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A ^B ^S ^T ^R ^A ^C ^T Calcium, phosphorus, sodium, carbonate, magnesium, and hydroxyproline were measured in iliac crest biopsies of 22 normal volunteers and 24 selected patients with renal osteodystrophy. Histologic classification revealed that 10 were mildly abnormal, 8 osteomalacic, and 6 osteofibrotic. Bone density measurements were performed on an additional 12 normal, 11 mildly abnormal, 6 osteomalacic, and 10 osteofibrotic subjects. The results revealed an increase in magnesium and a decrease in carbonate apparent in the minimal and osteomalacic lesions and a much greater change in osteofibrosis. The bone density was decreased in patients with osteofibrosis. These observations would appear to be explained by postulation of an impairment of the normal maturational process of bone whereby there is an increase in amorphous calcium phosphate and a decrease in apatite crystal. The data suggest that the maturational defect is present as soon as any abnormality can be identified histologically, is present to the same degree in osteomalacia, and is most severe in osteofibrosis. In comparison of two sets of six patients matched for age and duration of dialysis, neither acidosis nor vitamin D therapy appeared to influence the severity of the maturational defect.

INTRODUCTION

It has been demonstrated in rats that uremia results in ^a maturational defect in both collagen (1) and bone mineral (2). The failure to prevent this lesion by vitamin D therapy (2) has implied that this lesion was associated with secondary hyperparathyroidism or some as yet unidentified defect related to the accumulation of uremic toxins.

The present study identifies a decrease in bone carbonate and an increase in bone magnesium in all patients biopsied. These abnormalities are interpreted to be similar to the maturational defect found in rats. It

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is most severe in osteofibrosis, where it is associated with decreased density and appears to be uninfluenced by the degree of acidosis or by vitamin D therapy.

METHODS

All patients had chronic renal disease of long standing; 22 of 24 had been on repetitive hemodialysis for from 2 mo to $5\frac{1}{2}$ yr. Bone biopsies were taken 1 cm and 2 cm from the anterior superior iliac spine with an electric core drill' yielding a longitudinal biopsy approximately 0.4 cm in diameter and over ¹ cm in length, with a cortical strip of varying depth at one end.

Each biopsy was sliced in half, the spicular portion being placed in 10% buffered formaldehyde solution for the histological study and the spicular-cortical half weighed immediately and placed in 1:1 petroleum-ethyl ether solution for chemical analysis. After fat extraction the latter bone core was dried to an air-stable weight. True density (weight per unit volume of bone and matrix only) was obtained by the volume displacement method with heptane (bp 98° -99 $^{\circ}$ C) instead of water to prevent loss of carbonate and other soluble salts from the bone. The biopsy was attached to 6-0 silicon-coated silk suture, suspended, and weighed in the air, placed in heptane under vacuum until all trapped air was removed, and then resuspended, and weighed in heptane. True volume equals difference in weight per unit weight of heptane at ambient temperature. True density equals air-stable weight per unit true volume. Dry weight was obtained by heating to 100'C under vacuum for 12 h, after which the sample was frozen with liquid N2, powdered, and dried to an air-stable weight for the chemical determinations. The yield was normally 15-25 mg. The bone density measurements were not performed in the original 24 patients, but subsequently on 12 normals and 27 patients with renal osteodystrophy. Classification of the biopsies from which density is reported was based on the same histologic criteria used for the original set.

By methods described before (3), bone sodium was determined by neutron activation, bone carbonate by direct manometric analysis, and the whole sample subsequently used for measurement of phosphorus by the Techinicon AutoAnalyzer (Technicon. Instruments Corp., Tarrytown, N. Y.), and Ca and Mg by regular atomic absorption spectroscopic methods.

