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### Inhibition of Renal Tubular Sodium Reabsorption by Hypernatremia \*

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In a recent study (1), we confirmed the observation of Bresler (2) and Toussaint and Vereerstraeten (3) that tubular sodium reabsorption  $(T_{Na})$  in dogs increases following plasma sodium  $(P_{Na})$  elevation. However, when the increase in filtered sodium  $(F_{Na})$  that usually accompanied an elevated P<sub>Na</sub> was eliminated by reducing glomerular filtration rate (GFR) with an aortic clamp, hypernatremia appeared to inhibit rather than stimulate  $T_{Na}$ . Similar results have been obtained by Blythe and Welt (4), who found that sodium excretion during the infusion of hypertonic saline was above control when filtered sodium was reduced below control by inflating a balloon in the inferior vena cava. The micropuncture studies of Giebisch, Klose, and Windhager (5) in rats have also demonstrated that T<sub>Na</sub> is decreased by hypertonic saline infusions.

In all these studies the depression of  $T_{Na}$  was considered to be a consequence of hypernatremia per se. In each case, hypernatremia was induced by loading with hypertonic saline. The infusion of similar volumes of isotonic saline has been shown to inhibit  $T_{Na}$  to a comparable degree (6–9). Therefore, the studies cited do not clearly differentiate between volume expansion and hypernatremia as the cause of the decreased  $T_{Na}$ , nor, indeed, do they clearly establish that hyper-

† Postdoctoral fellow (5-F2-HE 16050) of the National Heart Institute.

‡Established Investigator of the American Heart Association. Address requests for reprints to Dr. Norman G. Levinsky, 15 Stoughton Street, Boston, Mass. 02118. natremia itself has any direct effect on  $T_{Na}$ . In the present experiments, we have attempted to isolate the effects of hypernatremia on  $T_{Na}$  by unilaterally elevating renal arterial  $P_{Na}$ , using the opposite kidney as a simultaneous control. Since any effect of volume expansion is presumably equal in the two kidneys, changes in  $T_{Na}$  in the experimental kidney relative to the control should be a function of hypernatremia per se. The results demonstrate that hypernatremia specifically inhibits  $T_{Na}$  by means of a direct intrarenal action.

#### Methods

Female mongrel dogs were anesthetized with pentobarbital, 30 mg per kg intravenously, and small supplementary doses were given as necessary to maintain light anesthesia. A sustaining infusion containing appropriate concentrations of inulin and p-aminohippurate (PAH) in 0.82% NaCl was given throughout each experiment at 8.0 ml per minute. The dogs were last fed about 20 hours before an experiment; water was usually allowed ad libitum. In some studies, water was withdrawn 18 hours before the experiment, and the dogs were given 5 U vasopressin tannate in oil intramuscularly 18 hours and again  $\frac{1}{2}$  hour before the experiment; aqueous Pitressin, 50 mU per kg per hour, was added to the sustaining infusion. Urine was collected separately from each kidney through polyethylene tubing that had been inserted into the ureters through a lower mid-line abdominal incision.

The experimental kidney was supplied with femoral arterial blood through a modification of the surgical procedure used by Goodman and Fuisz (10). The ipsilateral femoral artery was cannulated and connected to  $\frac{1}{4}$ -inch (i.d.) polyethylene tubing. Through a flank incision, the proximal segment of the artery to the experimental kidney was carefully dissected free from the surrounding connective tissue. After the intravenous administration of 50 mg of heparin, the renal artery was ligated at its origin and cannulated. Care was taken that the cannula was aligned properly with the direction of the artery and that the tip was proximal to the bifurcation. The renal arterial cannula was then connected to

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the tubing in the femoral artery, and circulation to the experimental kidney was thus completed. The period of renal ischemia during the creation of the shunt varied from 1 to 4 minutes.

Infusions into the shunt were accomplished by direct puncture of rubber tubing near its femoral end with the needle directed against the flow of blood to enhance mixing. Blood samples for the experimental kidney were obtained via a sidearm close to the renal end of the shunt. The length of the tubing between the site of infusion and the point at which blood samples were drawn was approximately 40 cm. Blood samples for the control kidney were obtained from a retention needle in the jugular vein.

