JCI The Journal of Clinical Investigation

Acute Parathyroid Hormone Response to Epinephrine In Vivo

Jan A. Fischer, ..., Jürg W. Blum, Ulrich Binswanger

J Clin Invest. 1973;52(10):2434-2440. https://doi.org/10.1172/JCI107434.

Research Article

The acute effects of epinephrine, norepinephrine, and isoproterenol on the plasma immunoreactive parathyroid hormone (iPTH) response were studied in 13 550-600 kg cows. Catecholamines were infused for 7.0 min. During epinephrine infusions at 0.08 μ mol/min iPTH increased from 0.48±0.12 (mean±SE, ng/ml) to 1.09±0.18 ng/ml (*P* < 0.02). Small increases in plasma free fatty acids and glucose could be detected with 0.08 μ mol/min epinephrine; the iPTH response to epinephrine was as sensitive as the free fatty acid and glucose responses and possibly of physiological importance. Plasma calcium (total and ionized) and magnesium did not change.

The responses were more pronounced at 0.8 μ mol/min epinephrine with a mean iPTH increase from 0.49±0.16 ng/ml to 1.74±0.35 ng/ml (*P* < 0.01). Small decreases in plasma calcium occurred at 0.8 μ mol/min epinephrine, but the plasma magnesium remained unchanged. However, when the plasma calcium was lowered with ethylene glycol bis(β -aminoethyl ether)-*N*, *N*'-tetraacetic acid (EGTA), a much more pronounced lowering of the plasma calcium was required to produce comparable increases of the plasma iPTH concentrations than when epinephrine was infused. It appears that epinephrine has a direct effect on the release of iPTH from the parathyroid glands.

Simultaneous infusions of calcium and epinephrine suppressed the stimulation by epinephrine. This points towards a common mechanism of the regulation of parathyroid hormone secretion caused by decreases in the extracellular [...]



Find the latest version:

https://jci.me/107434/pdf

Acute Parathyroid Hormone Response to Epinephrine In Vivo

JAN A. FISCHER, JÜRG W. BLUM, and ULRICH BINSWANGER

From the Research Laboratory for Calcium Metabolism, Department of Orthopedic Surgery, Department of Medicine, University of Zurich, 8008 Zurich, and the Department of Agriculture, Federal Institute of Technology, 8006 Zurich, Switzerland

A BSTRACT The acute effects of epinephrine, norepinephrine, and isoproterenol on the plasma immunoreactive parathyroid hormone (iPTH) response were studied in 13 550-600 kg cows. Catecholamines were infused for 7.0 min. During epinephrine infusions at 0.08 μ mol/min iPTH increased from 0.48±0.12 (mean±SE, ng/ml) to 1.09±0.18 ng/ml (P < 0.02). Small increases in plasma free fatty acids and glucose could be detected with 0.08 μ mol/min epinephrine; the iPTH response to epinephrine was as sensitive as the free fatty acid and glucose responses and possibly of physiological importance. Plasma calcium (total and ionized) and magnesium did not change.

The responses were more pronounced at 0.8 μ mol/min epinephrine with a mean iPTH increase from 0.49±0.16 ng/ml to 1.74±0.35 ng/ml (P < 0.01). Small decreases in plasma calcium occurred at 0.8 μ mol/min epinephrine, but the plasma magnesium remained unchanged. However, when the plasma calcium was lowered with ethylene glycol bis(β -aminoethyl ether)-N, N'-tetraacetic acid (EGTA), a much more pronounced lowering of the plasma calcium was required to produce comparable increases of the plasma iPTH concentrations than when epinephrine was infused. It appears that epinephrine has a direct effect on the release of iPTH from the parathyroid glands.

Simultaneous infusions of calcium and epinephrine suppressed the stimulation by epinephrine. This points towards a common mechanism of the regulation of parathyroid hormone secretion caused by decreases in the extracellular calcium concentration and/or alterations in the distribution of calcium within parathyroid cells following the administration of epinephrine.

The iPTH response to epinephrine was suppressed in the presence of propranolol. Isoproterenol was less active in raising iPTH than epinephrine, and norepinephrine was the least active. The stimulation by isoproterenol and the suppression by propranolol suggest beta adrenergic receptor sites within the parathyroid glands.

