Mitochondria in lung disease

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Mitochondria are a distinguishing feature of eukaryotic cells. Best known for their critical function in energy production via oxidative phosphorylation (OXPHOS), mitochondria are essential for nutrient and oxygen sensing and for the regulation of critical cellular processes, including cell death and inflammation. Such diverse functional roles for organelles that were once thought to be simple may be attributed to their distinct heteroplasmic genome, exclusive maternal lineage of inheritance, and ability to generate signals to communicate with other cellular organelles. Mitochondria are now thought of as one of the cell's most sophisticated and dynamic responsive sensing systems. Specific signatures of mitochondrial dysfunction that are associated with disease pathogenesis and/or progression are becoming increasingly important. In particular, the centrality of mitochondria in the pathological processes and clinical phenotypes associated with a range of lung diseases is emerging. Understanding the molecular mechanisms regulating the mitochondrial processes of lung cells will help to better define phenotypes and clinical manifestations associated with respiratory disease and to identify potential diagnostic and therapeutic targets.

Introduction

Maternally inherited and thought to be of bacterial descent, mitochondria are iconic double-membrane structures full of convoluted cristae that are present in nearly all cells and possess their own genome, transcriptome, and proteome. Revolutionary studies of bioenergetics in the 1950s, 1960s, and 1970s deemed mitochondria the "powerhouses" of the cell; however, while mitochondria throughout our bodies produce the bulk of the ATP needed for cells to live, our understanding of mitochondrial biology has undergone major transformations since the mysteries of oxidative phosphorylation (OXPHOS) were unraveled decades ago (1). In the last decade, the arrival of state-of-the-art high-throughput genomics, metabolomics, and complementary discoveries in signal transduction pathways have helped rapidly expand the list of genes encoding mitochondrial proteins as well as identify metabolites that are linked to human disease. It is becoming increasingly clear that abnormal mitochondrial signatures and mitochondrial dysfunction underlie the pathological mechanisms behind a plethora of lung diseases, including but not limited to chronic obstructive pulmonary disease (COPD), asthma, and lung cancer. It is also becoming clear that alterations in the mitochondrial genome, proteome, and metabolome may act not only as independent pathological processes, but also synergistically with existing pathological mechanisms to predispose, promote, or exacerbate lung disease. Given that lung diseases are among the leading causes of death worldwide, with four respiratory disease categories appearing in the worldwide top ten causes of mortality, together accounting for one in six deaths globally, the need for novel diagnostic and therapeutic approaches is critical (2). This article will discuss the multiple regulatory facets of mitochon-

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drial function in normal lung homeostasis and the prominence of mitochondrial dysfunction in acute and chronic lung injury and in lung disease. It will also highlight the concept that mitochondria are no longer thought of as simple, discrete, kidney bean-shaped energy factories, but are now believed to encompass a cell- and tissue-specific, dynamic organellar network that fuses, divides, and directs a vast array of functions central to cellular life, death, and differentiation (1).

Bioenergetics and nutrient sensing

The mammalian lung is composed of over forty different cell types, which are regionally and spatially localized throughout the organ and contain varying levels of mitochondria (3). Functionally, nearly every cell in the lung depends on mitochondrial metabolic activities, requiring a constant supply of energy from OXPHOS. Mitochondria are at the hub of cellular metabolism, regulating the continuous aerobic oxidation of fatty acids (FAs) and consuming the end products of glucose, glutamine, and amino acid degradation in order to aerobically produce ATP from oxygen and H₂O (Figure 1). The oxygen consumption rate of the lung is comparable to the oxygen consumption rates of other organs, including the intestine, pancreas, and spleen, but it is considerably lower than that of the brain, heart, and kidney (4). Likewise, the ATP content of the lung is similar to that of other organs, such as the brain, liver, and kidney (5), and is mostly dependent on mitochondrial sources (4).

Lung mitochondria preferentially use glucose-derived substrates, such as pyruvate, for oxidative energy production; however, other energy sources, including FAs, Krebs cycle intermediates, glycerol-3-phosphate, and glutamate, are also used, with the highest O_2 consumption rates achieved with succinate as a substrate (6). Lung mitochondria also have a unique advantageous metabolic adaptation to aerobic OXPHOS, owing to the fact that the lung possesses its own isoform of electron transport chain (ETC) complex IV, cytochrome *c* oxidase (COX subunit IV-2), which is present in all

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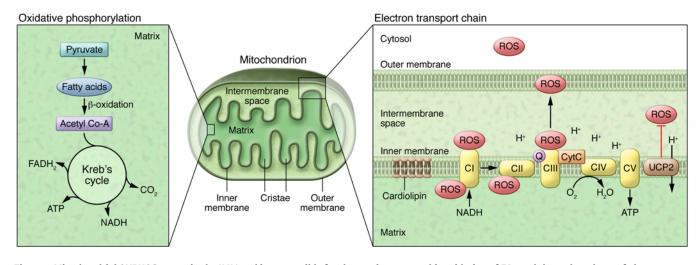


Figure 1. Mitochondrial OXPHOS occurs in the IMM and is responsible for the continuous aerobic oxidation of FAs and the end products of glucose, glutamine, and amino acid degradation in order to aerobically produce ATP from oxygen and H₂O.

lung cells, is oxygen sensitive, and renders lung COX two-fold more active (oxygen-binding) compared with COX in other tissues (7).

