

**Amendment history:**

- [Correction](#) (October 1971)

## The Dynamics of Glomerular Ultrafiltration in the Rat

Barry M. Brenner, ... , Julia L. Troy, Terrance M. Daugharty

*J Clin Invest.* 1971;[50\(8\)](#):1776-1780. <https://doi.org/10.1172/JCI106667>.

**Research Article**

Using a unique strain of Wistar rats endowed with glomeruli situated directly on the renal cortical surface, we measured glomerular capillary pressures using servo-nulling micropipette transducer techniques. Pressures in 12 glomerular capillaries from 7 rats averaged 60 cm H<sub>2</sub>O, or approximately 50% of mean systemic arterial values. Wave form characteristics for these glomerular capillaries were found to be remarkably similar to those of the central aorta. From similarly direct estimates of hydrostatic pressures in proximal tubules, and colloid osmotic pressures in systemic and efferent arteriolar plasmas, the net driving force for ultrafiltration was calculated. The average value of 14 cm H<sub>2</sub>O is lower by some two-thirds than the majority of estimates reported previously based on indirect techniques. Single nephron GFR (glomerular filtration rate) was also measured in these rats, thereby permitting calculation of the glomerular capillary ultrafiltration coefficient. The average value of 0.044 nl sec<sup>-1</sup> cm H<sub>2</sub>O<sup>-1</sup> glomerulus<sup>-1</sup> is at least fourfold greater than previous estimates derived from indirect observations.

**Find the latest version:**

<https://jci.me/106667/pdf>



# The Dynamics of Glomerular Ultrafiltration in the Rat

BARRY M. BRENNER, JULIA L. TROY, and TERRANCE M. DAUGHARTY

*From the Departments of Medicine, Veterans Administration Hospital, San Francisco, California 94121 and The University of California, San Francisco, San Francisco, California 94122*

**ABSTRACT** Using a unique strain of Wistar rats endowed with glomeruli situated directly on the renal cortical surface, we measured glomerular capillary pressures using servo-nulling micropipette transducer techniques. Pressures in 12 glomerular capillaries from 7 rats averaged 60 cm H<sub>2</sub>O, or approximately 50% of mean systemic arterial values. Wave form characteristics for these glomerular capillaries were found to be remarkably similar to those of the central aorta. From similarly direct estimates of hydrostatic pressures in proximal tubules, and colloid osmotic pressures in systemic and efferent arteriolar plasmas, the net driving force for ultrafiltration was calculated. The average value of 14 cm H<sub>2</sub>O is lower by some two-thirds than the majority of estimates reported previously based on indirect techniques. Single nephron GFR (glomerular filtration rate) was also measured in these rats, thereby permitting calculation of the glomerular capillary ultrafiltration coefficient. The average value of 0.044 nl sec<sup>-1</sup> cm H<sub>2</sub>O<sup>-1</sup> glomerulus<sup>-1</sup> is at least fourfold greater than previous estimates derived from indirect observations.

## INTRODUCTION

More than a century ago, Ludwig proposed that the initial event in the process of urine formation is the production of an ultrafiltrate of plasma across the glomerular capillary wall (1). Refinement of this view by Starling (2) to indicate that the mechanisms responsible for this ultrafiltrate formation are the same as those governing the movement of fluid across capillary membranes generally (namely the magnitude and direction of the imbalance of hydrostatic and colloid osmotic pressures across capillary walls) has to date received direct experimental confirmation only in nonmammalian species (3-5). That similar direct measurements have thus far not been performed in mammalian glomerular capillaries is due largely to the fact that glomeruli are rarely present as surface structures and are not therefore

This work was presented in part at the National Meeting of the American Federation for Clinical Research, Atlantic City, N. J., 1 May 1971.

