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# DIFFERENCES BETWEEN THE ACTIVITIES OF MATURE GRANULOCYTES IN LEUKEMIC AND NORMAL BLOOD<sup>1</sup>

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The abnormal appearance and number of granulocytes in the various leukemias suggest that phagocytosis by granulocytes in all stages of maturity might be so disturbed that leukemic patients might become more susceptible to bacterial infections than normal persons. Phagocytic activity among immature granulocytes is only slight but increases with maturity and becomes prominent among metamyelocytes and mature leukemic neutrophils (1-4). It has not been established, however, whether mature leukemic neutrophils are equal in phagocytic activity to normal mature neutrophils. Such a comparison has been attempted by Hirschberg (5) who concluded that the mature neutrophils in leukemic blood displayed less phagocytic activity than mature neutrophils in normal blood. Her results are open to question, however, because comparisons were made between phagocytic systems which were not uniform with respect to: 1) The number of phagocytes per unit volume; 2) quantity of opsonin; and 3) character of suspending medium. She also neglected to take into account the possible effect of antileukemic treatment on the phagocytic properties of leukocytes.

In the present study, the activity of mature leukemic granulocytes was measured in two ways: 1) Suspensions of leukemic and suspensions of normal leukocytes, containing approximately equal numbers of mature granulocytes, were inoculated with opsonized type II pneumococcus and the phagocytic activity of leukemic granulocytes was compared with that of normal granulocytes. 2) Heparinized leukemic blood was divided into two portions and the buffy coat removed from one to lower the count of mature neutrophils to normal

levels; each portion was inoculated with pathogenic staphylococci and the number of culturable staphylococci were counted before and after six hours' incubation.

The simultaneous performance of these two tests on specimens from a given patient provided information not only on the action of single mature granulocytes in a standardized medium, but also on the collective action of leukemic leukocytes in their own blood. Observations were also made: 1) On the effect of antileukemic therapy on phagocytosis by neutrophils; and 2) on the incidence of bacterial infection in the patients whose leukocytes were examined.

## METHODS AND MATERIALS

1. *Clinical material.* Patients were studied at the University Hospital or the Simpson Memorial Institute of the University of Michigan. The diagnosis of leukemia was established in each case by staff members of the Simpson Institute through laboratory examination of peripheral blood and bone marrow and clinical examination of the patient. The number of cases of each type of leukemia and the treatments are listed in the protocol. Roentgen irradiation was administered at the University Hospital under the supervision of Dr. Isadore Lampe.

Freshly drawn samples of venous blood obtained from these patients were divided into two portions. One portion was used to provide leukocytes for the test with opsonized pneumococci and one portion was inoculated with pathogenic staphylococci as described below.

2. *Measurement of leukocyte activity in a standard phagocytic system.* Leukocytes from patients with small buffy layers and from normal persons were separated from fresh venous blood (containing heparin 0.5 mg. per ml. as anticoagulant) by permitting the erythrocytes to sediment at 37° C. for 30 minutes in a glass tube tilted at 60 degrees; then the supernatant plasma was removed with a warm sterile pipette. Supernatant plasma obtained in this way usually contained at least as many leukocytes per cu. mm. as existed in the whole blood, but almost no erythrocytes. Sometimes it was necessary to centrifuge the supernate or the blood itself at 500 r.p.m. for 5 minutes to remove the erythrocytes. The leukocytes were sedimented only slightly at this speed. At first, only siliconized glassware was used, but later it was

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TABLE I

Antibacterial action of leukemic whole blood and of suspensions of leukemic leukocytes from patients with chronic granulocytic leukemia\*