Type II alveolar epithelial cells (AECs), which continuously release surfactant by exocytosis onto the epithelial cell surface, have approximately three times greater mitochondrial volume per cell than other lung cells, such as endothelial or type I AECs (8). Bronchial (ciliated) epithelial cells, which require mitochondria for cilia beating, as well as vascular smooth muscle cells (VSMCs) and alveolar macrophages (AMs) are also rich in mitochondria (9). During differentiation from type II to type I as a part of the normal physiological replacement or repair mechanism, type II AECs reduce the number and size of mitochondria (10), resulting in type I AECs having a lower energy demand with less COX expression (11). Type II AECs rely on mitochondria to produce acetyl-CoA for de novo FA synthesis, allowing for the generation of phospholipids that are needed to produce pulmonary surfactants. Irregularities in phospholipid production, such as an excessive increase in cardiolipin or abnormalities in the carnitine acyltransferase pathway, disrupt surfactant composition and, consequently, lung function (12). Under altered physiologic states, such as starvation, type II AECs rely on FAs as a source of energy (13). High rates of FA synthesis correlate with morphological transformations in lung mitochondria, and under conditions of cellular stress, mitochondria of type II AECs transform into lamellar bodies to facilitate more surfactant production (14). Such alterations in mitochondrial bioenergetic metabolism allow lung cells to adapt to cellular stresses; however, excessive or prolonged modifications to these processes may be pathogenic to normal lung function and have profound effects on the clinical indices of many lung diseases, including COPD, pulmonary hypertension (PH), asthma, cystic fibrosis (CF), and lung cancer, as discussed below.

COPD is a debilitating lung disease encompassing airway inflammation (chronic bronchitis), destruction of lung tissue (emphysema), and remodeling of the small airways (15). The pathogenesis of COPD involves aberrant inflammatory and dysregulated cellular responses of the lung to cigarette smoke (CS) exposure. In lung epithelial cells, cytotoxic exposure to CS reduces mitochondrial OXPHOS (15, 16), whereas treatment with nontoxic doses of CS increases mitochondrial metabolic activity (17, 18), inducing a metabolic shift from glucose (glycolysis) to palmitate (β -oxidation) metabolism (18). Loss of acetyl-CoA and the Krebs cycle intermediate succinate is observed in basal cells of smokers (19). Additionally, human airway smooth muscle (ASM) cells (20) and quadriceps, diaphragmatic, and external intercostal muscle of COPD (21) patients display altered OXPHOS with increased COX activity (22, 23) (Table 1).

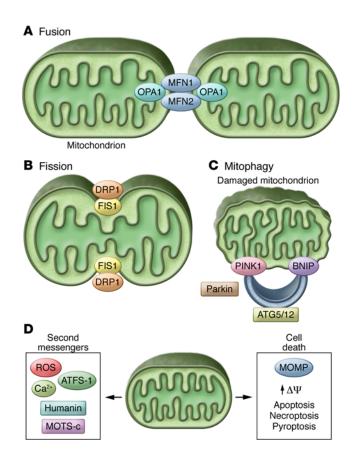
PH is characterized by obstruction of small pulmonary arteries, increased pulmonary arterial pressure, and lung vasculopathy. Vascular obstruction occurs as a consequence of excessive proliferation and apoptosis resistance of vascular cells. Chronic repression of mitochondrial metabolism, including decreased mitochondrial FA oxidation and transport (24), is associated with metabolic switching from mitochondrial-derived glucose oxidation to cytoplasmic-derived anaerobic glycolysis (25). The metabolic shift to anaerobic glycolysis contributes mechanistically to the apoptosisresistant, proliferative phenotype of PH. Such metabolic switching, a phenomenon also referred to as the Warburg effect (26), is also observed in non-small cell lung cancer (NSCLC), a type of lung cancer that accounts for 85% of all lung cancers and originates from bronchial/epithelial cells (27). In NSCLC, the switch from production of ATP via OXPHOS to anaerobic glycolysis is thought to be a more efficient way to produce ATP and other metabolic precursors in a hypoxic environment.

Oxygen-associated alterations in complex II and I in response to hyperoxia are also observed in models of bronchopulmonary dysplasia (BPD), a chronic lung disorder of infants and children who receive prolonged mechanical ventilation to treat respiratory distress syndrome (RDS) (28, 29). Similarly, asthma, a heterogeneous chronic inflammatory disease characterized by variable airway obstruction, airway remodeling, and bronchial hyperresponsiveness, is also associated with a reduction in OXPHOS, specifically decreased expression and activity of COX in bronchial epithelium (30), and increased Krebs cycle enzymatic activity in platelets (31). Absence of the lung-specific isoform of COX, COX4i2, results in