*Received for publication 10 May 1971.*

accessible for direct study. Recently however, a strain of Wistar rats with glomeruli situated on the renal surface has been identified in the laboratory of Dr. Klaus Thureau of the University of Munich. Using these rats<sup>1</sup> we have undertaken in the present study to characterize the transcapillary forces governing the formation of glomerular ultrafiltrate in the mammalian kidney.

## METHODS

Studies were performed on seven normally hypoprenic rats (six adults and one young rat) which were allowed free access to food and water. They were anesthetized with Inactin (100 mg/kg) and prepared for micropuncture as described previously (6).

Pressure measurements were obtained in capillaries of 12 different glomeruli, using continuous recording servo-nulling micropipette transducer techniques (7-9). Micropipettes with outer tip diameters of 2-3  $\mu$  and containing 1.5 M NaCl were used. Penetration of Bowman's Space and entry into single glomerular capillaries was performed under stereomicroscopic control ( $\times 210$ ). Hydraulic output from the servo-system was channeled via a strain gauge to a recorder. Accuracy, frequency response, and stability features of this servo-system will be described in detail elsewhere (Brenner et al. submitted for publication). In addition to glomerular capillary hydrostatic pressures ( $\bar{P}_{GC}$ )<sup>2</sup>, we also recorded pressures in Bowman's capsule ( $P_{BS}$ ), and in separate adjacent proximal tubules ( $P_T$ ), efferent arterioles ( $P_{EA}$ ) and third order branch peritubular capillaries ( $P_C$ ) in each rat.

To obtain similarly direct estimates of mean glomerular capillary colloid osmotic pressure ( $\bar{\pi}_{GC}$ ), protein concentrations in femoral arterial and efferent arteriolar blood plasmas were measured as recently described (6). Colloid osmotic pressures (COP) were calculated using the expression of Landis and Pappenheimer (10).  $\bar{\pi}_{GC}$  then was taken as one-half the sum of the calculated COP at each site. For estimation of the ultrafiltration coefficient ( $K_f$ ) for glomerular capillaries, single

<sup>1</sup> The generous gift to us of a number of adult rats of this unique strain by Professor Thureau is gratefully acknowledged.

<sup>2</sup> *Abbreviations used in this paper:*  $\bar{AP}$ , mean arterial pressure; COP, colloid osmotic pressure;  $K_f$ , ultrafiltration coefficient;  $P_{BS}$ , Bowman's Space pressure;  $P_C$ , peritubular capillary pressure;  $P_{EA}$ , efferent arteriolar pressure;  $\bar{P}_{GC}$ , mean glomerular capillary hydrostatic pressure;  $P_T$ , proximal tubule pressure;  $P_{UF}$ , net glomerular ultrafiltration pressure;  $\bar{\pi}_{GC}$ , mean glomerular capillary colloid osmotic pressure;  $\pi_{TF}$ , tubule fluid colloid osmotic pressure; SNGFR, single nephron glomerular filtration rates.

TABLE I  
A Summary of the Measured Determinants of Glomerular Ultrafiltration in Seven Munich-Wistar Rats

