

Activation of Rat Liver Perisinusoidal Lipocytes by Transforming Growth Factors Derived from Myofibroblastlike Cells

A Potential Mechanism of Self Perpetuation in Liver Fibrogenesis

Max G. Bachem, Dieter Meyer, Ralph Melchior, Klaus-Martin Sell, and Axel M. Gressner

Department of Clinical Chemistry and Central Laboratory, Philipps-University, Baldingerstrasse, D-3550 Marburg, Germany

Abstract

Rat liver perisinusoidal lipocytes (PL) cultured on uncoated plastic transform spontaneously within 6–10 d to myofibroblastlike cells (MFBIC). Parallel to the transformation the TGF α - and TGF β 1-mRNA expression increased and was highest in MFBIC. Competitive radioligand binding assays demonstrated that in contrast to untransformed PL the MFBIC synthesize and secrete transforming growth factor (TGF)- α (15 fmol/cell per 24 h) and predominantly the latent form of TGF β 1 (0.2 fmol/cell per 24 h). Medium conditioned by MFBIC (MFBcM) significantly stimulated PL proliferation with little effect on PL proteoglycan synthesis. By transient acidification of the MFBcM, known to activate the latent form of TGF β 1, the stimulatory effect on PL proteoglycan synthesis was enhanced and furthermore PL transformation (measured by expression of iso- α smooth muscle actin and loss of retinylpalmitate) was accelerated. Preincubation of this medium with neutralizing antibodies to TGF β resulted in (a) the growth inhibitory effect was converted to a growth stimulation and (b) the stimulatory effect on proteoglycan synthesis was abolished. In summary our data indicate that progressive activation of PL on plastic (transformation to MFBIC) leads to an enhanced expression of the TGF α - and TGF β 1-mRNAs and secretion of the corresponding proteins. Medium conditioned by MFBIC stimulates proliferation, transformation, and PG synthesis of untransformed PL. These mechanisms are suggested to be relevant in self perpetuation of liver fibrogenesis. (*J. Clin. Invest.* 1992; 89:19–27.) Key words: lipocytes • proliferation • transformation • proteoglycans • transforming growth factors

Introduction

In experimental (1, 2) and in human alcoholic liver injury (3) perisinusoidal lipocytes (PL)¹ (fat storing cells, Ito cells, stellate

cells) have the ability to proliferate strongly and transform via transitional cells (1) into myofibroblastlike cells (MFBIC) (3) showing dispersed nuclear chromatin (4); hypertrophy of rough endoplasmic reticulum (5, 6); reduction of number and size of fat droplets; decrease of retinoids; and long cytoplasmic extensions (5–7). PL cultured on uncoated plastic materials lose their differentiated phenotype and transform within 1–2 wk to highly proliferative and activated MFBIC (6, 7). PL and to an even higher amount, their transformed counterpart produce significant quantities of connective tissue components (6–9) and interestingly, a pattern of proteoglycan (PG) and collagen similar to that found in fibrotic extracellular matrix (for review see references 7 and 9).

Studies from others and our laboratory have shown recently that secretions of activated Kupffer cells and platelets stimulate proliferation of PL in culture (10–13) and also the synthesis of collagens (13), proteoglycans (14), and hyaluronan (15) by these cells. Potential candidates of the Kupffer cell and platelet-derived fibrogenic activity seem to be transforming growth factor α /epidermal growth factor (TGF α /EGF) and TGF β (7, 16–18). It was shown recently that (a) TGF β stimulates collagen- (16, 19) and proteoglycan synthesis in PL (20), (b) in acute CCl₄ induced liver injury the level of TGF β -mRNA rises (21), (c) PL in culture express the TGF β gene (22) and furthermore (d) TGF β gene expression is significantly enhanced during active fibrogenesis associated with liver disease in man (23). These results indicate a central role of TGF β in hepatic fibrogenesis. Furthermore, it was shown that TGF β 1 inhibited PL proliferation in a dose-dependent manner (19, 24), but significantly stimulated chondroitin sulfate and hyaluronate synthesis in these cells (untransformed PL) (20).

The current study described cell-cell interactions via soluble mediators between MFBIC and untransformed PL. Our results indicate that MFBIC synthesize and secrete soluble mediators and stimulate thereby PL proliferation (TGF α), transformation (TGF β), and proteoglycan synthesis (TGF β) in a paracrine way.

Methods

Materials

Collagenase H (clostridiopeptidase A, EC 3.4.24.3), trypsin (EC 3.4.21.4), and iso- α smooth muscle actin antibodies and FCS were from Boehringer Mannheim, GmbH, Mannheim, Germany. TGF β 1 (source human platelets) and TGF α (synthetic rat) were purchased from ICN Biochemicals, Cleveland, OH. [¹²⁵I]EGF (100 μ Ci/ μ g) was from Amersham Buchler, Braunschweig, Germany. [³⁵S]Sulfate (364.4–505.4 mCi/mmol) was from New England Nuclear, Boston, MA. [6-³H]Thymidine (15 Ci/mmol) was from New England Nuclear-DuPont (Dreieich, Germany). [¹²⁵I]TGF β 1 (1 μ Ci/ng) was from Paesel & Lorei, Frankfurt, Germany. Nycodenz (analytical grade) was from Nyegaard and Co. AS, Oslo, Norway. Calf thymus DNA (type I) and

Address correspondence to Dr. M. G. Bachem, Abt. f. Klin. Chemie u. Zentrallaboratorium, Philipps Universität, Baldingerstrasse, D-3550 Marburg, Germany.

Received for publication 10 July 1990 and in revised form 6 August 1991.

1. Abbreviations used in this paper: MFBcM, myofibroblastlike cell conditioned medium; MFBIC, myofibroblastlike cells (transformed perisinusoidal lipocytes); PG, proteoglycans; PL, perisinusoidal lipocytes; TGF α , transforming growth factor alpha; TGF β 1, transforming growth factor β 1.

J. Clin. Invest.

© The American Society for Clinical Investigation, Inc.

0021-9738/92/01/0019/09 \$2.00

Volume 89, January 1992, 19–27

pronase were from Sigma Chemical Co., Munich, Germany. Protein A-purified neutralizing antibodies against porcine and human TGF β 1 were supplied by R&D Systems, Minneapolis, MN. [32 P]dCTP (3,000 Ci/mmol), [32 P]ddATP (5,000 Ci/mmol) and the Hybond N blotting membrane were from Amersham/Buchler (Dreieich, Germany). The TGF α -cDNA probe (sp 65C17N3) was kindly provided by Dr. R. Derynck (Genentech, Inc., South San Francisco, CA), and the TGF β 1-cDNA probe (pTGF β 33) by Dr. P. Kondiah, National Institutes of Health, Bethesda, MD (25). Antibodies against vimentin, desmin, and bromodeoxyuridine were from Dakopatts, Hamburg, Germany.

Methods

PL ISOLATION AND CULTURE

Detailed procedures of the isolation and culture of PL from rats have been published previously by this laboratory (10, 12, 14, 15, 26). The purity of the PL preparations attained > 80% (nonvital hepatocytes, Kupffer cells, and endothelial cells represented the remaining 20%) as assessed by light microscopy using the presence of perinuclear lipid droplets and the typical cell shape as a marker. Viability assessed by trypan blue exclusion was > 80%. With the first (~ 8 h after seeding) and second (~ 20 h after seeding) medium change most of the contaminating cells (nonvital hepatocytes and endothelial cells) were removed, and the PL monolayers were essentially (> 97%) free of impurities. With the third medium change (~ 44 h after seeding) FCS in DME was reduced to 0.5 or 2% and aliquots of the conditioned media were added. Control and treated cultures received the same amount of FCS. 24 h after the first stimulation media were added again during a further medium change whereby cultures were labeled with radionuclide.

CULTURE OF MYOFIBROBLASTLIKE CELLS

Confluent PL in primary culture (6–7 d after seeding) were trypsinized in the presence of EDTA (0.05% wt/wt trypsin, 0.2% wt/wt EDTA) and seeded in DME containing 10% FCS.

