.5, and half maximum ratios were reached in 1–2 h. After ad-

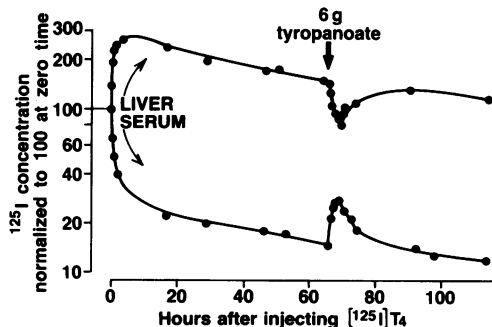


FIGURE 1 Effect of tyropanoate on labeled T_4 in liver and serum. All ^{125}I counts have been normalized to 100 at time of initial injection of labeled hormone.

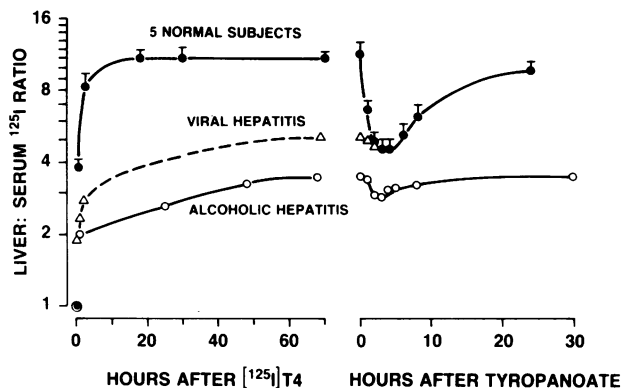


FIGURE 2 Effect of tyropanoate on hepatic T_4 content in normal subjects and in patients with liver disease. Liver:serum ^{125}I ratios, normalized to 1.0 at the time of $[^{125}\text{I}]\text{T}_4$ injection, are shown for five normal subjects (\bullet), one patient with alcoholic hepatitis (\circ), and one with viral hepatitis (Δ).

ministration of tyropanoate, the ratio fell in 2–4 h to a nadir that varied from 2.8 to 5.3.

Fig. 3 shows all five subjects plotted together, with the values for liver:serum ratios having been normalized to a value of 1.0 at the time of tyropanoate administration. Expressed in this manner, there was a remarkable uniformity in the pattern of decline in the liver:serum ratios. Also shown in this figure is the result in one subject who was retested with 3- and 9-g doses of tyropanoate; the change in liver:serum ratios was roughly proportional to dosage. In conjunction with the 9-g dose, serial counts were made over the left lower quadrant of the abdomen and over the right and left upper chest. In each site, concomitant with the rise in serum ^{125}I content and the fall in count rate over the liver, tyropanoate administration was followed by a rise in the count rate.

In the serum sample obtained at the peak of the tyropanoate effect, and subjected to column chro-

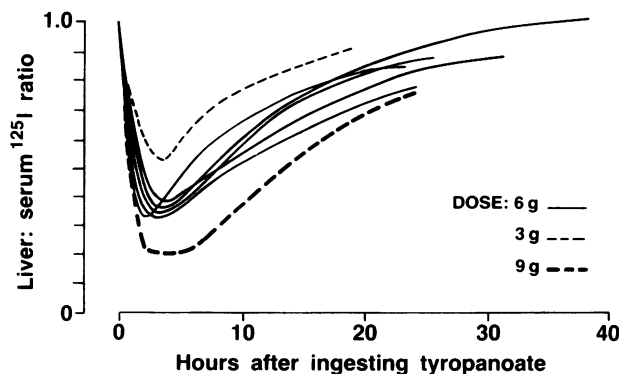


FIGURE 3 Effect of tyropanoate on hepatic $[^{125}\text{I}]\text{T}_4$ content. The liver:serum ratio at the time of tyropanoate administration has been normalized to a value of 1.0. The results in one subject who was restudied with 3- and 9-g doses of tyropanoate are shown by the light and heavy dashed lines, respectively.

matography, 90% of the radioactivity migrated in the expected location of T₄. This is confirmation that the radioactivity discharged from the liver is T₄ rather than a metabolic breakdown product. The results obtained by TCA precipitation provide additional confirmation. In a control sample of serum, to which a known amount of [¹²⁵I]T₄ had been added, 90% of the radioactivity was precipitated with TCA. In a serum sample drawn just before giving tyropanoate, the value was 92%, and in two samples taken near the peak of tyropanoate's effect, the values were 89 and 93%. This is evidence that breakdown of T₄ to iodide is not a major part of the response to tyropanoate administration.

