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A STUDY OF GLYCOLYSIS

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INTRODUCTION

Since Claude Bernard (1) first noted the disappearance of sugar from drawn blood in 1856 this phenomenon has been the subject of much interest and investigation. Following the discovery of insulin, studies on glycolysis were made with renewed interest in the hope that experiments in vitro might shed some light upon the rôle of insulin in carbohydrate metabolism. The rate of disappearance of sugar from blood in vitro and the rate of glycolysis are commonly considered to be synonymous, a custom which will be followed in this discussion. The accumulated data on glycolysis show a very striking lack of uniformity. The purpose of this report is to present a series of observations on glycolysis and to discuss some of the principal factors influencing the rate and the amount of sugar glycolyzed.

METHOD

Patients in the University Hospital served as subjects for the following experiments. They were unselected except that no patient with an abnormally high nonprotein nitrogen content of the blood was used. From each patient 10 cc. of blood was drawn two or two and one-half hours after a meal. This blood, with the exception of one-half cc. for the determination of the initial sugar content, was transferred immediately to a sterile 50 cc. Erlenmeyer flask containing a bent paper clip. The flask was loosely stoppered with sterile cotton, shaken for fifteen minutes to defibrinate the blood and incubated in a moist chamber at 37° C. The blood was well shaken before and after each of the sugar determinations, which were made at two hour intervals according to the method of Gibson, Mitchell and Larimer (2).

TABLE I

Total sugar glycolyzed in a series of bloods

Case	Diagnosis	Sex	Age		Total sugar glycolyzed: Hours				
				2	4	6	8	10	
			years	mgm. per 100 cc.	mgm. per 100 cc.	mgm. per 100 cc.	mgm. per 100 cc.	mgm. per 100 cc.	
1.	Myelogenous leukemia	M	59	98	128*				
2.	Myelogenous leukemia	M	48	94	158*				
3.	Diabetes mellitus	M	50	93	103	166	179	223*	
4.	Myelogenous leukemia	F	59	87	118*				
5.	Myelogenous leukemia	F	41	81	148	186*			
6.	Congenital heart disease	M	30	80	112*				
7.	Diabetes mellitus	M	49	75	143	215		242	
8.	Chronic endocarditis, cardiac decompensation, osteoar-					i			
	thritis	M	57	71	108	132*			
9.	Multiple fractures, empyema	M	56	69	129	149*			
10.	Gastric neurosis	М	51	68	100	128*			
11.	Malignant lymphomata,								
	Hodgkins type	F	20	67	128*				
12.	Arteriosclerosis, hypertrophy								
	of prostate	M	57	63	119	162*			
13.	Banti's disease, parotitis epi-								
	demic	F	22	63	110*				
14.	Septicopyemia	F	27	57	97	136*			
15.	Arsenical hepatitis		30	53	93	133			
16.	Hysteria	F	26	53	76	103*			
17.	Chronic myocarditis, hyper-								
4.0	tension	M	47	50	90	143			
18.	Mixed tumor of parotid gland	M	18	50	99	133			
40	with generalized metastases.	M	39	50	100	140	175*		
19.	Psychoneurosis	IVI	39	30	100	140	173		
20.	Pulmonary tuberculosis, tu- bercular peritonitis	M	31	50	103				
		M	43	49	87	117*			
21.	Mitral stenosis	M	23	48	95	130*			
22.	Arsenical hepatitis	F	30	48	83	127*			
23.	Peptic ulcer	J -	27	48 47	86	127*			
24.	Diabetes mellitus	M F	45	47	77	110*			
25.	Pernicious anemia	L,	43	41	''	110			
26.	Chronic progressive vascular	М	60.	46	91				
27	nephritis, chronic uremia	F	41	40	82	121*	1		
27.	Exophthalmic goiter	Г	41	***	02	121			
28.	Diabetes mellitus, adenoma of thyroid	м	54	44	96				
	inyloid	***	<u> </u>						