After the collection of three to five control periods, 20% saline was infused into the shunt at 0.5 to 3.0 ml per minute. After 5 to 10 minutes for equilibration, the "hypernatremic" periods were collected. In some experiments the degree of hypernatremia on the experimental side was then altered by changing the rate of the hypertonic infusion, and additional groups of hypernatremic periods were obtained. In some cases hypernatremic periods were collected with the shunt partially occluded by a clamp, so that clearance measurements could be made with sodium excretion from the experimental kidney equal to or only slightly greater than the value on the control side. In some experiments, additional control periods, during which no saline was infused into the shunt, and "isotonic" periods, during which 0.82% saline was infused into the shunt, were obtained between or after the groups of hypernatremic periods.

Inulin clearance  $(C_{In})$  was used as a measure of GFR, and  $F_{Na}$  was calculated as equal to  $P_{Na} \cdot C_{In}$  without a Donnan correction. The analytical methods used in this study have been listed elsewhere (6). Except where individual experiments are shown, each clearance value in this paper is the mean of three to seven consecutive clearance periods. Experiments were discarded if the filtration rates of the two kidneys differed by more than 10% in the initial control periods.

TABLE IProtocol of a representative experiment\*

Time   Exp.   Cont.   Exp. <t< th=""><th></th><th colspan="2">Сран</th><th colspan="2">Cīn</th><th colspan="2">P<sub>Na</sub></th><th colspan="2">F<sub>Na</sub></th><th colspan="2">U<sub>Na</sub>V</th><th colspan="2">T<sub>Na</sub></th></t<>		Сран		Cīn		P <sub>Na</sub>		F <sub>Na</sub>		U <sub>Na</sub> V		T <sub>Na</sub>	
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	Time	Exp.	Cont.	Exp.	Cont.	Exp.	Cont.	Exp.	Cont.	Exp.	Cont.	Exp.	Cont.
0 Primes given: 400 mg PAH, 600 mg inulin; infusion I started: 0.82% saline at 12 ml per minute   40 Ureters catheterized   110 Right femoral artery cannulated   120 Infusion I stopped and infusion II started: inulin 2.25 mg per ml, PAH 0.75 mg per ml, and aque: us Pitressin 2 mU per ml in .82% saline at 8 ml per minute   130 Right renal artery ligated   132 Right renal artery ligated   134 Right femoral-renal shunt completed   191-196 59 62 30.0 30.8 150 4.50 0.368 0.355 4.13 4.25   196-201 57 60 28.7 29.6 152 4.36 4.50 0.364 0.338 4.00 4.16   201-207 60 59 27.7 27.5 151 4.18 4.15 0.368 0.321 3.81 3.83   210 Infusion III started: 20% saline into shunt at 0.9 ml per minute 221-226 49 59 25.2 30.1 171 152 4.31 4.58 0.699 0.410 3.61 4.17   2206-231 57 57 24 0.71 163 <td< td=""><td></td><td colspan="2">ml/min</td><td colspan="2">ml/min</td><td colspan="2">mEq/L</td><td colspan="2">mEq/min</td><td colspan="2">mEq/min</td><td colspan="2">mEq/min</td></td<>		ml/min		ml/min		mEq/L		mEq/min		mEq/min		mEq/min	
40 Ureters catheterized   110 Right femoral artery cannulated   120 Infusion I stopped and infusion II started : inulin 2.25 mg per ml, PAH 0.75 mg per ml, and aque: us	0	Primes give		n: 400 mg PAH,		600 mg inulin; inf		nfusion I started :		0.82% saline at 12		ml per minute	