The pattern of change in hepatic extravascular radioactivity ($\Delta[L - S]$, defined in Methods) and in serum radioactivity in the five studies with 6 g tyropanoate is shown in Fig. 4. The proportionate change in serum radioactivity exceeded that in the liver, suggesting that the amount discharged was a smaller proportion of hepatic radioactivity than of serum radioactivity. Serum radioactivity also peaked first, suggesting that egress from serum to nonhepatic sites occurred and equalled the rate of ingress from liver 2–3 h after tyropanoate ingestion. If all the radioactivity leaving the liver enters the serum, the ratio between the changes in serum and hepatic radioactivity, at an early time point before significant ¹²⁵I has left the serum for nonhepatic sites, should equal the ratio between the hepatic T₄ distribution volume and the plasma volume. Table II summarizes the data at the first time point for which complete data are available, 1 h. The mean ratio was about 1.8, suggesting an hepatic distribution volume 1.8 times the plasma volume. Possible errors in this computation are considered in the Discussion.

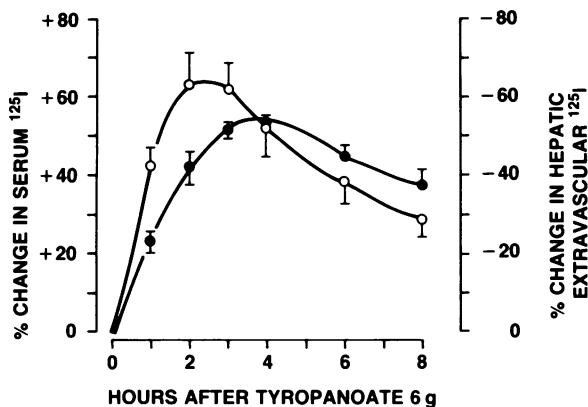


FIGURE 4 Changes in serum ¹²⁵I and in hepatic extravascular ¹²⁵I after tyropanoate. Values for serum radioactivity (○) are expressed as percent increase over the value at the time of tyropanoate ingestion. Hepatic extravascular radioactivity (●), estimated as described in Methods, is expressed as percent decrease from the value at the time of tyropanoate ingestion. Each symbol depicts mean and SE from five subjects.

TABLE II
Estimating the Ratio between Hepatic T₄ Distribution Volume and Plasma Volume

Subject	Dose of tyropanoate	ΔS	$\Delta(L - S)$	$\Delta S/\Delta(L - S)^*$
	g	% in 1 h	% in 1 h	
1	6	25.6	18.6	1.38
2	6	44.7	25.1	1.78
3	6	46.3	25.5	1.82
4	6	37.1	15.1	2.46
5	6	55.1	30.5	1.81
Mean \pm SE		41.8 \pm 5.0	23.0 \pm 2.7	1.85 \pm 0.17
1	3	21.5	13.4	1.60
1	9	67.0	36.7	1.83

* As discussed in the text, this ratio should be a function of the ratio between hepatic T₄ distribution volume and plasma volume.

Computing the percent fall in hepatic extravascular radioactivity, $\Delta(L - S)$, from the time of tyropanoate ingestion to the nadir, the mean value \pm SE was 55.3 \pm 2.0 (range, 52.2–60.7).

In selected serum samples, T₄ RIA was measured to determine whether changes in serum radioactivity were accompanied by similar changes in serum-stable T₄ concentrations. Compared with values just before tyropanoate ingestion, samples taken 2 h later, in three subjects, showed a mean percent increase in ¹²⁵I concentration, \pm SD, of 68 \pm 8%, although the mean increase in stable T₄ was 67 \pm 15%. At 8 h after tyropanoate, in four subjects, ¹²⁵I concentration was still 27 \pm 4%, and stable T₄ concentration 31 \pm 8%, above pretyropanoate values. These changes in stable T₄ levels are comparable with those reported by Suzuki et al.,² who found a 25% increase in serum T₄ 12 h after a 3-g dose of tyropanoate.