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^{&#}x27;Myelotomiegerat, Institute F. Straumann, Waldenburg B.L., Switzerland.

| Classification (n) | Total bone Perimeter | | | Total bone area | Bone formation | | | |
|-----------------------|-----------------------------|------------|-------------|--------------------------|----------------------------|-------------------|----------------------------|--|
| | Forming | Resorbing | Neutral | Mineralized (osteoid) | L_F * \times | R _{BA} t | R_{BF} $=$ | |
| | $\%$ | $\%$ | $\%$ | $\%$ | $mm^2/mm^3 \times 10^{-4}$ | μ m/day | $mm^3/mm^3 \times 10^{-8}$ | |
| Normal (19) | 18.16 | 4.03 | 77.80 | 94.31 (5.69) | 0.52 | 0.31 | 1.50 | |
| | ± 7.96 | ± 1.98 | ± 8.20 | ± 7.72 | ± 0.20 | ± 0.08 | ± 0.62 | |
| Mild disease (10) | 34.77 | 5.66 | 59.55 | 93.70 (6.30) | 1.02 | 0.28 | 2.87 | |
| | \pm 9.90 | ± 1.75 | ± 10.57 | ± 2.02 | ± 0.25 | ± 0.08 | ± 1.13 | |
| P vs. normal | < 0.001 | < 0.05 | < 0.001 | N.S. | < 0.001 | N.S. | < 0.001 | |
| Osteomalacic (8) | 76.68 | 5.55 | 17.76 | 67.59(32.41) | 3.04 | 0.10 | 3.31 | |
| | \pm 9.80 | ± 2.02 | \pm 9.30 | ±16.52 | ± 1.26 | ± 0.04 | ± 0.62 | |
| P vs. normal | < 0.001 | N.S. | < 0.001 | < 0.001 | < 0.001 | N.S. | ${<}0.001$ | |
| Osteofibrotic (6) | 61.18 | 8.60 | 31.88 | 87.90 (12.10) | 1.91 | 0.69 | 12.98 | |
| | ± 6.78 | ± 2.98 | ± 6.17 | ± 4.39 | ± 0.65 | ± 0.29 | ± 7.47 | |
| P vs. normal | < 0.001 | < 0.001 | < 0.001 | N.S. | ${<}0.001$ | < 0.001 | < 0.001 | |
| P vs. malacic | ${<}0.01$ | ${<}0.05$ | ${<}0.01$ | ${<}0.01$ | N.S. | < 0.001 | < 0.001 | |

TABLE ^I Histological Classification of Human Iliac Crest Biopsies

Values are the mean ± 1 SD. P values are by Student t distribution for grouped data, $P < 0.05$ being considered significantly different.

* Length of forming surface per total biopsy area.

 \ddagger Rate of bone apposition (width of tetracycline label per day).

§ Rate of bone formation $(L_F \times R_{BA})$.

Bone hydroxyproline (OH-proline) was measured by an adaptation of Hosley Olson, Horton, Michelsen, and Atkins' automated analysis of urinary OH-proline (4). The OHproline in the bone was released by autoclaving the hydrolysate used for the Ca, Mg, and P. determinations with an equal amount of ⁸ N HCl. An aliquot was titrated with NaOH to about pH 6.0, with methyl red as an indicator, and diluted to a volume such that OH-proline was in the 0.5-3.0 μ g/ml range and the Cl content was kept below 0.4 N to prevent color interference. The sensitivity of the AutoAnalyzer method was expanded by changing the sample line to $1.2 \text{ cm}^3/\text{min}$, the air line to $0.6 \text{ cm}^3/\text{min}$ and replacing the 40-min time delay coil with a cooling-jacketed mixing coil. In our experience the stock OH-proline standard was not stable in 0.001 N HCl, but was stable for over a yr at room temperature in 0.5 N HCl, the diluted standards being neutralized as were the samples. Optical density of the 2.0 μ g/ml standard was over 0.3. Duplicate samples, different determination, varied less than 5%.

Vitamin D treatment and acidosis were studied in two sets of six patients each, a treated set from Vancouver, Canada, and a nontreated set selected from Seattle to match the vitamin D-treated patients by age and duration of disease.

