

Human Neutrophil Elastase Modulates Platelet Function by Limited Proteolysis of Membrane Glycoproteins

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Abstract

During blood coagulation human polymorphonuclear leukocytes release elastase in amounts that can exceed 100 nmol/liter. We therefore studied the effect of elastase on platelet structure and function. Physiologic concentrations of elastase specifically inhibited thrombin-induced platelet aggregation and ristocetin-induced agglutination of washed platelets in a time- and dose-dependent manner. This was associated with a decrease in the number of high affinity thrombin binding sites on the platelet surface (analysis by "Ligand" program) from 31 per platelet to 12 per platelet ($P < 0.05$). As analyzed by sodium dodecyl sulfate-polyacrylamide gel electrophoresis, treatment of ^3H -labeled platelets with elastase resulted in a decrease in the percent glycoprotein at 130,000–150,000 M_r and an increase in the percent protein at $M_r = 102,000$. The supernatant from elastase-treated platelets contained a $M_r = 88,000$ glycoprotein not found in the supernatant from untreated platelets. Immunoprecipitation studies with monoclonal antiglycoprotein Ib demonstrated that treatment of whole platelets with physiologic concentrations of elastase resulted in proteolytic cleavage of glycoprotein Ib. Elastase treatment of glycoprotein immunoisolated with monoclonal antiglycoprotein Ib antibody resulted in formation of a glycopeptide with the same electrophoretic mobility as the $M_r = 102,000$ membrane-related glycopeptide. In contrast, analysis by Western blot technique using antiglycoprotein IIb and IIIa antibodies demonstrated that elastase did not degrade glycoproteins IIb or IIIa.

We conclude that elastase inhibition of thrombin-induced platelet stimulation is accompanied by (a) a reduction in the number of thrombin binding sites per platelet and (b) proteolysis of glycoprotein Ib.

Introduction

Human polymorphonuclear leukocyte (PMN)¹ elastase is the major neutral protease released by human PMN during blood

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1. Abbreviations used in this paper: DTT, dithiothreitol; GPA, tosyl-Gly-L-Pro-L-Arg-paranitroanalide; PBS, phosphate-buffered saline, 100 mM

coagulation, or activation by soluble immune complexes, C5a, or endotoxin (1–3). Elastase is released into the circulation in acute leukemia and in septicemia (4). Several studies have indicated that the decrease in the concentration of coagulation factors seen in these conditions may be the result of direct proteolysis by leukocyte elastase (5). Leukocyte elastase also inactivates human antithrombin III (6), α_2 -plasmin inhibitor, and C1 inactivator (7). In circulating blood leukocyte elastase is inactivated mainly by α_1 -antitrypsin (8); however, this inhibitor is readily inactivated by oxidizing agents such as those generated by phagocytosing PMN (9, 10).

Several studies suggest a potential role for neutrophils in the modulation of hemostasis and thrombosis. Arterial thrombi are characterized by the presence of large numbers of PMN lining the perimeter of platelet aggregates (11–13). Marcus et al. (14) and Maclouf et al. (15) have documented that platelet-derived arachidonate or 12-hydroperoxy-eicosatetraenoic acid stimulates the biosynthesis of PMN 5,12-dihydroxyeicosatetraenoic acid. This suggests that platelet-derived arachidonate may serve as a precursor for the PMN-derived eicosanoid leukotriene B₄, a powerful chemoattractant and secretagogue (15–17). Additionally, platelet factor 4, a cationic protein liberated by platelets during the release reaction, has been demonstrated to stimulate PMN elastase activity against elastin (18–20).

The present report details studies on the effect of human leukocyte elastase on platelet structure and function. Our findings indicate that physiologic concentrations of elastase specifically inhibit thrombin-induced platelet aggregation and serotonin release as well as ristocetin-induced platelet agglutination. Loss of thrombin responsiveness is accompanied by a reduction in the number of high affinity thrombin binding sites per platelet and by elastase-mediated proteolysis of platelet membrane glycoprotein Ib.

Methods

Human PMN elastase. Human PMN elastase was purified from outdated leukocyte concentrates as described (3, 7). The concentration of elastase was determined by its extinction coefficient of 9.85 (21). The purified elastase demonstrated one protein-staining band when analyzed by SDS-polyacrylamide gel electrophoresis (PAGE) following reduction with dithiothreitol (DTT) (22). The specific activity of the elastase was 92% by active site titration using the titrant Ac-Ala-Ala-Anval-ONp (a gift of Dr. J. Powers, Georgia Institute of Technology, Atlanta, GA) as described by Powers and Gupton (23).

Alpha-1-antitrypsin. α_1 -Antitrypsin was purified from plasma as

NaCl, 38 mM Na₂HPO₄, 12.5 mM NaH₂PO₄, pH 7.4; PBS-Triton, PBS containing 0.5% (vol/vol) Triton X-100, 0.02% NaN₃, 0.1% bovine serum albumin; PBS-Tween, PBS containing 0.5% (vol/vol) Tween 20; PAGE, polyacrylamide gel electrophoresis; PMN, polymorphonuclear leukocyte; PRP, platelet-rich plasma; Tris, 0.1 M Tris-HCl, 1.25 mM CaCl₂, pH 8.3.

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previously described and its concentration determined by its extinction coefficient of 5.3 (24). The final inhibitor preparation was free of other contaminating plasma protease inhibitors as analyzed by double diffusion in agarose gels with specific antibodies against α_2 -macroglobulin, C \bar{I} inactivator, antithrombin-III, inter- α -trypsin inhibitor, or chymotrypsin inhibitor obtained from Calbiochem-Behring Corp., La Jolla, CA.

Platelet aggregation studies. Platelet-rich plasma (PRP) and washed platelet suspensions were prepared by the technique of Mustard et al. (25) as modified by Weksler et al. (26, 27). Platelet aggregation studies were performed at 37°C with 0.4-ml aliquots of PRP or washed platelets (2×10^8 /ml) stirred at 1,000 rpm. Platelet aggregation was performed in a Payton 600BD dual-channel aggregometer (Payton Assoc., Scarborough, Ontario, Canada) and recorded with a Soltec 1242 (Sun Valley, CA) dual channel strip chart recorder (28). Aggregation was tested with threshold doses of the following agents (expressed as final concentration in the cuvette): sodium arachidonate (Nu Chek Prep, Elysian Fields, MN), 0.25 mM; ADP (Sigma Chemical Co., St. Louis, MO), 8 μ M; fibrillar bovine collagen (Hormon-Chemie, Munich, Federal Republic of Germany), 2 μ g/ml; thrombin (the generous gift of Dr. John W. Fenton III, New York State Department of Health, Albany, NY); epinephrine (Parke Davis Co., New York), 5 μ g/ml. Washed platelets (2×10^8 /ml) were agglutinated with Ristocetin (Sigma Chemical Co.) 0.5 to 1.0 mg/ml plus VIII:VWF (Hemophil [Hyland Div., Travenol Laboratories, Inc., Costa Mesa, CA]; 0.1 to 0.2 μ g/ml).