Rat No.	Body wt (g)	Kidney wt (g)	Obs. No.	$\overline{AP}^a$ cm H <sub>2</sub> O	$P_T^b$ cm H <sub>2</sub> O	$P_{BS}^c$ cm H <sub>2</sub> O	$\overline{P}_{GC}^d$ cm H <sub>2</sub> O	[Protein] <sup>e</sup>		$\pi_{FA}^f$ cm H <sub>2</sub> O	$\pi_{EA}^f$ cm H <sub>2</sub> O	$\overline{\pi}_{GC}^g$ cm H <sub>2</sub> O	$P_{UF}^h$ cm H <sub>2</sub> O	SNGFR <sup>i</sup> nl/sec	$K_f^j$ nl/sec/ cm H <sub>2</sub> O/ glom
								F.A.	E.A.						
1	298	1.02	1 2	122	14.5	13	78	5.8	7.8	26.1	41.2	33.7	29.8	0.54 0.55	0.019
2	290	1.00	1 2 3	135 142	11 8	12 8 10	55 55	5.4	8.1	23.6	43.7	33.6	11.9	0.56 0.56	0.047
3	308	1.22	1 2	110	11	10	65	5.4	9.2	23.6	54.0	38.8	15.2	0.42 0.51	0.031
4	123	0.61	1 2 3 4 5	108 110 110 110 110	13	16 10 10 12.5	60 56 64 60 57.5	5.2	8.0	22.3	42.8	32.5	14.0	0.32 0.35	0.023
5	250	1.08	1 2 3 4	162 162 162 156		15 10 10		5.3	7.0	23.0	34.7	28.8	9.2	0.50 0.42	0.050
6	290	1.51	1 2	108	10	10	62	6.0	9.3	27.4	55.0	41.2	10.8	0.44 0.47	0.042
7	252	0.92	1 2	160	14	13	60	5.2	9.3	22.8	55.0	38.9	7.1	0.77 0.62	0.098
Mean ±1 SE					11.6 0.8	11.3 0.6	60.1 2.1	5.5 0.1	8.4 0.3	24.1 0.7	46.6 3.0	35.4 1.7	14.0 2.8	0.46 0.06	0.044 0.010

a, mean arterial pressure; b, proximal tubule pressure; c, Bowman's Space pressure; d, mean glomerular capillary hydrostatic pressure; e, protein concentration in femoral arterial and efferent arteriolar blood plasmas; f, colloid osmotic pressures, calculated for values shown at e, using the Landis-Pappenheimer equation (10); g, mean glomerular capillary colloid osmotic pressure, calculated as  $\frac{\pi_{FA} + \pi_{EA}}{2}$ ; h, net ultrafiltration pressure, calculated as  $\overline{P}_{GC} - P_T - \overline{\pi}_{GC}$ ; i, single nephron glomerular filtration rate; j, ultrafiltration coefficient for these glomerular capillaries.

nephron glomerular filtration rates (SNGFR) also were measured in these rats, using free-flow micropuncture techniques. Standard analytical methods were employed (6).

## RESULTS

Pressures were measured in single capillaries of 12 superficial glomeruli from 7 normal Munich-Wistar rats (Table I). Values for  $\overline{P}_{GC}$  ranged from 49 to 78 cm H<sub>2</sub>O, with 10 of 12 pressures being between 55–65 cm H<sub>2</sub>O. As shown in Fig. 1 the wave form profile of the glomerular capillary pressure pulse is very similar to that of the central aorta. This preservation of the aortic wave form in these small diameter vessels makes it likely that their

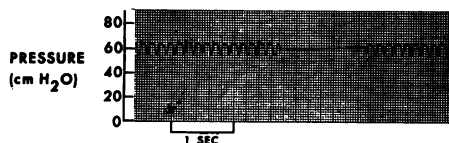


FIGURE 1 Characteristic pressure pattern in a surface glomerular capillary. Mean pressure is given by the horizontal line.

walls are relatively rigid. As shown in Fig. 2, the major changes in intrarenal vascular pressure occurred on either side of the glomerular capillary bed. An average fall in mean systemic arterial pressure ( $\overline{AP}$ ) of 51% occurred to the glomerular capillary. A second large pressure differential occurred between glomerular capillaries and surface efferent arterioles. Beyond the latter site the falls in pressure were small and gradual.

The relationship between the rate of formation of glomerular ultrafiltrate (GFR) and the responsible driving forces is given by the expression:

$$GFR = K_f(\overline{P}_{GC} - P_T - \overline{\pi}_{GC} + \pi_{TF}) \quad [1]$$

where  $K_f$  represents the ultrafiltration coefficient (ie, hydraulic conductivity per unit area  $\times$  glomerular capillary surface area),  $\overline{P}_{GC}$ ,  $P_T$ , and  $\overline{\pi}_{GC}$  are as defined above, and  $\pi_{TF}$  tubule fluid COP. Values for  $\pi_{TF}$  are very close to zero and may therefore be neglected.<sup>3</sup>

<sup>3</sup> Direct measurements of protein concentration in fluid from Bowman's Space of five glomeruli in four Munich-Wistar rats not otherwise studied yielded values below 200 mg/100 ml. The mean estimate of COP was 0.4 cm H<sub>2</sub>O.