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TABLE I-Continued

Case	Diagnosis	Sex	Age	Total sugar glycolyzed: Hours					
				2	4	6	8	10	
			years	mgm. per 100 cc.	mgm. per 100 cc.	mgm. per 100 cc.	mgm. per 100 cc.	mgm. per 100 cc.	
29.	Syphilitic aortitis, aortic in-		(2)			ł	100 11.	100	
30.	sufficiency	M	62	44	98	140*			
50.	rheumatic endocarditis, la-								
	tent syphilis	F	34	43	72	96*			
31.	Chronic cholecystitis	M	30	43	87	120*			
32.	Bronchopneumonia	M	53	42	82	120*			
33.		141	33	42	02	120			
٠٠.	toe	M	50	42	83		149*		
34.	Cardiac decompensation	M	61	42	102	132*	149		
35.	Malignant lymphomata,	171	01	72	102	132			
	Hodgkins type	M	43	41	69	100*			
36.	Pernicious anemia	F	59	40	74	93	149*	:	
37.	Diabetes mellitus	M	15	36	128	172	232		
38.	Chronic rheumatic endocardi-			00	120	1,2	232		
	tis	M	58	36	72		129*		
39.	Chronic interstitial nephritis.	M	42	35	77	146	164	203*	
40.	Tuberculosis	M	53	35	63	105*	104	203	
41.	Arteriosclerosis	M	71	34	71	97	145*		
42.	Visceroptosis	M	24	34	94	114	143		
43.	Pernicious anemia	М	68	34	82*	111			
44.	Lymphocytic leukemia	M	20	34	57	95*			
45.	Cardiospasm	M	59	33	78				
46.	Hypertension, cardiac decom-								
	pensation	М	64	33	58	90*			
47.	Catarrhal jaundice	М	39	33	80	114*			
48.	Subacute nephritis	М	34	32	66	119	175*		
49.	Cardiac decompensation	М	43	32	89	148	172*		
50.	Diabetes mellitus, gangrene of				0,	110	172		
	toe	M	57	31	72	107	175	252	
51.	Multiple leiomyomata of	İ				10.	1,0	202	
	uterus	F	28	31	78	115*			
52.	Chronic hypertrophic arthri-								
- 1	tis	M	42	30	64	93*			
53.	Psychoneurosis	F	37	29	83	132*			
54.	Pleurisy with effusion	M	24	28	54				
55.	Arteriosclerosis	M	68	28	60	108*			
56.	Angina pectoris, cardiac de-			İ					
	compensation	M	71	27	66	111		172*	

TABLE I-Continued

Case	Diagnosis	Sex	Age	Total sugar glycolyzed: Hours					
				2	4	6	8	10	
			years	mgm. per 100 cc.	mgm. per 100 cc.	mgm. per 100 cc.	mgm. per 100 cc.	mgm. per 100 cc.	
57.	Arsenical hepatitis	M	39	26	-60		İ		
58.	Carcinoma of stomach	M	61	26	76		l		
59.	Senile cataract	F	63	25	46	80			
60.	Diabetes mellitus	F	60	24	70	94	118	165	
61.	Chronic myocarditis, hyper-								
	tension	M	47	24	76		İ		
62.	Senile cataract	F	74	24	57	68	102*		
63.	Choroiditis	F	65	24	65	84	126*		
64.	Lymphocytic leukemia	F	24	23	64				
65.	Pulmonary tuberculosis, tu-								
	bercular peritonitis	M	31	23	54				
66.	Lymphocytic leukemia	M	20	23	64	126*			
67.	Polycythemia vera	М	44	22	62				
68.	Syphilis	M	63	22	44	65*			
69.	Myelogenous leukemia	F	66	22	79				
70.	Senile cataract	М	70	21	71	95	129*		
71.	Pernicious anemia	F	27	21	42				
72.	Pernicious anemia	м	68	21	71				
73.	Gangrene toe	М	50	20	64		117*		
74.	Gangrenous appendix, pelvic								
	abscess	м	47	20	85	126*			
75.	Latent syphilis, peptic ulcer	М	32	18	63				
76.	Diabetes mellitus	M	34	18	60	113	138	170	
77.	Adenoma of thyroid	F	32	18	85		138*		
78.	Chronic appendicitis	F	25	16	43	75	107	138*	
79.	Cardiac decompensation	м	43	15	47	101	140*		
80.	Diabetes mellitus, cardiac de-	-"-							
۱	compensation, chronic myo-		- 1	l					
	carditis	М	43	11	61	98	128*		

^{*} Initial blood sugar: the blood was completely glycolyzed when analyzed at the end of this period.

Stained smears were made from each specimen of the blood at the end of each experiment, and in no case were microorganisms found. Complete blood counts were obtained on the day of the study.