Bone histology was classified as mildly abnormal, osteomalacic, or osteofibrotic on the basis of measurements of total bone area and total bone surface. Area measurements give percent mineralized or osteoid per bone area or per biopsy area. Linear measurements were made of bone perimeter forming, neutral, and resorbing. Tetracycline labeling was used to calculate the rate of bone apposition $(R_{BA})^2$ in millimeters $\times 10^{-4}$ per day, the length of form-

² Abbreviations used in this paper: ACP, amorphous calcium phosphate; HA, crystalline OH-apatite; L_F , length of forming surface; R_{BA} , bone apposition rate; R_{BF} , bone formation rate.

ing surface (L_F) (square millimeters per total biopsy area), and the bone formation rate (RBF) (cubic millimeter $\times 10^{-8}$ per cubic millimeter per day) (5). Histologic results are
expressed as the mean ± 1 SD. P values are by the Student t test, grouped data, $P < 0.05$ indicating a significant difference.

RESULTS

Histological classification of mild, osteomalacic, and osteofibrotic (Table I) was based on the selection of patients with as well-defined a histological type of lesion as possible. Patients were ambulatory and asymptomatic. Normals were volunteers whose ages were between 30 and 60, roughly the age group of the patients.

The histological study of the normals showed $94.31 \pm$ 7.72% of the total tissue area mineralized and 5.69% osteoid, with $18.16 \pm 7.96\%$ of the bone perimeter forming, $4.03 \pm 1.98\%$ resorbing, and $77.80 \pm 8.20\%$ neutral. RBA by tetracycline labeling was 3.04 ± 0.79 mm $\times 10^{-4}$ / day, while the L_F (forming perimeter per total area) was 0.52 ± 0.20 mm²/mm³. The R_{BA} times L_F was the RBF and was 1.50 ± 0.62 mm⁸ \times $10^{-8}/\text{mm}^3/\text{day}$ for normals.

The mild lesion (10 patients) was characterized by a significant increase in percent of forming perimeter, 34.77 \pm 9.90%, $P < 0.001$, a consequent decrease in neutral, $59.55 \pm 10.57\%$, $P < 0.001$, with a marginal increase in resorbing, $5.66 \pm 1.75\%$, $P < 0.05$, and no change in the mineralized-osteoid percentage of total tissue area, $93.70 \pm 2.02\% - 6.30\%$. The RBA was normal, 2.78 \pm 0.81, but the greater L_F, 1.02 \pm 0.25, $P < 0.001$,

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resulted in a significant increase in RBF, 2.87 ± 1.13 , $P \leq$ 0.001.

In osteomalacia (eight patients) the most characteristic change was a large increase in osteoid, 32.41%, with concomitant decrease in percent tissue area mineralized, $67.59 \pm 16.52\%$, $P < 0.001$. This change was reflected in increased forming perimeter, $76.68 \pm 9.80\%$, $P < 0.001$, and decreased neutral, 17.76 \pm 9.30%, P < 0.001, with no change in the resorbing, $5.55 \pm 2.02\%$. RBA decreased significantly, 1.08 ± 0.45 , $P < 0.001$, but the marked increase in L_F, 3.04 ± 1.26 , $P < 0.001$, resulted in an increased RBF, 3.31 ± 0.62 , $P < 0.001$.

The fibrotic lesion (six patients) showed very different changes. The percent perimeter forming was increased about midway between the mild and malacic lesion, 61.18 \pm 6.78%, $P < 0.001$. However, the resorbing perimeter was also increased, $8.60 \pm 2.98\%$, $P \leq$ 0.001. These histologic observations were confirmed by measurement of bone formation. The L_F was increased, 1.91 \pm 0.65, P < 0.001, as was the R_{BA}, 6.90 \pm 2.94, P < 0.001. These increases resulted in a marked increase in RBF to 12.98 \pm 7.47, $P \le 0.001$, as compared to normal or osteomalacic rates.