110 Right femoral artery cannulated   120 Infusion I stopped and infusion II started : inulin 2.25 mg per ml, PAH 0.75 mg per ml, and aque: us Pitressin 2 mU per ml in .82% saline at 8 ml per minute   130 Right renal artery ligated   132 Right femoral-renal shunt completed   191-196 59 62 30.0 30.8 150 4.50 4.62 0.368 0.355 4.13 4.25   196-201 57 60 28.7 29.6 152 4.36 4.50 0.364 0.338 4.00 4.16   201-207 60 59 27.7 27.5 151 4.18 4.15 0.368 0.321 3.81 3.83   210 Infusion III started: 20% saline into shunt at 0.9 ml per minute 221-226 49 59 25.2 30.1 171 152 4.31 4.58 0.699 0.410 3.61 4.17   226-231 57 57 24 0.27 163 153 3.91 4.15 0.711 0.414 3.20 3.74	40	Ureters cath		neterized								-	
120 Infusion I stopped and infusion II started : inulin 2.25 mg per ml, PAH 0.75 mg per ml, and aque: us Pitressin 2 mU per ml in .82% saline at 8 ml per minute   130 Right renal artery ligated   132 Right femoral-renal shunt completed   191-196 59 62 30.0 30.8 150 4.50 4.62 0.368 0.355 4.13 4.25   196-201 57 60 28.7 29.6 152 4.36 4.50 0.364 0.338 4.00 4.16   201-207 60 59 27.7 27.5 151 4.18 4.15 0.368 0.321 3.81 3.83   210 Infusion III started : 20% saline into shunt at 0.9 ml per minute 221-226 49 59 25.2 30.1 171 152 4.31 4.58 0.699 0.410 3.61 4.17   226-231 57 57 24 0.27 1.63 3.91 4.15 0.711 0.414 3.20 3.74	110	Right femora		al artery cannul		ated							
Pitressin 2 mU per ml in .82% saline at 8 ml per minute     130   Right renal artery ligated     132   Right femoral-renal shunt completed     191-196   59   62   30.0   30.8   150   4.50   4.62   0.368   0.355   4.13   4.25     196-201   57   60   28.7   29.6   152   4.36   4.50   0.364   0.338   4.00   4.16     201-207   60   59   27.7   27.5   151   4.18   4.15   0.368   0.321   3.81   3.83     210   Infusion III started: 20% saline into shunt at 0.9 ml per minute   221-226   49   59   25.2   30.1   171   152   4.31   4.58   0.699   0.410   3.61   4.17     226-231   57   57   24   0   77   163   3.91   4.15   0.711   0.414   3.20   3.74	120	Infusion I stopped and infusion II started : inulin 2.25 mg per ml, PAH 0.75 mg per ml, and aque, us											
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$		Pitressin 2 mU per ml in .82% saline at 8 ml per minute											
132Right femoral-renal shunt completed $191-196$ 596230.030.81504.504.620.3680.3554.134.25 $196-201$ 576028.729.61524.364.500.3640.3384.004.16 $201-207$ 605927.727.51514.184.150.3680.3213.813.83 $210$ Infusion III started: 20% saline into shunt at 0.9 ml per minute $221-226$ 495925.230.11711524.314.580.6990.4103.614.17 $226-231$ 57572402711631533.914.150.7110.4143.203.74	130	Right renal artery ligated											
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	132	Righ	t femo	ral-renal	shunt co	ompletee	đ						
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	191–196	59	62	30.0	30.8	•	150	4.50	4.62	0.368	0.355	4.13	4.25
201-207   60   59   27.7   27.5   151   4.18   4.15   0.368   0.321   3.81   3.83     210   Infusion III started: 20% saline into shunt at 0.9 ml per minute     221-226   49   59   25.2   30.1   171   152   4.31   4.58   0.699   0.410   3.61   4.17     226-231   57   57   24   0   27   1   163   153   3.91   4.15   0.711   0.414   3.20   3.74	196–201	57	60	28.7	29.6		152	4.36	4.50	0.364	0.338	4.00	4.16
210   Infusion III started: 20% saline into shunt at 0.9 ml per minute     221-226   49   59   25.2   30.1   171   152   4.31   4.58   0.699   0.410   3.61   4.17     226-231   57   57   24.0   27.1   163   153   3.91   4.15   0.711   0.414   3.20   3.74	201–207	60	59	27.7	27.5		151	4.18	4.15	0.368	0.321	3.81	3.83