INDIRECT IMMUNOPEROXIDASE (IMMUNOFLUORESCENCE) STAINING

Cultures were washed twice with ice-cold PBS-Dulbecco, then fixed using alcohol/acetic acid 95:5 (vol/vol). After three washes with PBS-Dulbecco unspecific binding was blocked with 1:2 diluted calf serum (30 min, 22°C). The cells were next reacted with anti-iso- α smooth muscle actin (1:50) in PBS containing 1% BSA for 1 h at 22°C on a rotating platform. After three washes in PBS cells were reacted with peroxidase-conjugated anti-mouse IgG (1:50) in PBS containing 1% BSA or alternatively with fluorescein isothiocyanate-conjugated anti-mouse IgG (1:50) in PBS containing 1% BSA. After three washes in PBS the FITC treated cells were viewed under a microscope with an epifluorescence attachment. Cells treated with peroxidase-conjugated anti-mouse IgG were stained using 3,3'-diaminobenzidine tetrahydrochloride as substrate and hydrogen peroxide 0.001%, nickel chloride, and cobalt chloride in distilled water as intensifier. Thereafter, cells were washed with distilled water and then viewed under a microscope equipped with an optical grid and phase contrast. In cultures serving as negative control the first antibody was omitted.

PRODUCTION OF CONDITIONED MEDIA

After washing the near confluent monolayers with DME, fresh DME was conditioned for 24 h in the absence of FCS by subconfluent-confluent PL in primary culture (3, 6, and 10 d after seeding) and MFBIC (secondary culture after 1–3 passages). Conditioned media were removed sterilely, centrifuged (450 g, 7 min, 4°C) to remove cell debris and finally clarified by passing through a 0.45- μ m filter (Millex-GS; Millipore Corp., Bedford, MA). These media were used for all subsequent studies.

One half of the appropriate medium was dialyzed for 48 h against 100 vol distilled water at 4°C in tubings with a molecular cut-off 3.5 kD, lyophilized, and resolved in DME or binding buffer. The other half of the medium was acidified (pH 2.0) with 5 M HCl for 30 min at 4°C and neutralized with 5 M NaOH prior dialysis. In some experiments

the MFBcM was dialyzed against DME without lyophilization. Before the medium was added to the cultures it was sterilized by passing through a 0.22- μ m pore size filter (Millex-GS; Millipore Corp.).

DETERMINATION OF CELL PROLIFERATION

DNA measurement. DNA content of the cultures was measured fluorometrically using calf thymus DNA as a standard (27).

[3 H]Thymidine incorporation. Thymidine incorporation was measured as previously described (10, 12, 24) by TCA precipitation of cells that had been treated for 24 h with [3 H]thymidine (0.5 μ Ci/ml medium) beginning on the third day after seeding.

Bromodeoxyuridine incorporation. PL and MFBIC were seeded with a density of 10^4 cells/cm 2 in DME with 10% FCS in six well plates. 24 h after seeding cells were washed twice with DME and incubated for 48 h in DME alone or with 0.5, 2, and 10% FCS and for 24 h with bromodeoxyuridine (5×10^{-5} M final concentration). At the end of the culture period monolayers were washed twice with ice-cold PBS (pH 7.4) and then fixed with ice-cold ethanol/acetic acid 95:5 (vol/vol). Thereafter, the cells were incubated for 1 h at 21°C with antibromodeoxyuridine immunoglobulins (dilution 1/20 with PBS) and desoxyribonuclease (5 IU/ml) for DNA denaturation. After washing three times with PBS (pH 7.4) cells were incubated with peroxidase-conjugated rabbit anti-mouse immunoglobulins (dilution 1/50) for 2 h at 21°C. Thereafter cells were washed again three times with PBS and nuclei which had taken up bromodeoxyuridine were stained with DAB as substrate and hydrogen peroxide 0.001%, nickel chloride, and cobalt chloride in water as intensifier. Cells were counted using an Olympus phase contrast microscope equipped with a 1-mm 2 optical grid.

Cell numeration. Cells were counted after trypsination in a standard hemocytometer or alternatively using an Olympus phase contrast microscope equipped with a 1-mm 2 optical grid. With the second procedure cells were counted after fixation and indirect immunoperoxidase staining.

DETERMINATION OF THE SYNTHESIS OF TOTAL MEDIUM PROTEOGLYCANS

The synthesis of sulfated proteoglycans (PG) was determined by the incorporation of [35 S]sulfate (20 μ Ci/ml medium) during a labeling period of 24 h (details described elsewhere [12, 14, 20, 26]). Proteoglycan synthesis was expressed as radioactivity on the basis of DNA.

NEUTRALIZATION OF TGF β ACTIVITY

To bind and neutralize active TGF β 1 anti-TGF β IgG (2.5–50 μ g/ml medium) was added to transiently acidified MFBcM and incubated for 1 h at 21°C on a rotating platform. In a control experiment anti-TGF β 1 IgG was incubated with pure TGF β 1.

MOLECULAR HYBRIDIZATION AND BLOTTING TECHNIQUES

Total RNA from PL and MFBIC was isolated as described (28). For slot blot analysis, 20 μ g RNA was blotted on Hybond N membranes using a Millipore slot blot chamber. The measurement of ribosomal S6 gene expression was used as internal standard. The 0.8-kb probe for TGF β 1 was obtained by Sac I-Pvu II digestion of the pTGF β 33 plasmid, subsequent separation by gel electrophoresis, elution of the 0.8-kb fragment and labeling with (32 P)-dCTP. For hybridization of TGF α the 1.4 kb Eco RI fragment of sp 65C17N3 was used and labeled as described above. Hybridization was carried out in a buffer containing 50% formamide and Denhardt's solution at 42°C. Because TGF α -cDNA was a human probe, washing conditions were low stringent. The blots were exposed for 4–6 d at –80°C to Hyperfilm MP (Amersham Buchler, Braunschweig, Germany) using an intensifying screen.

COMPETITIVE RADIOLIGAND ASSAY TO QUANTITATE EGF/TGF α AND TGF β

Subconfluent MFBIC in six-well culture plates and skin fibroblasts in six-well culture plates were washed with binding buffer (128 mmol NaCl, 5 mmol KCl, 1.2 mmol CaCl $_2$, 1.2 mmol MgSO $_4$, 50 mmol Hepes, pH 7.5, and 5 mg/ml BSA) and incubated for ~ 30 min at 37°C

in binding buffer. Receptor binding was performed in 0.5-ml binding buffer per well. After equilibration for 10 min at 20°C appropriate concentrations of [¹²⁵I]TGFβ1 or [¹²⁵I]EGF, and either unlabeled peptide or dialyzed 10-fold concentrated conditioned media, were added. Incubations proceeded for 3 h at 20°C on a shaking platform and were terminated by aspiration of the binding buffer and washing the monolayers three times with ice-cold binding buffer.

Bound radioactivity was liberated with 1% Triton X-100 in PBS and measured in a gamma-counter. Nonspecific binding was determined at a 200-fold excess of cold ligand and subtracted from total binding. Nonspecific binding of [¹²⁵I]TGFβ1 never exceeded 10%, of [¹²⁵I]EGF 5% of total binding. All binding experiments were done in duplicate and repeated at least three times.

CELLULAR RETINYL-PALMITATE CONTENT

All the following steps were performed in the dark. PL cultures in six-well culture plates were washed twice in the dark with PBS. After washing, the retinoids were extracted with methanol/ethylacetate 1:1 (vol/vol) (1 ml per culture well, 2 h at 4°C). After centrifugation (10 min at 600 g) the transparent extraction fluid was used for HPLC. Retinyl-palmitate was determined in 100-μl aliquots by reversed phase HPLC using a RPC18 analytical column (Waters Associates, Milford, MA), developed at 1.3 ml/min for 30 min with methanol/ethylacetate/water 510:140:25 (vol/vol/vol) and fluorescence detection (Ex. 338 nm, Em. 425 nm). The retinyl-palmitate concentration was determined by comparing the peak area of the cell extracts with external standards of retinyl-palmitate in methanol/ethylacetate.

STATISTICAL ANALYSIS

Data are presented as mean±SD. Differences between groups were tested using variance analysis (Scheffé multiple range test). An α-level of *P* < 0.05 was considered statistically significant.