RESULTS AND DISCUSSION

The rates of glycolysis in 80 specimens of blood from patients with widely different diseases are recorded in Table I. It will be observed

that the amount of sugar glycolyzed during a two hour period of observation varied from 11 mgm. to 98 mgm. per 100 cc. of blood. The variation in the glycolytic activity of different bloods is more evident in this table than in previously reported data. There are instances, notably those reported by Falcon-Lesses (3) and by Schmitz and Glover (4), in which a large amount of sugar was glycolyzed. Likewise there are observations in which the rate of glycolysis was very low (Birchard (5), White and Watson (6), John (7), and Lemann and Liles (8)). There is in this study no correlation of the age or sex of the patient and the rate of glycolysis.

An examination of the methods used will do much to explain the variation in results obtained by different investigators. The most important cause for the diversified results in glycolytic studies has been the use of anticoagulants. Bürger (9) has demonstrated that the addition of citrate or oxalate affects the rate of glycolysis. Data obtained in this laboratory showed further that the glycolytic power of blood is retarded in proportion to the concentration of citrate or oxalate. Fluoride arrests glycolysis, as has been demonstrated by Major (10), and by Dickens and Simer (11). Hence all data obtained with blood to which oxalate, citrate or fluoride has been added cannot be compared with results obtained without the use of these anticoagulants. Defibrination or heparinization of blood does not affect the rate of glycolysis (3) (4) (12).

Of other extrinsic factors which might influence the rate of glycolysis, temperature is of great importance. Claude Bernard (1) employed a temperature of 15° C., Lemann and Liles (8) kept their specimens of blood between 9° and 11° C., Denis and Giles (13), Mauriac (14), and Stammers (15) all used "room temperature." Thalhimer and Perry (16), Cajori and Crouter (17), Katayama (18), Negelein (19) and Mackenzie (20) used a temperature of 37–38° C. Bürger (9) demonstrated that the optimum temperature for glycolysis was 37° C., an observation which has been confirmed in this laboratory. It is to be regretted that the data on the rate of glycolysis given by different investigators often cannot be compared because standard temperature conditions were not maintained.

The influence of the initial sugar concentration on the rate of glycolysis has been the subject of much discussion. Some investigators (5) (7) (8) (21) have taken as the rate of glycolysis the percentage of the initial sugar content which is glycolyzed in two hours. This is arbitrary and illogical since the absolute rate of glycolysis does not appear to be dependent upon the initial sugar concentration. In the author's blood specimens the initial sugar content varied from 65 mgm. per 100 cc. (Case 68) to 1084 mgm. per 100 cc. (Case 37), but the absolute rate of glycolysis in the blood of the latter patient with its excessive amount of sugar was well within the limits of normal for this series. Further, it was observed (Table II) that the rate of glycolysis

TABLE II

Glycolysis of excessively high blood sugar

(Case H. M.)

Hours	Blood sugar	Sugar glycolyzed
	mgm. per 100 cc.	mgm. per 100 cc.
. 0	800	0
2.0	725	75
4.0	655	145
6.0	585	215
10.0	558	242
18.5	455	345
23.0	312	488
41.5	120	680
65.5	45	755
72.0	. 0	800

continues to be about the same for six hours, even though the concentration of sugar is decreasing steadily. The decrease in the rate of glycolysis which occurs after six hours is probably due to mechanical and chemical damage to the blood cells. Cajori and Crouter (17) and Macleod (22) have demonstrated that exogenous glucose is glycolyzed at the same rate as endogenous blood sugar. This fact has been confirmed repeatedly in this laboratory. As additional proof of the lack of influence of the initial blood sugar content on the rate of glycolysis Table III is inserted to call attention to the glycolytic rate of the blood of a patient with diabetes mellitus both before and during management by diet and insulin.

The effect of insulin on the rate of glycolysis is also of interest. Three cases are presented (Table IV) to demonstrate the effect of insulin. In these cases, blood was withdrawn from the vein of one

TABLE III

Glycolysis before and during diabetic management
(Case D. F. B.)