INTRODUCTION

The most important factors regulating parathyroid hormone (PTH)¹ secretion are the plasma calcium and magnesium concentrations. There is an inverse relationship between the responses of immunoreactive parathyroid hormone (iPTH) and changes in plasma calcium and magnesium (1–5). A decrease in the plasma ionized calcium concentration (Ca⁺⁺) rather than a fall in plasma total calcium (Ca) is responsible for the acute iPTH response in the cow in vivo (5).

There is evidence from the study of parathyroid explants in vitro that the addition of catecholamines, dibutyryl cyclic adenosine 3',5'-monophosphate (cyclic AMP), and thyrocalcitonin to the incubation medium stimulates PTH secretion (6–9). The recent demonstration in homogenates of parathyroid glands of adenylate cyclase activity that is inhibited by calcium and stimulated by thyrocalcitonin suggests that adenylate cyclase may mediate the effects of calcium on PTH secretion (10, 11). In view of the current concept that the actions

The Journal of Clinical Investigation Volume 52 October 1973 · 2434-2440

This work was presented in part at the National Meeting of the American Federation for Clinical Research, Atlantic City, N. J., 29 April 1973.

Received for publication 12 April 1973 and in revised form 4 June 1973.

¹ Abbreviations used in this paper: Ca, concentration of total plasma calcium; Ca⁺⁺, concentration of ionized plasma calcium; cyclic AMP, cyclic adenosine 3',5'-monophosphate; EGTA, ethylene glycol bis(β -aminoethyl ether)-N,N'-tetraacetic acid; iPTH, concentration of plasma immunoreactive parathyroid hormone; Mg, concentration of plasma magnesium; PTH, parathyroid hormone.

of beta-adrenergic agents are mediated by increases in cyclic AMP formation in target tissues (12) we have investigated the effects of epinephrine, norepinephrine, and isoproterenol on the regulation of PTH secretion in vivo in the cow.

METHODS

Experimental design. Experiments were performed in 13 fasting 5-10 yr old, lactating Brown Swiss cows. They weighed 550-600 kg, and they were kept on a roughage diet supplemented with cornsilage. Starting at 9 a.m. blood was drawn from one jugular vein, and catecholamines or ethylene glycol bis (β -aminoethyl ether)-N,N'-tetraacetic acid (EGTA) were infused into the contralateral vein through indwelling catheters. The animals were not anesthetized. 15 ml of blood was drawn aneorobically without stasis into 20 ml plastic syringes containing 30 U.S.P.U. of heparin (Hoffmann-La Roche, Inc., Basel, Switzerland). 5 ml of blood was immediately placed under oil, and the plasma was separated and used for the determination of the ionized plasma calcium concentrations (Ca⁺⁺); 4 U.S.P. U of heparin per milliliter did not affect the determination of Ca⁺⁺ and was used to prevent coagulation (13, 14). The remaining 10 ml of blood was put on ice, and the plasma was separated, frozen, and subsequently used for all the other determinations.

After four base-line specimens of blood were obtained at 2-min time intervals, catecholamines dissolved in 0.9% NaCl or EGTA neutralized with NaOH (pH 7.4) were infused for 7.0 min with a peristaltic pump (Vario-Perpex II, W. Meyer, Lucerne, Switzerland). Additional blood samples were obtained at 2, 4, 6, 7, 8, 10, 13, 16 min after the start of the infusions. When the effects of epinephrine were compared with epinephrine-plus-calcium and epinephrine-plus-propranolol, epinephrine was infused from time "0" to 7.0 min and from 60.0 to 67.0 min.

Propranolol or calcium was infused through a second catheter driven by a second identical peristaltic pump. Propranolol dissolved in 0.9% NaCl was infused at 70 μ mol/min from 56.0 to 68.0 min. Calcium was infused as 10% calciumborogluconate at 6.7 mmol/min from 38.0 to 45.0 min and at 2.0 mmol/min for the remainder of the experiments.

Results were either expressed as mean values (with SEM) or as the mean of the incremental or decremental (Δ) changes of samples taken at 6, 7, and 8 min after the start of the epinephrine infusions (6-8' Δ).

Before and during the infusion with catecholamines the heart rate was estimated at 1-2 min time intervals.