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Table 1. Mitochondrial bioenergetics in lung health and disease							
Name Bioenergetics/metabolism/	Function /OXPHOS	Role in the lung	Pathological effect	Disease association			
Aerobic glycolysis	Oxygen consumption ATP production mROS production	Used by lung epithelial cells Used by AMs for phagocytosis, specifically M2-polarized macrophages	Excess mROS Inflammation Cell death	Less mitochondrial-derived glucose oxidation in PH (25) CS reduces mitochondrial OXPHOS (15, 16) Decreased state 3 OXPHOS (142) in COPD patient quadriceps, but increased in the diaphragm and external intercostal muscles of COPD patients (23) Less OXPHOS in lung cancer cells (26) LPS stimulation reduces OXPHOS to aerobic glycolysis (84)			
Anaerobic glycolysis	Generation of ATP Production of lactate	Used by M1 macrophages Metabolic switching may alter lung cell phenotypes	Inflammation Acetyl-CoA accumulation Lipid accumulation	PH is associated with a metabolic switch to anaerobic glycolysis (25) COPD peripheral muscles undergo a shift from oxidative to glycolytic energy metabolism, whereas the opposite is observed in the diaphragm (143) Elevated concentrations of lactic acid in fibrotic lung tissue (144) Platelets from asthmatic individuals rely less on glycolysis (31) CF patient fibroblasts have increased activity of glycolysis (34)			
Complex I ETC	Electron transport Release 0 ₂ ⁻ into the mitochondrial matrix	Needed for efficient OXPHOS	mROS	Decreased in BPD models (29) Downregulated in CF (33) Downregulated in COPD (20)			
Complex IV (cytochrome <i>c</i> oxidase) ETC	Oxygen consumption Release O ₂ ⁻ into the mitochondrial matrix	Low expression in type I AECs (11) Lung-specific isoform (7) required for maximal airway responsiveness (7)	mROS Cell death	Increased in COPD patients (22, 23) Subunit IV increased in the lung epithelial cells from patients with idiopathic interstitial pneumonias (145) COX4i2 may be important in the pathogenesis of asthma (7) Absence of COX4i2 results in lung pathology that worsens over time with impaired airway constriction and reduced airway responsiveness (7) Reduction in bronchial epithelium in asthma (30) Increased by LPS (146)			
FA oxidation	Substrate for OXPHOS Needed to produce ATP	Used by type II AECs to make phospholipids for surfactant production Used by M2-polarized macrophages	High rates of FA synthesis in type II cells correlate with morphological transformations (13) Palmitate used by type II cells under altered physiologic states (13)	Decreased mitochondrial FA oxidation with associated intracellular lipid accumulation in PH (24) Decreased FA oxidation in CF (34) Increase in palmitic FA in COPD (147) CS increases palmitate (β-oxidation) metabolism (13) Increase in oleic and decreases in eicosapentaenoic and FAs in asthma (147)			
Krebs cycle	The oxidation of acetate to supply ATP for OXPHOS	Source of energy via the oxidation of pyruvate, FAs, and amino acids such as glutamine Intermediates are essential for anabolic and glutathione metabolism	LPS stimulation reduces the expression of Krebs cycle enzymes	Loss of acetyl-CoA and succinate is observed in basal cells of smokers (19) Platelets from asthmatic individuals have increased Krebs cycle enzymatic activity (31)			

Table 1. Mitochondrial bioenergetics in lung health and disease



reduced airway responsiveness and a lung pathology that worsens over time, thus highlighting the potential importance of COX4i2 in the pathogenesis of asthma (7). Loss of the complex I OXPHOS proteins CISD1 and MT-ND4 (32), decreased complex I activity (33), and decreased FA oxidation (34) are also associated with CF, a lethal autosomal recessive disease associated with abnormal transport of chloride and sodium ions across the epithelium, leading to viscous airway secretions (Table 1).

Mitochondrial dynamics and biogenesis

Changes in bioenergetic processes, such as those observed in the lung diseases described above, may alter mitochondrial shape, movement, and cellular interactions. Mitochondria form a dynamic interconnected intracellular network, changing cellular location via cytoskeletal motors and altering size and shape in response to the metabolic needs of the cell. Mitochondria undergo membrane remodeling through cycles of fusion and division (35); the balance of these processes controls mitochondrial structure and metabolism as well as the cell cycle and results in the intermixing of the mitochondrial population in the cell both during normal mitochondrial turnover in homeostatic physiology and in response to mitochondrial or cellular stress (35). Increased fusion or reduced fission promotes the formation of elongated mitochondrial networks, whereas increased fission or reduced fusion causes mitochondrial fragmentation. Cells that primarily use OXPHOS metabolism, such as type II AECs, have more fusion and more elongated mitochondrial networks (15), whereas the mitochondria in cells that are more glycolytic and less reliant on OXPHOS, such as lung microvascular endothelial cells, appear more punctate (36).

Figure 2. Mitochondrial fission, fusion, mitophagy, and cell death. Mitochondrial biogenesis and mitophagy allow cells to quickly replace metabolically dysfunctional mitochondria with fresh, undamaged organelles. (A) Mitochondrial fusion is mediated by the dynamin-related GTPases MFN1 and MFN2 at the OMM and by OPA1 in the IMM. (B) Mitochondrial fission requires the recruitment of DRP1 from the cytosol to receptors on the OIMM (FIS1, MFF, MID49, and MID51), which causes constriction of the mitochondria and eventual division of the organelle in two. (C) Metabolically active cells, such as type II AECs, have developed robust programs to maintain mitochondrial quality. Damaged or defective mitochondria are removed via mitophagy, which is regulated by PINK1, BNIP, Parkin, and ATG5/12. (D)Mitochondrial-derived second messengers trigger a series of stress response pathways that provide both short-term and long-term benefits in increased stress resistance and longevity. However, excessive activation of these pathways may ultimately become detrimental to the cell, leading to the activation of programmed cell death pathways, including apoptosis, necroptosis, and pyroptosis.