Results

Characterization of the PL in culture during transformation to MFBIC. The PL which were isolated from rat liver and seeded on uncoated culture wells had a purity of > 97% after the second medium change. The purity had been assessed by light-, fluorescence-, and electron microscopy, respectively. The relative amount of Kupffer cells which was assessed additionally by staining for endogenous peroxidase and by phagocytosis of latex beads was < 1% and decreased further during culture. Within the first 3 d after seeding the cells (untransformed PL) appeared homogenous and showed a characteristic morphology with lipid droplets surrounding the nucleus (Fig. 1 *a*). During the next several days the cells transdifferentiated spontaneously into MFBIC. This transdifferentiation was associated with a cell spreading, the development of long cytoplasmic extensions, a loss of fat droplets, and the appearance of one or two nucleoli (Fig. 1 *b*). Furthermore the desmin and iso-α smooth muscle actin expression (Fig. 1 *c*) increased. Transmission electron microscopy indicated that the size of the fat droplets de-

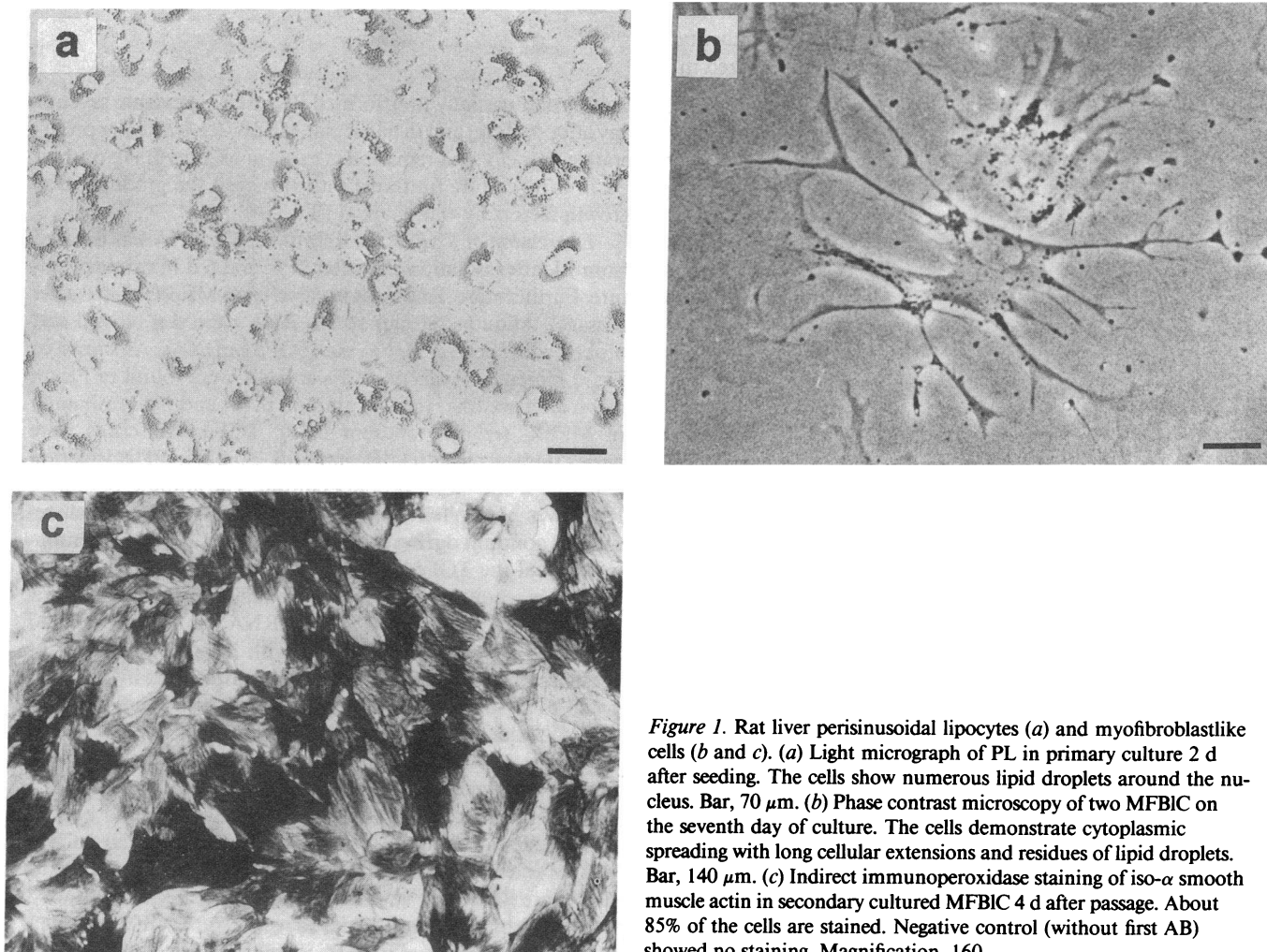


Figure 1. Rat liver perisinusoidal lipocytes (*a*) and myofibroblastlike cells (*b* and *c*). (*a*) Light micrograph of PL in primary culture 2 d after seeding. The cells show numerous lipid droplets around the nucleus. Bar, 70 μm. (*b*) Phase contrast microscopy of two MFBIC on the seventh day of culture. The cells demonstrate cytoplasmic spreading with long cellular extensions and residues of lipid droplets. Bar, 140 μm. (*c*) Indirect immunoperoxidase staining of iso-α smooth muscle actin in secondary cultured MFBIC 4 d after passage. About 85% of the cells are stained. Negative control (without first AB) showed no staining. Magnification, 160.

Table I. Effect of FCS on PL and MFBIC Proliferation

	DME 0 FCS	DME with 0.5% FCS	DME with 2% FCS	DME with 10% FCS
PL (3 d)				
(% BrdUrd +)	0.1±0.1	2.0±1.0	13±4.8	46±6.2
PL (7 d)				
(% BrdUrd +)	10±1.6	23±4.0	38±5.3	58±7.7
MFBIC				
(% BrdUrd +)	21±4.6	29±8.7	44±7.6	60±15.0

Rat liver PL and MFBIC were seeded in DME with 10% FCS (density, 10^4 cells/cm²) on 6er plates. In PL (3 d) and MFBIC fetal calf serum was reduced at the first day after seeding to the indicated concentrations (0, 0.5, 2, and 10% respectively); in PL (7 d) fetal calf serum was reduced at the fifth day after seeding. 1 d after the reduction of the FCS serum cells were labeled without a further medium change for 24 h with 5×10^{-5} M bromodeoxyuridine (BrdUrd). Thereafter cultures were stopped by fixation using alcohol/acetic acid (95:5, vol/vol). Cell proliferation was measured by indirect immunoperoxidase staining of nuclei which had incorporated BrdUrd using BrdUrd-mAb and peroxidase-conjugated anti-mouse IgG (both diluted with PBS 1/50). Cells were counted using a phase contrast microscope equipped with an optical grid. 10 10-mm² squares/culture were counted. Data represent mean±SD of cells which had incorporated BrdUrd (% BrdUrd +).

creased during transformation and that the MFBIC developed both, a hypertrophy of rough endoplasmic reticulum and a hypodense dispersed nuclear chromatin as signs of highly active protein synthesis. Observations which are summarized in

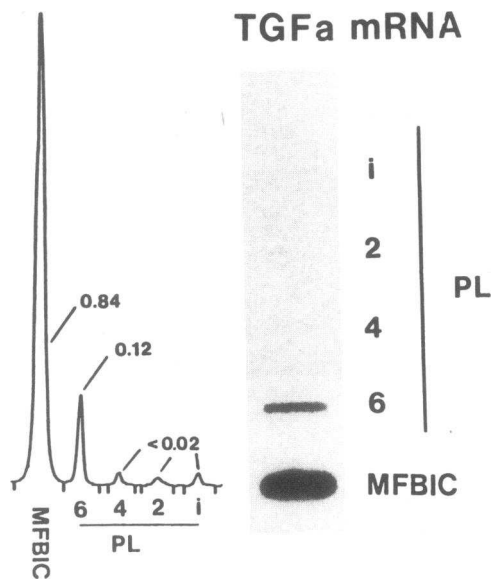


Figure 2. Expression of TGF α -mRNA in perisinusoidal lipocytes and myfibroblastlike cells. RNA was prepared from PL after isolation (i) and after 2, 4, and 6 d in primary culture and from MFBIC (6 d after passage). Aliquots (20 μ g) of the isolated RNA were slot blotted and probed with a human TGF α -cDNA. The relative amount of TGF α -mRNA was quantitated using a laser densitometer (LKB 2202 Utroskan). The amount of blotted RNA was controlled by the mRNA of the S6-gene (see Fig. 3). Northern hybridization resulted in a single band with the size of 4.6–4.8 kb (data not shown). (PL, perisinusoidal lipocytes; i, RNA from PL after isolation; 2, 4, 6, RNA isolated from PL after 2, 4 and 6 d in primary culture; MFBIC, myfibroblastlike cells).