Materials. Isoproterenol HCl was obtained from Winthrop Products Co., Surbiton-on-Thames, England, and propranolol was a gift from Imperial Chemical Industries, Macclesfield, England. Epinephrine and norepinephrine were purchased from Fluka AG, Buchs, Switzerland, and EGTA was purchased from the City Chemical Corp., New York.

Methods of analysis. Immunoreactive parathyroid hormone was determined in multiple dilutions in plasma by radioimmunoassay according to Arnaud, Tsao, and Littledike (15). The following alterations were made: Highly purified 1-84 amino acid bovine PTH extracted from parathyroid glands (Wilson Laboratories, Chicago, Ill.; Lot 147865) was used for iodination with ¹³⁰I (The Radiochemical Centre, Amersham, England) and as standard. Chicken 14 M anti-bovine PTH serum was used at a final concentration of 1: 1,500. Dilution curves using hyperparathyroid



FIGURE 1 Effects of 0.08 and 0.8 μ mol/min epinephrine infused from time "0" to 7.0 min on plasma iPTH, Ca, Ca⁺⁺, glucose, and free fatty acid (FFA) levels. Each value represents the mean±SEM of six separate experiments. Open symbols represent statistically significant changes from the mean of four preinfusion levels ($\bigcirc P < 0.05$, $\square P < 0.01$, $\triangle P < 0.001$), closed symbols ($\bullet P > 0.05$).

plasma and 1-84 bovine PTH were practically superimposable indicating immunological similarity (16). Furthermore, the affinity of chicken 14 M anti-bovine PTH serum for synthetic 1-34 bovine PTH (17) (Beckman Instruments, Inc., Palo Alto, Calif.) is practically identical (on the basis of protein) to its affinity for 1-84 bovine PTH extracted from bovine parathyroid glands. This suggests that this antiserum reacts well with the biologically active region of the PTH molecule (amino terminal).² All samples from one experiment were analyzed in the same assay. The coefficient of variation of a measurement within the same assay was 12% and between assays 14%.

Calcium was determined by automatic EGTA titration using calcein as an indicator (Marius Calcium Titrator, Utrecht, The Netherlands) (coefficient of variation 0.8%). Ca⁺⁺ was measured by a flow-through electrode system (Orion Research, Inc., Cambridge, Mass.; Model 99-20)

^a Arnaud, C. D. Personal communication.

Epinephrine and Parathyroid Hormone 2435

(coefficient of variation 0.7%) (13, 14). Changes in pH in response to the various infusions did not exceed 0.06 units; the mean changes were 0.00 (SEM 0.004) (n = 58) and therefore did not affect the changes in Ca⁺⁺. Magnesium was measured by atomic absorption flame spectrophotometry (Perkin-Elmer Corp., Norwalk, Conn.; Model 305) (coefficient of variation 1.3%). Glucose was measured by an oxidase method (coefficient of variation 1.1%) (18), and the free fatty acid concentrations were assayed by a colorimetric technique (coefficient of variation 5.0%) (19).

Statistical analysis was done by paired t test and by regression analysis (20).

RESULTS

The acute immunoreactive parathyroid hormone rcsponse to epinephrine. Epinephrine was infused for 7.0 min, and it briskly increased the concentration of iPTH in the plasma with peak values occurring between 4 and 8 min (Fig. 1). iPTH responses were compared to the known effects of epinephrine on free fatty acid and glucose release. For this purpose epinephrine was infused at a relatively low dose of 0.08 μ mol/min into six cows. iPTH and free fatty acids increased significantly during the infusions (P < 0.05), whereas no significant increase in plasma glucose levels could be detected until after the end of the epinephrine infusions. The heart rate did not change. Plasma Ca and Mg concentrations remained unchanged.

In contrast, when on a different day $0.8 \ \mu mol/min$ epinephrine was infused into the same animals the iPTH response was more pronounced together with a delayed

but significant (P < 0.001) increase in plasma glucose and free fatty acid levels. Due to the infusions, there was an increase in the heart rate from 60 to 80 up to 110 per minute. The plasma Mg did not change. The plasma Ca decreased significantly by 0.20 mg/100 ml (P < 0.02) (Ca⁺⁺ by 0.16 mg/100 ml, (P < 0.001)) at the end of the epinephrine infusions.