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	210	210 Infusion III started : 20% saline into shunt at 0.9 ml per minute											
226-231 57 57 24.0 27.1 163 153 3.01 4.15 0.711 0.414 3.20 3.74	221-226	49	59	25.2	30.1	171	152	4.31	4.58	0.699	0.410	3.61	4.17
	226-231	57	57	24.0	27.1	163	153	3.91	4.15	0.711	0.414	3.20	3.74
231-236 62 59 24.4 28.8 167 154 4.07 4.44 0.722 0.442 3.35 4.00	231-236	62	59	24.4	28.8	167	154	4.07	4.44	0.722	0.442	3.35	4.00
243 Shunt constricted with clamp	243	Shur	nt const	tricted w	ith clam	D							
248-253 53 53 24.1 28.3 180 157 4.34 4.44 0.652 0.499 3.69 3.94	248-253	53	53	24.1	28.3	180	157	4.34	4.44	0.652	0.499	3.69	3.94
253-258 53 59 23.6 30.4 183 158 4.32 4.80 0.668 0.537 3.65 4.26	253-258	53	59	23.6	30.4	183	158	4.32	4.80	0.668	0.537	3.65	4.26
258-263 59 25.4 29.1 182 160 4.62 4.66 0.672 0.537 3.95 4.12	258-263		59	25.4	29.1	182	160	4.62	4.66	0.672	0.537	3.95	4.12
263-268 56 59 23.1 27.9 183 161 4.23 4.49 0.674 0.537 3.56 3.95	263-268	56	59	23.1	27.9	183	161	4.23	4.49	0.674	0.537	3.56	3.95
269 Clamp around shunt released	269	Clan	ip arou	nd shun	t released	1							
270 Infusion III stopped and infusion IV started: 0.82% NaCl into shunt at 2.9 ml per minute	270	Infus	sion III	stopped	l and infi	usion IV	/ started	: 0.82%	NaCl int	o shunt at	t 2.9 ml pe	er minute	2
288-294 54 52 24.4 25.1 158 157 3.86 3.94 0.214 0.195 3.65 3.74	288-294	54	52	24.4	25.1	158	157	3.86	3.94	0.214	0.195	3.65	3.74
294-301 48 52 23.3 26.7 157 159 3.66 4.25 0.186 0.190 3.47 4.06	294-301	48	52	23.3	26.7	157	159	3.66	4.25	0.186	0.190	3.47	4.06
301-309 47 48 22.9 24.3 154 159 3.53 3.86 0.172 0.171 3.36 3.69	301-309	47	48	22.9	24.3	154	159	3.53	3.86	0.172	0.171	3.36	3.69
309-316 46 48 23.2 22.8 154 159 3.57 3.63 0.160 0.155 3.41 3.47	309-316	46	48	23.2	22.8	154	159	3.57	3.63	0.160	0.155	3.41	3.47
325 Infusion IV stopped; infusion III restarted into the shunt at 1.5 ml per minute, shunt constricted	325	Infus	sion IV	stopped	; infusio	n III res	started in	ito the s	hunt at 1	.5 ml per	minute. sl	hunt con	stricted
with clamp		with	clamp	••						•	,		
337-342 33 48 15.8 24.9 250 165 3.95 4.11 0.761 0.345 3.19 3.76	337-342	33	48	15.8	24.9	250	165	3.95	4.11	0.761	0.345	3.19	3.76
342-348 35 50 17.4 25.6 249 169 4.33 4.32 0.850 0.422 3.48 3.90	342-348	35	50	17.4	25.6	249	169	4.33	4.32	0.850	0.422	3.48	3.90
354-359 38 41 17.9 22.4 240 174 4.30 3.90 1.080 0.543 3.22 3.36	354-359	38	41	17.9	22.4	240	174	4.30	3.90	1.080	0.543	3.22	3.36
359-364 35 43 18.1 24.8 248 177 4.49 4.39 1.100 0.603 3.39 3.79	359-364	35	43	18.1	24.8	248	177	4.49	4.39	1.100	0.603	3.39	3.79
365 Clamp around shunt released	365	Clan	ip arou	nd shun	t released	1 L							
366 Infusion III stopped : infusion IV restarted into shunt at 2.9 ml per minute	366	Infus	sion III	stopped	1: infusio	n IV re	started i	nto shur	nt at 2.9 r	nl per mi	nute		
380-385 39 41 17.4 22.3 173 175 3.01 3.90 0.263 0.325 2.75 3.57	380385	39	41	17.4	22.3	173	175	3.01	3.90	0.263	0.325	2.75	3.57
385-390 41 41 17.9 21.6 170 172 3.04 3.72 0.234 0.298 2.81 3.42	385-390	41	41	17.9	21.6	170	172	3.04	3.72	0.234	0.298	2.81	3.42
390-398 31 41 16.3 21.9 171 173 2.79 3.79 0.167 0.239 2.62 3.55	390–398	31	41	16.3	21.9	171	173	2.79	3.79	0.167	0.239	2.62	3.55