## CHRONIC GRANULOCYTIC LEUKEMIA

NAME AGE DURATION	DATE	PERIPHERAL BLOOD								RINGS SUSPENSION OF LEUKOCYTES: APPROXIMATELY 1,000 MATURE NEUTROPHILS (BANDS AND SEGS) PER MM <sup>3</sup>						TREATMENT
		WBC X1000	DIFFERENTIAL				MATURE NEUTRO- PHILS PER MM <sup>3</sup>	CULTURABLE STAPHYLOCOCCI (COLONIES PER ML.)		PER CENT OF MATURE NEUTROPHILS PHAGOCYTOZING PNEUMOCOCCI			AVERAGE NUMBER BACTERIA PER PHAGOCYTTIC CELL			
			S	B	I	O		BEFORE	AFTER 6 HOURS	S/B	PATIENT	CONTROL	PATIENT	CONTROL		
R.F. 57 6 mo.	3/24/53	476.	13	9	73	5	105,000	16,800	106	90/10	11	100	7.0	10.5	X-Ray to spleen begun this date	
	3/27/53	364.	30	14	54	2	160,000	18,000	3,600	55/45	25	100	3.2	8.1	3rd day of X-Ray to spleen	
	4/2/53	148.	52	33	8	2	125,000	18,600	848	48/52	78	100	2.8	14.5	1 week after X-Ray	
M.W. 39 10 mo.	3/19/53	8.	50	0	0	50	4,000	22,000	4,400	100/0	100	100	16.0	16.0	Sustained response to P32 given July 1952 when WBC was 99,000	
	4/24/53	410.	18	39	39	4	240,000	4,800	1,560	60/40	48	100	8.9	3.8	None	
H.R. 21 4 mo.	4/29/53	370.	21	14	55	10	130,000	20,000	694	80/20	4	90	10.0	9.0	Urethane started 4/28/53	
	5/5/53	379.	35	29	33	3	255,000	18,000	648	55/45	18	48	4.5	4.7	1 week of urethane	
	5/11/53	338.	45	23	30	2	230,000	9,600	32	20/80	11	100	0.7	9.5	Urethane stopped preceding day	
E.J. 6 mo.	12/15/52	430.5	29	33	26	12	275,000	21,000	1,018		38	100	3.3	9.0		
	12/23/52	222.	28	33	31	8	140,000				74	100	13.5	17.7	P32 on 12/17/52	
T.B. 26 6 mo.	1/12/53	85.	51	6	33	10	49,000				67/33	66	67	9.9	23.3	P32 July, August, October, and December 1952
	2/9/53	143.	48	14	28	10	92,000	18,000	688	84/16	28	100	9.8	10.5	P32 this date	
	4/10/53	62.	54	26	10	10	50,000	18,800	1,640	88/12	60	90	5.0	10.7	Total body irradiation 3/30/53 to 4/10/53	
A.E. 3 yr.	2/20/53	19.	76	0	10	14	15,000	20,600	12,000	100/0	21	86	1.3	20.1	No recent treatment	
	3/6/53	11.	72	2	0	26	9,000	22,000	6,200	95/5	95	95	16.2	16.6	X-Ray to spleen 2/20/53 to 3/6/53 (6 wk.)	
M.P. 55 9 mo.	1/12/53	730.	16	28	55	1	300,000	13,680	276	64/36	15	100	19.5	25.0	None	
	1/14/53	800.	18	34	65	3	400,000	19,600	588		0	90	0.0	15.0		
H.P.	3/31/53	37.	9	13	42	38	8,100	18,000	6,000	60/40	66	94	16.5	13.0	Total body X-Ray started 3/28/53 and completed 4/6/53	
M.S. 51 3 yr.	4/29/53	340.	28	6	71	5	115,000	18,800	560	40/60	15	89	3.0	9.0	Received 10 mg. TEM on day before tests	
C.S. 48 3 yr.	3/20/53	82.	45	18	23	14	51,000	20,160	940	90/10	50	100	5.0	15.0	P32 3/19/53	
E.S. 56 2 yr.	8/12/52	23.	58	17	15	20	14,900				80	83	6.6	10.3	P32 1/14/52	
F.O. 45 2 yr.	7/7/52	39.7	38	52	1	9					75/25	69	96	11.1	12.4	No recent therapy
	7/11/52	25.1	35	50	4	11					65/35	55	85	5.5	6.0	Total body X-Ray started 7/9/52
	9/16/52	140.	5	0	95	0	7,000	18,360	162						Receiving total body X-Ray 9/10/52 to 9/29/52	
	9/25/52	151.5	6	3	91	0	13,600	28,080	3,840						Receiving total body X-Ray	
	10/3/52	187.	1	3	96	0	7,500	28,000	5,960							
H.W. 51 6 mo.	7/28/52	100.	40	16	39	5					70/30	5	44	14.5	7.3	TEM 5 mg. daily July 24, 25, 26, 1952
	10/16/52	72.5	46	18	32	6	45,000	25,800	7,800							
M.L. 53 2 yr.	12/15/52	174.5	38	31	37	6	110,000	21,600	8,400	50/50	39	100	9.0	14.0	Receiving total body X-Ray started on 12/10/52	
	1/30/53	174.	29	28	35	8	102,000	22,000	9,000	40/60	52	82	4.0	7.0	No treatment - last X-Ray completed 6 weeks before	
	2/4/53	209.	57	23	20	10	120,000	18,000	6,600							
	3/17/53	268.5	31	28	40	1	182,000	18,240	970	40/60	40	100	4.0	25.0	X-Ray therapy to Rt. humerus 2/11/53 to 2/17/53	
	3/19/53	220.	22	22	58	0	97,000	17,800	2,600	50/50	15	82	3.0	4.2	X-Ray to spleen started previous day	
	3/31/53	16.5	45	32	16	7	12,700	19,800	8,400	80/10	67	94	7.0	13.0	X-Ray to spleen completed 1 week previously	
J.D. 62 24 yr.	10/13/52	197.	34	31	30	5	130,000	10,800	80						P32 9/5/52	
	10/15/52	163.	32	16	52	0	81,000	18,000	1,080						TEM 3 mg. this date, also 10/16/52 and 10/17/52	
	10/21/52	83.	47	28	24	3	65,000	24,600	600							
G.B. 3 mo.	12/4/52	45.5	50	10	23	17	27,000	29,000	13,200						P32 12/3/52	
L.B. 42 4 mo.	11/24/52	114.	24	18	38	20	46,000	24,240	6,800						None	
	12/2/52	113.5	40	5	53	2	50,000	1,000	12						Receiving urethane 3 gm. daily	
	2/4/53	129.	21	18	37	24	52,000	18,000	1,568						Receiving urethane	
W.H. 33 1 mo.	9/20/52	282.	31	17	42	10	140,000	18,480	848						None	
	10/27/52	24.8	40	22	14	24	16,000	13,200	4,080						P32 on 9/21/52	
N.D. 59 3 yr.	3/11/53	80.	40	3	18	39	33,000	19,200	5,000						X-Ray to spleen 2/27/53 to 3/8/53	
E.H. 48 2 yr.	5/20/52	83.4	7	51	42	0	48,000	16,308	16,758						X-Ray to spleen 4/10/52 to 4/16/52	

\* Whole bloods were inoculated with pathogenic staphylococci, and leukocyte suspensions with opsonized type 2 pneumococci (S = segmented neutrophils, B = band neutrophils, I = immature neutrophils, O = leukocytes other than neutrophils; S/B = ratio of segmented to band forms.)

found that comparable results were obtained with clean glassware which had not been treated with silicone. Leukocytes from patients with high counts were easily separated by pipetting the thick buffy coat from heparinized venous blood. The plasma suspensions of leukocytes were centrifuged at 1,000 r.p.m. for 5 minutes. The supernatant plasma was then removed and replaced by Ringer's solution containing 0.5 mg. heparin per ml. The Ringer-heparin solution was added in amounts necessary to give a final count of approximately 8,000 mature neutrophils per cu. mm. No attempt was made to remove the immature neutrophils also present in the suspension. Mature neutrophils were defined as band forms and segmented forms. Immature neutrophils were defined as metamyelocytes, myelocytes, progranulocytes, and myeloblasts. All cells were identified according to the criteria reported by the committee for clarification of the nomenclature of cells (6).