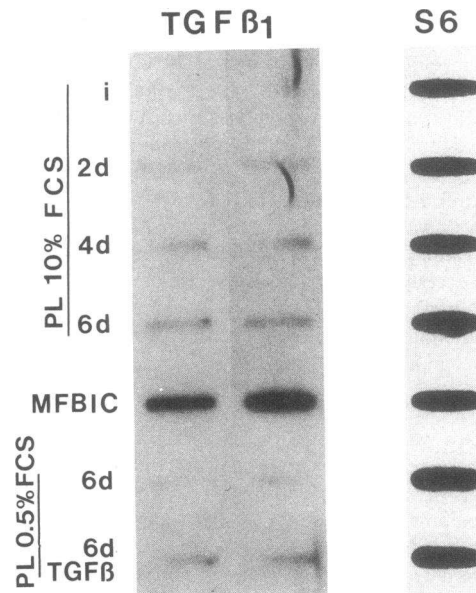


Figure 3. Expression of TGF β 1-mRNA in perisinusoidal lipocytes and myfibroblastlike cells. RNA was prepared from PL and MFBIC grown in the presence of 10% FCS (according to Fig. 2) or from PL grown for 6 d in the presence of 0.5% FCS with and without 1 ng/ml TGF β 1. The amount of blotted RNA (20 μ g) was controlled by the mRNA of the S6-gene. Northern blot analyses resulted in a single band slightly below the 28 S ribosomal RNA (data not shown). (PL, perisinusoidal lipocyte; MFBIC, myfibroblastlike cell; FCS, fetal calf serum).

Table I that partially transformed PL (primary culture seventh day after seeding) and in particular MFBIC have a higher proliferative capacity and require less exogenous growth factor than PL in early culture (untransformed) suggest the production of growth factors by transformed PL.

Expression of TGF α - and TGF β -mRNA. RNA was isolated from PL after isolation and after 2, 4, and 6 d in primary culture. Furthermore, RNA was isolated from MFBIC (3–6 d after passage). Aliquots (20 μ g) of this RNA were slot blotted and probed with specific TGF α - and TGF β 1-cDNAs. As shown in Fig. 2 increased levels of TGF α -mRNA were found in PL on day 6 after seeding (relative amount 0.12) and predominantly in MFBIC (relative amount 0.84). TGF α transcripts were nearly undetectable (relative amount < 0.02) in PL at isolation and after 2 and 4 d in primary culture. The amount of blotted RNA was controlled by the RNA of the S6-gene which remained constant during transformation (Fig. 3). Fig. 3 demonstrates that the TGF β 1 transcripts were first detected in PL 2 days after seeding and increased gradually in primary culture. The highest expression of TGF β 1-mRNA was found in MFBIC (Fig. 3). If PL were treated for 6 d with 1 ng/ml TGF β 1 in the presence of 0.5% FCS the TGF β 1 transcripts were higher compared to unstimulated cells (Fig. 3). This result indicates that TGF β 1 induced its own mRNA expression in PL.

TGF α and TGF β quantification. The TGF α and TGF β 1 protein concentration in media conditioned by PL and in MFBICM was determined by competitive radioligand binding assays and calculated by comparison of dilution curves with that of standard curves. Representative binding curves for TGF α are shown in Fig. 4, for TGF β 1 in Fig. 5. Competition of [¹²⁵I]EGF binding was achieved using rat TGF α (standard curve), competition of [¹²⁵I]TGF β binding was achieved using

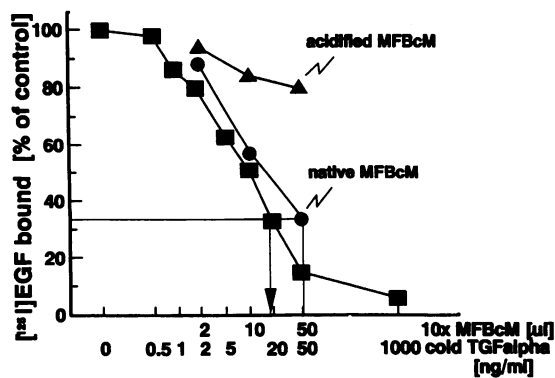


Figure 4. Measurement of TGF α protein concentration in MFBCM by a competitive radioligand binding assay. The media were conditioned for 24 h by MFBC in the absence of FCS. Prior receptor binding aliquots of the media were acidified (pH 2) with 5 M HCl for 30 min and neutralized with 5 M NaOH prior dialysis against binding buffer. TGF α concentration was determined by competition of various volume equivalents of 10-fold concentrated MFBCM with [125 I]EGF binding to confluent skin fibroblasts (●—● native MFBCM, ▲—▲ acidified MFBCM). Recombinant rat TGF α was used as cold ligand (■—■). The native MFBCM displayed a higher competition compared to the acidified MFBCM. In this representative recording competition of 50 μ l 10-fold concentrated native MFBCM diluted with 450 μ l binding buffer was equivalent 18 ng cold TGF α . (MFBCM, medium conditioned by myofibroblastlike cells; TGF α , transforming growth factor α ; EGF, epidermal growth factor).

human TGF β 1. In previous experiments, incubation time and temperature were optimized (data not shown). Based on these data, binding was performed at 20°C for 3 h. Under these conditions, confluent monolayers of skin fibroblasts (3×10^4 cells/cm 2) incubated with 20 fmol (0.46 ng) [125 I]TGF β /0.5 ml for 3 h at 20°C bind 4–6% of the input radioactivity.

By sequential 24-h medium collections from PL in primary culture during 10 d it was shown that TGF β synthesis starts in parallel to transformation at day 5–6 after seeding, increased with culture time (transformation) and was highest in media conditioned by MFBC (Table II). However, native MFBCM caused little [125 I]TGF β competition, acidified MFBCM (dialyzed against binding buffer) resulted in high competition (Fig. 5). Based on the data of three different binding studies the native MFBCM was calculated to contain 0.15 ng/ml (5.7 pM) active TGF β 1, whereas acidified MFBCM was contained 1.6 ± 0.2 ng/ml (61 ± 7.7 pM) active TGF β 1. Furthermore we calculated that a single MFBC produced ~ 0.2 fmol TGF β 1 within 24 h. The fact that latent TGF β does not bind to its receptor (data not shown) suggests that most of the TGF β in the native MFBCM is in the inactive form. By transient acidification the latent form of TGF β 1 was converted to the active (mature) form. In contrast to TGF β 1 active TGF α was higher in native media. Native MFBCM was calculated to contain 22.5 ± 6.6 ng/ml (4 ± 1.2 nM) TGF α ($n = 3$) and a single MFBC was calculated to produce ~ 15 fmol TGF α /24 h. In media conditioned by PL TGF α was not detected (Table II).

Effect of conditioned media on PL proliferation. Media conditioned by PL between the second and fifth day after seeding or by MFBC after the first to third passage were tested on PL for their mitogenic activity. PL conditioned medium (PLCM) neither stimulated nor suppressed PL growth measured by DNA, cell count, and [3 H]thymidine incorporation (data not shown). In contrast MFBCM stimulated PL proliferation (Fig.