The possibility remained that small decreases in plasma Ca were the reason for the increase in iPTH. This was tested with additional experiments in which the responses to epinephrine were compared with EGTA. Fig. 2 shows values for plasma iPTH (6-8' Δ) plotted as a function of corresponding values for plasma Ca (and Ca⁺⁺) (6-8' Δ) for responses to 0.04-1.60 µmol/min epinephrine and 0.25-6.0 mmol/min EGTA. There was a highly significant (P < 0.001) negative, linear correlation between plasma Ca and iPTH (6-8' Δ) responses in the experiments with EGTA. The plasma Ca concentrations decreased slightly in the majority of the experiments with epinephrine. Visual inspection of this plot shows almost complete discrimination between responses to epinephrine and to EGTA.

Effects of calcium and propranolol on the acute immunoreactive parathyroid hormone response to epinephrine. The results are summarized in Table I and in Fig. 3.

 $0.8 \ \mu mol/min$ epinephrine were infused into four cows from time "0" to 7.0 min and again from 60.0 to 67.0 min. The two infusions of epinephrine produced es-



FIGURE 2 Plasma iPTH (6-8' Δ) as a function of plasma Ca and Ca⁺⁺ (6-8' Δ) in 27 experiments (13 cows) infused with 0.04-1.60 μ mol/min epinephrine (\bullet), and in 11 experiments (8 cows) infused with 0.25 to 6.0 mmol/min EGTA from time "0" to 7.0 min (\bigcirc) (correlation coefficients, *r*, for iPTH responses to EGTA = -0.94 with Ca and = -0.93 with Ca⁺⁺, (P < 0.001).

2436 J. A. Fischer, J. W. Blum, and U. Binswanger

		Bas	al*		1st	epinephrine	infusion,‡ (6–	8' Δ)		60 n	nin		2nd	epinephrine in	ifusion,‡ (6-8'	(ک ا
	iPTH	Ca	Ca++	Mg	iPTH	Ca	Ca++	Mg	iPTH	Ca	Ca++	Mg	iPTH	Ca	Ca++	Mg
	lm/gn	mg/100	mg/100	mg/100	ng/ml	mg/100	mg/100	mg/100	1m/Bu	mg/100 ml	mg/100 ml	mg/100 ml	ng/ml	mg/100 ml	mg/100 ml	mg/100 ml
<u>,</u>	1. 00.00	1.4:1-1-0-0-0	111	1111		1111										
	1. IIU at		2 0.2	1 00	0.64	-010	c	-0.07	0.48	10.60	3.96	1.77	0.65	-0.02	0.02	0.05
- ~	0.60	0.31	3.61	2.26	10.0	01.0	-0.17	-0.01	0.72	9.03	3.59	2.18	0.74	-0.02	-0.02	-0.06
1 თ	0.81	9.01	3.63	2.18	0.90	0	0.11	-0.01	0.66	9.01	3.59	2.18	1.57	-0.12	-0.08	0
4	0.25	10.53	4.81	2.20	0.43	-0.16	-0.08	-0.04	0.17	10.58	4.93	2.16	0.33	0.09	0.07	-0.01
Mean	0.56	9.91	3.99	2.14	0.74	-0.07	-0.03	-0.02	0.51	9.81	4.02	2.07	0.82	-0.02	0	-0.01
SE	0.12	0.44	0.28	0.08	0.12	0.04	0.06	0.01	0.12	0.45	0.32	0.10	0.26	0.04	0.03	0.02
Group	II: calci	ium infus	ion from	38 to 68	min (see	text)										
1	0.18	10.68	5.02	2.35	0.44	-0.10	-0.05	-0.05	0.10	13.96	6.45	2.13	0	-0.06	-0.08	-0.01
2	0.46	9.43	4.29	1.70	1.43	0	0	0	0.16	11.83	4.99	1.62	0.02	0.04	0.02	0.02
ŝ	0.44	9.68	4.29	1.94	0.58	-0.12	-0.06	-0.01	0.34	11.73	5.16	1.81	-0.06	-0.09	-0.03	-0.02
4	1.03	10.10	3.93	2.19	0.70	-0.01	0.01	0.01	0.54	11.33	5.10	2.14	0	-0.04	-0.02	0
Mean	0.53	9.97	4.38	2.05	0.78	- 0.06	-0.03	-0.01	0.29	12.21	5.42	1.93	-0.01	-0.04	-0.03	0
SE	0.18	0.27	0.23	0.14	0.22	0.03	0.02	0.01	0.10	0.59	0.34	0.13	0.02	0.03	0.02	0.01
Group	III: pro	pranolol	infusion	(70 µmol	/min) fro	m 56 to 68	min									
-	0.81	9.70	3.53	2.18	1.42	-0.22	-0.01	0	0.66	9.78	3.73	2.18	0.04	-0.16	-0.08	-0.02
2	0.75	9.01	3.59	2.19	2.06	-0.12	-0.08	-0.05	0.69	9.00	3.61	2.08	0.09	-0.10	-0.10	-0.09
3	0.60	10.10	4.48	2.27	1.33	-0.15	-0.09	-0.01	0.40	10.10	4.44	2.34	0.05	-0.15	-0.05	0.02
4	0.18	10.13	4.19	1.92	0.91	-0.01	0.01	-0.02	0.19	10.25	4.28	1.88	0.23	-0.08	0.03	0.05
5	0.22	9.90	4.24	2.33	0.43	0	-0.02	-0.02	0.21	9.82	4.19	2.23	0.19	-0.22	0.01	-0.02
Mean	0.51	9.77	4.01	2.18	1.23	-0.10	-0.04	-0.02	0.43	9.79	4.05	2.14	0.12	-0.14	-0.04	-0.01
SE	0.13	0.20	0.19	0.07	0.27	0.04	0.02	0.01	0.11	0.22	0.16	0.08	0.04	0.02	0.03	0.02
* Mean ‡ Epine	of four phrine v	samples. was infus	ed at 0.8	#mol/m	in from ti	me "0" to	7.0 min (1	st infusion) and froi	m 60.0 to	67.0 mir	1 (2nd inf	usion).			