Many chemical parameters were tested and correlations sought because the iliac crest biopsy presents problems not found in pure cortical bone. Depth of cortex varies greatly along the iliac crest so that the ratio of cortical to spicular bone in a biopsy depends much more on location than on state of disease. Therefore, wherever possible, ratios of related measurements were used rather than relationship by weight or volume, for the latter is often disproportionately affected by varying cortical-spicular combinations. For example, between varying cortical-spicular biopsies from the same normal individual, a $7-8\%$ variation in $CO₃/g$ will

drop to $1-2\%$ as $CO₃/Ca$; a $9-10\%$ Mg/g variation will drop to 3% as Mg/Ca. Calcium and phosphorus were compared by weight, volume, and OHproline. No reference base is superior to weight, the variance being about 5%. The Ca/P ratio, on the other hand, varies only about 1% . OH-proline, representing collagen matrix, is constant throughout the biopsy and therefore relates better to true volume than to weight, reducing ^a 6% variance to 2%.

Of the many chemical parameters studied, the only consistent abnormalities found for the histological classes were a decrease of bone carbonate and an increase in bone magnesium (Table II, Fig. 1). Bone CO_s (meq/g) decreased from a normal of 1.589 ± 0.178 to $1.417 \pm$ 0.177, $P < 0.02$, for the mild lesion; 1.360 \pm 0.137, $P < 0.01$, for the malacic; and 1.104 \pm 0.217, $P < 0.001$, for the fibrotic. The more consistent COs/Ca ratio decreased from $166.9 \pm 7.8 \times 10^{-8}$ for normals to $141.1 \pm 12.6 \times 10^{-3}$, $P < 0.001$, for mild; 145.5 ± 16.0 \times 10⁻³, $P \le 0.001$, for malacic; and 126.3±12.3 \times 10^{-3} , $P \le 0.001$, for fibrotic. Bone Mg (meq/g) increased from a normal of 0.1798 ± 0.0258 to 0.2497 ± 0.0282 , $P \leq$ 0.001, for the mild; 0.2486 ± 0.0410 , $P \le 0.001$, for the osteomalacic; and 0.3101 ± 0.0543 , $P < 0.001$, for the osteofibrotic. With the Mg/Ca ratio, the normal of $19.00\pm2.93\times10^{-3}$ increased to $25.02\pm3.47\times10^{-3}$, $P \leq$ 0.001, in mild disease; $28.00 \pm 5.48 \times 10^{-8}$, $P < 0.001$, in osteomalacia; and $36.18 \pm 4.42 \times 10^{-3}$, $P < 0.001$, in osteofibrosis. Both abnormalities were so much greater in osteofibrosis than in osteomalacia that a statistical difference exists between the two, $P \leq 0.05$ for the decrease in CO₃/C_a or $P < 0.02$ for the increase in Mg/C_a.

After true density measurements were begun, 11 patients with mild disease, 6 with osteomalacia, and 10 with osteofibrosis were identified histologically. The

| | | Chemical Parameters of Human Iliac Crest Biopsies Grouped by Histological Classification | | | | | | | | |
|-----------------------------|-----------------|--|-------------|-----------------|-------------|--------------|---------------|--------------|--------------|--|
| Histological class (n) | CO ₃ | CO _a /Ca | Ca | PO ₄ | Ca/P | Mg | Mg/Ca | $OH-P$ | Na | |
| | meq/g | | meq/g | meq/g | mmol/mmol | meq/g | | mmol/g | meq/g | |
| Normal (19) | 1.539 | 0.1669 | 9.507 | 8.283 | 1.720 | 0.1798 | 0.01900 | 0.2862 | 0.2911 | |
| | ±0.178 | ± 0.0078 | ±0.860 | ± 0.736 | ± 0.077 | ± 0.0258 | ± 0.00293 | ± 0.0356 | ± 0.0120 | |
| Mild(10) | 1.417 | 0.1411 | 10.020 | 8.604 | 1.746 | 0.2497 | 0.02502 | 0.2786 | 0.3007 | |
| | ±0.177 | ± 0.0126 | ±0.569 | ±0.450 | ± 0.035 | ± 0.0282 | ± 0.00347 | ± 0.0145 | ± 0.0153 | |
| P vs. normal | < 0.02 | < 0.001 | NS | NS | NS | < 0.001 | < 0.001 | NS | NS. | |
| Osteomalacic (8) | 1.360 | 0.1455 | 9.249 | 8.083 | 1.709 | 0.2486 | 0.02800 | 0.2889 | 0.2945 | |
| | ± 0.137 | ± 0.0160 | ± 0.567 | ± 0.377 | ± 0.052 | ± 0.0410 | ± 0.00548 | ± 0.0322 | ±0.0286 | |
| P vs. normal | < 0.01 | < 0.001 | NS | $_{\rm NS}$ | NS | < 0.001 | < 0.001 | NS | NS | |
| Osteofibrotic (6) | 1.086 | 0.1263 | 8.624 | 7.414 | 1.738 | 0.3101 | 0.03618 | 0.3094 | 0.2817 | |
| | ±0.189 | ± 0.0123 | ±1.431 | ±0.888 | ±0.104 | ±0.0543 | ± 0.00442 | ± 0.0412 | ± 0.0178 | |
| P vs. normal | < 0.001 | < 0.001 | NS | < 0.05 | NS | < 0.001 | < 0.001 | NS | NS | |
| P vs. mild | 0.01 | 0.02 | < 0.02 | < 0.01 | NS | < 0.02 | < 0.001 | < 0.05 | < 0.05 | |
| P vs. malacic | < 0.02 | $<$ 0.05 | NS | NS | NS | < 0.05 | < 0.02 | $_{\rm NS}$ | NS | |