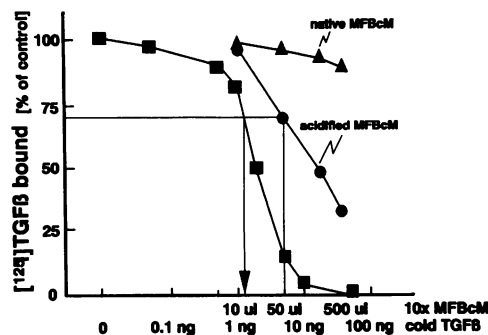


Figure 5. Measurement of TGF β protein concentration in native and transiently acidified MFBCM by competitive radioligand binding assay. Media were conditioned and treated as described in Fig. 4. TGF β concentration was determined by competition of 50 μ l of 10-fold concentrated MFBCM with [125 I]TGF β 1 binding to confluent skin fibroblasts (▲—▲ native MFBCM, ●—● acidified MFBCM). Purified TGF β from human platelets was used as standard (■—■). The acidified MFBCM displayed a higher competition compared to the native MFBCM. Interestingly, the slope of the acidified MFBCM was not exactly parallel to that of the standard, suggesting some nonspecific binding of [125 I]TGF β 1 in the presence of high concentrations of MFBCM. For calculation (Table II) the competition of [125 I]TGF β 1 binding by 50 μ l 10-fold concentrated media was used. In this representative recording competition of 50 μ l 10-fold concentrated aMFBCM diluted with 450 μ l binding buffer was equivalent 1.2 ng cold TGF β 1. (MFBCM, medium conditioned by myofibroblastlike cells; TGF β 1, transforming growth factor β 1).

6). 10% of the MFBCM (0.02 ml 10-fold concentrated MFBCM added to 1.98 ml DME with 0.5% FCS) stimulated [3 H]-thymidine incorporation in primary cultured PL to 2.03-fold of control (Fig. 6) and DNA content to 1.60-fold of control. However, a higher dose of MFBCM inhibited PL proliferation ([3 H]thymidine incorporation at 30% MFBCM 1.3-fold and at

Table II. TGF α and TGF β 1 Production Rate

	PL 3rd d	PL 6th d	TC 10th d	MFBLC secondary culture
TGF α (fmol/cell/d)	n.d.	n.d.	8	15
TGF β 1 (fmol/cell/d)	n.d.	0.02	0.13	0.20

Media were conditioned for 24 h in the absence of FCS by rat liver PL (third and sixth day after seeding), partially transformed PL (TC: 10th day after seeding) and MFBLC (secondary culture, completely transformed). After removing the conditioned media cells were trypsinized and counted in a standard hemocytometer. TGF α was measured in native, TGF β 1 in transiently acidified media by competitive radioligand binding assay (see Methods). In native media conditioned by PL 3 and 6 d after seeding TGF β 1 was not detectable. In native media conditioned by TC and MFBLC TGF β 1 was about one-tenth of the concentration obtained with transiently acidified media. Data points represent the mean TGF α and TGF β 1 production rate per cell, which was calculated on the basis of TGFs concentration in conditioned media and cell count (at least two different cultures). (PL, perisinusoidal lipocyte; TC, transitional cell; MFBLC, myofibroblastlike cell; TGF α , transforming growth factor α ; TGF β 1, transforming growth factor β 1; n.d., not detectable).

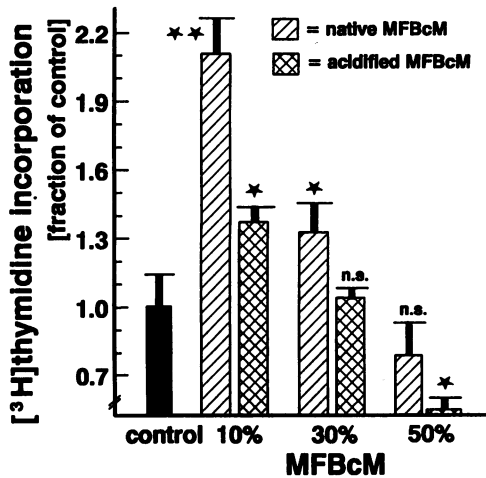


Figure 6. Effect of native and transiently acidified MFBcM on PL proliferation. Media were conditioned for 24 h by MFBcM in the absence of FCS. One-half of the MFBcM was acidified (pH 2) with 5 M HCl for 30 min and thereafter neutralized with 5 M NaOH to pH 7.4. Before conditioned media were added, both the acidified and native media were dialyzed against DME. Control media (DME with 2% FCS) were not dialyzed. Each culture (control and media stimulated) received DME with 2% FCS. The media were added to PL in primary culture in the appropriate concentrations (10, 30, and 50%) on the second and third day after seeding. Cells were labeled with 0.5 μ Ci [3 H]thymidine/ml medium for 24 h between the third and fourth culture day. Mean values + SD of three independent experiments, each with $n = 4$. Statistically significant differences between control and treated cultures are indicated by stars (* $P < 0.05$; ** $P < 0.01$; MFBcM, medium conditioned by myofibroblastlike cells).

50% MFBcM 0.73-fold of control) suggesting that, beside mitogens, potent growth inhibitors also must be present in MFBcM. Transient acidification of the MFBcM resulted in a depressed [3 H]thymidine incorporation (10% MFBcM, 2.03-fold vs. 1.4-fold; 50% MFBcM, 0.8-fold vs. 0.5-fold).

We have shown previously that TGF β 1 acts as a potent growth inhibitor of the EGF/TGF α stimulated PL proliferation (24). Interestingly, the dose response curve of native MFBcM is parallel to that of pure TGF β 1 in the presence of 20-ng/ml EGF or TGF α .

Effect of conditioned media on PL transformation. Light and phase contrast microscopy of PL treated with acidified MFBcM suggested a stimulated PL transformation (flat cell shape, faster reduction of the number, and the size of fat droplets). As the expression of the cytoskeletal protein iso- α smooth muscle actin, and the reduction of cellular retinyl-palmitate in PL are the best indicators for objective measurement of transformation we determined these two parameters at different time intervals after seeding. As shown in Fig. 7 *b* 20% acidified MFBcM in DME with 10% FCS stimulated the expression of iso- α smooth muscle actin. Compared to the control, the difference was most pronounced 11 d after seeding. At that time interval, 70% of the PL treated with acidified MFBcM were iso- α smooth muscle actin positive; in the control cultures only 27% of the cells were iso- α smooth muscle actin positive. The total cell number was not significantly affected by 20% acidified MFBcM (Fig. 7 *a*). Furthermore, as demonstrated in Fig. 8, transiently acidified MFBcM added to PL in primary culture stimulated the loss of cellular retinyl-palmitate. The difference

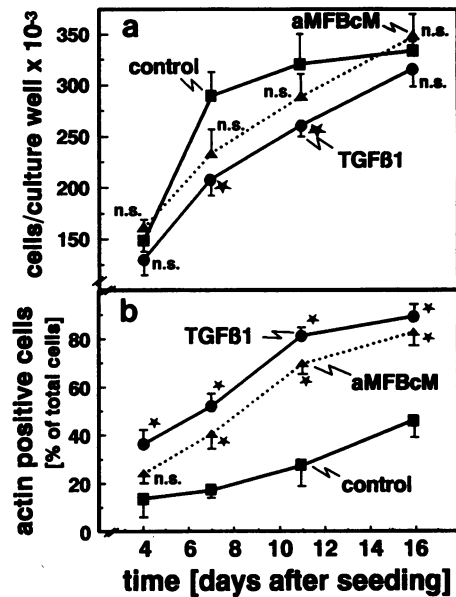


Figure 7. Effects of TGF β 1 and transiently acidified medium conditioned by myofibroblastlike cells on perisinusoidal lipocyte growth (cell numeration) and expression of iso- α smooth muscle actin. Seeding density of PL was 0.08×10^5 cells/cm 2 . The control as well as the treated cells were seeded and cultured in DME with 10% FCS. Beginning on the second day after seeding one-third of the cultures received 1 ng/ml TGF β 1 (circles), one-third received 40 μ l transiently acidified 10-fold concentrated MFBcM (final concentration of MFBcM 40%) (triangles) and one-third received only 10% FCS (squares). On day 4, 7, 11, and 16 after seeding part of the cultures were stopped by fixation using alcohol/acetic acid 95/5 (vol/vol); the remainder was stimulated again with TGF β 1 or aMFBcM. After immunoperoxidase staining for iso- α smooth muscle actin each culture well was examined at least 10 times in different squares of 1 mm 2 each. (a) PL growth was determined by cell numeration using an Olympus phase contrast microscope equipped with a 1-mm 2 optic grid. All cells visible by phase contrast microscopy (positive and negative stained) were counted. The data points represent the average of counts on triplicate cultures (mean \pm SD) and the results were expressed as the number of cells/culture well. (b) Transformation was assessed by expression of iso- α smooth muscle actin. After fixation the cells were stained using anti-iso- α smooth muscle actin, peroxidase conjugated anti-mouse IgG and DAB. The data points represent the average of iso- α smooth muscle actin positive cells as a percentage of the total cell number (mean \pm SD). (aMFBcM, transiently acidified medium conditioned by myofibroblast-like cells; TGF β 1, transforming growth factor β 1; n.s., not significant; * $P < 0.05$ significant difference between control and treated cultures).

between the cultures treated with a MFBcM and the control was significant after 6 d in culture.