esium Responses to Epinephrine and to 20 A Ma TABLE I H F? 1 5 140

Epinephrine and Parathyroid Hormone

2437



FIGURE 3 The iPTH results of Table I are summarized. Plasma iPTH (mean±SEM of samples taken at 6, 7, 8 min) during 1st (\bigcirc) and 2nd (\bullet) epinephrine infusions. No addition refers to group I, the addition of calcium to group II and the addition of propranolol to group III. P values indicate statistically significant decreases of plasma iPTH during epinephrine-plus-calcium and epinephrine-plus-propranolol infusions (2nd infusion) in comparison to epinephrine alone (1st infusion).

sentially the same iPTH responses. Plasma calcium and magnesium concentrations $(6-8'\Delta)$ did not change significantly. In different groups of cows 0.8 μ mol/min epinephrine were infused from time "0" to 7.0 min in the absence of and from 60.0 to 67.0 minutes in the presence of a raised plasma calcium concentration or of propranolol.

The plasma calcium was increased with an infusion of 6.7 mmol calcium/min and kept at a plateau with an in-

fusion of 2.0 mmol/min for the remainder of the experiment. Plasma Ca was raised significantly from 9.97 ± 0.27 (mean \pm SE, mg/100 ml) to 12.21 ± 0.59 mg/100 ml (P < 0.02) and the plasma Ca⁺⁺ from 4.38 ± 0.23 to 5.42 ± 0.34 mg/100 ml (P < 0.01). As a consequence the plasma iPTH was decreased but not significantly (P < 0.1). When epinephrine was infused from 60.0 to 67.0 min in the presence of a raised plasma Ca, iPTH did not change. The difference of the epinephrine response (mean $6-8'\Delta$) in the absence and in the presence of a raised plasma Ca level was significant (P < 0.05). The plasma Mg did not change.

When propranolol was infused at 70 μ mol/min, the iPTH at 60 min was decreased but not significantly (P < 0.2). In the presence of epinephrine there was a small increase in iPTH of 0.12±0.04 ng/ml, which contrasted with a larger increase of 1.23±0.27 ng/ml (mean 6-8' Δ) in the absence of propranolol. The difference of the iPTH response was significant (P < 0.025).

The acute immunoreactive parathyroid hormone response to isoproterenol and norepinephrine. At 0.08 μ mol/min isoproterenol was inactive, but at 0.8 μ mol/ min the iPTH levels increased significantly (P < 0.01) (Fig. 4). Isoproterenol had a somewhat lower potency



FIGURE 4 Effects of 0.08 and 0.8 μ mol/min isoproterenol infused from time "0" to 7.0 min on plasma iPTH, Ca, Ca⁺⁺, and Mg levels. Each value represents the mean±SEM of five separate experiments. Open symbols represent statistically significant changes from the mean of four preinfusion levels ($\bigcirc P < 0.05$, $\square P < 0.01$), closed symbols ($\oplus P > 0.05$).