\* Abbreviations: Exp. = experimental kidney; cont. = control kidney;  $C_{PAH}$  = clearance of *p*-aminohippurate;  $C_{In}$  = clearance of inulin;  $P_{Na}$  = plasma sodium;  $F_{Na}$  = filtered sodium;  $U_{Na}V$  = excreted sodium;  $T_{Na}$  = tubular sodium reabsorption.

Fre				Cīn		P <sub>Na</sub>		F <sub>Ns</sub>		UnaV	
, r	no.	Collection periods	Infusion*	Exp.	Cont.	Exp.	Cont.	Exp.	Cont.	Exp.	Cont.
			ml/min	ml/min		mEq/L		mEq/min		mEq/min	
	1.	Control	0	25	28		152	3.78	4.18	0.109	0.114
		Hypernatremic	1.2	25	31	189	165	4.60	5.13	0.563	0.525
	2.	Control	0	41	37		150	6.18	5.63	0.130	0.125
		Hypernatremic <sup>†</sup>	1.2	23	41	221	176	5.08	7.16	0.245	0.205
	3.	Control	0	31	34		144	4.46	4.83	0.178	0.221
		Hypernatremic	0.5	25	32	166	148	4.20	4.70	0.245	0.226
	4.	Control	0	25	25		152	3.85	3.82	0.157	0.143
		Hypernatremic <sup>†</sup>		17	25	213	170	3.59	4.16	0.399	0.240
		Hypernatremic <sup>†</sup>		14	24	238	184	3.36	4.31	0.585	0.576
	5.	Control	0	38	42		160	6.02	6.63	0.201	0.243
		Isotonic	3.0	39	42	164	158	6.39	6.62	0.263	0.271
		Isotonic	3.0	36	43	163	158	5.94	6.83	0.243	0.416
		Hypernatremic <sup>†</sup>	3.0	25	41	238	177	5.95	7.31	1.38	0.987
	6.	Control	0	40	43		152	6.13	6.48	0.242	0.272
		Hypernatremic	0.5	37	43	160	153	5.94	6.55	0.465	0.465
		Isotonic	2.9	43	46	151	151	6.45	6.91	0.453	0.475
		Hypernatremic	0.6	37	41	184	155	6.73	6.33	1.19	0.600
		Control	0	42	47		157	6.58	7.37	0.460	0.494
	7.	Control	0	28	31		147	4.15	4.49	0.261	0.249
		Hypernatremic <sup>†</sup>	0.7	27	30	173	156	4.60	4.62	0.521	0.384
		Isotonic	2.8	27	30	153	153	4.21	4.60	0.239	0.241
		Hypernatremic <sup>†</sup>	0.9	24	30	193	164	4.57	4.88	0.699	0.490
		Isotonic	2.8	25	31	161	162	4.05	5.02	0.284	0.335
		Control	0	28	31		163	4.49	4.96	0.221	0.253
		Hypernatremic	1.2	24	32	196	169	4.71	5.44	0.745	0.399
		Hypernatremic	1.7	23	29	220	174	5.10	5.01	1.511	0.577
		Control	Ō	16	20		178	2.77	3.54	0.246	0.289
	8.	Control	Ō	29	29		151	4.35	4.42	0.367	0.338
	•••	Hypernatremic	0.9	25	29	167	153	4.10	4.39	0.710	0.422
		Hypernatremict	0.9	24	29	182	159	4.38	4.60	0.667	0.527
		Isotonic	2.9	$\overline{24}$	25	156	159	3.66	3.92	0.183	0.178
		Hypernatremict	1.5	17	24	247	171	4.27	4.18	0.948	0.478
		Isotonic		17	$\overline{22}$	171	173	2.95	3.81	0.221	0.287

TABLE II Summary of all experiments\*

\* Abbreviations as in Table I. Infusion = rate of infusion into shunt. Twenty per cent saline was infused during hypernatremic periods, 0.82% saline during isotonic periods. Each value in this Table is the mean of three to seven clearance periods.

† Shunt constricted with clamp during collection of this group of periods.