In the two patients with liver disease, hepatic T₄ uptake, as judged by liver:serum ¹²⁵I ratios, was much less than in the normal subjects (Fig. 2). Tyropanoate caused very little discharge from the liver of the patient with alcoholic hepatitis. Hepatic discharge was probably also minimal in the patient with viral hepatitis, although data are incomplete as the result of poor patient cooperation. These results indicate, as would be expected, that the storage pool affected by tyropanoate is markedly diminished in severe liver disease.

Effects of ipodate. Fig. 5 shows the effect of another cholecystographic agent, ipodate (Oragrafin), on labeled T₄ in liver and serum in subject 2; similar results were obtained in subject 1. Because of a report that ipodate may inhibit the binding of thyroxine to plasma proteins (11), the free T₄ fraction in each serum sample was determined by equilibrium dialysis, allowing estimation of the serum concentration of free

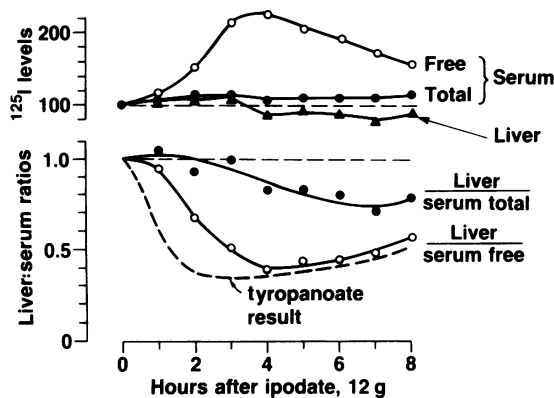


FIGURE 5 Effect of ipodate on hepatic T_4 content. All ^{125}I levels have been normalized to a value of 100 at the time of ipodate administration. The top half of the figure depicts changes in the serum, both free and total, and in the liver. The bottom half of the figure depicts the ratios of liver to serum-total and of liver to serum-free counts.

labeled T_4 . During the 8-h period after the administration of a quadruple dose of ipodate, 12 g, there were only small changes in total serum radioactivity and in the liver radioactivity, so that the ratio of hepatic to serum-total radioactivity showed only a modest decline after ipodate. There was, however, a major rise in the free T_4 counts, reaching a peak 4 h after administration of ipodate. Therefore, the ratio of hepatic to serum-free radioactivity underwent a major decline, with a temporal course reminiscent of the result with tyropanoate (Fig. 5). Thus, it appears that ipodate also can discharge T_4 from binding sites in the liver, but this is obscured by its simultaneous effect of displacing the hormone from its binding sites on plasma proteins.

Effects of BSP. These experiments were performed to provide a further comparison with our findings in rat liver slices, where BSP was as effective as cholecystographic agents in inhibiting T_4 uptake (4). With the small dose, 5 mg/kg, any effect was smaller than the variability of the methods. The larger dose, 20 mg/kg, caused a rise in serum radioactivity and a fall in hepatic radioactivity in all three subjects tested. The maximum changes in hepatic T_4 content, by Eq. 1, were 26, 20, and 13% in the three subjects.

DISCUSSION

It is well known that the liver plays an important role in the distribution of thyroid hormones. Cavalieri and Searle (9) have speculated that the hepatic pool of T_4 acts as a buffer to modulate abrupt changes in the level of circulating hormone. Other workers have shown that alterations in the storage pool of thyroid hormone occur in a variety of pathophysiologic settings. Nicoloff and Dowling (12) and Busnardo et al. (13) have reported that hepatic accumulation of labeled T_4 is increased

in subjects with hyperthyroidism and decreased in hypothyroid subjects. Patients with hepatic disease are known to have a reduction in hepatic T_4 content (9, 12, 14, 15), presumably as the result of a decrease in the functioning mass of liver. In euthyroid subjects with normal liver function, the hepatic T_4 distribution volume is a function of the percent free T_4 in plasma, and hence is decreased when levels of plasma T_4 binding globulin are increased by estrogen treatment (16). Conversely, the hepatic T_4 distribution volume is increased when T_4 binding activity in plasma is decreased, as it is in patients with congenitally low levels of T_4 binding globulin (12, 14, 16, 17), in subjects given androgens to lower T_4 binding globulin levels (18), and in subjects given amounts of salicylates sufficient to displace T_4 from its binding proteins (19). Thus, the expected effect of a compound that increases percent free T_4 , such as ipodate, would be to increase liver:serum T_4 ratios; its failure to do so in the present study suggests an additional effect of ipodate on hepatic T_4 uptake.