Effect of conditioned media on fat storing cell PG synthesis. To detect potential fibrogenic mediators stimulating PG synthesis in conditioned media these media were added in appropriate concentrations (10, 30, 50%) to PL in primary culture grown in DME with reduced (0.5%) FCS. Native and transiently acidified PLcM did not influence PG synthesis, whereas MFBcM stimulated PG synthesis measured as [35 S]sulfate incorporation based on DNA (Fig. 9). Maximal stimulation of PG synthesis per single cell was seen with native MFBcM at dilution 1/2 (1.45-fold of control), and with acidified MFBcM at dilution 1/10 (2.2-fold of control), respectively (Fig. 9).

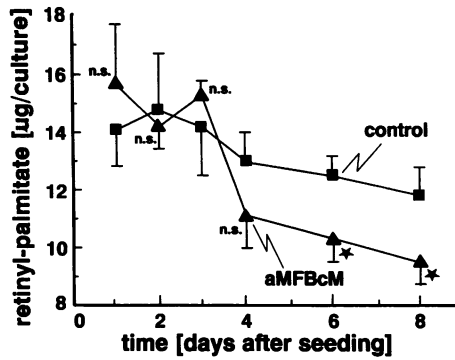


Figure 8. Effect of transiently acidified MFBcM on PL transformation (loss of retinyl-palmitate). The control as well as the treated cells were seeded and cultured in DME with 10% FCS. Beginning on the first day after seeding one-half of the cultures received transiently acidified MFBcM (final concentration of MFBcM, 40%) (triangles) and the other half received only 10% FCS (squares). On day 1, 2, 3, 4, 6, and 8 after seeding part of the cultures were stopped by extraction of retinoids with methanol/ethylacetate. Retinyl-palmitate was determined by HPLC using fluorescence detection. (aMFBcM, transiently acidified medium conditioned for 24 h by myofibroblastlike cells; n.s., not significant; *significant difference ($P < 0.05$) between control and treated cultures).

Identification of TGFs as fibrogenic mediators produced by MFBcM. The activation of the growth inhibitory and PG synthesis stimulatory effect by transient acidification of the MFBcM (whereby TGF β 1 is activated) suggested that TGF β 1 is responsible for these effects. To verify this hypothesis MFBcM were

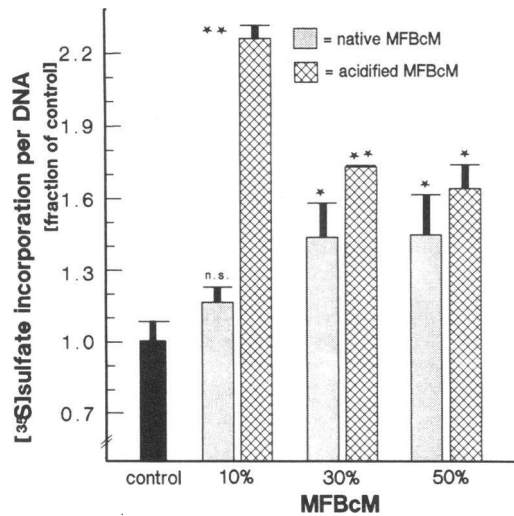


Figure 9. Effects of native and transiently acidified media conditioned by myofibroblastlike cells on perisinusoidal lipocyte proteoglycan synthesis. Conditioned media were treated as described in Fig. 6 and added to PL monolayers in the appropriate concentrations at the third and fourth day after seeding. Control and treated cultures received DME with 0.5% FCS. Between the fourth and fifth culture day cells were exposed for 24 h to 20 μ Ci [35 S]sulfate/ml medium. Proteoglycan synthesis was determined by liquid scintillation counting after binding of the proteoglycans to DEAE-sephacel. Values are expressed as mean \pm SD of three cultures on the basis of DNA. (MFBcM, medium conditioned for 24 h by myofibroblastlike cells; n.s., not significant; * $P < 0.05$, ** $P < 0.01$, significant difference between control and treated cultures).

preincubated with TGF β neutralizing antibodies before addition to PL. To demonstrate the neutralizing potency of anti-TGF β IgG 2 ng/ml pure human TGF β 1 was incubated for 1 h at 22°C with increasing doses of anti-TGF β (2.5–50 μ g/ml) before adding to PL monolayers. The growth inhibitory activity and PG synthesis stimulating activity of these 2 ng/ml TGF β 1 could be neutralized completely by 12.5 μ g/ml anti-TGF β (data not shown). As outlined in Fig. 10 a the PG synthesis stimulating activity of 20 and 50% acidified MFBcM (2.1-fold and 1.6-fold of control) was neutralized by preincubation of the medium with 12.5 μ g/ml anti-TGF β 1. Furthermore, the growth inhibitory activity of 40% transiently acidified MFBcM was abolished by preincubation with 2.5 μ g/ml anti-TGF β and even converted to a growth stimulation by higher concentrations of anti-TGF β (Fig. 10 b). These results not only serve to further confirm the presence of TGF β in MFBcM, but also provide evidence about the presence of one or more mitogens in MFBcM.

Discussion

Using cell culture systems, it has previously been shown that Kupffer cells stimulate via soluble mediators rat liver PL proliferation (10, 11, 13) and extracellular matrix synthesis by these cells (13–15, 17). Similar results, i.e., an enhanced PL proliferation and PG synthesis were obtained using platelet lysate as stimulus (12). Concerning the nature of the macrophage- and platelet-derived factor(s) stimulating PL EGF/TGF α was shown to stimulate proliferation (24), whereas TGF β 1 inhibited PL proliferation (19, 24) but significantly stimulated PL

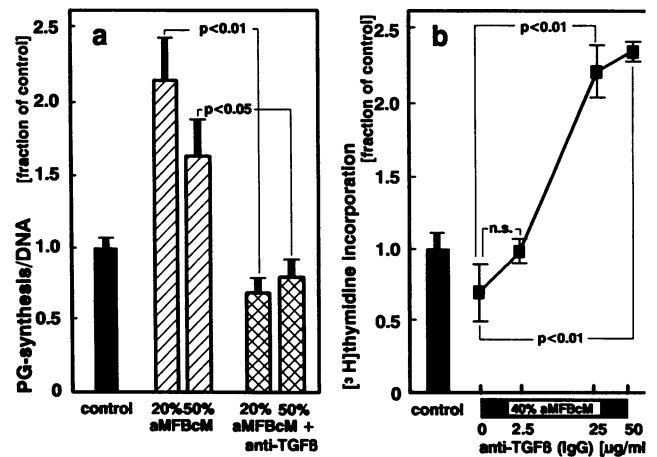


Figure 10. Effect of neutralizing anti-TGF β 1 IgG on MFBcM induced stimulation of PG synthesis (a) and inhibition of PL proliferation (b). Media were conditioned and acidified as described in Fig. 6. Anti-TGF β IgG (2.5, 12.5, 25, and 50 μ g/ml) was added to the media and incubated for 1 h at 21°C on a rotating platform. Thereafter, these media were added to PL monolayers in the appropriate concentrations at the third and fourth day after seeding. (a) The effect 12.5 μ g/ml anti-TGF β 1 on stimulated PG synthesis is shown. Control and treated cultures received DME with 0.5% FCS. Cells were exposed for 24 h to 20 μ Ci [35 S]sulfate/ml medium and proteoglycans were measured as described in Fig. 9. Mean values \pm SD of a representative experiment with $n = 4$. (b) Control and treated cultures received DMI with 2% FCS. Cells were labeled with 0.5 μ Ci [3 H]thymidine/ml medium for 24 h between the fourth and fifth culture day. Mean values \pm SD of a representative experiment with $n = 4$.

collagen- (16, 19) and PG synthesis (20). In combination, both TGFs seem to be potent fibrogenic mediators enhancing predominantly chondroitin sulfate and hyaluronan synthesis by PL (7, 20). Very recently, TGF β was identified as the prominent mediator elaborated by Kupffer cells stimulating PL collagen (17) and PG synthesis (18). Conclusions drawn from these results suggest that in liver injury infiltrating platelets and activated Kupffer cells elaborate transforming growth factors, acting as fibrogenic mediators initiating in a paracrine way not only PL proliferation and transformation but also the synthesis of PG, hyaluronan, and collagen (for review see references 7 and 9).