#### Results

The detailed protocol of a single experiment is presented in Table I. In the control periods filtered sodium was equal in the two kidneys; sodium excretion was slightly higher on the experimental side. Hypertonic saline (20%) was then infused into the right femoral-renal arterial shunt at about 0.9 ml per minute, elevating  $P_{Na}$  in the shunt blood 10 to 20 mEq per L above systemic P<sub>Na</sub>. After one set of hypernatremic periods at the spontaneous rates of sodium excretion had been obtained, excretion from the experimental kidney was brought closer to the rate of the control kidney by partial occlusion of the shunt, and a second set of hypernatremic periods was obtained. Despite modest but definite reduction of  $F_{Na}$  in the experimental kidney during both of these collections, sodium excretion from the experimental kidney exceeded that from the control by 0.130 to 0.300 mEq per minute. The clamp was released and the 20% saline infusion stopped. A 0.82% concentration of saline was then infused into the shunt at 2.9 ml per minute, a rate well in excess of that at which the 20% saline had been given. In the isotonic periods, which were next collected, sodium excretion was approximately equal in the two kidneys, although a modest reduction in  $F_{Na}$ of the experimental kidney was still present. The isotonic infusion was then stopped, 20% NaCl was restarted into the shunt at nearly twice the earlier rate, and the shunt was again partially occluded. The reduction in GFR of the experimental kidney due to clamping of the shunt compensated almost exactly for the elevation of 65 to 85 mEq per L in  $P_{Na}$  in the shunt blood, so that filtered sodium was approximately equal in the two kidneys. Mean sodium excretion from the experimental kidney now exceeded mean excretion from the control kidney by 0.475 mEq per minute. Finally the clamp was released, and isotonic saline was infused into the shunt in place of hypertonic. The GFR of the experimental kidney did not quite return to control in the ensuing isotonic periods;  $F_{Na}$  and sodium excretion were both lower on the experimental side. Thus, during all three sets of hypernatremic periods, sodium excretion from the experimental kidney was significantly greater than that from the control kidney, despite equal or decreased  $F_{Na}$  in the experimental kidney. This increase in sodium excretion did not occur when a larger volume of isotonic saline was infused into the shunt.

The results of all eight experiments are summarized in Table II. In seven studies, sodium excretion from the experimental kidney was elevated during the infusion of hypertonic saline, despite a lower  $F_{Na}$ . In the remaining experiment (no. 6), sodium excretion in the first set of hypernatremic periods was equal when  $F_{Na}$  was 0.6 mEq per minute lower in the experimental kidney; in the second group of hypernatremic periods, experimental sodium excretion exceeded control excretion by 0.59 mEq per minute when filtered sodium was only 0.4 mEq per minute greater on that side. These results are illustrated in Figure 1. Each point represents the mean difference, experimental minus control kidney, in filtered and excreted sodium during one group of periods. Eleven control, seven isotonic, and fifteen hypernatremic periods are plotted. In the control periods, during which there was no infusion into the shunt,  $F_{Na}$  on the experimental side was higher than on the control side in one, the same in two, and 0.3 to 2.1 mEq per minute lower in eight collections. Differences in sodium excretion between the kidneys were slight, varying from + 0.05 to - 0.05 mEq per minute; on the whole, differences in excretion paralleled differences in F<sub>Na</sub>, so that mean excretion was slightly less on the experimental side. The results of the isotonic periods, during which 0.82% saline was infused into the shunt, were similar to those obtained during control collections. Sodium excretion from the experimental kidney was generally slightly lower than that from the control kidney; filtered sodium was 0.2 to 1.0 mEq per minute lower on the experimental side. In nine of fifteen hypernatremic collections, sodium excretion from the experimental kidney was 0.04 to 0.34 mEq per



FIG. 1. DIFFERENCE IN SODIUM EXCRETION BETWEEN THE KIDNEYS PLOTTED AGAINST THE CORRESPONDING DIFFERENCE IN FILTERED SODIUM. Each point represents the mean of three or more periods, taken without infusion (control) or during infusion of 0.82% saline (isotonic) or 20% saline (hypernatremic) into the shunt supplying blood to the experimental kidney.



FIG. 2. COMPARISON OF FRACTIONAL SODIUM REAB-SORPTION ( $T_{Nn}$ ) IN THE TWO KIDNEYS, WITHOUT INFU-SION (CONTROL) OR DURING INFUSION OF 0.82% SALINE (ISOTONIC) OR 20% SALINE INTO THE SHUNT SUPPLYING BLOOD TO THE EXPERIMENTAL KIDNEY. Relative filtered sodium ( $F_{Nn}$ ) in the two kidneys is shown on the abscissa. Each point is the mean of three or more periods.