Tyropanoate, ipodate, and BSP, the three compounds shown to discharge T_4 from liver in this study, are members of a larger group of organic anions taken up by the liver. Our results, therefore, imply a relationship between accumulation of thyroid hormones and of other organic anions by the liver. The findings also raise the possibility that other factors, both endogenous and exogenous, might alter hepatic hormone content and thus regulate the effects of thyroid hormones on hepatic function. Two possible mechanisms for the effects we have demonstrated can be considered: inhibition of binding to cytosol proteins and inhibition of transport through the plasma membrane.

Several groups of workers have identified specific proteins in the cytosol of liver cells that bind thyroid hormones, employing hepatic tissue from several species, including rat (20–22), dog (23), and man (24). Lichter et al. (25) have studied cytosol thyroid hormone binding proteins with particular emphasis on the organic anion binding proteins, Y (ligandin) and Z. In extensive earlier studies, these cytosol proteins were shown to bind bilirubin, BSP, and cholecystographic agents (26), and were found in all species tested, including rats, mice, guinea pigs, rhesus monkeys, sheep, and man. Lichter et al. (25) were able to show that both T_4 and T_3 are bound by both Y and Z, and are displaced from these proteins by BSP and by bilirubin. However, there was prominent binding of the thyroid hormones to other cytosol proteins; the major portion of T_4 was bound to a protein (or proteins) migrating near, but distinct from, albumin on Sephadex G-100. BSP and bilirubin displaced T_4 from this protein as well as from Y and Z. Dillman et al. (22) also described a rat liver cytosol protein that could bind T_4 , and that had an affinity for BSP about 1:10,000 of that

for T_4 ; the ratio of BSP to T_4 in the displacement studies by Lichter et al. (25) may have been in the same range. Although Davis et al. (23), in their study of canine liver cytosol proteins, did not see prominent binding of BSP to the cytosol T_4 -binding protein, they did not specifically study the ability of large amounts of BSP to displace T_4 from its specific protein.

Thus, the available evidence suggests that high concentrations of the organic anions, BSP and bilirubin, can displace T_4 from binding to cytosol proteins, even though the major protein involved may not be the Y or Z protein. Our own studies in liver slices (4) have shown that cholecystographic agents, including tyropanoate and ipodate, share with BSP and bilirubin the ability to displace T_4 from cells. The interaction of organic anions with hepatic T_4 binding proteins may be weak in man, as it is in rats; the molar ratio between our standard dose of tyropanoate (6 g, 9.0 mmol) and an estimate of hepatic T_4 content (270 μ g, 0.35 μ mol) is in the neighborhood of 25,000:1. The effective dose of BSP, 20 mg/kg, would represent \sim 2 mmol, yielding a BSP: T_4 ratio of 6,000:1.

In addition to cytosol binding proteins, it is possible that plasma membrane transport plays a role in the accumulation of cholecystographic agents and T_4 by hepatocytes. Scharschmidt et al. (27) have demonstrated in the rat that there is a carrier-membrane transport system for the hepatic uptake of organic anions such as BSP, bilirubin, and indocyanine green. The agents are mutually competitive ligands for this process. Receptors for BSP have been identified in liver plasma membranes (28). Pliam and Goldfine (29) have demonstrated specific thyroid hormone binding sites in rat liver plasma membranes. It seems conceivable, therefore, that competitive inhibition of inward transport through the plasma membrane is another mechanism of tyropanoate's effects; to explain discharge of accumulated hormone by this mechanism would also require that exit from the cell be less inhibited than entry, and that exit be fairly rapid.

The two possible mechanisms have quite different implications for the intracellular fate of T_4 . If cholecystographic agents interfere primarily with membrane transport, this could result in a lowering of intracellular free T_4 levels. On the other hand, if the primary effect is to inhibit binding of T_4 already within the cell to cytosol proteins, cholecystographic agents should transiently increase the intracellular free T_4 levels. Further study is clearly indicated to sort through these possibilities and to determine the effect of gallbladder agents upon intracellular free T_4 levels.

Our data in patients with liver disease, although limited, suggest that the dischargeable hepatic T_4 pool was almost abolished. Also, there was a slower rate of initial equilibration between count rates over the liver and serum 125 I concentration (Fig. 2); this may reflect

uptake into chest wall tissues that accumulate T_4 at the slow rate characteristic of the arm (30), leg (9, 17), or muscle tissue in general (31).