If PL are cultured on uncoated plastic materials they lose their normal phenotype (4) and transform within 1–2 wk to highly active MFBIC showing reduced number and size of fat droplets; hypertrophy of rough ER; long cytoplasmic extensions; and pronounced positive staining for iso- α smooth muscle actin (6, 7, 29). The current study demonstrates that these transformed cells (MFBIC) have the capacity to produce soluble mediators stimulating thereby PL proliferation, transformation, and enhanced PG synthesis. Medium conditioned by MFBIC stimulated PL proliferation with little effect on PG synthesis. Transient acidification of this medium, whereby latent TGF β is activated (30–32), inhibited PL proliferation but enhanced PG synthesis. These results suggest that TGF β (mainly in the inactive form) together with mitogenic stimuli are present in MFBcM. By competitive radioligand binding assays, TGF β was detectable in PLcM in increasing amounts beginning from day 6 after seeding and particularly in MFBcM (secondary culture). By a further experiment, evidence has been obtained for relevant TGF β -like activity in MFBcM. The biological activity of the acidified MFBIC conditioned media (inhibition of PL proliferation and stimulation of PG synthesis) could be neutralized by TGF β antibodies.

Because MFBIC produce the latent form of TGF β , activation is required before receptor binding. In general, latent TGF β may be activated by extreme, unphysiological pH (pH < 2 or > 12) (30–32), by several proteases (31), and by glycosidases (33). The nature and magnitude of the TGF β mediated PL stimulation may be regulated by the TGF β production rate; the balance of activation/inactivation; and the expression of TGF β receptors on cell surface. Because essentially all cell types of the liver seem to have receptors for TGF β the TGF β activating mechanism is considered to be the major regulatory step. However, the mechanism of TGF β activation in PL cultures and in injured liver remains to be established. Recently obtained results showing that alpha2-macroglobulin is secreted by PL in increasing amounts beginning with the fifth day after seeding (34) and data indicating that alpha2-macroglobulin may bind with and inactivate TGF β (35, 36), suggest a potential role of alpha2-macroglobulin as a scavenger for active TGF β at sites of liver injury. Interestingly, heparin, which is related in its structure to the PG heparan sulfate predominantly produced by parenchymal cells (37), may liberate active TGF β from its complex with alpha2-macroglobulin (38). The pathobiochemical relevance of this mechanism is unclear. Furthermore, binding of TGF β to fibronectin (39) and subsequent dissociation on acidic conditions at sites of tissue injury and inflammation may be relevant in the development of liver fibrogenesis.

The result that PL proliferation is stimulated by MFBcM preincubated with neutralizing anti-TGF β IgG suggests the syn-

thesis of one or more mitogens in addition to TGF β by MFBIC. The radioligand binding assay indicates that significant amounts of EGF or TGF α must be present in MFBcM. Because the EGF cell surface receptor binds both growth factors with similar affinity (40), the radioligand binding assay is unable to differentiate between EGF and TGF α . However, the expression of TGF α -mRNA in MFBIC suggests that at least one factor with mitogenic activity is TGF α . Preliminary data suggest that insulinlike growth factor 1 (somatomedin C) may be a second candidate of the MFBIC-derived mitogenic activity (41, 42). Platelet-derived growth factor (PDGF-BB and PDGF-AB) a further potent mitogen for PL was not detected in cellfree supernatants of passaged PL (43).

In conclusion, our data together with previously reported results (7, 10–15, 17, 44) suggest that in culture on plain plastic and even more important in liver injury a cascade of stimulatory mechanisms act on PL. In early primary culture and in the early stages of tissue response to liver injury the release of TGF β 1 from Kupffer cells (17, 18) and platelets (12) results in an augmented transformation of PL to MFBIC. Thereafter, activated PL (MFBIC) itself produces fibrogenic mediators and thereby stimulates proliferation (TGF α), transformation (TGF β) and proteoglycan synthesis (TGF β) of untransformed PL. These findings also imply that autocrine stimulation of transformed PL (MFBIC) may occur in vivo via secretion of these cytokines. However, further in vivo and in vitro studies should confirm this hypothesis.

Acknowledgments

Expert technical assistance was provided by Brigitte Heitmann and Lothar Scheckel.

This work was supported by grants from Deutsche Forschungsgemeinschaft (Gr 463/7-2) and Kempkes Stiftung, Marburg, who supported M. G. Bachem.

References

1. Mak, K., M. A. Leo, and C. S. Lieber. 1984. Alcoholic liver injury in baboons: transformation of lipocytes to transitional cells. *Gastroenterology*. 87:188–200.
2. Ogawa, K., J. I. Suzuki, M. Narasaki, and M. Mori. 1985. Healing of focal injury in the rat liver. *Am. J. Pathol.* 119:158–167.
3. McGee, J. O., and R. S. Patrick. 1972. The role of perisinusoidal cells in hepatic fibrogenesis. *Lab. Invest.* 26:429–440.
4. Friedman, S. L., F. J. Roll, J. Boyles, D. M. Arenson, and D. M. Bissell. 1989. Maintenance of differentiated phenotype of cultured rat hepatic lipocytes by basement membrane matrix. *J. Biol. Chem.* 264:10756–10762.
5. Mendoza-Figueroa, T., C. Argüello, and W. Kuri-Harcuch. 1988. Culture of proliferating and differentiating fat storing cells in 3T3-conditioned medium. *Biol. Cell.* 64:29–38.
6. Geerts, A., R. Vrijnsen, and J. Rauterberg. 1989. In vitro differentiation of fat-storing cells parallels marked increase of collagen synthesis and secretion. *J. Hepatol. (Amst.)* 9:59–68.
7. Gressner, A. M., and M. G. Bachem. 1990. Cellular sources of noncollagenous matrix proteins: role of fat-storing cells in fibrogenesis. *Semin. Liv. Dis.* 10:30–46.
8. Milani, S., H. Herbst, D. Schuppan, E. G. Hahn, and H. Stein. 1989. In situ hybridization for procollagen types I, II and IV mRNA in normal and fibrotic rat liver: evidence for predominant expression in nonparenchymal liver cells. *Hepatology*. 10:84–92.
9. Friedman, S. L. 1990. Cellular sources of collagen and regulation of collagen production in liver. *Semin. Liv. Dis.* 10:20–29.
10. Zerbe, O., and A. M. Gressner. 1988. Proliferation of fat storing cells is stimulated by secretion of Kupffer cells from normal and injured liver. *Exp. Mol. Pathol.* 49:87–101.
11. Shiratori, Y., A. Geerts, T. Ichida, T. Kawase, and E. Wisse. 1986. Kupffer cells from CCl₄-induced fibrotic livers stimulate proliferation of fat-storing cells. *J. Hepatol. (Amst.)* 3:294–303.