Mitochondrial fusion is mediated by the dynamin-related GTPases mitofusin 1 and 2 (MFN1/2) at the outer mitochondrial membrane (OMM) and by the dynamin-related protein optic atrophy 1 (OPA1) at the inner mitochondrial membrane (IMM). Mitochondrial fission requires the recruitment of dynamin-related protein 1 (DRP1) from the cytosol to receptors (mitochondrial fission protein 1 [FIS1], mitochondrial fission factor [MFF], and mitochondrial elongation factors 1 and 2 [MID51 and MID49]) (ref. 35 and Figure 2). Hyperfusion has been documented in COPD and in lung cancer (17, 18, 37, 38), whereas loss of fusion, specifically loss of MFN2, has been associated with lung cancer (38) and PH (39). The role of mitochondrial fission and fusion in lung cancer may be microenvironment specific; increased fission may allow cancer cells to proliferate rapidly and invade the surrounding tissue, while increased fusion may allow for cell survival during times of stress or drug toxicity (27). Similarly, mitochondrial distribution within lung endothelial cells influences mitochondrial signaling in PH with the perinuclear clustering of mitochondria associated with the regulation of hypoxia-sensitive genes (40).

Fission and fusion are intimately linked to the formation of new mitochondria and allow the cell to maintain a healthy pool of mitochondria. Mitochondria are not formed de novo, but mitochondrial biogenesis results from the growth and division of preexisting mitochondria (41). Mitochondrial biogenesis is regulated mostly at the transcriptional level and requires the coordinated expression of both nuclear-encoded and mitochondrialencoded proteins, including mitochondrial transcription factor A (TFAM), PPAR coactivator-1 α (PGC-1 α), AMPK, and nuclear respiratory factors 1 and 2 (NRF-1/2) (41). Mitochondrial biogen-

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Name	Function	Role in the lung	Pathological effect	Disease association				
Mitochondria dynamics, biogenesis, and mitophagy								
Fusion/fission	Elongated mitochondrial networks Efficient OXPHOS Exchange of mtDNA	Cells that primarily use OXPHOS metabolism, such as type II AECs have elongated mitochondrial networks (15)	Increased fission may allow cancer cells to proliferate rapidly and invade into the surrounding tissue, while increased fusion may allow for cell survival during times of stress or drug toxicity (27)	 Human lung cancers have decreased MFN2 and increased OPA1 and DRP1 (38) Activation of fission (148) and downregulation of fusion in PH (39) Increased fission and hyperfusion in COPD (15, 17, 18, 37) Perinuclear clustering of mitochondria associated with the regulation of hypoxia-sensitive genes in lung (40) 				
Biogenesis	Production of new mitochondria Activated in response to stress or environmental stimuli	Occurs in distal lung cells (42), in smooth muscle of small blood vessels, and in inflammatory cells of the alveolar region (9) during growth or times of high- energy demand or stress	Inducible process that rescues mice from lethal sepsis (44) PGC1- α and TFAM increased after <i>S. aureus</i> sepsis in the distal lung (44)	Increased in ALI, pneumonia, hyperoxia (43) Increased upon <i>S. aureus</i> -associated sepsis (44), may be associated with resolution of lung injury (42) Increased in bronchial smooth muscle remodeling in asthma (46) Increased in lung cancer (47) Reduced in COPD, which may be associated with a significantly lower body mass index and less muscle mass (48)				
Mitophagy	Removal of damaged mitochondria Isolation of mtDAMPs	Occurs in lung epithelial cells, fibroblasts, and AMs	Increased after <i>S. aureus</i> sepsis in the distal lung (44)	Low and defective mitophagy promote fibrosis (54, 55) Increased in COPD models and COPD patients (15, 50) Defective mitophagy lead to CS stress–induced lung cellular senescence (50, 53) PINK1-induced mitophagy associated with pulmonary vascular remodeling and PH (51, 114) <i>S. aureus</i> infection upregulates PINK1 to induce ALI (52)				

Table 2. Mitochondrial dynamics in lung health and disease

esis can occur in distal lung cells, including type II AECs (42), in small blood vessel SMCs, and in inflammatory cells of the alveolar region (9) and is thought to arise during growth, conditions of high-energy demand, or cellular stress. In the parenchyma, type II AECs initiate mitochondrial biogenesis during acute lung injury (ALI), pneumonia, hyperoxic lung injury (43), and Staphylococcus aureus-associated sepsis (44). ALI, acute respiratory distress syndrome (ARDS), and sepsis remain significant sources of morbidity and mortality in the critically ill patient population (45). ALI and ARDS result from the inflammatory response of the lung to both direct and indirect insults and are characterized by severe hypoxemia, hypercapnia, diffuse infiltration visible in the chest x-ray, and a substantial reduction in pulmonary compliance (45). Mitochondrial biogenesis is increased in bronchial smooth muscle remodeling in asthma (46) and in lung cancer; however, in lung cancer, it is unclear whether these alterations contribute to tumorigenesis or are a consequence of carcinogenesis (47). In contrast, loss of mitochondrial biogenesis is associated with COPD, which may be associated with a significantly lower body mass index and lower muscle mass (ref. 48 and Table 2).

Mitophagy

Metabolically active cells, such as type II AECs, have developed robust programs of mitochondrial quality control consisting of mitochondrial biogenesis and mitochondrial removal. Damaged or defective mitochondria are removed by selective encapsulation into double-membraned autophagosomes that are delivered to the lysosome for degradation, a process called mitophagy (15). To date, the best-documented mitophagy regulators are PTENinduced kinase 1 (PINK1), which is expressed at low levels in healthy mitochondria (mitochondria with normal mitochondrial membrane potential [$\Delta \psi$ m]), the BH-3 only BCL2 protein BNIP3, and the E3 ubiquitin ligase Parkin. When the $\Delta \psi$ m is low (i.e., under conditions of stress), damaged, depolarized mitochondria stabilize PINK1 or BNIP3, which accumulates on the OMM and recruits Parkin or the autophagy protein LC3B, respectively (15). Parkin ubiquitinates various OMM proteins, including MFN1/2, and recruits autophagosomes (Figure 2 and ref. 49). Mitochondrial biogenesis and mitophagy allow cells to quickly replace metabolically dysfunctional mitochondria before energy failure (43).