The threshold stimulus determined for individual platelet preparations was defined as the lowest concentration that caused a 90% or greater increase in light transmission at 5 min when added to stirred platelets at 37°C (28). Elastase inhibition of aggregation by the above agents was tested by adding various concentrations of elastase to platelets for various times at 37°C. After inhibiting elastase with α_1 -antitrypsin (α_1 -antitrypsin:elastase molar ratio 5:1), a previously determined threshold dose of the aggregating agent was added, and aggregation was recorded.

Thrombin-induced secretion of [14 C]serotonin by platelets. This was measured by standard technique (27, 29). Before elastase treatment, at least 80% of total [14 C]serotonin was released from platelets upon stimulation with high concentrations of thrombin (≥ 100 nM).

Labeling of [125 I]-thrombin. α -Thrombin was labeled with [125 I] by the method of Martin et al. (30). Titration of [125 I]-thrombin with tosylglycyl-L-proline-L-arginine-paranitroanalide (GPA, Vega Fox Biochemicals, Tucson, AZ) was modified from the method of Kirchof et al. (31). GPA was dissolved in methanol (14 mg/ml) and diluted 10-fold with Tris 0.1 M, CaCl $_2$ 1.25 mM, pH 8.3 containing bovine serum albumin 1 mg/ml (Tris buffer). Serial dilutions of [125 I]- α -thrombin in Tris buffer (0.15 ml) were mixed with GPA solution. Color development at 405 nm was followed at 10 min intervals in a Titertek Multiscan photometer (Flow Laboratories, Rockville, MD). The change in absorbance per minute in the thrombin-containing wells was subtracted from the values obtained in wells containing no thrombin. The change in absorbance per minute was used to calculate the concentrations of [125 I]-thrombin using a standard curve simultaneously generated by the serial dilution of unlabeled α -thrombin, the concentration of which had been determined by active site titration with *p*-nitrophenyl-*p'*-guanidinobenzoate-HCl (32). The specific radioactivity of the [125 I]-thrombin was ~ 5 Ci/mmol.

Binding measurements. Measurement of [125 I]-thrombin binding to platelets was modified from the oil centrifugation method of Martin et al. (30). Whole human blood was added to tubes containing EDTA 100 mM in 0.15 M phosphate-buffered saline, pH 7.0 (19:1 vol/vol) and centrifuged at 160 *g* for 15 min at 23°C. The PRP was collected and centrifuged at 1,067 *g* for 15 min. The sedimented platelets were resuspended in 15 ml Tris-HCl (75 mM), KCl (100 mM), sodium citrate (12 mM), and EDTA (5 mM) pH 6.3. The platelets were washed twice more and resuspended in NaCl 136 mM, Tris-HCl 25 mM, glucose 5 mM, EDTA 1 mM, pH 7.4 containing 5 mg/ml bovine serum albumin (Sigma Chemical Co., essentially fatty acid free). Platelets ($1.5\text{--}7 \times 10^9$ /ml) were equilibrated in this buffer for 10 min at 23°C before the addition of thrombin. Incubation with thrombin

was for 10 min at 23°C unless otherwise specified. Platelets then were separated from buffer (0.3 ml) by layering onto 0.5 ml of a 6.7:3.3 mixture of Dow 550 (Dow Chemical Co., Midland, MI) and Dow 556 silicone oil (sp gr 1.040; Dexter Corp., Olean, NY) (33) and by centrifugation in a 1.5-ml conical polypropylene centrifuge tube (12,000 *g* for 2 min) in an Eppendorf centrifuge. Two 0.1-ml aliquots of the aqueous phase were transferred to counting vials. The tube was carefully inverted, drained, and the bottom of the tube containing the platelet pellet was sheared from the tube and counted in a Searle 1185 gamma counter (Searle Radiographics, Inc., Des Plaines, IL). The maximum binding of thrombin to platelets was reached within 5 min after addition of thrombin; binding remained maximum for at least 30 min after addition of thrombin. Of added [14 C]sorbitol (Amersham Corp., Arlington Heights, IL), $<0.013\%$ was trapped in the pellet. This would result in an error in the bound/free ratio of $<2\%$ at the highest concentrations of thrombin, so that corrections were not made.

Data was analyzed with the "Ligand" program for fitting multiple binding sites data (Biomedical Computing Technology Information Center, Vanderbilt University, Nashville, TN) developed by Munson and Rodbard (34). The Ligand program was used to correct for nonspecific binding using both a one-site and two-site model.

3 H-Labeling of platelet surface glycoproteins. Platelet surface glycoproteins were labeled with tritium by modification of established methods (35, 36). Washed platelets from 150 ml of whole blood were resuspended at 0°C in 1.3 ml of 137 mM NaCl, 2.7 mM KCl, 1.2 mM NaHCO $_3$, 0.4 mM NaH $_2$ PO $_4$, 1 mM MgCl $_2$, 2 mM CaCl $_2$, 0.1% glucose, 0.35% bovine serum albumin, pH 7.35. Sodium metaperiodate (2 mM) 0.15 ml was added to the platelet suspension prior to incubating in the dark for 10 min at 0°C. The suspension was diluted with buffer, sedimented by centrifugation at 1,067 *g* for 10 min and washed once more. The platelets resuspended in 1.5 ml buffer at 23°C were mixed with 2.5 mCi [3 H]NaBH $_4$ (New England Nuclear, Boston, MA) in 0.01 ml NaOH (0.01 M). After 30 min, cold buffer (25 ml) was added and the platelets centrifuged at 1,067 *g* for 10 min. The platelets were resuspended, and centrifuged twice more. Aliquots were taken for study by aggregometry as detailed above.

Treatment of 3 H-labeled platelets with elastase. The 3 H-labeled platelet suspensions (2×10^9 platelets/ml) were exposed to either elastase (0–300 nM) or to elastase buffer (0.05 M sodium acetate, 0.45 M NaCl, pH 5.5) for various time intervals at 23°C. α_1 -Antitrypsin was added to inhibit elastase (α_1 -antitrypsin:elastase molar ratio ≥ 5 :1) and the platelets centrifuged at 1,067 *g* for 10 min. Each supernatant was removed and the platelet pellet washed and resuspended in 0.05 ml buffer. Aliquots were taken for study by aggregometry as detailed above.