A type II pneumococcus, highly virulent for mice and containing a large capsule rendering it resistant to phagocytosis in the absence of specific opsonin *in vitro*,

was subcultured on rabbit blood-agar plates. At the time of each test, freshly grown cultures of the pneumococcus were removed from the surface of the plate with a platinum wire and the bacteria suspended in Ringer's solution to give a density equal to that of the No. 1 tube of the MacFarland nephelometer. The pneumococcal suspension was distributed in 0.1 ml. quantities among six glass test tubes measuring 15 × 100. Rabbit sera containing high titers of specific type II pneumococcal antibody were used to opsonize the pneumococcus by adding 0.02 ml. of antiserum to each of two tubes containing pneumococcal suspension. Pneumococci in two other tubes were similarly treated with antiserum diluted in Ringer's solution; those in the other two tubes with normal non-immune rabbit serum. The dilute antiserum was used to cover the possibility that diminished phagocytic activity of leukemic cells might not be apparent in the presence of excess antibody. The mixtures of serum and pneumococci were incubated in a water bath at 37° C. for 30 minutes. Then 0.1 ml. of leukemic cells suspended in Ringer's was added to one of each of the three pairs of

TABLE II

*Antibacterial action of leukemic whole blood and suspensions of leukemic leukocytes from patients with acute and subacute granulocytic leukemia\**

## ACUTE AND SUBACUTE GRANULOCYTIC LEUKEMIAS

NAME AGE DURATION	DATE	PERIPHERAL BLOOD							RINGER'S SUSPENSION OF LEUKOCYTES: APPROXIMATELY 8,000 MATURE NEUTROPHILS (BANDS AND SEGGS) PER MM <sup>3</sup>				TYPE OF GRANULOCYTIC LEUKEMIA	TREATMENT	
		WBC X1000	Differential				MATURE NEUTRO- PHILS PER MM <sup>3</sup>	CULTURABLE STAPHYLOCOCCI (COLONIES PER ML.)		PER CENT OF MATURE NEUTROPHILS PHAGOCYTOSING PNEUMOCOCCI		AVERAGE NUMBER BACTERIA PER PHAGOCYTIC CELL			
			S	B	I	O		BEFORE	AFTER 6 HOURS	PATIENT	CONTROL	PATIENT			CONTROL
D.W. 22 5 mo.	7/5/52	6.1	11	0	65	24				75	82	6.7	11.1	SA;SL	None
E.L. 73 3 wk.	3/6/53	15.6	39	1	1	59	6,000	22,200	4,800	92	95	11.5	16.6	SA;SL	None
E.T.	3/5/53	3.96	59	9	11	30	2,400	20,000	150	51	60	3.8	3.0	SA;SL	?
I.B. 22 4 mo.	11/15/52	5.6	16	3	81	0				77	82	4.5	7.0	SA;SL	None
G.S. 69 5 mo.	3/24/53	227.	20	2	62	16				68	100	4.1	10.5	Acute	None
R.V. 55 9 mo.	3/19/53	32.5	22	6	70	2				98	82	9.9	4.2	Acute	Cortisone
F.S. 17 2 wk.	3/31/53	4.5	18	3	38	41	900	20,400	13,200	100	100	40.0	10.0	Acute	None
G.H. 25 4 mo.	8/15/52	11.	15	4	74	7				58	44	5.3	6.0	Acute	ACTH 25 mg. for 2 days
	10/13/52	49.75	11	6	73	10	8,500	18,000	4,800						None
	10/15/52	127.	6	2	90	2	10,000	29,000	6,000						None
	10/21/52	60.5	6	4	88	2	6,000	24,880	14,400						None
F.H. 76 6 mo.	10/23/52	17.	28	36	19	17	11,500	24,000	2,400					SA;SL	
P.C. 58 7 mo.		26.5	16	14	4	66	7,900	28,000	17,600	97	88	15.0	6.0	SA	
F.C. 59 1 mo.	1/28/53	145.	2	1	91	6	4,600	24,000	17,600						Cortisone 75 mg. - 6 H.

\* (S.A. = Subacute, SL = Subleukemic; abbreviations of cells same as Table I; L = Lymphocytes).

TABLE III

*Antibacterial action of leukemic whole blood and suspensions of leukocytes from patients with chronic lymphocytic leukemia\**