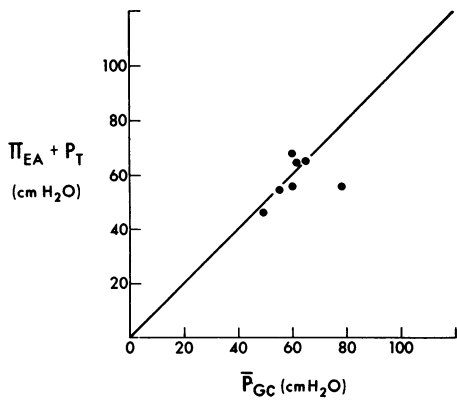


FIGURE 3 Comparison of values for  $\bar{P}_{GC}$  and  $(\pi_{EA} + P_T)$ . Filtration equilibrium is given by the line of identity. Symbols denote individual animals.

workers (17, 18), that COP rises as a linear function of glomerular capillary length. The more likely possibility, however, given the extreme leakiness of these glomerular capillaries to water, is that glomerular capillary plasma COP rises exponentially, approaching equilibrium conditions very near the beginning of the capillary segment. This is supported by the fact that even for extrarenal capillaries which have far lower  $K_f$  values that these glomerular capillaries  $40 \times 10^{-8}$  ml/sec per cm  $H_2O/cm^2$  net fluid movement follows an exponential profile (20). Given these conditions the bulk of the glomerular ultrafiltrate would be formed across a relatively small fraction of the estimated available glomerular capillary surface area. Thus although the remaining fraction of the total capillary surface area would likely contribute little to ultrafiltrate formation, this additional length of capillary is still represented in the total surface area estimate used in the calculation of  $K_f$ . It is for this reason that we have preferred to express our values for  $K_f$  in the present study in the manner given in Table I.

It is not yet possible to estimate precisely where along the length of the glomerular capillary plasma COP reaches a value which, when added to  $P_T$ , closely approximates  $\bar{P}_{GC}$  and thereby blunts further ultrafiltrate formation. That this equilibration does obtain however, is clearly demonstrated by the findings in the present study. Fig. 3 is an identity plot of the relationship between the measured value for  $\bar{P}_{GC}$  and the sum of  $\pi_{EA} + P_T$  for each of the seven rats studied. The finding of nearly identical values for these opposing pressures in six of seven rats (for all,  $\pi_{EA} + P_T/\bar{P}_{GC} = 0.97$ ) provides evidence, in accord with earlier predictions (21), that filtration equilibrium normally obtains before entry of glomerular capillary blood into the efferent arteriole.

## ACKNOWLEDGMENTS

We are grateful to Dr. Eugene M. Renkin and Dr. Joseph P. Gilmore for the opportunity to read unpublished material and to Dr. Curt A. Wiederhielm for rewarding discussions. Support for these studies was provided by the Veterans Administration (01/1073.1), National Institutes of Health (AM 13888) and American Hoechst Corp., Somerville, N. J.