SDS-PAGE. Samples were mixed with an equal volume of 0.06 M Tris-HCl, 2% SDS, 5% glycerol, 0.001% bromphenol blue, and 2% DTT; solubilized by heating at 10°C for 3 min, and stored (-70°C). SDS-slab gel electrophoresis was performed by the method of Laemmli (37) using a 3.8% stacking gel and a 9% separation gel. Gels were fixed in 25% isopropanol and 10% acetic acid prior to staining with Coomassie Blue (0.1%) and radioautography (38). Densitometry was performed at 633 nm using an LKB Ultrosan Laser Densitometer (LKB Instruments, Inc., Rockville, MD) and a 2210 2-channel recorder.

Immunoenzymatic analysis of the effect of elastase on intact platelet glycoproteins IIb and IIIa. Platelets (10^{10} /ml) were treated with elastase (80 nM) or buffer prior to the addition of α_1 -antitrypsin as detailed above. Following SDS-PAGE of unreduced samples (3 μ g/lane), proteins were transferred onto nitrocellulose by standard technique (39). After electroblotting, the nitrocellulose was incubated overnight at 4°C in phosphate-buffered saline (PBS, 100 mM NaCl, 38 mM Na $_2$ HPO $_4$, 12.5 mM NaH $_2$ PO $_4$, pH 7.4) containing 5% bovine serum albumin (Sigma Chemical Co., essentially fatty acid free) and 3 mM Na $_2$ S $_2$ O $_3$. The nitrocellulose was incubated an additional 30 min in PBS containing 0.5% (vol/vol) Tween 20 (PBS-Tween), and the blocked nitrocellulose membrane then was incubated with primary antibody (rabbit anti-human glycoprotein IIb or IIIa) (40) diluted 1:100 in PBS-Tween for 1 h at 37°C with gentle rocking. The membrane was washed three

times with PBS-Tween and incubated with biotinylated goat anti-rabbit IgG (Enzo Biochem, New York) diluted 1:1000 in PBS-Tween for 1 h at 37°C with gentle rocking. The membrane was washed three times with PBS-Tween, rinsed once in PBS, and incubated 30 min at 23°C in PBS containing 5 mM trisodium EDTA, 2% bovine serum albumin, and 0.05% (vol/vol) Triton X-100. Avidin-acid phosphatase complex (Enzo Biochem) diluted 1:200 in PBS, 5 mM trisodium EDTA was incubated with the membrane (0.08 ml/cm²) for 1 h at 23°C before washing 5 times for 5 min each in 10 mM K₂HPO₄, 0.5 M NaCl, 1 mM EDTA, pH 6.5, containing 2% bovine serum albumin and 0.5% Triton X-100. The membrane was rinsed twice (2 min per rinse) in a clean dish with 0.2 M acetic acid, 0.2 M sodium acetate, pH 5.8. To detect the complex the membrane was incubated 5 min with 1 mM Naphthol AS-MX phosphate (Enzo Biochem), 0.2 M sodium acetate, pH 5.8, before adding 1/100 vol of Fast violet B salt (Enzo Biochem) 4 mg/ml in 0.2 M sodium acetate, pH 5.8. Color development at 23°C was first observed after 5–20 min. After 1 to 4 h, the membrane was removed from the dye solution, rinsed in tapwater, and air dried. The color of the bands on some nitrocellulose membranes faded upon drying. The addition of water or buffer (neutral or slightly acidic) restored color.

Immunoprecipitation of platelet membrane proteins. This was modified from the techniques of Kessler (41) and Coller et al. (42). Washed platelets were prepared from 150 ml freshly drawn blood and radiolabeled with [³H]NaBH₄ as detailed above. The platelets (1.4 × 10¹⁰/ml) were divided into two 0.7-ml portions and treated with elastase (80 nM) or buffer for 3 min at 37°C. α₁-Antitrypsin (0.2 μM) was added and the samples centrifuged at 12,000 g for 30 s.

The precipitates were resuspended in 1.0 ml PBS, pH 7.4, containing 1% Triton X-100, 1 mM leupeptin (Sigma Chemical Co.) and 3 mM NaN₃ at 4°C for 15 min with sonication (three times for 15–20 s, power reading 3.5 with a 4½-in. microprobe and microsonicator, Kontes Glass Co., Vineland, NJ). The samples were centrifuged at 5,000 g for 15 min at 4°C, and the supernatant recentrifuged at 40,000 g for 1 h at 4°C.

Immuno beads (1.0 ml) coated with rabbit anti-mouse immunoglobulin (10 mg/ml, Bio-Rad Laboratories, Richmond, CA) were washed three times in PBS containing 0.5% Triton X-100, 0.02% NaN₃, 0.1% bovine serum albumin, pH 8.4 (PBS-Triton), by spinning at 1,500 g for 2 min and resuspending in 1.0 ml after each wash. The beads were divided into two 0.5-ml portions and mixed with 1.0 ml of either normal mouse ascites fluid (Cappel Laboratories, Cochranville, PA) diluted 1:1000 in PBS-Tween or desiccated ascites fluid containing monoclonal antiglycoprotein Ib (AP-1, a generous gift of Thomas Kunicki; Blood Center of Southeastern Wisconsin) (43) reconstituted in PBS containing 1 mg/ml BSA, and then similarly diluted 1:1000 in PBS-Tween.

After incubating for 40 min at 23°C and 10 min at 37°C, samples were washed four times in PBS-Triton and resuspended in 0.5 ml buffer. The solubilized platelet proteins (0.1 ml) and the antibody coated Immuno beads (0.1 ml) were incubated 60 h at 4°C with mixing and centrifuged 12,000 g for 2 min. The pellet was washed three times in PBS-Triton as above. To release the bound antigen, the pellet was mixed with 0.1 ml of a solution consisting of one part 3.3% SDS, 6 mM N-ethylmaleimide, and one part 1% SDS, 12.5 mM Tris-HCl, 20% glycerol, 0.025% Bromphenol blue, pH 6.8, and heated to 100°C for 3 min. Samples were centrifuged at 12,000 g for 2 min and the supernatants stored (–70°C). Samples (10,000 dpm/lane) were analyzed by electrophoresis as described above.