Hours	Before diab	etic management	During diabetic management			
	Blood sugar	Sugar glycolyzed	Blood sugar	Sugar glycolyzed		
	mgm. per 100 cc.	mgm. per 100 cc.	mgm. per 100 cc.	mgm. per 100 cc.		
0	372	0	277	0		
2	341	31	239	38		
4	300	72	198	79		
6	265	107	140	137		
8	197	175	118	159		
10	120	252	73	204		
12	102	270	34	243		
14	67	305				

arm and, without removing the needle, insulin was injected directly into the vein. After three minutes blood was withdrawn from the vein of the other arm. Glycolysis was allowed to proceed in the usual way. It will be observed that the rates of glycolysis are parallel and that there is no indication of an increased glycolytic rate due to the direct action of insulin when it is added to the blood in this manner. These results are in accord with the work of Eadie, Macleod and Noble (23), who found no evidence of a modified glycolytic rate in the blood of dogs drawn before and after the subcutaneous administration of Raab (24), Bierry, Rathery and Kourilsky (25), Ducceschi (26), Hepburn and Latchford (27), and Lemann and Liles (8) noted no change in the amount of glycolysis after insulin. Mauriac and Aubertin (28) concluded that insulin acts without changing the glycolytic power of the tissues. The authors' results do not support the work of Achard (29), who maintained that insulin acts by decreasing hyperglycemia and increasing the rate of glycolysis, nor of Thalhimer and Perry (16), who stated that the amount of glycolysis varies directly with the administration of insulin.

Under standard conditions of technique there are marked variations in the rate of glycolysis, as is evident from a study of Table I. The plasma is the only blood element which is incapable of displaying any glycolytic activity; if incubated under sterile conditions for several days, its original sugar content remains unchanged. Plasma is not

TABLE IV Influence of insulin on glycolysis

Case L. P.	After insulin	Blood Sugar sed sugar glycolyzed	er mgm. per mgm. per	277	257	212		135 142				
ű	Before insulin	Blood Sugar sugar glycolyzed	mgm. per mgm. per 100 cc.		257 18	215 60		137 138				
	After insulin	Sugar	mgm. per 100 cc.	0		20	110	125	155	190	233	
Case B. J.	After	Blood	mgm. per 100 cc.	350		285	225	210	180	145	102	
Case	Case Before insulin	Sugar glycolyzed	mgm. per 100 cc.	0		33	8	118	138	175	218	
	Before	Blood sugar	mgm. per 100 cc.	320		287	230	202	182	145	102	
	After insulin	Sugar glycolyzed	mgm. per 100 cc.		32	85	110	175	257	270	300	
). F. B.	After	Blood	mgm. per 100 cc.	372	340	287	262	197	115	102	72	
Case D	Case D. F. B. insulin	Sugar	mgm. per 100 cc.	0	36	77	112	180	257	275	310	319
	Before insulin	Blood	mgm. per 100 cc.	377	341	300	265	197	120	102	29	28
	Hours			0	7	4	9	· •	10	12	14	16

essential as a carrier of glucose for the cells; the author has shown that Tyrode's solution may be substituted for it without affecting the glycolytic rate, and Kawashima (30) has substituted physiological salt solution and Locke's solution with equal success. Rona and Doblin (31) and von Noorden (32) attributed some glycolytic power to blood serum, but it is conceded by Irving (12), Milne and Peters (33) and Macleod (22) that neither blood serum nor plasma is able to glycolyze any of the sugar which they contain.

The erythrocytes are the cause for the disappearance of much of the sugar in vitro. Bloods containing large numbers of erythrocytes glycolyze sugar more rapidly than bloods containing fewer red blood corpuscles, other things being equal. The results of a typical observation of the rate of glycolysis in a normal blood and in the same blood when the cellular elements are diluted with its own serum are presented in Table V. In Table VI the glycolytic rates of anemic bloods

TABLE V

Effect of diluting blood on glycolysis

(Case H. B.)

		i, 46 per cent per cent plasma	"Modified" bl corpuscles, 70	ood, 30 per cent per cent plasma	"Modified" blood, 23 per cent corpuscles, 77 per cent plasma		
Hours	Blood sugar	Sugar glycolyzed	Blood sugar	Sugar glycolyzed	Blood sugar	Sugar glycolyzed	
	mgm. per 100 cc.	mgm. per 100 cc.	mgm. per 100 cc.	mgm. per 100 cc.	mgm. per 100 cc.	mgm. per 100 cc.	
0	155	0	159	0	186	0	
2	122	33	130	29	162	24	
4	77	78	96	63	137	49	

TABLE VI

Effect of concentrating anemic blood on glycolysis
(Case B. A.)