minute greater than from the control despite a  $F_{Na}$  0.02 to 2.1 mEq per minute lower in the hypernatremic kidney. In three hypernatremic collections, the experimental kidney excreted sodium at the same rate as the control side in spite of a  $F_{Na}$  0.5 to 1.0 mEq per minute less than the control value. In the three remaining hypernatremic collections, both filtered and excreted sodium were greater on the experimental side, but the difference in sodium excretion was 0.2 to 0.8 mEq per minute more than the difference in  $F_{Na}$ . The mean differences in  $F_{Na}$  between the kidneys (experimental - control) in the control, isotonic, and hypernatremic periods, respectively, were -0.235, -0.580, and -0.506 mEq per minute. These numbers are not significantly different from one another. The mean differences in excretion were -0.015, -0.045, and +0.251 mEq per minute in the control, isotonic, and hypernatremic periods, respectively. The hypernatremic value differs statistically from each of the other two (p < 0.01 by t test); the control and isotonic values do not differ from each other.

The effect of hypernatremia on  $T_{Na}$  is illustrated in Figure 2. Fractional reabsorption of sodium in the experimental kidney, divided by the value for the control kidney  $[(exp. T_{Na}/F_{Na})/(cont.$  $T_{Na}/F_{Na}$ ], is shown on the ordinate for the control, isotonic, and hypernatremic collections of all eight experiments. During the control and isotonic collections, fractional reabsorption was approximately equal in the two kidneys; the mean ratios were 0.99 and 1.0, respectively. As shown in the third column, during the hypernatremic periods the mean reabsorption ratio fell to 0.93, a value significantly different (p < 0.01) from both the control and the isotonic ratios. The decrease in fractional T<sub>Na</sub> in the experimental kidney during hypernatremia is not due to differences in relative  $F_{Na}$  of the two kidneys in the hypernatremic as compared to the other periods. Inspection of the abscissa of Figure 2 shows that the ratio of  $F_{Na}$  in the kidneys ( $F_{Na} \exp./cont.$ ) was similar during the control (0.94), isotonic (0.89), and hypernatremic (0.91) collections, and this was confirmed by statistical analysis.

#### Discussion

These studies were designed to isolate the effect of hypernatremia on  $T_{Na}$ . It was found that elevation of  $P_{Na}$  depresses  $T_{Na}$ , so that Na excretion is increased approximately 0.3 mEq perminute in a hypernatremic kidney as compared to a control at the same  $F_{Na}$  (Figure 1). The decrease in  $T_{Na}$  is equivalent to approximately 7% of  $F_{Na}$  (Figure 2). The depression of  $T_{Na}$  in the studies of Blythe and Welt (4) in dogs and of Giebisch, Klose, and Windhager (5) in rats is roughly comparable to that found in the present study. In our previous experiments (1), there was a smaller effect of hypertonic loading (sodium excretion from both kidneys was increased 0.18 mEq per minute at a given  $F_{Na}$ ).

The experimental kidney functioned normally, and there is no reason to believe that the shunting technique introduced an artifact that depressed  $T_{Na}$ . During control and isotonic collections  $T_{Na}$ in the two kidneys was quite comparable (Figure 2). Since control, isotonic, and hypernatremic periods were intermixed throughout most experiments, gradual deterioration of the experimental kidney cannot account for the decrease in  $T_{Na}$ during the hypernatremic collections. The results were comparable in the experiments in which GFR fell spontaneously and those in which it was necessary to clamp the shunt to reduce GFR during the hypernatremic periods. Hence, the clamping procedure itself does not depress fractional  $T_{Na}$ . During control and isotonic periods, sodium excretion from the experimental kidney was decreased when GFR was lower on the experimental side, demonstrating that reduction of GFR per se does not account for the increased excretion during hypernatremia (Figure 1).