CHRONIC LYMPHOCYTTIC LEUKEMIA

NAME AGE DURATION	DATE	PERIPHERAL BLOOD							RINGERS SUSPENSION OF LEUKOCYTES: APPROXIMATELY 8,000 MATURE NEUTROPHILS (BANDS AND SEGGS) PER MM <sup>3</sup>				TREATMENT	
		WBC X1000	S	B	I	L	MATURE NEUTRO- PHILS PER MM <sup>3</sup>	CULTURABLE STAPHYLOCOCCI (COLONIES PER ML.)		PER CENT OF MATURE NEUTROPHILS PHAGOCYTOSING PNEUMOCOCCI		AVERAGE NUMBER BACTERIA PER PHAGOCYTTIC CELL		
								BEFORE	AFTER 6 HOURS	PATIENT	CONTROL	PATIENT		CONTROL
W. T. 48 1 mo.	11/21/52	128.	1	0	0	99	1,280	26,400	13,200	85	46	5.1	12.2	None
J. P.	4/24/53	352.5	<1	0	0	99+	<3,500	23,400	16,800	100	100	20	12	X-Ray to spleen 4/23/53 to 5/5/53
H. C. 74 7 yr.	12/2/52	24.	23	0	1	76				100	100	10.9	7.2	Receiving X-Ray to nodes 11/3/52 to 12/13/52
J. H. 67 2 yr.	3/5/53 4/17/53	400. 280.	1 3	0 0	0 0	99 97	4,000 8,400	23,400 18,600	14,400 6,600	76 92	92	9.5	8.6	X-Ray to regional node 2/23/53 to 3/5/53 6 weeks after completion of X-Ray
A. V.	3/31/53	51.	9	0	0	91	4,590	17,000	9,600	88	94	17.6	13	
J. H. 61 2 mo.	3/3/53	186.	8	0	0	92	15,000	19,600	8,160	82	100	15	15	X-Ray to nodes 2/25/53 to 3/9/53
A. R. 83 2 1/2 yr.	12/1/52	8.7	41	0	0	59	3,600	25,600	2,640	100	100	>40	15	None within 6 months
J. O. 84 2 yr.	4/21/53	15.3	24	0	2	74	3,500	18,144	14,994					Total body X-Ray 4/17/52 to 4/25/52

\* Abbreviations same as Tables I and II.

TABLE IV

*Antibacterial action of leukemic whole blood and suspensions of leukocytes from patients with acute and subacute lymphocytic leukemia\**

ACUTE AND SUBACUTE LYMPHOCYTTIC AND MONOCYTTIC LEUKEMIA

NAME AGE DURATION	DATE	PERIPHERAL BLOOD							RINGERS SUSPENSION OF LEUKOCYTES: APPROXIMATELY 8,000 MATURE NEUTROPHILS (BANDS AND SEGGS) PER MM <sup>3</sup>				TYPE OF LEUKEMIA	TREATMENT		
		WBC X1000	SB	M	PM	B	L	MATURE NEUTRO- PHILS PER MM <sup>3</sup>	CULTURABLE STAPHYLOCOCCI (COLONIES PER ML.)		PER CENT OF MATURE NEUTROPHILS PHAGOCYTOSING PNEUMOCOCCI				AVERAGE NUMBER BACTERIA PER PHAGOCYTTIC CELL	
									BEFORE	AFTER 6 HOURS	PATIENT	CONTROL			PATIENT	CONTROL
H. T. 34 2 mo.	4/15/53	120.	0	0	50	43	7	0	18,000	21,600	0	100	I	15	Acute Monocytic	None
	4/17/53	51.	0	0	25	66	9	0	12,000	9,600	0	100	I	12.5		
	4/29/53	12.	0	0	30	64	6				0-Promon. active	89	I	9.		
C. K.	5/5/53	1.2	2	0	0	30	69	24	368	2,400					Subacute Subleukemic Monocytic	None
W. S. 58 2 wk.	9/26/53	5.35	17	13	10	38	22	920 +650 mono- cytes	738	348					Acute Subleukemic Monocytic	None
A. Gh	4/10/53	19.	3	0	0	0	97	570	824	11,600	90	90	33	10.7	Subacute Subleukemic Lymphocytic	None
A. G. 80	12/23/52	2.1	8	0	0	0	92	168	1,368	3,600					Subacute Subleukemic Lymphocytic	None

SB: Mature neutrophils  
M: Monocytes  
PM: Promonocytes  
B: Blasts  
L: Lymphocytes  
I: Innumerable

tubes and 0.1 ml. of normal cell suspension to each of the corresponding three tubes. All tubes were sealed with paraffin and the mixtures of leukocytes and opsonized pneumococci were rotated in a machine at 6 r.p.m. for 15 minutes. The tubes were opened and a drop of the con-

tents spread on a glass coverslip. The film was dried, stained with Wright's Stain, and mounted on a glass slide. Phagocytic activity was determined as follows: Approximately 100 mature neutrophils were counted; the number which had ingested pneumococci were re-

corded and subsequently referred to as "Per cent phagocytosis." The total number of intracellular pneumococci were also counted and the average number of pneumococci ingested were recorded for those cells which contained intracellular organisms. Results in Tables I, II, III, and IV are given for undiluted opsonin because the results with diluted opsonin were confirmatory.

3. *Measurements of the antibacterial action of whole leukemic blood.* The purpose of this experiment was to determine the capacity of whole leukemic blood inoculated with pathogenic staphylococci to lower the bacterial count after 6 hours incubation at 37° C. Pathogenic staphylococci were selected as the test organism not only because they produced most bacterial infections in leukemic patients, but also because they multiplied rapidly in fresh plasma or in leukocyte-free whole blood incubated at 37° C. Although this test has been regarded as a measure of bactericidal activity, it has been demonstrated that the decrease in number of bacterial colonies results from phagocytosis and intracellular clumping, rather than from killing of staphylococci (7). When leukemic or normal phagocytes are disrupted with distilled water or by raising the pH above 10, the dispersed cocci grow separately instead of in clumps and the number of colonies increases greatly in the agar plates. The alkaline pH also breaks up the extracellular clumps of coagulase positive staphylococci which are present in plasma.

The strain of staphylococcus (B) used in all experi-

ments had been isolated from human empyema fluid. It clotted human plasma, clumped instantly in fibrinogen solution, and on blood agar produced hemolysis and a golden yellow pigment. It was maintained in a smooth phase by frequent subculture on blood agar. For routine tests, the staphylococci were grown for 18 hours in tryptose phosphate broth and then resuspended in saline to give a density equal to that of the No. 1 tube of the MacFarland nephelometer. The suspension of staphylococci was then diluted in saline either to 10<sup>-4</sup> or 10<sup>-3</sup> depending on whether heavy or light inoculums were desired. Because of the large errors involved in reaching a given dilution by pipetting, numbers of bacteria were selected for inoculation that could be counted in agar plates without serial dilution.