TABLE II

Values are the mean ± 1 SD. P values are by the Student t test, grouped data, $P < 0.05$ indicating a significant difference. Weight values are per gram air dried, fat-free, powdered bone.

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true density of the air-stable, defatted bone was found to be 1.976 \pm 0.068 for 12 normals with a C.V. of 3.4%. Patients with mild disease showed a density of $1.916 \pm$ 0.084, C.V. 4.4% , while the density of osteomalacia was 1.961 \pm 0.050, C.V. 2.5%. The fibrotic lesion was associated with a density of 1.897 ± 0.035 , C.V. 3.4% , significantly decreased from normal, $P < 0.01$, and also from the malacic group, $P \le 0.01$ (Fig. 2).

Table III shows the results of the chemical and histological studies of iliac crest biopsies from a set of six vitamin D-treated dialysis patients from Vancouver, Canada, and a comparable set, by age and duration of dialysis, of nontreated patients from Seattle. Dialysate concentrations and total hours of dialysis were the same, but in the Seattle patients dialysis was performed three times weekly, while the Vancouver patients dialyzed twice weekly, resulting in a lower ongoing serum $HCO₃$ level, 18.0 ± 1.5 vs 23.0 ± 1.7 , $P < 0.01$.

The vitamin D-treated patients showed a decrease in osteoid, $9.00 \pm 5.5\%$ vs. $16.4 \pm 9.6\%$, $P < 0.02$, with a concomitant increase in the percent bone mineralized. There was a decrease in the percent bone perimeter forming, $38.3 \pm 14.2\%$ vs $65.7 \pm 11.1\%$, $P < 0.001$. Bone resorption by percent perimeter showed no change.

The chemical measurements showed a slight increase in bone calcium per gram, approaching significance for the vitamin D-treated patients. There was a decrease in

FIGURE 1 Comparison of CO₃/Ca and Mg/Ca ratios in patients with mild, osteomalacic, and osteofibrotic renal osteodystrophy. Figures signify the mean±1 SD.

FIGURE 2 Comparison of true bone density in patients with mild, osteomalacic, and osteofibrotic renal osteodystrophy. Figures signify the mean ± 1 SD.

OH-proline per gram of the same significance, which is probably only the reflection of the additional weight per biopsy unit. The only significant difference is an increase in the Ca/P ratio for the patients under vitamin D treatment, 1.804 ± 0.040 vs 1.683 ± 0.093 , $P < 0.02$, for the untreated set. Bone sodium, carbonate, and magnesium were also measured but showed no differences.