Immunoisolation of platelet membrane glycoprotein Ib was modified from the technique of McEver et al. (44). Immuno beads were coated with monoclonal antiglycoprotein Ib as detailed above. Solubilized ³H-labeled platelet membrane proteins (0.5 ml), prepared as above, and antibody coated Immuno beads (0.5 ml) were incubated 60 h at 4°C with mixing and centrifuged at 12,000 g for 2 min. The Immuno beads were washed three times in PBS-Triton before washing in 0.15 M NaCl, 0.02 M Tris-HCl, 1% Triton X-100; then 1 M NaCl, 1% Triton X-100, pH 7.4; then 0.1 M Tris-HCl, 1% Triton X-100, pH 7.4, and

finally in 0.15 M NaCl, 0.02 M Tris-HCl, 1% Triton X-100, pH 7.4. Bound protein was eluted with 0.05 M diethylamine, 1% Triton X-100, pH 11.5. The eluate was neutralized with 2 M Tris-HCl, 1% Triton X-100, pH 7.4, dialyzed against 0.15 M NaCl, 0.02 M Tris-HCl, 2.5 mM EDTA, and stored at 4°C. Aliquots were incubated with either elastase (5 nM) or buffer for various times at 37°C prior to heating (100°C) for 3 min with an equal volume of 0.06 M Tris-HCl, 2% SDS, 5% glycerol, 0.001% bromphenol blue, and 2% DTT. All samples were stored (–70°C) prior to electrophoresis (10,000 dpm/lane).

Results

Elastase inhibition of thrombin-induced platelet aggregation. Elastase inhibited thrombin-induced platelet aggregation in a time- and dose-dependent manner. In Fig. 1, washed platelets (2 × 10⁸/ml) were stirred with either elastase (10 nM; 30,000 molecules/platelet) or with buffer for 1 min at 37°C. A threshold dose of thrombin (1 nM) then was added. The elastase-treated platelets failed to aggregate, whereas the buffer-treated platelets responded normally. The concentration of elastase chosen was below that documented to appear during blood coagulation (28–140 nM) (3). Although aggregation of elastase-treated platelets did not occur after exposure to thrombin, examination by phase microscopy revealed that shape change had occurred. In subsequent experiments, the inhibition of thrombin-induced platelet aggregation was found to be both time and dose dependent (Fig. 2). Elastase inhibition of ristocetin-mediated agglutination paralleled the inhibition of thrombin stimulation. In contrast, similar concentrations of elastase did not inhibit platelet aggregation by ADP, epinephrine, arachidonic acid, or collagen.

To explore whether the action of elastase was due either to a direct interaction with platelets or to proteolytic inactivation of thrombin, α₁-antitrypsin was used to inhibit elastase activity. In Fig. 3 (bottom) elastase (10 nM) was incubated with washed platelets for 1 min at 37°C. After inhibition of elastase by addition of α₁-antitrypsin (64 nM), thrombin (1 nM) was added. Aggregation did not occur. These experiments suggested that elastase inhibited aggregation by acting on the platelet rather than on thrombin. In a separate study (Fig. 3, top tracing) α₁-antitrypsin (64 nM) was added to washed platelets prior to addition of elastase. Elastase was then added and the mixture incubated for 1 min (37°C). Subsequent addition of thrombin (1 nM) resulted in aggregation. This study demon-

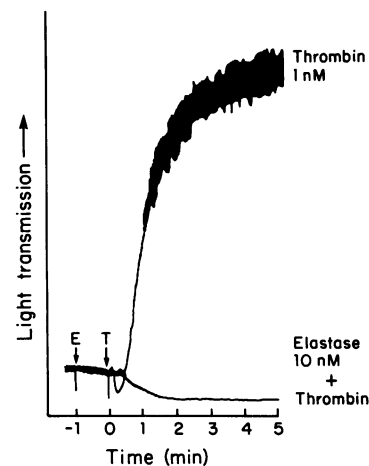


Figure 1. Elastase inhibition of thrombin-induced platelet aggregation. Washed platelets (2 × 10⁸/ml) were stirred with (bottom tracing) or without (top tracing) elastase (10 nM; 30,000 molecules/platelet) for 1 min at 37°C. Thrombin (1 nM) was then added. Aggregation was recorded as an increase in light transmission.

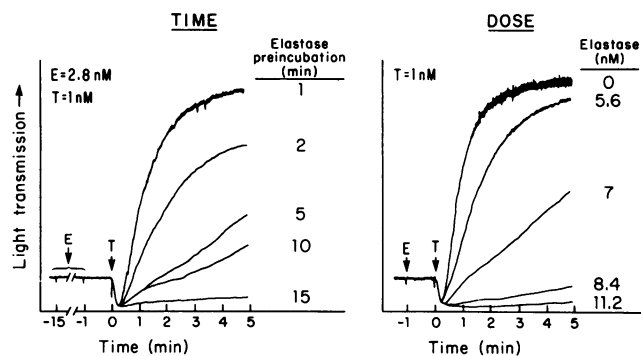


Figure 2. Time- and dose-dependent inhibition of thrombin-induced platelet aggregation by elastase. (A) Time dependence. Washed platelets (2×10^8 /ml) were stirred with elastase (E) 2.8 nM (a lower concentration than in Fig. 1) at 37°C for 1–15 min prior to the addition of a 1 nM threshold dose of thrombin (T). (B) Dose dependence. Washed platelets (2×10^8 /ml) were stirred with elastase (0–11.2 nM) for 1 min at 37°C prior to the addition of thrombin (1 nM). Aggregation was recorded by the increase in light transmission over time. Aggregation study shown is a representative study from four separate time and dose experiments.

strates that an excess of α_1 -antitrypsin can prevent elastase-mediated inhibition of thrombin-induced platelet aggregation. Although α_1 -antitrypsin has been reported to inhibit thrombin (45), in concurrent control studies platelet aggregation was not inhibited by mixing similar concentrations of α_1 -antitrypsin with platelets prior to addition of thrombin.