Leukemic whole blood used in these experiments was part of that drawn to provide leukocytes for the standardized phagocytic system in Section 1 and permitted simultaneous performance of both procedures on aliquots from the same sample. The portion used for inoculation with staphylococci was divided into equal aliquots; the leukocyte count was lowered considerably in one of the aliquots by pipetting off the buffy coat. Two and one half ml. of blood with buffy coat intact and 2.5 ml. of blood with buffy coat removed were each inoculated with 0.5 ml. of staphylococcal suspension and incubated for 6 hours at 37° C. in tubes measuring 38 × 180 mm. The tubes were not agitated because it was found that agitation for a

TABLE V

*Antibacterial action of leukemic whole blood measured by its ability to reduce the number of culturable staphylococci (coagulase positive) during 6 hours incubation: Effect of lowering to approximately normal the numbers of mature granulocytes by removing the leukemic buffy coat (Compare with Table VA which gives results for normal blood)*

Patient	NUMBER MATURE GRANULOCYTES (SEGS. AND BANDS) PER MM <sup>3</sup>	BUFFY COAT REMOVED	STAPHYLOCOCCI CULTURED BEFORE AND AFTER 6 HOURS INCUBATION IN LEUKEMIC WHOLE BLOOD (COLONIES PER ML.)			
			LARGE INOCULUM		SMALL INOCULUM	
			BEFORE	AFTER 6 HOURS	BEFORE	AFTER 6 HOURS
R. F. 3/24/53	160,000	no	18,000	3,600	790	8
	7,000 0	yes Plasma	22,000	26,000	712 840	810 9,600
4/2/53	125,000	no	18,600	828	1,170	10
	9,000	yes Plasma	16,400 20,000	16,800 Innumerable	546	650
E. J. 12/15/52	275,000	no	21,000	10,000	516	10
	6,500	yes Plasma	30,000 unable	32,400 to obtain	1,058 cell-free	1,640 plasma
L. G. 11/24/52	46,000	no	24,240	6,600	1,376	112
	10,500	yes Plasma	24,480	20,800	1,440 1,200	634 18,000
M. L. 3/17/53	162,000	no	18,240	970	620	8
	4,200	yes Plasma	16,800	21,000	658 1,024	1,336 16,400
3/31/53	12,700	no	19,800	8,400	920	288
	4,000	yes Plasma	18,000	18,000	1,058 1,018	1,158 2,400
A. A. E. 2/20/53	15,000	no	20,600	12,000	1,008	330
	5,700	yes Plasma	20,800	19,700	1,188 1,018	4,000 20,000
M. P. 1/12/53	300,000	no	13,680	476	652	28
	26,000	yes Plasma	29,400 unable	20,400 to obtain	cell-free	plasma

TABLE VA

**ANTIBACTERIAL ACTION OF NORMAL BLOOD MEASURED BY ITS ABILITY TO  
REDUCE THE NUMBER OF CULTURABLE STAPHYLOCOCCI COAGULASE POSITIVE  
DURING 6 HOUR INCUBATION**

		NEUTROPHILS PER MM <sup>3</sup>	STAPHYLOCOCCI CULTURED BEFORE AND AFTER 6 HOURS INCUBATION IN NORMAL WHOLE BLOOD OR PLASMA (COLONIES PER ML.)			
			LARGE INOCULUM		SMALL INOCULUM	
			BEFORE	AFTER 6 HOURS	BEFORE	AFTER 6 HOURS
1.	Blood Plasma	5,500 0	24,000	3,880	840 768	52 6,960
2.	Blood Plasma	5,000 0	20,400	6,600	1,096	130
3.	Blood Plasma	4,500 0	18,000	6,240	912 1,152	224 22,000
4.	Blood Plasma	6,500 0	17,600 10,000	3,400 40,000	260 364	24 9,840
5.	Blood Plasma	4,500 0	21,000	10,200	456 232	120 9,000
6.	Blood Plasma	5,500 0	24,400	10,320	1,440	24,000
7.	Blood Plasma	5,000 0	20,400 3,840	3,600 9,000	558 686	56 2,400
8.	Blood Plasma	3,000 0	22,800	7,200	1,080 692	368 14,400
9.	Blood Plasma	6,300 0	23,000 20,400	1,200 49,000	860 428	16 6,480
10.	Blood Plasma	3,500 0	20,400 20,000	4,200 39,600	496 500	58 3,100
11.	Blood Plasma	5,300 0	22,000	2,400	672 424	42 5,200
12.	Blood Plasma	3,600 0	12,000 9,600	3,600 40,000	440 364	272 7,840

period as long as 6 hours was very destructive to the leukocytes. Before incubation, and immediately after removal from the incubator, the tubes were shaken to disperse the leukocytes and bacteria, and 0.5 ml. of the blood was removed each time and incorporated into agar pour plates and incubated at 37° C. for 24 hours. The bacterial colonies were then counted. Normal whole blood was inoculated with staphylococci and treated in the same way. Results were not recorded if heavy growth failed to develop in cell-free plasma controls.

4. *Phagocytic activity of normal leukocytes in leukemic plasma.* The phagocytic system described in Section 2 was useful for demonstrating an antileukocyte factor in leukemic plasma. The procedure was identical except that normal leukocytes were resuspended in leukemic plasma instead of Ringer's solution. The leukemic and normal control plasmas had been collected on the dates indicated in Table VI and stored at 4° C. until 5/19/53 and 5/21/53 when they were removed for the experiments. In this way all plasmas were tested simultaneously against normal leukocytes obtained from a given sample of blood.