It is assumed that all the variables (other than hypernatremia) that undoubtedly changed during the course of these experiments affected  $T_{Na}$  in both kidneys equally. In particular, any depression of T<sub>Na</sub> induced by volume loading (6-9) presumably was equal in the two kidneys. Hence, the depression of  $T_{Na}$  in the experimental kidney during hypernatremia represents a specific effect of the elevated P<sub>Na</sub>. The decrease in T<sub>Na</sub> during hypertonic saline infusion noted in previous studies (1, 4, 5) may represent the combined effects of volume loading and hypernatremia, each of which independently depressed T<sub>Na</sub>. Alternatively, the data are consistent with the possibility that loading with hypertonic saline does not induce a "volume loading" effect on  $T_{Na}$  comparable to that induced by isotonic loading. The micropuncture studies of Giebisch and his colleagues (5) demonstrate that hypertonic saline infusion depresses distal sodium reabsorption but does not decrease proximal or loop of Henle sodium transfer, provided sodium excretion is less than 12% of  $F_{Na}$ . The micropuncture experiments of Cortney, Mylle, and Gottschalk (11), however, indicate that isotonic saline loading decreases fractional sodium reabsorption in the proximal tubule. It may be, therefore, that the volume loading effect is proximal whereas the effect of hypernatremia per se is exerted on distal tubular sodium transfer. The depression of proximal  $T_{Na}$  noted by Giebisch and co-workers (5) when sodium excretion was in excess of 12% might represent a volume loading effect added to the action of hypernatremia itself in depressing distal T<sub>Na</sub>. It should be noted that all these micropuncture data were obtained in rats and may not be applicable in detail to the interpretation of our clearance experiments in dogs.

Our experiments show that T<sub>Na</sub> can be de-

pressed unilaterally by hypernatremia. This indicates that hypernatremia affects sodium transfer by a local action within the kidney, not indirectly via a systemic factor such as a hormone. Giebisch and co-workers (5) suggested that the decrease in distal sodium reabsorption in their experiments during hypernatremia was probably caused by a reduction in circulating mineralocorticoids. Our data indicate that this explanation cannot be correct, as least in the dog.

One possible mechanism, suggested by Blythe and Welt (4), is that back diffusion of sodium is enhanced because a steeper concentration gradient for passive diffusion of sodium from the interstitium into the tubular lumen exists during hypernatremia. The increased back diffusion might be sufficient to significantly diminish net transfer of sodium from lumen to interstitium, if the active transfer capacity in that direction were limited. The data of Giebisch and associates (5) demonstrating decreased net transfer of sodium in the distal tubule during hypernatremia would be compatible with this view, since it is known that sodium reabsorptive capacity in this segment is limited (12). It has recently been demonstrated that the sodium permeability of the mucosal surface of toad bladder (13) and the corresponding outer surface of frog skin (14) decreases as the sodium concentration in the bathing medium is increased. A similar effect of increased tubular fluid sodium concentration on the permeability of the luminal surface of the distal tubular epithelium to sodium might limit sodium reabsorption and account for the effect of hypernatremia on  $T_{Na}$ .

Thurau (15) has suggested that angiotensin release is stimulated by a high concentration of sodium in the tubular fluid passing the early distal segment in which the macula densa is located. Micropuncture data in rats (5) indicate that during hypertonic saline loading sodium concentration in early distal tubular fluid is probably in-(Tubular fluid/plasma sodium ratios creased. are unchanged from those in noninfused rats, but plasma sodium is higher in the loaded rats.) Several lines of evidence (16-18) suggest that angiotensin may directly inhibit T<sub>Na</sub>. Thus it is possible that the depression of T<sub>Na</sub> by hypernatremia is mediated by intrarenal release and action of angiotensin. The amount released would be small enough so that no significant concentration would reach the contralateral kidney through the systemic blood. No direct evidence for this mechanism was obtained in our experiments. It is interesting, however, that in each of the six hypernatremic periods in which the shunt was not clamped, GFR in the experimental kidney relative to the control fell as compared to the preceding control or isotonic period. The magnitude of this fall in these six periods was 3, 4, 3, 1, 5, and 4 ml per minute (Table I). It is conceivable that this fall in GFR due to hypernatremia per se is the result of intrarenal arteriolar vasoconstriction induced by angiotensin. Obviously, it may also represent a direct effect of hypernatremia on arterioles. Many alternative mechanisms can be proposed to account for the decrease in  $T_{Na}$  and GFR; final conclusions will depend on further data.

#### Summary

1. To study the effects of hypernatremia per se on sodium excretion, hypertonic saline was infused into one renal artery of dogs by means of a femoral to renal arterial shunt.

2. Even when filtered sodium in the experimental kidney was reduced below that in the control, sodium excretion from the experimental kidney was increased above that on the control side. The depression of tubular sodium reabsorption by hypernatremia was equivalent to about 7% of the filtered load.

3. The data indicate that hypernatremia per se decreases tubular reabsorption of sodium independently of any effect of systemic volume loading. This effect of hypernatremia is exerted directly on the kidney.

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