Elastase inhibition of thrombin-induced secretion of [^{14}C]serotonin by platelets. Elastase inhibited thrombin-induced [^{14}C]serotonin release from platelets. Examples of a standard curve of thrombin-induced serotonin release and the effect of a 1-min (37°C) incubation of platelets with various concentrations of elastase (0.9–90 nM) prior to thrombin addition are shown in Fig. 4. Compared with untreated platelets, a washed platelet suspension pretreated with elastase was less sensitive to thrombin-induced [^{14}C]serotonin secretion. As a result the curve of serotonin release shifted to the right. The concentration of thrombin required to liberate 50% of maximally releasable [^{14}C]serotonin from control platelets was 2.5 nM. After platelets were preincubated with elastase (0.9 nM), the concentration of thrombin required was 4 nM. Increasing the concentration of elastase resulted in a progressive shift to the right, suggesting that the inhibitory effect of elastase on thrombin-induced

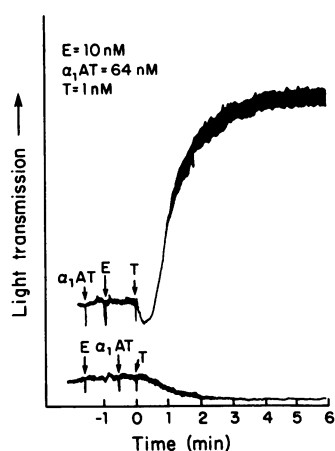


Figure 3. Inhibition of elastase by α_1 -antitrypsin. (top) Washed platelets (2×10^8 /ml) were stirred with α_1 -antitrypsin ($\alpha_1\text{AT}$) 64 nM for 1 min at 37°C . Elastase (E) 10 nM was then added for ~ 1 min before the addition of thrombin (T) 1 nM; (bottom) washed platelets were stirred with elastase (10 nM) for ~ 1 min at 37°C . α_1 -Antitrypsin (64 nM) was then added ~ 1 min before the addition of thrombin (1 nM). Aggregation was recorded as the increase in light transmission.

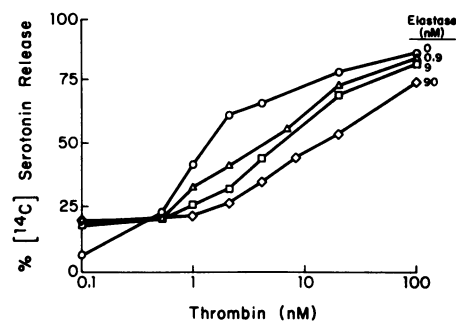


Figure 4. Inhibition of thrombin-induced [^{14}C]serotonin release from platelets by elastase. Washed platelets (2×10^8 /ml) containing [^{14}C]serotonin were mixed with elastase (0.9–90 nM) or buffer for 1 min at 37°C . α_1 -Antitrypsin was added (inhibitor:elastase molar ratio $\geq 1.3:1$) and the platelets were incubated with various concentrations of thrombin as described in Methods. Percent ^{14}C release is plotted as a function of thrombin concentration.

serotonin release was dose dependent. The concentration of thrombin required to liberate 50% of [^{14}C]serotonin from platelets incubated with 9 nM elastase was 6 nM; and with 90 nM elastase, 15 nM.

Effect of elastase on the binding of thrombin to platelets.

Treatment with concentrations of elastase sufficient to inhibit thrombin-induced platelet aggregation altered the binding of thrombin to platelets (Fig. 5). The Scatchard plot of thrombin binding before elastase treatment (Fig. 5, closed circles) was nonlinear. As analyzed by the method of Munson and Rodbard (34), these findings suggested two classes of binding sites (F ratio test of two-site versus one-site model: $F = 22.8$; $P < 0.001$). Before elastase treatment there were 31 ± 83 high affinity receptors per platelet ($K_d = 0.093 \pm 0.054$ nM) and $1,500 \pm 300$ low affinity receptors ($K_d = 122 \pm 250$ nM). After elastase treatment (Fig. 5, open circles), the steep portion of the curve is attenuated, and the presence of high affinity binding sites is less apparent. Analyzed as above, there were 5.4 ± 6.4 high affinity receptors per platelet ($K_d = 0.042 \pm 0.003$ nM) after elastase treatment and $1,100 \pm 4,300$ low affinity receptors ($K_d = 212 \pm 250$ nM).

The results of seven experiments of this type are summarized in Table I, demonstrating that the number of high affinity binding sites per platelet was diminished from 31 ± 10 before elastase exposure to 12 ± 13 after elastase treatment ($P < 0.05$). The K_d for the high affinity receptor was 0.10 ± 0.06 nM; the decrease in the K_d after elastase treatment was not statistically significant. Neither the number of low affinity thrombin binding sites/platelet ($4,500 \pm 5,100$) nor the K_d (98 ± 90 nM) was significantly altered by incubation with elastase under these conditions.

In three of the seven thrombin binding experiments summarized in Table I the number of elastase molecules per platelet was approximately 6×10^3 (the number of molecules/platelet in Fig. 1 was 1.5×10^4). Two of the three incubations resulted in a reduction in the number of high affinity receptors $> 25\%$. With an increase in both the ratio of molecules of elastase/platelet to $\sim 2.2 \times 10^4$ and in the incubation time to 20 min, all four subsequent determinations resulted in reductions of $> 45\%$ in the number of high affinity receptors per platelet. In two of these four experiments high affinity receptors were no longer detectable after elastase treatment.