## RESULTS

*Chronic granulocytic leukemia*

Pneumococci were phagocytosed by fewer mature leukemic neutrophils than by normal neutrophils in most experiments. Phagocytosis equal to that of the normal control was observed among leukocytes from only 2 of 14 leukemic patients (A. E. 3/6/53 and M. W.) at a time when both had experienced marked clinical and hematologic remission from successful therapy. In two other cases (T. B. 1/12/53 and E. S. 8/12/53) the per cent phagocytosis by mature neutrophils approached normal, but the average number of intracellular pneumococci per phagocytic cell was markedly lower than normal. Phagocytes from 7 of the 14 patients contained at one or more times an average number of intracellular pneumococci equal to or greater than the average observed in

normal phagocytes serving as controls. Two of these patients were those who had undergone remission after treatment and whose leukocytes were all as active as normal. Only a fraction of the leukocytes from the other five patients were phagocytic, but the phagocytic cells were, on the average, as active as normal or more so. In addition to the two whose leukocyte function was normal during remission, three patients (M. L. 3/31/53, E. J. 12/23/53, R. F. 4/2/53) and perhaps a fourth (T. B. 4/10/53) manifested improvement of leukocyte function after treatment with x-ray or P<sub>32</sub>. A sharp fall in phagocytic activity was observed in leukocytes from patient (M. R.) after the start of treatment with urethane.

Both band and segmented cells were regarded as mature neutrophils. Numerous reports have disclosed that there is no significant difference between the phagocytic activity of band forms and segmented forms from normal persons (2-4, 8). Phagocytosis by metamyelocytes was occasionally observed but rarely if ever occurred among myelocytes or younger cells in our preparations. Distinction between band and segmented forms was sometimes difficult when the cell contained numerous cocci. For this reason no attempt was made to compare quantitatively the relative phagocytic power of band cells and segmented cells. Table I discloses that in chronic granulocytic leukemia the per cent phagocytosis was usually much less than the per cent of segmented neutrophils in a given suspension. It is clear, therefore, that phagocytosis by segmented neutrophils was impaired in chronic granulocytic leukemia.

Samples of blood from each of the 12 patients whose mature neutrophils showed impaired phagocytosis of pneumococci reduced the number of culturable staphylococci in 6 hours as readily as normal blood. Bloods from a total of 19 of 20 patients lowered the total staphylococcal count as effectively, or more so, than normal blood. The one exception (E. H.) was a patient whose blood contained mostly band neutrophils. Although his blood did not lower the bacterial count, it prevented growth.

The observed impairment of phagocytosis by individual neutrophils in chronic granulocytic leukemia, as measured by the phagocytosis of pneumococci, was also demonstrated when the total

neutrophil count of whole leukemic blood was reduced to normal. Thus, in bloods from seven patients, removal of the buffy coat not only reduced the neutrophil count to levels approaching normal, but also rendered the blood incapable of lowering significantly the count of staphylococci (Table V).

#### *Acute and subacute granulocytic leukemia*

The per cent phagocytosis by mature neutrophils was at least normal or nearly normal, in eight of nine patients (Table II). In five of these eight leukemic patients, the average number of pneumococci per phagocytic cell was normal or greater. Only in the suspension made from the cells of one patient (G. S.) were both the per cent phagocytosis and average number of intracellular bacteria reduced when compared to the normal control. He was the only patient of the nine whose total leukocyte count in the peripheral blood was greatly elevated when the test was performed. In three patients (P. C., F. S., and R. V.) there was a great increase in phagocytic activity over normal as judged both by per cent phagocytosis and number of bacteria per cell. Neutrophils from patient F. S. were so heavily packed with pneumococci that the bacteria could not be counted. All bloods from patients with acute or subacute granulocytic leukemia lowered the count of inoculated staphylococci as well as normal blood. The blood from two patients reduced the number of culturable staphylococci despite the presence of only 2,400 (patient E. T.) and 900 (patient F. S.) mature neutrophils per cu. mm.

#### *Chronic lymphocytic leukemia* (Table III)

The per cent phagocytosis by mature neutrophils from five patients was essentially normal. The per cent phagocytosis by leukocytes from two patients (J. H. and J. M.) was reduced, but not by much. Phagocytes from five of the seven patients contained more intracellular pneumococci than normal, from one patient (J. M.) the same as normal, and only those from patient W. Y., less than normal. Two of the patients (J. P. and H. C.) whose leukocytes were more active than normal were receiving x-ray therapy during the periods when the tests were made. Patient J. M., whose leukocyte function was slightly reduced, was also receiving x-ray therapy.



Although the count of mature neutrophils was 4,000 per cu. mm. or less in the blood of five of seven patients, normal or nearly normal reduction in numbers of inoculated staphylococci was observed in each case. The blood of patient A. R., containing only 3,600 mature neutrophils per cu. mm., reduced the number of culturable staphylococci by almost 90 per cent or more than that produced by normal blood. His leukocytes were also exceedingly active in the standardized phagocytic system against pneumococci.

*Acute and subacute monocytic and lymphocytic leukemias (Table IV)*

It was possible to test the neutrophils of only one patient (A. G.) for phagocytosis of pneumococci. These were more active than normal. Some of the promonocytes (per cent not determined) of patient H. T. showed tremendous phagocytic activity and engulfed so many pneumococci that the bacteria could not be counted in each cell.

These phagocytic promonocytes were apparently so active that they restrained the growth of inoculated staphylococci and even lowered the count on one occasion (4/17/53). Similar activity of monocytes or promonocytes may be evident in the case of W. S. whose blood reduced a small inoculum of staphylococci by nearly 50 per cent. The number of phagocytes in the bloods of the other three patients in this group were too low to restrain staphylococcal multiplication.