Hours	Whole blood, 2 79 per	1 per cent corpuscles, cent plasma	"Modified" blood, 39 per cent corpuscles 61 per cent plasma				
	Blood sugar	Sugar glycolyzed	Blood sugar	Sugar glycolyzed			
	mgm. per 100 cc.	mgm. per 100 cc.	mgm. per 100 cc.	mgm. per 100 cc.			
0	136	0	132	0			
2	115	21	88	44			
4	94	42	51	81			

are compared with the rates observed in the same specimens after removal of enough serum to bring the cellular concentration approximately to normal. These experiments confirm Harrop's (34) demonstration that the erythrocytes have glycolytic power, and indicate that their concentration is important. These results are in accord with those reported by Kawashima (30) and by Doyon and Morel (35). Falcon-Lesses (3) and Cook and Somogyi (36) reached a similar conclusion after studies of the glycolytic rate in erythremia.

Not only is the concentration of erythrocytes a factor in glycolysis, but the nature of the red blood corpuscles is important. It has been demonstrated (37) that in diseases of the hematopoietic system characterized by great bone marrow activity and the releasing into the blood stream of great numbers of young and even immature red blood corpuscles there is a marked increase in the amount of sugar glycolyzed. In this connection it is interesting to note that Harrop (34), Morawitz (38) and Derra (39) have reported the increased use of oxygen by immature erythrocytes. No attempt was made in this study to correlate the rate of glycolysis and the oxygen consumption but, in view of the conclusions drawn by Glover, Daland and Schmitz (40) regarding the oxygen utilization and glycolytic rate of normal and leukemic leucocytes, such a study is desirable.

Of importance, also, in the study of glycolysis is the glycolytic power of the leukocytes. Although they exist in blood in relatively small numbers, they have a high glycolytic power, as may be demonstrated by the increased rate of disappearance of sugar accompanying slight Table I shows the high glycolytic rate characteristic leukocytosis. of leukemia. This observation has also been made by Glover, Daland and Schmitz (40), Falcon-Lesses (3), and Lepine and Boulud (41). However, as pointed out by Bürger (9), Falcon-Lesses (3) and Schmitz and Glover (4) in their observations of chronic myelogenous leukemia and lymphatic leukemia, the amount of sugar glycolyzed is not necessarily in proportion to the total number of leucocytes present. suggests that the various types of leucocytes have different glycolytic abilities, and that it is the young leucocytes (particularly the cells of the myelocytic series) which possess the greatest glycolytic power. This is revealed very clearly in Table I and has been emphasized by all investigators studying glycolysis in leukemic bloods. Lymphocytes, on the other hand, possess a lower glycolytic activity, as is demonstrated by the fact that the blood in lymphatic leukemia utilizes much less sugar per unit of time, although the number of white blood cells may be as great as in myeloid leukemia. Mauriac (42), Bürger (9), Macleod (43) and the author have shown that polymorphonuclear leucocytes are endowed with a much greater glycolytic power than mononuclear leucocytes.

It may be postulated that the endothelial cells have glycolytic ability, but these cells occur in the blood in such small numbers that they can be of very little importance in the utilization of sugar. Blood platelets which Warburg (44) states have a measurable oxygen consumption can scarcely be much of a factor in glycolysis, especially if they are effectively removed with the fibrin during defibrination as Harrop (34) believes.

SUMMARY AND CONCLUSIONS

- 1. A study of the rate of glycolysis in bloods from 80 different patients with widely different diseases is presented.
- 2. There is no correlation of the age or sex of the patients and the rate of glycolysis.
- 3. The glycolytic rate is markedly influenced by temperature. Glycolytic studies should be performed at 37° C.—the optimum temperature for the disappearance of sugar from blood in vitro.
- 4. Citrates and oxalates, when used as anticoagulants, retard glycolysis in drawn blood in proportion to the amount in which they have been added.

Fluorides arrest glycolysis.

Defibrination or heparinization does not interfere with the rate of glycolysis, and blood so treated should be used for glycolytic studies.

- 5. The rate of glycolysis bears no relationship to the initial glucose content of the blood.
 - 6. Glycolysis in vitro is not affected by insulin.
- 7. Both erythrocytes and leucocytes are responsible for the glycolytic power of blood, and their number, type, age, and physiological integrity are important factors.
- 8. Glycolysis is a complex process involving certain variables which must be controlled if the results are to be correct.

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