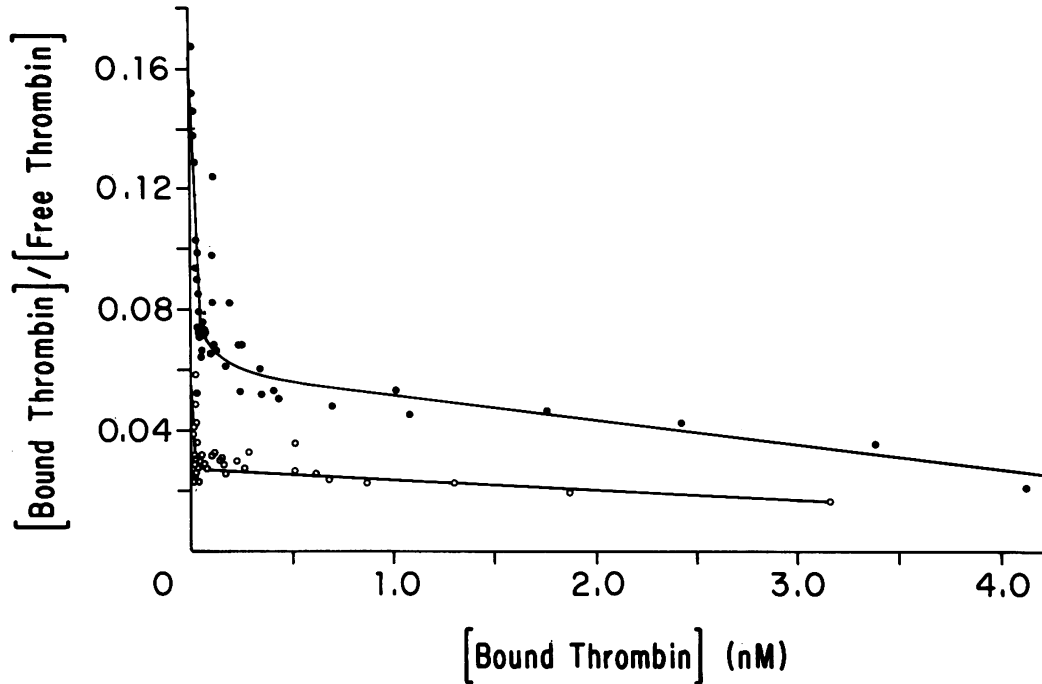


Figure 5. Scatchard plot analyses of the binding of thrombin to normal platelets (●) and elastase treated platelets (○). Washed platelets (2×10^9 /ml) were incubated with elastase (74 nM; 2.23×10^4 molecules/platelet) or buffer for 20 min at 37°C. After adding α_1 -antitrypsin (0.3 μ M), the platelets were equilibrated for 15 min at 23°C, diluted (3×10^9 /ml), and incubated with various concentrations of 125 I-thrombin (0–200 nM) for 10 min at 23°C. Bound and free thrombin were measured as described in the methods. Data points represent uncorrected binding from a single experiment. Lines

were derived from the formula (46) $B = \sum_{n=1}^2 \frac{FR_n}{F + N_n}$, where B is the concentration of receptor-bound thrombin, F is the concentration of free thrombin, R_n is the total concentration of receptor_n (occupied and unoccupied), and K_n is the equilibrium dissociation constant of receptors belonging to binding class n . Values for the constants R_n and K_n are obtained from the Ligand program described in Methods.

Effect of elastase on platelet surface glycoproteins. Human leukocyte elastase (0.8 μ M) was incubated with washed 3 H-labeled platelets and the results analyzed after SDS-PAGE by fluorography (Fig. 6) and densitometry. In comparison with untreated platelets (Fig. 6 a), there was a marked difference in platelet surface membrane glycoproteins after treatment with elastase (Fig. 6 b). First, elastase treatment resulted in a decrease in the relative percentage of glycoprotein in the range of $M_r = 130,000$ to 150,000. Proteins in this range include glycoproteins Ib and IIb (47). Concomitant with this was a relative increase in glycoprotein at $M_r = 102,000$.

The supernatants from both elastase-treated and untreated platelets were similarly analyzed. Untreated platelets (Fig. 6 c) showed one band at $M_r = 131,000$ believed to be glyocalicin (47). Compared with control platelets the supernatant from the elastase-treated platelets (Fig. 6 d) showed new protein bands at approximate molecular weights of 88,000, 35,000, and 29,000. Protein at $M_r = 88,000$ is in the region of glyco-

protein V (48, 49). However, the quantity of protein at this molecular weight raises the possibility that at least part of the protein was formed by elastase cleavage of a higher molecular weight glycoprotein precursor.

To explore further the action of elastase on platelet surface glycoproteins, 3 H-labeled platelets (10^{10} /ml) were incubated with elastase (50 nM; 3,000 molecules elastase/platelet), a 16-fold lesser concentration than used in Fig. 6. As demonstrated in Fig. 7 (lanes 1–5) the relative quantity of platelet surface glycoprotein at molecular weight 130,000 to 140,000 was markedly reduced within 30 s of incubation; additional reduction in protein at this molecular weight occurred with prolonged

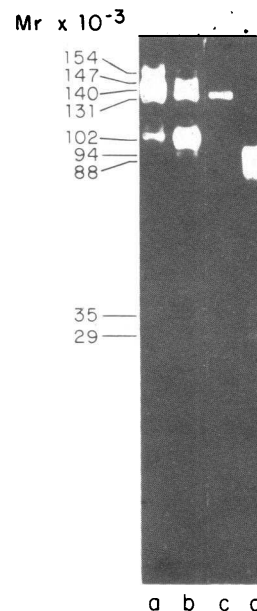


Figure 6. Effect of elastase on platelet surface sialoglycoproteins. 3 H-surface labeled platelets (2×10^9 /ml) were incubated with or without elastase (0.8 μ M) for 1 min at 23°C. α_1 -Antitrypsin (4 μ M) was then added to inhibit elastase activity. The platelets were sedimented, the supernatant harvested, and the platelets washed and resuspended in buffer (10^{10} /ml). Samples (10^5 dpm/lane) of the resuspended pellets and supernatants were reduced and analyzed by SDS-PAGE and fluorography. (a) Control platelets not treated with elastase; (b) platelets after treatment with elastase; (c) supernatant from control platelets not treated with elastase; (d) supernatant from elastase-treated platelets.

Table 1. Platelet Thrombin Receptors: Effect of Elastase

Platelets	High affinity receptor		Low affinity receptor	
	Sites/platelet*	K_d nM	Sites/platelet	K_d nM
Normal‡	31±10§	0.10±0.06	4,600±5,100	98±90
Elastase treated	12±13§	0.037±0.05	3,800±8,300	77±96

* The number of thrombin binding sites/platelet and K_d were calculated by analysis of seven binding experiments using the "Ligand" binding program as detailed in Methods. Values are means±standard deviation.

‡ Platelets (2×10^9 /ml) were treated with 14–74 nM elastase (0.6 to 2.2×10^4 molecules/platelet) or with buffer alone for 1–20 minutes at 37°C.

§ $P < 0.05$ by Wilcoxon signed-ranked test.

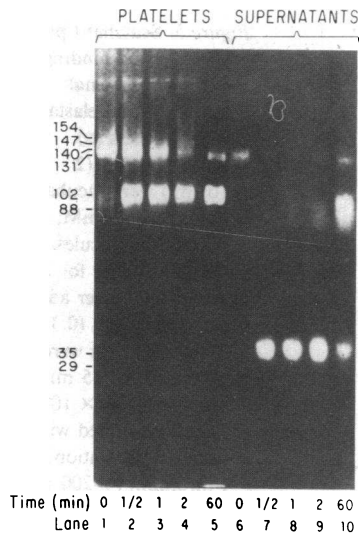


Figure 7. Alteration in glycoprotein structure by elastase as a function of time. Labeled platelets ($10^{10}/\text{ml}$) were incubated with elastase (50 nM) at 23°C. At the times indicated 0.1 ml aliquots were mixed with α_1 -antitrypsin (1 μM) to inhibit elastase activity. The platelets were sedimented by centrifugation and the supernatants removed and harvested. The platelets, resuspended in 50 μl and the supernatants (each $\sim 80 \mu\text{l}$) were mixed with equal volumes of SDS-DTT and heated 100°C for 3 min.