*Phagocytic activity of normal leukocytes in leukemic plasmas (Table VI)*

The plasmas obtained from patient M. L. (chronic granulocytic) and from patient F. S. (acute granulocytic) abolished the phagocytic activity of normal leukocytes obtained from two donors. These plasmas also damaged the neutrophils, producing severe lysis (plasma M. L.) or pyknosis (plasma F. S.). The plasmas of three other patients (A. G., subacute subleukemic

TABLE VI

## PHAGOCYTTIC ACTIVITY OF NORMAL NEUTROPHILES SUSPENDED IN LEUKEMIC PLASMAS

All plasmas were stored at 4° C. and later tested simultaneously with each sample of normal cells.

SOURCE OF LEUKEMIC PLASMA			PER CENT OF NEUTROPHILES SUSPENDED IN LEUKEMIC PLASMA WHICH PHAGOCYTOSED OPSONIZED TYPE 2 PNEUMOCOCCI		APPEARANCE OF CELLS
CASE	DATE	TYPE OF LEUKEMIA	NORMAL CELLS J.F.	NORMAL CELLS A.B.	
A.G.	4/10/53	Subacute subleukemic lymphocytic	72	50	Lysis (of some cells)
M.L.	3/31/53	Chronic granulocytic	0	0	Lysis of all cells
F.S.	3/31/53	Acute granulocytic	2	0	Pyknosis of all cells
H.T.	4/29/53	Acute monocytic	64	62	Normal
A.E.	3/6/53	Chronic granulocytic	50	65	Normal
E.L.	3/6/53	Subacute subleukemic granulocytic	68	86	Normal
J.P.	4/24/53	Chronic lymphocytic	64	88	Normal
U.Y.	2/24/53	Chronic lymphocytic	65	87	Normal
M.W.	3/19/53	Chronic granulocytic	75	70	Normal
M.R.	5/5/53	Chronic granulocytic		90	Normal
M.S.	4/29/53	Chronic granulocytic		90	Normal
Normal Control. A				90	
Normal Control. B				96	
Normal Control. C			100		

TABLE VII

## INCIDENCE OF BACTERIAL INFECTIONS IN LEUKEMIC PATIENTS

Infections were all staphylococcal and consisted of mild furuncles, pharyngitis, and a generalized pustular dermatitis.

TYPE OF LEUKEMIA	TOTAL NUMBER OF CASES	TOTAL NUMBER WITH PROVEN BACTERIAL INFECTION
Chronic Granulocytic	24	2
Acute Granulocytic	8	3
Chronic Lymphocytic	10	0
Acute Lymphocytic	2	0
Acute Monocytic	3	0
TOTAL	47	5

lymphocytic; H. T., acute monocytic; A. E., chronic granulocytic) also seemed to reduce phagocytosis of normal leukocytes, but not by much. The plasma of A. G. produced slight cytolysis.

## DISCUSSION

There are at least two possible explanations for the decreased phagocytic activity of mature neutrophils in chronic granulocytic leukemia, observed in this study. One possible explanation is that there is an inherent defect in the leukemic cell. The other is that leukemic plasma contains a factor which is injurious to the neutrophil. Whatever the cause, the decreased phagocytic activity could not have been anticipated by morphologic study because the nuclear configuration and cytoplasmic structure of most of the mature leukemic neutrophils were normal. The existence of functional changes without morphologic abnormality in neutrophils has also been suggested by observations on leukocytes removed from patients with diabetes and anemia. Recently, Martin, McKinney, Green, and Becker (9) have reported that leukocytes from diabetics cannot oxidize glucose normally *in vitro* unless insulin is provided. In anemia, Berry and Spies (10) have found that neutrophils, instead of showing disturbed activity, possess greater phagocytic capacities than normal cells.

This state of reduced phagocytic capacity is not a universal feature of the leukemic process, however. In chronic lymphocytic leukemia, in acute granulocytic leukemia, and perhaps in acute lymphocytic leukemias, the individual function of the

granulocytes was not usually disturbed. Because of the diminished total numbers of cells, however, the total antibacterial activity of whole blood in these conditions was sometimes less than that of blood from patients with chronic granulocytic leukemia and even less than normal.

It does not seem likely that the decreased activity of neutrophils in granulocytic leukemia was due to irradiation because leukocytes from patients who never received irradiation prior to the study displayed impaired phagocytic activity. Moreover, irradiation appears to enhance rather than depress phagocytic activity. Thus leukocytes from some patients with lymphocytic leukemia receiving Roentgen irradiation displayed greater phagocytosis than normal. In animals, irradiation has also produced greater phagocytosis than normal by neutrophils (11). There is little question that in several patients with granulocytic leukemia, radiation therapy actually produced an improvement in phagocytic activity. The effect of urethane appeared to differ from that of irradiation in the one patient studied because there was a decrease instead of increase in phagocytic activity.

Although the function of the individual cell is impaired in chronic granulocytic leukemia, the excess number of granulocytes raises the total antibacterial defense of the whole blood to normal. Since tissues receive their granulocytes from the blood, it is not surprising that the cellular morphology of the inflammatory reaction in infected tissues is normal in granulocytic leukemia. Jaffé (12) found that in these patients the cells comprising the inflammatory reaction had the appear-

ance of normal neutrophils and they were present in normal numbers or greater.