The role of mitophagy in lung disease is complex. In some cases, mitophagy-related processes appear to be pathogenic, whereas in others, these processes are protective. Specifically, COPD patients have increased mitochondrial fission and increased mitophagy (15). CS also induces PINK1/Parkin-associated mitophagy, which regulates mitochondrial ROS (mROS) production and cellular senescence in primary human bronchial epithelial cells (50), and CS-induced mitophagy regulates necroptosis in lung epithelial cells and in experimental COPD murine models (15). PINK1-induced mitophagy triggers pulmonary vascular remodel-

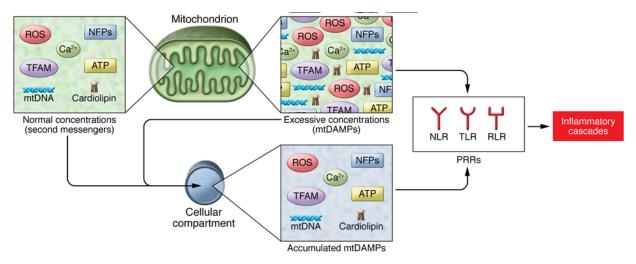


Figure 3. mtDAMPs. A wide variety of mitochondrial-derived molecules, which at normal physiological concentrations act as second messengers in the lung, can also behave as mtDAMPs when produced in excess or in an alternative cellular compartment. DAMPs primarily activate PRRs, including RLRs, TLRs, and NLRs, resulting in the induction of inflammatory cascades.

ing and PH (51), while S. aureus infection increases PINK1 to induce ALI (52). Conversely, defective mitophagy leads to CS-induced cellular senescence in human lung fibroblasts and small AECs (53), and loss of PINK1 and defective mitophagy promote pulmonary fibrosis (PF) in animal models and in human idiopathic pulmonary fibrosis (IPF) (54, 55). PF is characterized by irreversible destruction of lung architecture, abnormal wound healing, and deposition of extracellular matrix (ECM) proteins, leading to disruption of gas exchange and death from respiratory failure. Lung fibrosis is either idiopathic (54) or arises from exposure to environmental toxins, such as fibers, asbestos, metals, pesticides, chemotherapeutic drugs, viruses, or radiotherapy. While the pathogenic role of mitophagy in lung disease is perplexing, the differential role of mitophagy in specific cell types in the lung and in diseases such as COPD and IPF may help to explain the differences in obvious clinical, radiological, and pathologic features and may offer novel routes for therapeutic intervention or biomarker development.

Genetic regulation

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Mitochondrial biogenesis and mitophagy also allow cells to quickly replace or segregate mitochondria with pathogenic damaged mitochondrial DNA (mtDNA) away from the rest of the cell. Mammalian mtDNA encodes 37 genes, 24 of which are dedicated to processing 13 key genes essential to OXPHOS and energy production, while others encode transfer RNAs (tRNAs) and rRNAs essential for expression of these genes (35). Until a few years ago, it was assumed that mtDNA was homoplasmic for a single mtDNA genotype. However, with the arrival of deep sequencing, it has become clear that lowlevel heteroplasmy in mtDNA is present in most tissues, including the lung (56), where a portion is maternally inherited and another portion is presumed to arise from de novo acquired mutations (35). mtDNA is 3 to 10 times more susceptible to oxidative DNA damage than nuclear DNA (43). Chronic oxidation of over 50% of mtDNA without prompt repair results in a reduction of mtDNA copy number, loss of OXPHOS, and altered mitochondrial dynamics (43). Such persistent mtDNA damage is lethal to some cell populations, whereas oxidant-induced alterations in mtDNA/protein stability may acutely influence the behavior of other cell populations, thereby facilitating cellular adaptation (43). Pathogenic defects in or loss of mtDNA is associated with a number of lung diseases, including COPD (57), PH (58), lung cancer (59), asthma (60), and IPF (ref. 61 and Table 3). Asthma is not considered a mitochondrial syndrome per se; however, maternal inheritance is a risk factor for asthma and other atopic diseases (62), and mitochondrial haplogroups are associated with increased serum IgE levels (63, 64). Similarly, inherited mtDNA haplotypes may also predispose or confer susceptibility to COPD (ref. 65 and Table 2).

Second messenger signaling

Recent technological advances have identified hundreds of mitochondrial proteins that vary in a cell-specific and tissuedependent manner; however, the biochemical functions of the majority of these proteins remain unknown (1). With the other 1,500 or so proteins that make up the mitochondrion encoded by the nucleus, mitochondria have developed a symbiotic codependency on the nuclear genome and must signal to it in a retrograde fashion to ensure survival and adaptation. The best example of such signaling is the generation of mROS where, under normal physiological conditions, basal ROS released from the mitochondria acts as a second messenger to maintain cellular homeostasis (66). mROS production is tightly regulated by the ETC and antioxidant systems in mitochondria and is generated by a one-electron reduction of molecular O, to yield superoxide (O₂) (ref. 43 and Figure 1). mROS production leads to translocation of NRF2, resulting in expression of antioxidant and antiinflammatory proteins, such as the mitochondrial sirtuins (SIRT3, -4, and -5), which directly regulate expression of antioxidant genes. Genetic ablation of Nrf2 enhances susceptibility to CS-induced emphysema (67) and bleomycin-induced PF (68) in mice. SIRT5 is upregulated by CS in lung epithelial cells (69), SIRT3 regulates cell proliferation and apoptosis in NSCLC cells (70), and Sirt4 knockout mice spontaneously develop lung tumors (71), suggesting a key role for mitochondrial antioxidant systems in lung disease.