Samples were electrophoresed (20,000 dpm/lane) and fluorograms of the platelets and of the corresponding supernatants were obtained by exposure of the gel to the film for 30 days. (Lanes 1–5 contain $\sim 3 \mu\text{l}$ of a 1:10 dilution of the platelet pellet after mixing with SDS-DTT. Lanes 5–9 contain 5 μl of undiluted platelet supernatant after mixing with SDS-DTT; lane 10 contains 3.5 μl .)

incubation. Concurrently, there was an increase in the concentration of radiolabeled protein at $M_r = 102,000$, suggesting that it is a cleavage product of the $M_r = 140,000$ glycoprotein.

The supernatants from platelets after elastase treatment were similarly analyzed (Fig. 7, lanes 6–10). The supernatants prior to elastase treatment showed a single band at $M_r = 131,000$ believed to be glyocalicin as in Fig. 6. Within 30 s of incubation with elastase, protein at $M_r = 131,000$ is reduced, and a new band appears at $M_r = 35,000$. With longer incubation times, protein is observed at $\sim M_r = 88,000$, migrating ahead of the $M_r = 102,000$ band observed in lanes 1–5. At 60 min a faint band at $M_r = 29,000$ appears.

In experiments not shown, incubation of platelets with elastase in the presence of EDTA (1 mM) or antipain (0.2 mM) gave similar results, suggesting that calcium activated protease was not an essential component of this interaction (50).

Immunoenzymatic analysis of the effect of elastase on glycoproteins IIb and IIIa. The effects of elastase on platelet glycoproteins IIb and IIIa were analyzed by Western blot technique followed by immunoenzymatic detection of the glycoprotein with avidin-acid phosphatase complex (Fig. 8). The treatment of platelets with concentrations of elastase sufficient to inhibit thrombin-induced platelet aggregation did not alter the molecular weight or the apparent quantity of platelet glycoproteins IIb and IIIa. Glycoprotein IIb is detected at $M_r = 136,000$ (Fig. 8 a). Treatment of platelets with elastase does not alter its molecular weight (Fig. 8 b). Likewise, glycoprotein IIIa is apparent at $M_r = 106,000$ both before and after elastase treatment (Fig. 8 c, d). The lighter band at $M_r = 122,000$ in Fig. 8 c, d is in the position that glycoprotein IIIa is found after reduction. Similar analysis of samples after reduction with dithiothreitol confirmed that treatment of platelets with elastase did not alter the electrophoretic mobility of glycoproteins IIb and IIIa (data not shown).

Immunoprecipitation of glycoprotein Ib. Platelets were ra-

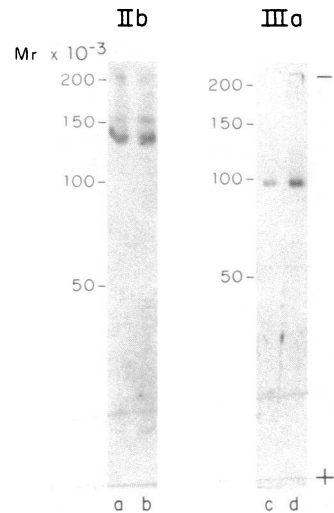


Figure 8. Immunoenzymatic analysis of the effect of elastase on platelet glycoproteins IIb and IIIa. Washed platelets ($8.9 \times 10^9/\text{ml}$) were incubated with or without elastase (80 nM; 5,400 molecules/platelet) for 1 min at 37°C. α_1 -Antitrypsin 0.2 μM then was added to inhibit elastase activity. The platelets were washed, resuspended in buffer, and solubilized in SDS. After electrophoresis and electrotransfer, samples (3 $\mu\text{g}/\text{lane}$) were analyzed by immunoenzymatic detection with avidin-acid

phosphatase complex as in Methods. (Left) Detection of glycoprotein IIb in elastase-treated and untreated platelets is shown in lanes a and b, respectively. (Right) Detection of glycoprotein IIIa in elastase-treated and untreated platelets is shown in lanes c and d, respectively.

diolabeled, solubilized with Triton X-100 and incubated with Immunobeads coated with either AP-1 or normal mouse IgG. Approximately 23% of the radioactivity from solubilized platelet membranes not pretreated with elastase bound to the AP-1 coated beads. In contrast, after elastase treatment of washed platelets, 0.9% of the radioactivity from the solubilized platelet membrane bound to the AP-1 coated beads. Control beads coated with normal mouse IgG bound less than 0.5% of total radioactivity both before and after elastase treatment.

The immunoprecipitate formed by the AP-1 antibody with ^3H -labeled platelet membranes was analyzed by polyacrylamide gel electrophoresis (10,000 dpm/lane) after solubilization in SDS. As shown by fluorography, the control bead immunoprecipitate from either solubilized normal platelets (Fig. 9 c)

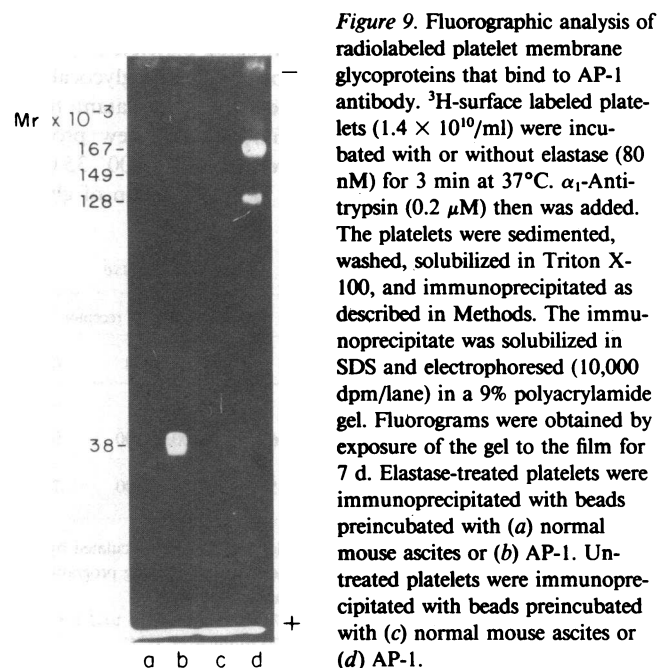


Figure 9. Fluorographic analysis of radiolabeled platelet membrane glycoproteins that bind to AP-1 antibody. ^3H -surface labeled platelets ($1.4 \times 10^{10}/\text{ml}$) were incubated with or without elastase (80 nM) for 3 min at 37°C. α_1 -Antitrypsin (0.2 μM) then was added. The platelets were sedimented, washed, solubilized in Triton X-100, and immunoprecipitated as described in Methods. The immunoprecipitate was solubilized in SDS and electrophoresed (10,000 dpm/lane) in a 9% polyacrylamide gel. Fluorograms were obtained by exposure of the gel to the film for 7 d. Elastase-treated platelets were immunoprecipitated with beads preincubated with (a) normal mouse ascites or (b) AP-1. Untreated platelets were immunoprecipitated with beads preincubated with (c) normal mouse ascites or (d) AP-1.