## REFERENCES

- Ludwig, C. 1843. *In* Beitrage zur Lehre vom Mechanismus der Harnsekretion. Elwert'sche Universitäts- und Verlagsbuchhandlung, N. G., Marburg, Germany. 42.
- Starling, E. H. 1899. The glomerular functions of the kidney. *J. Physiol.* 24: 317.
- Hayman, J. M. Jr. 1927. Estimations of afferent arteriole and glomerular capillary pressures in frog kidney. *Amer. J. Physiol.* 79: 389.
- White, H. L. 1929. Observations on the nature of glomerular activity. *Amer. J. Physiol.* 90: 689.
- Wind, F. 1937. Versuche zur unmittelbaren Bestimmung des Flüssigkeitsaustritts aus den Blutkapillaren des Mesenterium und des Nierenglomerulus bei Kaltbluter. I. Mitteilung. *Arch. Exp. Pathol. Pharmacol.* 186: 161.
- Brenner, B. M., and J. H. Galla. 1971. Influence of post-glomerular hematocrit and protein concentration on rat nephron fluid transfer. *Amer. J. Physiol.* 220: 148.
- Wiederhielm, C. A., J. W. Woodbury, S. Kirk, and R. F. Rushmer. 1964. Pulsatile pressures in the microcirculation of frog's mesentery. *Amer. J. Physiol.* 207: 173.
- Falchuk, K. H., B. M. Brenner, M. Tadokoro, and R. W. Berliner. 1971. Oncotic and hydrostatic pressures in peritubular capillaries and fluid reabsorption by the proximal tubule. *Amer. J. Physiol.* 220: 1427.
- Brenner, B. M., T. M. Daugharty, and J. L. Troy. 1971. Continuous pressure measurements in renal tubules and postglomerular vessels recorded simultaneously using dual servo-controlled transducers. *Clin. Res.* 19: 526. (Abstr.)
- Landis, E. M., and J. R. Pappenheimer. 1963. Exchange of substances through the capillary walls. *Hand. Physiol.* 2: 961.
- Gertz, K. H., J. A. Mangos, G. Braun, and H. D. Pagel. 1966. Pressure in the glomerular capillaries of the rat kidney and its relation to arterial blood pressure. *Pfluegers Arch.* 288: 369.
- Koch, K. M., T. Dume, H. H. Krause, and B. Ochwadt. 1967. Intratubular Druck, glomerularer Capillardruck, und Glomerulumfiltrat während Mannit—Diurse. *Pfluegers Arch.* 295: 72.
- Gottschalk, C. W., and M. Mylle. 1956. Micropuncture study of pressures in proximal tubules and peritubular capillaries of the rat kidney and their relation to ureteral and renal venous pressures. *Amer. J. Physiol.* 185: 430.
- Selkurt, E. E., P. Deetjen, and H. Brechtelsbauer. 1965. Tubular pressure gradients and filtration dynamics during urinary stop flow in the rat. *Pfluegers Arch.* 286: 19.
- Hayslett, J. P., D. T. Domoto, M. Kashgarian, and F. H. Epstein. 1970. Role of physical factors in the natriuresis induced by acetylcholine. *Amer. J. Physiol.* 218: 880.
- Andreucci, V. E., R. C. Blantz, J. Herrera-Acosta, F. C. Rector, Jr., and D. W. Seldin. 1971. Effect of partial ureteral obstruction and acute volume expansion on glomerular pressure, effective filtration pressure, and single neph-

- ron glomerular filtration rate in the rat. *Clin. Res.* 19: 524. (Abstr.)
17. Pappenheimer, J. R., E. M. Renkin, and L. M. Borrero. 1951. Filtration, diffusion and molecular sieving through peripheral capillary membranes. *Amer. J. Physiol.* 167: 13.
  18. Renkin, E. M., and J. P. Gilmore. Glomerular filtration. *Handb. Physiol.* In Press.
  19. Kirkman, H., and R. E. Stowell. 1942. Renal filtration surface in the albino rat. *Anat. Rec.* 82: 373.
  20. Lee, J. S., L. H. Smaje, and B. W. Zweifach. 1971. Fluid movement in occluded single capillaries of rabbit omentum. *Circ. Res.* 28: 358.
  21. Smith, H. W., H. Chasis, W. Goldring, and H. H. Ranges. 1940. Glomerular dynamics in the normal human kidney. *J. Clin. Invest.* 19: 751.