DISCUSSION

The normal process of bone maturation involves the cellular elaboration of amorphous calcium phosphate (ACP), which is metastable in relation to crystalline OH-apatite (HA) (6). Transformation of ACP to HA progresses with age. In growing rat bone, ^a 70% ratio of ACP to HA at ⁸ days decreases to approximately 36% ACP at ⁶⁰ days and thereafter (7). This declining ratio of ACP to HA is also characterized by an increasing Ca/P ratio (7) and an increased bone density (8, 9). The physical characteristics of this maturation process result in a progressive increase in crystal surface fluid interface because HA crystals are needle-shaped in contrast to the spherical shape of ACP (10), and because the mean crystal size increases until 40 days of age (11). Comparison by gas absorption of the surface area of ACP to that of HA reveals that the

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| | | Vitamin D-treated $(n = 6)$ | Untreated $(n = 6)$ | | | |
|---|-------|--------------------------------|------------------------|--------------|------------------------|--|
| Parameter | Mean | SD | Mean | SD | \boldsymbol{p} value | |
| Bone mineralized, $\%$ | 91.0 | \pm 5.5 | 83.6 | ± 9.6 | ${<}0.02$ | |
| osteoid, $\%$ | 9.0 | \pm 5.5 | 16.4 | ± 9.6 | < 0.02 | |
| Bone perimeter, forming, $\%$ | 38.3 | ± 14.2 | 65.7 | ± 11.1 | < 0.001 | |
| resorbing, $\%$ | 8.5 | ± 1.9 | 7.4 | ± 23 | NS | |
| R_{BF} mm ³ /mm ³ \times 10 ⁻⁸ | 4.0 | ± 4.4 | 12.4 | \pm 9.9 | NS | |
| Bone calcium, meq/g | 9.650 | ± 1.016 | 8.390 | ± 1.275 | NS (<0.10) | |
| Phosphate, meq/g | 8.010 | ± 0.697 | 7.389 | ± 0.884 | NS. | |
| Ca/P molar ratio | 1.804 | ± 0.040 | 1.683 | ± 0.093 | < 0.02 | |
| Carbonate, meq/g | 1.342 | ± 0.153 | 1.193 | ± 0.277 | NS. | |
| $CO3/Ca$ ratio | 0.139 | ± 0.005 | 0.141 | ± 0.018 | NS | |
| Magnesium, meq/g | 0.290 | ± 0.046 | 0.294 | ± 0.059 | NS. | |
| Mg/Ca ratio | 0.030 | ± 0.0025 | 0.0353 | ± 0.0059 | NS (< 0.10) | |
| Sodium, meq/g | 0.283 | ± 0.015 | 0.278 | ± 0.016 | NS. | |
| OH-Proline, $mmol/g$ | 0.275 | ± 0.018 | 0.312 | ± 0.041 | NS (< 0.10) | |
| Serum bicarbonate, meg/liter | 18.0 | ± 1.5 | 23.0 | ± 1.7 | 0.01 | |

TABLE III Histological and Chemical Measurements of Iliac Crest Biopsies of Vit. D- Treated and Untreated Dialysis Patients

Patients were matched by age and duration of dialysis.

HA has 2-4 times the surface area, reflecting differences in geometry and particle size (12).

Coincident with bone crystal maturation is the cellular elaboration of collagen. This collagen matrix has decreased solubility as it becomes mineralized (13, 14).

While there is presently no direct evidence regarding the carbonate content of ACP, indirect evidence implies that ^a higher ratio of ACP to HA is associated with a decrease in carbonate content. As previously noted, the normal aging process is associated with decreasing ACP and increasing HA. In studies of mineral maturation in the chick embryo, a corresponding progressive increase was seen in bone carbonate from 0.6 mmol/g ash at 9 days to 1.38 mmol/g ash at six mo $(15).$

Magnesium is known to stabilize ACP and to inhibit the formation of HA in vitro (16). Perhaps more important to the interpretation of the increased Mg herein reported is that it has been found to be surface-located in relation to crystal lattice (17). Mg content has not been studied as a function of bone maturation.

In the uremic state there is considerable evidence of altered bone metabolism. In in vitro studies of bone from patients with uremia, increased O2 uptake, lactate production, and bone formation resorption have been observed (18). In rats with chronic renal insufficiency a maturational defect has been characterized as an increase in soluble collagen and a decrease in bone density (1, 2). In studies of human necropsy material, decreased bone carbonate (19), Ca/P ratio (19, 20), and decreased bone density (19) have been reported. Since the advent of atomic absorption, increased Mg has been reported in necropsy material (21, 22), but contrary to our findings, normal Mg has been reported in biopsy material (23).