It has previously been shown that a major effect of cholecystographic agents is to block the peripheral conversion of T_4 to T_3 (1-5), an effect shared by tyropanoate.² Indeed, Wu et al. (32) have used repeated doses of ipodate in the treatment of hyperthyroidism. A decreased hepatic uptake of T_4 is one potential mechanism for decreased T_3 formation; Jennings et al. (33) have recently suggested that decreased hepatic uptake of T_4 , rather than decreased activity of a converting enzyme, is responsible for the decreased rate of T_3 production that occurs in fasting. However, when the time-course we have demonstrated for tyropanoate's effect on hepatic T_4 content (maximum at 4 h, dissipated by 24 h) is compared with that for inhibition of T_3 formation (persisting beyond 3 d²), it seems likely that the two effects are separate.

There has been disagreement in the literature concerning the magnitude of the hepatic storage pool for thyroid hormones. Nicoloff and Dowling (30) calculated that the liver represents about 14% of the total nonthyroidal pool of T_4 ; a similar estimate was made by Busnardo et al. (13). Cavalieri and Searle, however, reported values twice as large for this pool, claiming that nearly one-third of the total nonthyroidal pool of T_4 is located in the liver (9, 17). The figure offered by Oppenheimer et al. (14) is in close agreement; they report that 29% of the total extrathyroidal pool is in the liver, and similar values are given by some other workers (15, 16).

Further support for the larger hepatic pool size derives from extensive studies performed both in sheep (34, 35) and in man (36) by Irvine, who concludes that the liver contributes 30% of the total pool of extrathyroidal T_4 in man (31). The smaller estimates derive from models in which the interstitial fluid (ISF) is viewed as a separately equilibrating compartment, whose size is estimated by the early passage of radioactivity into the arm (30) or thigh (13). Irvine has, however, demonstrated that the rate of change in the specific activity of lymph in animals injected with labeled hormone varies from organ to organ, so that the ISF specific activity in each organ is about the same as the total tissue specific activity (35), i.e., transfer of T_4 from plasma to ISF is the rate-limiting step in tissue entry. This negates the assumption by Nicoloff and Dowling (12) of a distinct ISF pool; moreover, their assumption that the liver has an ISF compartment corresponding to the early change in arm:plasma count rate ratios results in assigning a large component of hepatic T_4 to the interstitial space. Evidence that the hepatic interstitial T_4 pool is small has been provided in autoradiographic studies by Oppenheimer et al. (14, 37) and in direct postmortem measurements of

hepatic interstitial T_4 in sheep where the ratio, hepatic ISF T_4 :hepatic plasma T_4 , was <0.2 (34).

Expressing the data of Cavalieri and Searle (9, 17), Oppenheimer (14), and Irvine (31) as a ratio of hepatic T_4 content to plasma T_4 content (T_4 content = volume $\times T_4$ concentration), the individual values range from 0.97 to 1.85 with median values between 1.3 and 1.4. Our average estimate, 1.8 (Table II), is higher. Because there were no evident errors in our estimation of plasma radioactivity, the major error that could elevate the ratio would be an underestimate of the proportion of radioactivity discharged from the liver ($\Delta[L - S]$ in Table II). It is possible that the segment of liver counted is not representative, and that portions of the liver distant from the probe discharge more completely; there is no noninvasive method to check this possibility, although gamma camera images of the liver, performed in subject 2, showed a uniform loss of ^{125}I after tyropanoate. A more well-defined source of error is the contribution of chest wall radioactivity to "liver" counts. In one subject, placing the probe over the upper chest gave a count rate about one-tenth that over the liver, before tyropanoate ingestion. Using this value to correct for the chest wall contribution in each subject, the mean value for $\Delta(L - S)$ in Table II becomes 26%, and the mean ratio falls to 1.67. Finally, some of the ^{125}I entering serum may have come from tissues other than the liver. Discharge of T_4 from the kidney by tyropanoate seems particularly likely because we have shown that cholecystographic agents inhibit T_4 uptake by rat kidney slices;⁴ the kidney has transport systems (38) and cytosol binding proteins for cholecystographic agents (39) similar to those found in liver, and has been shown to concentrate T_4 in man (37). Assuming that the renal pool is one-sixth the size of the hepatic pool, this would further reduce the estimate of the liver space alone to 1.4 times the plasma volume, more closely agreeing with earlier workers. It does seem unlikely that other tissues are responsible for much of T_4 discharge. Counting over the upper chest, there was actually a rise in radioactivity after tyropanoate; in studies not reported here, we have observed similar increases in radioactivity over the arm and thigh after tyropanoate ingestion. In each site, the rise has been compatible with the rise in plasma radioactivity, and there was little net change in extravascular radioactivity in the chest wall, arm, or thigh. In other words, tyropanoate did not appear to cause significant discharge of T_4 from skeletal muscle, the major repository of extravascular T_4 beside liver and kidney (31).