12. Bachem, M. G., R. Melchior, and A. M. Gressner. 1989. The role of thrombocytes in liver fibrogenesis: effects of platelet lysate and thrombocyte-derived growth factors on the mitogenic activity and glycosaminoglycan synthesis of cultured rat liver fat storing cells. *J. Clin. Chem. Clin. Biochem.* 27:555-565.
13. Friedman, S. L., and M. J. Arthur. 1989. Activation of cultured rat hepatic lipocytes by Kupffer cell conditioned medium. Direct enhancement of matrix synthesis and stimulation of cell proliferation via induction of platelet-derived growth factor receptors. *J. Clin. Invest.* 84:1780-1785.
14. Gressner, A. M., and O. Zerbe. 1987. Kupffer cell-mediated induction of synthesis and secretion of proteoglycans by rat liver fat storing cells in culture. *J. Hepatol. (Amst.)* 5:299-310.
15. Gressner, A. M., and R. Haarmann. 1988. Regulation of hyaluronate synthesis in rat liver fat storing cell cultures by Kupffer cells. *J. Hepatol. (Amst.)* 7:310-318.
16. Matsuoka, M., N. T. Pham, and H. Tsukamoto. 1989. Differential effects of interleukin-1 alpha, tumor necrosis factor alpha, and transforming growth factor beta 1 on cell proliferation and collagen formation by cultured fat-storing cells. *Liver* 9:71-78.
17. Matsuoka, M., and H. Tsukamoto. 1990. Stimulation of hepatic lipocyte collagen production by Kupffer cell-derived transforming growth factor β : implication for a pathogenetic role in alcoholic liver fibrogenesis. *Hepatology* 11:599-605.
18. Meyer, D. H., M. G. Bachem, and A. M. Gressner. 1990. Modulation of hepatic lipocyte proteoglycan synthesis and proliferation by Kupffer cell-derived transforming growth factors type β 1 and type α . *Biochem. Biophys. Res. Commun.* 171:1122-1129.
19. Davis, B. H. 1988. Transforming growth factor β responsiveness is modulated by extracellular collagen matrix during hepatic Ito cell culture. *J. Cell. Physiol.* 136:547-553.
20. Bachem, M. G., U. Riess, R. Melchior, K. M. Sell, and A. M. Gressner. 1989. Transforming growth factors (TGF alpha and TGF beta 1) stimulate chondroitin sulfate and hyaluronate synthesis in cultured rat liver fat storing cells. *FEBS (Fed. Eur. Biochem. Soc.) Lett.* 257:134-137.
21. Czaja, K. C., L. Flanders, M. A. Biempica, M. A. Zern, and F. R. Weiner. 1988. Differential expression of tumor necrosis factor-alpha and transforming growth factor- β 1 in acute liver injury. *Hepatology* 8:1234. (Abstr.)
22. Weiner, F. R., M.-A. Giambone, M. J. Czaja, A. Shah, G. Annoni, S. Takahashi, M. Eghbali, and M. A. Zern. 1990. Ito-cell gene expression and collagen regulation. *Hepatology* 11:111-117.
23. Annoni, G., M. J. Czaja, F. R. Weiner, and M. A. Zern. 1988. Increased transforming growth factor- β 1 (TGF β 1) gene expression in human liver disease. *Hepatology* 8:1227. (Abstr.)
24. Bachem, M. G., U. Riess, and A. M. Gressner. 1989. Liver fat storing cell proliferation is stimulated by epidermal growth factor/transforming growth factor alpha and inhibited by transforming growth factor beta. *Biochem. Biophys. Res. Commun.* 162:708-714.
25. Kondaiiah, P., E. Van-Obberghen-Schilling, R. L. Ludwig, R. Dhar, M. B. Sporn, and A. B. Roberts. 1988. cDNA cloning of porcine transforming growth factor-beta 1 mRNAs. Evidence for alternate splicing and polyadenylation. *J. Biol. Chem.* 263:18313-18317.
26. Schäfer, S., O. Zerbe, and A. M. Gressner. 1987. The synthesis of proteoglycans in fat storing cells of rat liver. *Hepatology* 7:680-687.
27. Labarca, C., and K. Paigen. 1980. A simple, rapid, and sensitive DNA assay procedure. *Anal. Biochem.* 102:344-352.
28. Chomczynski, P., and N. Sacchi. 1987. Single-step method of RNA isolation by acid guanidinium-thiocyanate-phenol-chloroform extraction. *Anal. Biochem.* 162:156-159.
29. Ballardini, G., M. Fallani, G. Biagini, F. B. Bianchi, and E. Pisi. 1988. Desmin and actin in the identification of Ito cells and in monitoring their evolution to myofibroblasts in experimental liver fibrosis. *Virchows Archiv B Cell Pathol.* 56:45-49.
30. Wakefield, L. M., D. M. Smith, K. C. Flanders, and M. B. Sporn. 1988. Latent transforming growth factor beta from human platelets. A high molecular weight complex containing precursor sequences. *J. Biol. Chem.* 263:7646-7654.
31. Lyons, R. M., J. Keski-Oja, and H. L. Moses. 1988. Proteolytic activation of latent transforming growth factor beta from fibroblast-conditioned medium. *J. Cell Biol.* 106:1659-1665.
32. Miyazono, K., U. Hellman, C. Wernstedt, and C. H. Heldin. 1988. Latent high molecular weight complex of transforming growth factor beta 1. Purification from human platelets and structural characterization. *J. Biol. Chem.* 263:6407-6415.
33. Miyazono, K., and C. H. Heldin. 1989. Role for carbohydrate structures in TGF-beta1 latency. *Nature (Lond.)* 338:158-160.
34. Andus, T., G. Ramadori, and P. C. Heinrich. 1987. Cultured Ito cells of rat liver express the alpha2-macroglobulin gene. *Eur. J. Biochem.* 168:641-646.
35. Huang, S. S., P. O'Grady, and J. S. Huang. 1988. Human transforming growth factor beta-alpha 2-macroglobulin complex is a latent form of transforming growth factor beta. *J. Biol. Chem.* 263:1535-1541.
36. O'Connor-McCourt, M. D., and L. M. Wakefield. 1987. Latent transforming growth factor beta in serum. A specific complex with alpha 2-macroglobulin. *J. Biol. Chem.* 262:14090-14099.
37. Prinz, R., U. Klein, P. R. Sudhakaran, W. Sinn, K. Ullrich, and K. von Figura. 1980. Metabolism of sulfated glycosaminoglycans in rat hepatocytes. Synthesis of heparan sulfate and distribution into cellular and extracellular pools. *Biochim. Biophys. Acta.* 630:402-413.
38. McCaffrey, T. A., D. J. Falcone, C. F. Brayton, L. Agarwal, F. G. P. Welt, and B. B. Weksler. 1989. Transforming growth factor- β activity is potentiated by heparin via dissociation of the transforming growth factor- β /alpha2-macroglobulin inactive complex. *J. Cell Biol.* 109:441-448.
39. Mooradian, D. L., R. C. Lucas, J. A. Weatherbee, and L. T. Furcht. 1989. Transforming growth factor- β 1 binds to immobilized fibronectin. *J. Cell Biochem.* 41:189-200.
40. Bachem, M. G., D. Meyer, W. Schäfer, and A. M. Gressner. 1989. Evaluation of fibrogenic polypeptide mediators stimulating fat storing cell proliferation and/or proteoglycan synthesis. In *The Liver, Metabolism and Aging*. Vol. 13. K. W. Woodhouse, C. Yelland, and O. F. W. James. Eurage, Rijswijk, Netherlands, 169-178.
41. Bachem, M. G., W. Schäfer, and A. M. Gressner. 1989. Untersuchungen zur Rezeptorbindung und mitogenen Aktivität von Insulin und Insulin like Growth Factor I an Ito-Zellen. *Z. Gastroenterol.* 27:17. (Abstr.)
42. Pinzani, M., and D. C. Aron. 1989. Rat liver fat-storing cells secrete insulin-like growth factor I and its binding protein. *Hepatology* 10:635. (Abstr.)
43. Pinzani, M., T. C. Knauss, G. F. Pierce, P. Hsieh, W. Kenney, G. R. Dubyak, and H. E. Abboud. 1990. Mitogenic signals for platelet-derived growth factor isoforms in rat liver fat-storing cells. *Hepatology* 12:919. (Abstr.)
44. Nakatsukasa, H., R. P. Evarts, C.-C. Hsia, and S. S. Thorgeirsson. 1990. Transforming growth factor- β 1 and type I procollagen transcripts during regeneration and early fibrosis of rat liver. *Lab. Invest.* 63:171-180.