2438 J. A. Fischer, J. W. Blum, and U. Binswanger



FIGURE 5 Effects of 0.8 and 4.6 μ mol/min norepinephrine infused from time "0" to 7.0 min on plasma iPTH, Ca, Ca⁺⁺, and Mg levels. Each value represents the mean±SEM of five separate experiments. Open symbols represent statistically significant changes from the mean of four preinfusion levels ($\bigcirc P < 0.05$, $\square P < 0.01$), closed symbols ($\bullet P >$ 0.05).

than epinephrine in sharply raising iPTH levels. Plasma Ca and Mg did not change significantly during the infusions.

Norepinephrine produced significant iPTH responses at 4.6 μ mol/min (P < 0.01), and at 0.8 μ mol/min it was inactive (Fig. 5). Norepinephrine was less active on a molar basis in raising iPTH than either epinephrine or isoproterenol. Plasma Mg and Ca did not change significantly during the infusions.

DISCUSSION

This study demonstrates that infusions with catecholamines in vivo result in a sharp rise in plasma iPTH. The fact that the antiserum used in the measurement of iPTH was directed against the biologically active amino terminal region of bovine PTH (17) suggests that the secretion of PTH is increased. However, because the measurements have not been performed in the venous effluent of the parathyroid glands but in peripheral plasma samples, a metabolic alteration of the circulating iPTH peptides cannot be ruled out.

The increase in iPTH was demonstrated at a dose level of epinephrine $(0.08 \ \mu mol/min)$ that enhanced the release of free fatty acids and of glucose only slightly (21). The heart rate did not change; it appears unlikely that hemodynamic alterations affected the iPTH response to epinephrine at the relatively low dose used. The plasma Ca (and Ca⁺⁺) and Mg did not change.

When higher amounts of epinephrine (0.8 µmol/ min) were infused the iPTH response was more pronounced, and it was followed by a delayed and marked rise in free fatty acids and glucose and a moderate rise in the heart rate. There were small decreases of the plasma ionized and total calcium and no changes of the plasma magnesium concentrations. More important, however, is the demonstration that decreases of plasma Ca (and Ca⁺⁺) produced by epinephrine infusions increased iPTH much more than comparable decreases of plasma calcium produced by EGTA. In addition to inducing small decreases in the plasma calcium concentrations, epinephrine appears to be able to raise iPTH directly. This is supported by the stimulation of the iPTH secretion with epinephrine seen in vitro in parathyroid explant cultures (6, 7).

The presence of a rich adrenergic nerve supply to the parathyroid glands (22, 23) suggests a possible role for the autonomic nervous system in the physiological regulation of PTH secretion. A direct neural mechanism has been demonstrated for the beta adrenergic-mediated control of glucagon secretion (24). The existence of a similar mechanism in parathyroid glands is possible because iPTH responses to epinephrine can occur without marked decreases of the plasma ionized and total calcium and no changes of the plasma magnesium concentrations. Epinephrine was more active on a molar basis than isoproterenol in sharply raising iPTH in vivo, and norepinephrine was the least active. The stimulation by the beta specific agent isoproterenol and the suppression of the iPTH response to epinephrine by the betablocking agent propranolol suggest a beta adrenergic receptor site within parathyroid glands.

The action of beta adrenergic agents are probably mediated by increases in cyclic AMP. Ball et al. (12) have demonstrated that plasma cyclic AMP increased in response to infusions with epinephrine, isoproterenol, or norepinephrine, but no increase occurred in the presence of propranolol. These observations are consistent with the concept that the action of beta adrenergic agents are mediated by increases in cyclic AMP in target tissues. This is supported by the results obtained in vitro by Abe and Sherwood (8, 25), who demonstrated that the quantity of iPTH released into the incubation medium as a result of low calcium and magnesium concentration paralleled the release of cyclic AMP. The demonstration by Dufresne and Gitelman (10) and by Matsuzaki and Dumont (11) of adenvlate cyclase activity in homogenates of parathyroid glands that is inhibited by calcium suggests that adenylate cyclase may mediate the known effects of calcium on PTH secretion. Simultaneous infusions of calciumborogluconate abolished the effects of epinephrine, suggesting that concentration changes or translocation of calcium inside parathyroid cells may modulate the response of iPTH to epinephrine. This is a distinct possibility in view of the inhibition by calcium of the specific binding of adrenergic catecholamines to a subcellular fraction from cardiac muscle (26). It points towards a common mechanism of the stimulation of PTH secretion caused by decreases in extracellular calcium concentration and/or by epinephrine and of the suppression of PTH secretion with calcium.