There are other possible errors in our method that could lead to an underestimate of the hepatic T_4 space. First, tyropanoate could discharge ^{125}I into bile so that

only a portion of T_4 lost from the liver entered serum. Also, from the nature of the curve in Fig. 6, it seems likely that some of the ^{125}I entering serum from liver had already been lost to other extravascular sites by 1 h, and that ratios computed at earlier time intervals would have been higher and would more accurately reflect relative distribution volumes. It is of course possible that estimates in the literature are low. In particular, some of the values for plasma volume reported by Cavalieri and Searle (9, 17) and by Oppenheimer et al. (14) seem large.

The only agent previously shown to displace labeled T_4 from the liver (40) with a concomitant rise in serum radioactivity (40, 41) is diethyl ether; a similar redistribution of T_4 may be caused by halothane (42) and isoflurane (43). The published data concerning the effects of ether anesthesia on radioactivity in serum, liver, and thigh in subjects who had received [^{131}I] T_4 60 h earlier (40) can be interpreted as showing some discharge from the thigh as well as the liver,⁵ in contrast to tyropanoate which does not discharge T_4 from the thigh. We speculate, therefore, that the effect of ether has a different mechanism than the effect of tyropanoate, and, thus, that the present studies may be the first example in man of a direct effect on hepatic T_4 content. Agents inhibiting hepatic storage of T_4 may provide a useful tool for investigating the effects of disease and physiologic variables on hepatic storage. From considerations discussed above, our standard protocol provides an approximate estimate of the distribution volume ratio between liver and serum. This method could be applied, for example, to determine whether fasting alters hepatic storage of T_4 in man, as it apparently does in rats (33).

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⁵ Oyama et al. (40) reported a mean rise of 55% in serum [^{131}I] T_4 and no change in radioactivity in the thigh, after 40 min of ether anesthesia. If, as reported by Cavalieri (9), 30% of the thigh counts represent intravascular radioactivity, the failure of thigh counts to change when serum radioactivity rose implies some loss of extravascular radioactivity from the thigh; substituting thigh radioactivity for liver radioactivity in Eq. 1, and assuming that 30% of initial thigh radioactivity was intravascular, the data of Oyama et al. (40) indicate a 25% decline in thigh extravascular T_4 content during anesthesia. A firm conclusion cannot be reached, however, because ether may also cause changes in regional blood flow and in plasma volume. Using labeled human serum albumin, we have shown in two subjects that tyropanoate does not alter plasma volume, nor the amount of externally detected labeled albumin in liver and thigh.

⁴ Green, W. L., and G. Bellamy. Unpublished observations.