Table 3. mtDAMPs in the lung						
Name mtDAMPS	Function	Role in the lung	Pathological effect	Disease association		
mROS	Second messenger Regulates antioxidant responses, DNA damage responses, iron metabolism, cell proliferation, survival, and differentiation	Oxidative burst in AMs	Activates the NLRP3 inflammasome	Increased in ALI (40) Increased mROS in the PF (54) Increased in COPD (20) Increased in asthma (104) Increased in PH (108) Increased in lung cancer (106)		
ATP	Energy Coenzyme	Optimizes airway surface layer hydration, mucus composition, and mucociliary clearance (93)	Activates P2X7 to induce IL-1β (94); ATP levels increased in the BALF of mice treated with bleomycin (97)	ATP levels in BALF are elevated in patients with COPD (95) Increased levels of extracellular ATP are observed in the BALF of asthmatics and in mice sensitized with OVA and alum (96)		
NFPs	Share similarities with bacterial NFPs and are potent immune system activators (99)		Chemoattractants for neutrophils	Induces severe hypotension in rodent models (99) May link among trauma, SIRS, and cardiovascular collapse (99)		
Calcium	Second messenger		Several NLRP3 inflammasome activators mobilize Ca ²⁺	Increased Ca²+ (78) in PH Altered calcium secretion and homeostasis in CF (33)		
Cardiolipin	Mitochondrial-specific phospholipid Tethers members of the ETC to the IMM	Activates inflammasome- mediated immune responses (100) and generates lipid mediators after ALI (12)	In extracellular space directly, activates inflammasome (100) Externalization during mitophagy (49)	Elevated in BALF of individuals with pneumonia (12) Intratracheal injection of cardiolipin results in lower lung compliance with higher elastance and resistance (12) LYCAT altered in IPF (102). Cardiolipin elevated in smoking COPD patients (101) Generates lipid mediators during ALI (12)		
mtDNA	24 genes dedicated to processing 13 key genes essential to OXPHOS and energy production Cytoplasmic and extracellular danger signal	Activates the NLRP3 inflammasome intracellularly (86, 88) and activates neutrophils, vascular endothelial cells (90), and AMs extracellularly (40, 91)	Stimulates TGF-β1 from AECs (92) Activates the NLRP3 inflammasome (86, 88) and cGAS (89)	Circulating mtDNA as plasma biomarker in medical ICU patients (45) mtDNA increased in a human IPF (61) and murine PQ model of PF (92) mtDNA haplogroups confer susceptibility to COPD (65) mtDNA haplogroups influence atopic diathesis (64) mtDNA abnormalities and asthma (60, 62, 63) mtDNA mutations and PH (58) mtDNA mutations and lung cancer (59)		
TFAM	Mitochondrial transcription factor Mitochondrial structural protein	Increased in response to stress in the lung (9, 42)	Increased by LPS (146) Intravenous treatment of TFAM increases inflammation in rat lung (149)	Decreased in the lung tissues in COPD (48) Increased after <i>S. aureus</i> sepsis (44)		

Table 3. mtDAMPs in the lung

Mitochondria may also actively regulate homeostasis at the cellular and organismal level via peptides encoded within their genome (72, 73) or via the mitochondrial unfolded protein response (UPR) pathway (74); however, little is known about the function of these processes in the lung. Mitochondria are key regulators of the second messengers calcium (Ca²⁺) and iron (Fe), which control a diverse range of cellular processes, including mROS production. The formation of mitochondrial iron-sulfur (Fe-S) clusters is essential for many of the ETC complexes (complexes I and II) and other enzymes important for mitochondrial metabolism (75). Similarly, Ca²⁺ mobilization and the activation of Ca²⁺-binding proteins con-

trol a diverse range of cellular processes, including mitochondrial biogenesis (76). Loss or overload of mitochondrial Fe or Ca^{2+} can lead to mitochondrial dysfunction (77) and is associated with PH (76, 78), NSCLC (77), asthma (78, 79), and CF (33).

The activation of mitochondrial-derived second messengers may trigger a protective or hormetic response that provides both short-term benefits and the potential for long-term benefits in increased stress resistance and longevity (80). However, excessive activation of these pathways may ultimately become detrimental to the cell. Mitochondria consistently play a vital role in stress responses and programmed cell death pathways. The decline of $\Delta \psi m$, constitutive opening of mitochondrial pores, arrest of OXPHOS, interruption of mitochondrial protein import, and leakage of cytochrome *c* into the cytoplasm have all been associated with cell death pathways. Mitochondria regulate four forms of cell death, including (a) extrinsic apoptosis, (b) intrinsic apoptosis, (c) necrosis/necroptosis (15), and (d) pyroptosis, all of which have been documented in lung cells in various models of lung disease (15, 81–83).