The recent description of a 12 yr old boy with familial pheochromocytoma and mild hypercalcemia, whose serum calcium returned to normal levels after adrenalectomy suggests that he might temporarily have suffered from a new form of secondary hypercalcemic hyperparathyroidism caused by the raised epinephrine levels in the blood. The possibility that the pheochromocytoma of this patient secreted a substance with PTH-like activity cannot, of course, be ruled out (27).

ACKNOWLEDGMENTS

We are indebted to Dr. Claude Arnaud, Mayo Clinic, Rochester, Minn. for the generous gift of the antiserum and to Bayer Pharma AG, Zurich, Switzerland for the Trasylol used in the radioimmunoassay for PTH. The free fatty acid determinations were kindly performed by Dr. J. Zapf, Department of Medicine, University of Zurich. We are grateful to Dr. M. Berchtold, School of Veterinary

Epinephrine and Parathyroid Hormone 2439

Medicine, University of Zurich, who kindly supplied us with cows.

The excellent and precise technical assistance of Miss M. Balzer, Miss M. Eschler, Mrs. K. Wuethrich, Mrs. E. Tammilehto, and Mr. E Wasser is gratefully acknowledged.

This work was supported by grants from the Schweizerische Nationalfonds zur Förderung der wissenschaftlichen Forschung (Nr. 3.807.72), by the Sandoz-Stiftung zur Förderung der biologisch-medizinischen Wissenschaften, by the Emil-Barrell-Stiftung of Hoffman-La Roche Co., by the EMDO Stiftung, by the Hartmann-Müller-Stiftung, and by the Stiftung für wissenschaftliche Forschung an der Universität Zurich.

REFERENCES

- 1. Sherwood, L. M., J. T. Potts, Jr., A. D. Care, G. P. Mayer, and G. D. Aurbach. 1966. Evaluation by radioimmunoassay of factors controlling the secretion of parathyroid hormone. Intravenous infusions of calcium and ethylenediamine tetraacetic acid in the cow and goat. *Nature (Lond.).* 209: 52.
- Sherwood, L. M., J. T. Potts, Jr., A. D. Care, G. P. Mayer, and G. D. Aurbach. 1966. Perfusion of isolated parathyroid gland of the goat and sheep. *Nature* (Lond.). 209: 55.
- Sherwood, L. M., G. P. Mayer, C. F. Ramberg, Jr., D. S. Kronfeld, G. D. Aurbach, and J. T. Potts, Jr. 1968. Regulation of parathyroid hormone secretion: proportional control by calcium, lack of effect of phosphate. *Endocrinology.* 83: 1043.
- 4. Arnaud, C. D., T. Littledike, and H. S. Tsao. 1970. Calcium homeostasis and the simultaneous measurement of calcitonin and parathyroid hormone in the pig. *In* Calcitonin 1969. S. Taylor and G. Foster, editors. W. Heinemann Ltd., London. 95.
- 5. Fischer, J. A., U. Binswanger, and J. W. Blum. 1973. The acute parathyroid hormone response to changes in ionized calcium during phosphate infusions in the cow. *Eur. J. Clin. Invest.* 3: 151.
- Sherwood, L. M., and M. Abe. 1972. Adrenergic receptors and the release of parathyroid hormone. J. Clin. Invest. 51: 88a. (Abstr.)
- 7. Williams, G. A., G. K. Hargis, E. N. Bowser, W. J. Henderson, and N. J. Martinez. 1973. Evidence for a role of adenosine 3',5'-monophosphate in parathyroid hormone release. *Endocrinology*. 92: 687.
- 8. Abe, M., and L. M. Sherwood. 1972. Regulation of parathyroid hormone secretion by adenyl cyclase. *Biochem. Biophys. Res. Commun.* 48: 396.
- Fischer, J. A., S. B. Oldham, G. W. Sizemore, and C. D. Arnaud. 1971. Calcitonin stimulation of parathyroid hormone secretion in vitro. *Horm. Metab. Res.* 3: 223.
- Dufresne, L. R., and H. J. Gitelman. 1972. A possible role of adenyl cyclase in the regulation of parathyroid activity by calcium. *In* Calcium, Parathyroid Hormone and the Calcitonins. R. V. Talmage and P. L. Munson, editors. Excerpta Medica Foundation, Amsterdam. 202.