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REFERENCES

1. Bürgi, H., C. Wimpfheimer, A. Burger, W. Zaunbauer, H. Rösler, and T. Lemarchand-Beraud. 1976. Changes of circulating thyroxine, triiodothyronine and reverse triiodothyronine after radiographic contrast agents. *J. Clin. Endocrinol. Metab.* **43**: 1203–1210.
2. Wu, S.-Y., I. J. Chopra, D. H. Solomon, and L. R. Bennett. 1978. Changes in circulating iodothyronines in euthyroid and hyperthyroid subjects given ipodate (Oragrafin), an agent for oral cholecystography. *J. Clin. Endocrinol. Metab.* **46**: 691–697.
3. Kaplan, M. M., and R. D. Utiger. 1978. Iodothyronine metabolism in rat liver homogenates. *J. Clin. Invest.* **61**: 459–471.
4. Green, W. L., and G. Bellamy. 1977. Role of organic anion transport systems in uptake and metabolism of thyroid hormones by rat liver slices. Program of the 53rd Annual Meeting of the American Thyroid Association, Cleveland, Ohio. TR-16. (Abstr.).
5. Chopra, I. J. 1978. Inhibition of outer ring monodeiodination of T_4 and reverse T_3 (rT_3) by some radiocontrast agents. *Clin. Res.* **26**: 303A. (Abstr.).
6. Green, W. L. 1972. Separation of iodo compounds in serum by chromatography on Sephadex columns. *J. Chromatogr.* **72**: 83–91.
7. Lepp, A., H. Pena, V. Hoxie, and L. Oliver. 1965. Protein-bound iodine determination with stable end point. *Am. J. Clin. Pathol.* **44**: 331–341.
8. Sterling, K., and M. A. Brenner. 1966. Free thyroxine in human serum: simplified measurement with the aid of magnesium precipitation. *J. Clin. Invest.* **45**: 153–163.
9. Cavalieri, R. R., and G. L. Searle. 1965. The role of the liver in the distribution of ^{131}I thyroxine in man. *In* Current Topics in Thyroid Research: Proceedings of the Fifth International Conference. C. Cassano and M. Andreoli, editors. Academic Press, Inc., New York. 336–345.
10. McChesney, E. W. 1971. Routes and rates of excretion of radiocontrast agents. *In* International Encyclopedia of Pharmacology and Therapeutics. Section 76. Radiocontrast Agents. P. K. Knoeffel, editor. Pergamon Press Ltd., Oxford. **1**: 335–344.
11. Cristofori, F. C., and G. G. Duncan. 1964. Effect of a new orally administered cholecystographic compound on the I^{131} -triiodothyronine red-cell-uptake test of thyroid function. *N. Engl. J. Med.* **271**: 564–565.
12. Nicoloff, J. T., and J. T. Dowling. 1968. Studies of peripheral thyroxine distribution in thyrotoxicosis and hypothyroidism. *J. Clin. Invest.* **47**: 2000–2015.
13. Busnardo, B., M. E. Girelli, L. Varotto, and A. Giacobbi. 1974. Computer analysis of early thyroxine distribution in thyrotoxicosis and hypothyroidism. *Horm. Metab. Res.* **6**: 202–207.
14. Oppenheimer, J. H., G. Bernstein, and J. Hasen. 1967. Estimation of rapidly exchangeable cellular thyroxine from the plasma disappearance curves of simultaneously administered thyroxine- ^{131}I and albumin- ^{125}I . *J. Clin. Invest.* **46**: 762–777.
15. Yumoto, Y., T. Namba, and H. Yamamoto. 1970. Kinetic analysis of thyroxine outside thyroid with ^{131}I -thyroxine by aid of computer—with special reference to liver diseases. *Acta Med. Okayama.* **24**: 457–470.
16. Musa, B. U., R. S. Kumar, and J. T. Dowling. 1969. Role of thyroxine-binding globulin in the early distribution of thyroxine and triiodothyronine. *J. Clin. Endocrinol. Metab.* **29**: 667–674.
17. Cavalieri, R. R., and G. L. Searle. 1966. The kinetics of distribution between plasma and liver of ^{131}I -labeled L-thyroxine in man: observations of subjects with normal and decreased serum thyroxine-binding globulin. *J. Clin. Invest.* **45**: 939–949.
18. Zaninovich, A. A., R. Volpe, R. J. Soto, and C. Ezrin. 1969. Accumulation and release of thyroid hormones by the normal and diseased human liver. *Acta Endocrinol.* **60**: 412–422.
19. Musa, B. U., R. S. Kumar, and J. T. Dowling. 1968. Effects of salicylates on the distribution and early plasma disappearance of thyroxine in man. *J. Clin. Endocrinol. Metab.* **28**: 1461–1464.
20. Hamada, S., K. Torizuka, T. Miyake, and M. Fukase. 1970. Specific binding proteins of thyroxine and triiodothyronine in liver soluble proteins. *Biochim. Biophys. Acta.* **201**: 479–492.
21. Spaulding, S. W., and P. J. Davis. 1971. Thyroxine binding to soluble proteins in rat liver and its sex dependence. *Biochim. Biophys. Acta.* **229**: 279–283.
22. Dillman, W., M. I. Surks, and J. H. Oppenheimer. 1974. Quantitative aspects of iodothyronine binding by cytosol proteins of rat liver and kidney. *Endocrinology.* **95**: 492–498.
23. Davis, P. J., B. S. Handwerker, and F. Glaser. 1974. Physical properties of a dog liver and kidney cytosol protein that binds thyroid hormone. *J. Biol. Chem.* **219**: 6208–6217.
24. Hamada, S., and M. Fukase. 1976. Demonstration and some properties of cytosol-binding proteins for thyroxine and triiodothyronine in human liver. *J. Clin. Endocrinol. Metab.* **42**: 302–308.
25. Lichter, M., G. Fleischner, R. Kirsch, A. J. Levi, K. Kamisaka, and I. M. Arias. 1976. Ligandin and Z protein in binding of thyroid hormones by the liver. *Am. J. Physiol.* **230**: 1113–1120.
26. Levi, A. J., Z. Gatmaitin, and I. M. Arias. 1969. Two hepatic cytoplasmic protein fractions, Y and Z, and their possible role in the hepatic uptake of bilirubin, sulfobromophthalein, and other anions. *J. Clin. Invest.* **48**: 2156–2167.
27. Scharschmidt, B. F., J. G. Waggoner, and P. D. Berk. 1975. Hepatic organic anion uptake in the rat. *J. Clin. Invest.* **56**: 1280–1292.
28. Reichen, J., B. L. Blitzer, and P. D. Berk. 1977. The binding of organic anionic dyes to hepatocellular plasma membranes. *Clin. Res.* **25**: 468A. (Abstr.).
29. Pliam, N. B., and I. D. Goldfine. 1977. High affinity thyroid hormone binding sites on purified rat liver plasma membranes. *Biochem. Biophys. Res. Commun.* **79**: 166–172.
30. Nicoloff, J. T., and J. T. Dowling. 1968. Estimation of thyroxine distribution in man. *J. Clin. Invest.* **47**: 26–37.
31. Irvine, C. H. G. 1975. Four compartment model of thyroxine metabolism. *In* Thyroid Hormone Metabolism. W. A. Harland and J. S. Orr, editors. Academic Press, Inc., New York. 163–188.
32. Wu, S.-Y., I. J. Chopra, D. H. Solomon, and D. E. Johnson. 1978. The effect of repeated administration of ipodate (Oragrafin) in hyperthyroidism. *J. Clin. Endocrinol. Metab.* **47**: 1358–1362.
33. Jennings, A. S., D. C. Ferguson, and R. D. Utiger. 1979. Regulation of the conversion of thyroxine to triiodothyronine in the perfused rat liver. *J. Clin. Invest.* **64**: 1614–1623.
34. Irvine, C. H. G. 1974. Concentration of thyroxine in cellular and extracellular tissues of the sheep and the