mtDAMPs

A wide variety of mitochondrial-derived molecules, which at normal physiological concentrations act as second messengers in the lung, can also behave as mitochondrial damage-associated molecular patterns (mtDAMPs) when produced in excess or in an alternative cellular compartment. DAMPs arise from endogenous molecules secreted or released from intracellular or extracellular sources as a result of tissue injury and primarily activate pathogen recognition receptors (PRRs), including retinoic acid inducible gene-like (RIG-1-like) receptors (RLRs), TLRs, and nuclear oligomerization domain-like (NOD-like) receptors (NLRs), resulting in the induction of inflammatory cascades (84). Oxidized, fragmented mtDNA released from damaged mitochondria in response to stress or injury (85, 86) is one of the most important mtDAMPs needed to regulate innate immunity (Figure 3). Oxidized mtDNA is thought to act as a sentinel molecule in the cell, such that before an externally applied oxidant stress rises to a level that threatens the nuclear genome with mutation, oxidative mtDNA damage triggers death of the affected cell and promotes the propagation of signals to alert neighboring and roaming cells (43). We have previously shown that mtDNA released from dysfunctional mitochondria in response to stress and/or infection activates the NLRP3 inflammasome (86) and that circulating extracellular mtDNA is associated with mortality in medical intensive care unit (ICU) patients (45). Others have replicated these findings (87, 88) and shown that mtDNA also activates the DNA sensor cyclic GMP-AMP synthase (cGAS) (89) intracellularly and activates neutrophils (87), vascular endothelial cells (90), and AMs extracellularly (91). Injection of mitochondrial lysates in the rat causes lung inflammation (87), and mtDNA released as a result of injury in murine models of PF stimulates TGF-\u00df1 release from AECs (92). These findings suggest that mtDNA is a fundamental signaling molecule in the lung for the regulation and initiation of inflammation (Table 3).

ATP can also act as a mtDAMP in the lung. Normal lung function requires ATP release by AECs to optimize airway surface layer hydration, mucus composition, and mucociliary clearance (93). However, excessive ATP release from dying or damaged cells acts as a DAMP and is recognized by the purinergic receptor P2X7, which is expressed specifically on cells of the immune system and is involved in the release of IL-1 β (94). ATP levels are increased in bronchoalveolar lavage fluid (BALF) of COPD (95) and asthma patients as well as in murine asthma (96) and PF (bleomycin) models (97), all of which are associated with increased inflammation. TFAM is an integral regulator of mtDNA integrity that, when released from mitochondria, acts as a mtDAMP to regulate inflammatory responses (98). Similarly, *N*-formyl peptides (NFPs) originating from mitochondrial proteins act as chemoattractants for neutrophils and may link trauma, systemic inflammatory response syndrome (SIRS), and cardiovascular collapse (99). SIRS, a leading cause of death in ARDS patients, is a nonspecific inflammatory state caused by ischemia, inflammation, trauma, infection, or a combination of insults that triggers the release of inflammatory mediators from damaged lung tissue. Cardiolipin (Figure 1), which tethers members of the ETC to the IMM, directly activates inflammasome-mediated immune responses (100) and generates lipid mediators during ALI (12), when it is released into the cytosol or extracellular space. Cardiolipin concentrations are also elevated in BALF of individuals with pneumonia (12) and in smoking COPD patients (101), and mice given intratracheal injection of cardiolipin display lower lung compliance with greater elasticity and resistance (12). Finally, lysocardiolipin acyltransferase (LYCAT), a cardiolipin-remodeling enzyme, is significantly altered in lung tissues from patients with IPF (ref. 102 and Table 3).

mROS is the most universal and best-documented mtDAMP. The exact source of mROS may change with the mitochondrial stressor and particular disease state, but investigations into the source of mROS in many lung diseases is limited by the lack of highly specific in vivo mROS-sensing agents (103). That said, there is compelling evidence for a role of mROS in the pathogenesis of a number of lung diseases, including PF (54), COPD (20, 50), asthma (104), CF (105), lung cancer (106), BPD (107), and PH (108).

Mitochondria and inflammation

Lung inflammation is caused by pathogens or by exposure to toxins, pollutants, irritants, and allergens. The innate immune response of the lung relies on resident AMs for the detection of infectious agents, cellular stresses, or tissue damage. Bacterial or viral constituents as well as DAMPs secreted by lung epithelial cells ligate PRRs on AMs, causing AMs to secrete proinflammatory cytokines that activate alveolar epithelial receptors, which leads to recruitment of activated neutrophils. Mitochondria play a key role in the correct functioning of immune cells in the lung, including AMs and CD4+ Th2 and CD8+ (T cytotoxic) cells. Proinflammatory M1 macrophages exhibit robust glycolysis, and profibrotic/antiinflammatory M2-polarized macrophages increase oxygen consumption through the induction of mitochondrial biogenesis and FA oxidation (109). Mitochondria regulate mROS and AM phagocytosis in response to bacteria (110). Mitochondrial proteins and metabolites also interact and regulate TLR signaling (84), directly linking the activation of the downstream proinflammatory cytokines IL-1 β , IL-6, and TNF- α to mitochondrial function.