- 11. Matsuzaki, S., and J. E. Dumont. 1972. Effect of calcium ion on horse parathyroid gland adenyl cyclase. *Biochim. Biophys. Acta.* 284: 227.
- 12. Ball, J. H., N. I. Kaminsky, J. G. Hardman, A. E. Broadus, E. Sutherland, and G. W. Liddle. 1972. Effects of catecholamines and adrenergic-blocking agents on plasma and urinary cyclic nucleotides in man. J. Clin. Invest. 51: 2124.
- 13. Moore, E. W. 1970. Ionized calcium in normal serum, ultrafiltrates, and whole blood determined by ion-exchange electrodes. J. Clin. Invest. 49: 318.
- Studer, O., M. Knob, and U. Binswanger. 1972. Kalziumionenkonzentration im Serum. Schweiz. Med. Wochenschr. 102: 305.
- Arnaud, C. D., H. S. Tsao, and T. Littledike. 1971. Radioimmunoassay of human parathyroid hormone in serum. J. Clin. Invest. 50: 21.
- Arnaud, C. D., G. W. Sizemore, S. B. Oldham, J. A. Fischer, H. S. Tsao, and E. T. Littledike. 1971. Human parathyroid hormone: glandular and secreted molecular species. Am. J. Med. 50: 630.
- 17. Potts, J. T., Jr., G. W. Tregear, H. T. Keutmann, H. D. Niall, R. Sauer, L. J. Deftos, B. F. Dawson, M. L. Hogan, and G. D. Aurbach. 1971. Synthesis of a biologically active N-terminal tetratricontrapeptide of parathyroid hormone. *Proc. Natl. Acad. Sci. U. S. A.* 68: 63.
- Kadish, A. H., R. L. Litle, and J. C. Sternberg. 1968. A new and rapid method for the determination of glucose by measurement of rate of oxygen consumption. *Clin. Chem.* 14: 116.
- 19. Duncombe, W. G. 1963. The colorimetric micro-determination of long-chain fatty acids. *Biochem. J.* 88: 7.
- Diem, K., and C. Lentner. 1968. Documenta Geigy, Wissenschaftliche Tabellen. J. R. Geigy, Basel. 7th edition. 146.
- Steinberg, D. 1966. Catecholamine stimulation of fat mobilization and its metabolic consequences. *Pharmacol. Rev.* 18: 217.
- 22. Altenähr, E. 1971. Electron microscopical evidence for innervation of chief cells in human parathyroid gland. *Experientia* (*Basel*). 27: 1077.
- Yeghiayan, E., J. M. Rojo-Ortega, and J. Genest. 1972. Parathyroid vessel innervation: an ultrastructural study. J. Anat. 112: 137.
- 24. Marliss, E. B., L. Girardier, J. Seydoux, C. B. Wollheim, Y. Kanazawa, L. Orci, A. E. Renold, and D. Porte, Jr. 1973. Glucagon release induced by pancreatic nerve stimulation in the dog. J. Clin. Invest. 52: 1246.
- 25. Sherwood, L. M., and M. Abe. 1972. Magnesium ion, parathyroid hormone secretion and cyclic 3',5'-AMP. *Clin. Res.* 20: 756. (Abstr.)
- Lefkovitz, R. J., G. W. G. Sharp, and E. Haber. 1973. Specific binding of beta-adrenergic catecholamines to a subcellular fraction from cardiac muscle. J. Biol. Chem. 248: 342.
- Swinton, N. W., Jr., E. P. Clerkin, and L. D. Flint. 1972. Hypercalcemia and familial pheochromocytoma. Correction after adrenalectomy. Ann. Intern. Med. 76: 455.