- rate of equilibration of labeled thyroxine. *Endocrinology*. **94**: 1060–1071.
35. Irvine, C. H. G., and M. W. Simpson-Morgan. 1974. Relative rates of transcapillary movement of free thyroxine, protein-bound thyroxine, thyroxine-binding proteins, and albumin. *J. Clin. Invest.* **54**: 156–164.
 36. Irvine, C. H. G. 1968. Effect of exercise on thyroxine degradation in athletes and non-athletes. *J. Clin. Endocrinol. Metab.* **28**: 942–948.
 37. Oppenheimer, J. H., M. I. Surks, and H. L. Schwartz. 1969. The metabolic significance of exchangeable cellular thyroxine. *Recent Prog. Horm. Res.* **25**: 381–414.
 38. Bárány, E. H. 1973. The liver-like anion transport system in rabbit kidney, uvea and choroid plexus. II. Efficiency of acidic drugs and other anions as inhibitors. *Acta Physiol. Scand.* **88**: 491–504.
 39. Kirsch, R., G. Fleischner, K. Kamisaka, and I. M. Arias. 1975. Structural and functional studies of ligandin, a major renal organic anion-binding protein. *J. Clin. Invest.* **55**: 1009–1019.
 40. Oyama, T., S. Shibata, and A. Matsuki. 1969. Thyroxine distribution during ether and thiopental anesthesia in man. *Anesth. Analg.* **48**: 1–6.
 41. Fore, W., P. Kohler, and J. Wynn. 1966. Rapid redistribution of serum thyroxine during ether anesthesia. *J. Clin. Endocrinol. Metab.* **26**: 821–830.
 42. Oyama, T., S. Shibata, S. Matsuki, and T. Kodo. 1969. Thyroxine distribution during halothane anesthesia in man. *Anesth. Analg.* **48**: 715–719.
 43. Oyama, T., P. Latto, D. A. Holaday, and H. Chang. 1975. Effect of isoflurane anaesthesia and surgery on thyroid function in man. *Can. Anaesth. Soc. J.* **22**: 474–477.