The IMM ETC uncoupling protein 2 (UCP2) regulates NLRmediated caspase-1 activation through the stimulation of lipid synthesis in macrophages with implications for sepsis-induced lung injury (111). Activation of NLRP3 signaling, a key regulator of IL-1 β and IL-18 secretion, is regulated by mitochondrial pathways (84, 112), and the NLRP3 inflammasome adaptor (ASC) and UCP2 regulate hypoxia-induced PH in mice (51, 113, 114). NLRP3 inflammasome activation by mROS in bronchial epithelial cells is required for allergic inflammation (104) and in AMs contributes to mechanical stretch-induced lung inflammation and injury (115). Caspase-1-dependent IL-1 β secretion is critical for host defence against *Chlamydia pneumoniae* lung infection (116), and mitochondrial Ca²⁺-dependent NLRP3 activation exacerbates the *Pseudomonas aeruginosa*-driven inflammatory response in CF (117).

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NLRP3 signaling is also regulated by the mitochondrial RLR signaling protein MAVS. RLR signaling resulting in the production of type I IFNs and other proinflammatory cytokines that promote adaptive antiviral immunity is regulated by mtDNA and a number of mitochondrial proteins (84, 118). AMs detect respiratory syncytial virus (RSV) via MAVS, and loss of MAVS may underlie the development of RSV-induced severe lung inflammation (118). MAVS also regulates the response of lung mast cells to influenza A virus (IAV) (119) and may be responsible for CS enhancement of virus-induced pulmonary innate immune and remodeling responses in mice (120).

Mitochondria also play a role in the adaptive immune response of lymphocytes in the airway and lung parenchyma. Specifically, mitochondria regulate antigen processing and presentation and localize to the immune synapse during T cell activation (121). Mitochondrial metabolism also maintains the memory T cell phenotype (122) and dictates the different inflammatory and suppressive CD4⁺ Th cells (109). A balanced Th1 and Th2 response is suited to the immune challenge, and a dysregulated response is linked to a variety of chronic inflammatory lung conditions such as asthma and chronic bronchitis (123). Nuclear factor of activated T cells (NFAT) regulates mitochondrial remodeling and contributes to apoptosis resistance in PH and cancer (124), and mitochondrialderived proteins modulate BALF eosinophilia by regulating both eosinophil apoptosis and Th2-type cytokine production (125).

Reciprocally, inflammatory pathways also regulate mitochondrial function (84). Bacterial pathogens hijack the mitochondrial cell death machinery of host cells, and viral infection alters the mitochondrial proteome, increases mROS and mitochondrial biogenesis, and attenuates mitochondrial lipid β -oxidation (84). Examples of such modulation in the lung include the following: influenza infection increasing long-chain acylcarnitine secretion from mitochondria, which in turn inhibit the surface adsorption of pulmonary surfactant, thereby increasing the risk for lung injury (126); LPS activating MAPK kinase 3 (MKK3), which regulates mitochondrial biogenesis and mitophagy in sepsis-induced lung injury (127); *S. aureus* decreasing cardiolipin availability in pneumonia models (52); and RSV infection increasing lung mitochondrial bioenergetics (128).

Therapeutic targeting of mitochondria in lung disease

Given the central role of mitochondria and mROS in human disease, several natural antioxidants, such as vitamin C, vitamin E, and curcumin, have been investigated both in vitro and in vivo; however, most of these were found to be ineffective in attenuating mROS production in response to an environmental stimulus or in patients with lung disease (129–132). While there has been considerable advancement in the development of mitochondria-targeted small molecule antioxidants (3) and alternative approaches to targeting mROS (133–137) appear promising, such mROS-targeted therapeutic approaches must be used with caution. mROS behave as a cytoprotective agent that leaves cells less susceptible to subsequent perturbations. This response, termed mitohormesis, is being rapidly dissected in many model systems and must be considered in the design of all mitochondrial-targeted therapeutics for the treatment of lung disease (66). Other mitochondrial-targeted therapeutic strategies that could potentially be used to treat lung diseases include the use of metabolic modulating compounds such as dicholoroacetate (138), histone deacetylase inhibitors (139, 140), and fission inhibitors. Stimulating adaptive mitochondrial biogenesis and mitophagy may be a useful adjuvant therapy for ALI in sepsis (44), and transfer of mitochondria from bone marrowderived mesenchymal stem cells to injured alveolar epithelium may be beneficial in ARDS, asthma, or COPD (141).

Conclusion

Lung diseases are among the leading causes of death worldwide, with lung infections, lung cancer, and COPD together accounting for over 9.5 million deaths in 2008 (2). In the next two decades, it is predicted that the proportion of deaths and disability arising from chronic lung diseases will rise significantly, yet few advances have been made to effectively treat the majority of lung diseases, with suboptimal therapeutic options eliciting only modest improvements in disease symptoms. Mitochondrial dysfunction is rapidly advancing as a key pathological feature that appears early and consistently in the development of lung disease. It is clear that abnormal mitochondrial signatures, including metabolic switching, altered mitochondrial biogenesis and mitophagy, increased occurrence of mtDNA mutations, abnormalities in mitochondrial-derived signaling, and the activation of mtDAMPs, play a substantial role in a number of lung diseases. While the pathogenic role of each of these mitochondrial processes in the lung remains complex, precise differential mitochondrial signatures in specific lung cell types may help to elucidate the clinical, radiological, and pathological alterations associated with each disease. To be able to identify such disease-associated mitochondrial signatures, a rigorous mapping of mitochondrial turnover and dynamics, mitochondrial metabolic activity, mtDNA sequence heterogeneity, and profiling of lung-specific mitochondrial proteins represent potential approaches that may bear dividends for a better understanding of normal physiologic and pathologic processes in the lung. Mitochondria therefore offer promising potential targets in the search for novel diagnostics and therapeutics in lung diseases.

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