Bacterial infections were not frequent among the patients in this report (Table VII). Only two patients with chronic granulocytic leukemia suffered from infection. These were both mild furuncles which were promptly localized. Three patients with acute granulocytic leukemia appeared to suffer bacterial infections. Two patients had a sore throat from which coagulase positive staphylococci were cultured. These bacteria are frequently present without producing infection and cannot be definitely regarded as the cause of the sore throats. A third patient experienced a generalized pustular eruption. All three cases recovered after receiving antibiotics. No bacterial infections were noted in 12 patients with lymphocytic leukemia. The relatively low incidence of severe infection in all patients studied may be correlated with observations on leukocyte function. In chronic granulocytic leukemia, an excess number of neutrophils offsets poor individual function. In lymphocytic leukemia, and in the acute leukemias, there appear to be enough normally active phagocytes for defense against bacterial infection even though the total numbers may be somewhat lower than normal. Thus, Jaffé (12) observed that patients with very low leukocyte counts could mobilize adequate numbers of neutrophils from isolated foci in the bone marrow to provide good cellular defense reactions to infection in the tissues. Other cells than granulocytes may also contribute significantly to cellular defense in leukemia. An example of this is provided by the patient with acute monocytic leukemia whose promonocytes were so actively phagocytic that whole blood containing them but no other phagocytes possessed total antibacterial properties equal to that of normal blood. The extreme susceptibility of certain patients with acute leukemia to bacterial infections appears to be a function of a marked decrease in total number of mature granulocytes rather than a decrease in function of individual cells.

The antiphagocytic and leukocyte-destroying properties of stored leukemic plasmas are recorded here as a preliminary observation of interest. The two plasmas most destructive for neutrophils were obtained from patients with granulocytic leukemia when their leukocyte counts were low. Yet the activity of their own leukocytes against staphylo-

cocci was good in the fresh whole blood from which the stored plasmas were obtained. In one of these patients, however, the total leukocyte count had fallen from 220,000 to 16,500 just before the plasma was obtained. It remains to be determined if the factor in leukemic plasma which is toxic for leukocytes is related to the leukotoxic substance demonstrated by Doan (13), and what role, if any, it may have in the pathogenesis of leukemia.

#### SUMMARY AND CONCLUSIONS

Phagocytic activity of leukocytes in 45 cases of leukemia was studied by two methods: 1) Suspensions of leukemic and suspensions of normal leukocytes, containing approximately equal numbers of mature granulocytes were inoculated with opsonized type II pneumococcus and the phagocytic activity of leukemic cells was compared to that of normal granulocytes; 2) Heparinized leukemic blood was divided into two portions and buffy coat removed from one to lower the count of mature neutrophils to normal levels. Each portion was inoculated with pathogenic staphylococci and the number of culturable staphylococci were counted before and after 6 hours incubation.

Pneumococci were phagocytosed less actively by mature granulocytes from chronic granulocytic leukemia than by normal granulocytes. In lymphocytic leukemia, phagocytosis of pneumococci by granulocytes was usually normal. Treatment of patients by irradiation sometimes restored normal phagocytosis in granulocytic leukemia.

Similarly, in chronic granulocytic leukemia, when buffy coat was removed to leave a count of 5,000 to 10,000 mature granulocytes per ml. of blood, the number of culturable staphylococci was often not significantly lowered. Yet normal bloods averaging 5,000 mature granulocytes markedly reduced the numbers of staphylococci. If buffy coat was not removed, blood from chronic granulocytic leukemia also reduced markedly the number of staphylococci.

In chronic granulocytic leukemia, phagocytosis by individual granulocytes was usually impaired but overproduction of granulocytes raised the total defense of the blood to normal. In chronic lymphocytic, and certain acute leukemias, individual function of mature granulocytes is usually undisturbed, but decrease in total numbers sometimes weakens antibacterial defense.

## REFERENCES

1. Hertzog, A. J., The phagocytic activity of human leukocytes with special reference to their type and maturity. *Am. J. Path.*, 1938, **14**, 595.
2. Strumia, M., and Boerner, F., Phagocytic activity of circulating cells in the various types of leukemia. *Am. J. Path.*, 1937, **13**, 335.
3. Huddleson, I. F., and Munger, M., Phagocytic activity of bone-marrow cells. *Proc. Soc. Exper. Biol. & Med.*, 1936, **35**, 27.
4. Jersild, M., Phagocytic activities of various types of leukocytes. *Acta med. Scandinav.*, 1948, Supplement 213, 238.
5. Hirschberg, N., Phagocytic activity in leukemia. *Am. J. Med. Sc.*, 1939, **197**, 706.
6. Condensation of the first two reports of the Committee for Clarification of the Nomenclature of Cells and Diseases of the Blood and Blood-forming Organs. *Blood*, 1949, **4**, 89.
7. Braude, A. I., and Feltes, J., Studies in the destruction of staphylococci by human leukocytes: effect of clumping of intracellular and extracellular bacteria on the results obtained with the agar-plate method. *J. Lab. & Clin. Med.*, 1953, **42**, 289.
8. Ponder, E., and Flinn, Z. M., Studies on the Arneht Count. I. The relation between phagocytosis and nuclear configuration. *Quart. J. Exper. Physiol.*, 1927, **16**, 207.
9. Martin, S. P., McKinney, G. R., Green, R., and Becker, C., The influence of glucose, fructose, and insulin on the metabolism of leukocytes of healthy and diabetic subjects. *J. Clin. Invest.*, 1953, **32**, 1171.
10. Berry, L. J., and Spies, T. D., Phagocytosis. *Medicine*, 1949, **28**, 239.
11. Esplin, D. W., Marcus, S., and Donaldson, D. M., Effects of x-irradiation and adrenalectomy on phagocytic activity. *J. Immunol.*, 1953, **70**, 454.
12. Jaffé, R. H., Morphology of the inflammatory defense reactions in leukemia. *Arch. Path.*, 1932, **14**, 177.
13. Doan, C. A., The recognition of a biologic differentiation in the white blood cells. *J.A.M.A.*, 1926, **86**, 1593.