It thus appears that in necropsy material (19), and now in our biopsy material, a progression of renal osteodystrophy is associated with decreasing carbonate content. This information does not imply that the complexity of the bone carbonate decrease is resolved. For many years it has been known that acidosis can reduce bone carbonate (24, 25). Bone carbonate can decrease 1.5% in 6 h with an acute acid load.8 This decrease is entirely as sodium carbonate. In dogs rendered acidotic for 5-10 days there is a 7% decrease in bone carbonate. Approximately 80% of this carbonate decrease is in excess of Na (3) and is presumably associated with calcium as judged by calciuresis (26). It is now also clear that progression of renal osteodystrophy from the minimal lesion to osteofibrosis is associated with a progressive decline in bone carbonate. Yet from the observations herein reported, more acidotic dialysis patients on chronic maintenance hemodialysis did not manifest greater decreases in bone carbonate than did mildly acidotic patients.

With regard to bone Mg there appears to be even

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greater uncertainty. It is clear that an increase in Mg is present in patients with minimal morphologic bone change and is greatly increased in osteofibrosis. It is tempting to consider the possibility that the presence of increased bone Mg contributes to the maturational defect because of its known capability of increasing the stability of ACP and inhibiting the formation of crystalline apatite (16). However, such a postulate permits no reasonable explanation for the presence of the increased Mg. It seems more likely that events at the cellular level result in increased metabolic activity and increased bone turnover, resulting in an increased ACP and decreased apatite crystals, probably of smaller size. Thus, if Mg is surface-located in the lattice, these events would result in the presence of higher Mg/Ca ratios. The inverse relationship of Mg and CO₃ content observed in the present study (Fig. 1) might be interpreted as having a common etiology. However, this inverse relationship does not always exist, as evidenced by the fact that in the dogs rendered acidotic for 7-10 days we failed to find in retrospective analysis any increase in Mg (3) , although the bone carbonate was decreased 7%.

The finding of a decreased density in osteofibrosis may well reflect the increased ACP-HA ratio seen in less mature bone $(2, 8, 9)$; however, some caution must be observed. An alternative could be that increased fibrous tissue in the marrow spaces of the osteofibrotic set could increase the true volume, particularly by replacing lipid, thereby decreasing density.

The data herein reported would appear to identify in human biopsy material the existence of a maturational defect previously found only in growing rats (2). These data extend the characterization of the maturational defect to include a decrease in carbonate and an increase in magnesium. The Mg increase could be solely the consequence of the presumed increase in surface area; however, the only evidence for such a speculation is its known concentration in the hydration shell around apatite crystals (17).

The histologic correlation with the maturational defect appears to be good. The lesser increases in RBF and resorption seen in the mild lesion are associated with smaller changes in bone CO_s and Mg. The very high R_{BF} and resorption seen in osteofibrosis are associated with the most severe alteration of bone CO_s and Mg and with a reduction in bone density.

Evidence as to the role of vitamin D deficiency in production of the maturational defect is conflicting. In rats fed ^a diet deficient in vitamin D the ACP content is increased and Ca/P ratio decreased (6). Yet in studies in uremic rats vitamin D therapy failed to influence the maturational defect (2), while in our biopsy data vitamin D therapy appeared to increase Ca/P ratios, but did not influence bone CO₃ or Mg.

The remaining known factor in renal osteodystrophy is hyperparathyroidism. The correlation of osteofibrosis with both hyperparathyroidism (27) and with the severity of the maturational defect is striking. Also, it seems likely that all of our patients have hyperparathyroidism (28, 29). However, the question of whether the maturational defect described in rats $(1, 2)$ and now in humans can be ascribed solely to hyperparathyroidism must remain unanswered at present. A possible alternative or additional factor from the accumulation of urenmic toxins may well have as yet undescribed effects on bone cellular metabolism.

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