or solubilized elastase-treated platelets (Fig. 9 *a*) moved with the buffer front. In contrast, the AP-1 immunoprecipitate from untreated platelets (Fig. 9 *d*) contained two protein bands, a dark band of approximate $M_r = 167,000$ and a light band at $M_r = 130,000$. These correspond to the reported molecular weights of glycoprotein Ib, 166,000, and glyocalicin, 135,000 (51–53). Proteins of $<20,000 M_r$ migrated with the buffer front so that the intensely reactive band of $M_r \sim 20,000$ reported by Collier et al. (42) was not observed. After treatment with elastase (Fig. 9 *b*), the relative intensity of the bands at $M_r 167,000$ and $130,000$ was reduced. Concomitant with this was the appearance of a protein band at $M_r \sim 35,000$, corresponding to the $M_r = 35,000$ band found in the supernatant of elastase-treated platelets. That this peptide contains the antigenic site for the AP-1 monoclonal antibody and is not merely adhering nonspecifically to the AP-1 coated beads remains to be proven.

These studies demonstrate that treatment of whole platelets with elastase results in the degradation of glycoprotein Ib.

Effect of elastase on glycoprotein Ib. Platelet membrane glycoprotein was immunopurified with Immunobeads coated with the AP-1 anti-glycoprotein Ib antibody. Human leukocyte elastase (5 nM) was incubated with immunopurified, ^3H -labeled glycoprotein and the mixtures were analyzed by SDS-PAGE after reduction with dithiothreitol. As shown by fluorography (Fig. 10), the molecular weight of the glycoprotein immunoisolate prior to elastase treatment was $\sim 140,000$. Upon incubation with elastase for 0.5 min, the relative intensity of the band at 140,000 was reduced and a new band appeared at $M_r \sim 94,000$. After one min, most of the radioactivity was found in the $M_r \sim 94,000$ band. No additional degradation was observed with incubation for up to 60 min (Fig. 10, lanes *a–e*). In the control experiment (Fig. 10, lanes *f–j*), incubation of the immunoisolate in the absence of elastase did not result in the formation of a lower molecular weight peptide.

These experiments demonstrated that elastase-mediated proteolysis of the $M_r = 140,000$ glycoprotein resulted in the formation of a $M_r \sim 94,000$ cleavage product. On SDS-PAGE

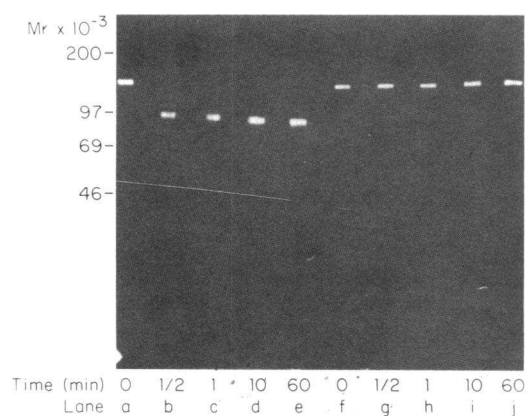


Figure 10. Effect of elastase on platelet glycoprotein Ib. Glycoprotein Ib was immunoprecipitated and eluted as described in Methods. The isolated glycoprotein was incubated for various times with 5 nM elastase (lanes *a–e*) or with buffer (lanes *f–j*) at 37°C prior to heating at 100°C for 3 min with an equal volume of 0.06 M Tris-HCl, 2% SDS, 5% glycerol, 0.001% Bromphenol blue, and 2% DTT. After electrophoresis in a 9% polyacrylamide gel, fluorograms were obtained by exposure of the gel to the film for 7 d.

this peptide comigrated with the $M_r \sim 102,000$ glycopeptide found in elastase-treated platelets (Fig. 6 *d*), and it migrated more slowly than the $M_r = 88,000$ glycopeptide appearing in the supernatant of elastase-treated platelets (gel not shown). These findings suggested that elastase-mediated proteolysis of membrane-bound glycoprotein Ib resulted in formation of a $M_r = 94,000$ to $102,000$ membrane-associated glycopeptide (Figs. 6, 7, and 10).

Glycoprotein V has a molecular weight of 68,000 to 85,000 (48, 49), raising the possibility that a portion of the $M_r = 88,000$ glycopeptide appearing in the supernatant of elastase-treated platelets is glycoprotein V. However, immunoprecipitation studies similar to those above using anti-glycoprotein V antibody (the kind gift of Dr. David Phillips, Gladstone Foundation Laboratories, San Francisco, CA) (54), demonstrated that elastase treatment did not increase the concentration of glycoprotein V in platelet supernatants (data not shown).

Discussion

These studies demonstrate that physiologic concentrations of human leukocyte elastase inhibit thrombin-induced platelet aggregation in a time- and dose-dependent manner. Leukocyte elastase also inhibits ristocetin-induced platelet agglutination and the release of [^{14}C]serotonin from platelets. This loss of functional activity is associated with (*a*) a reduction in the number of high affinity thrombin binding sites per platelet and (*b*) the proteolysis of platelet glycoprotein Ib.

The low concentration of elastase used to demonstrate inhibition of thrombin-induced platelet aggregation in this study indicates that platelets are highly susceptible to elastase and suggests that these findings are of physiologic relevance. As blood coagulates, polymorphonuclear leukocytes degranulate and release elastase (2, 3). Several potentially toxic reactive oxygen species also can be released by degranulated leukocytes (55). α_1 -Antitrypsin, the major plasma inhibitor of elastase, is particularly susceptible to oxidants that can react with the methionine at the reactive center of the inhibitor, thus inactivating its protease-inhibiting activity (10). In a cell contact system Campbell et al. (56) demonstrated that α_1 -antitrypsin inhibited about 80% of PMN-derived proteolytic activity. Although PMN-derived oxidants did not potentiate elastase-mediated proteolysis in this system, Weiss and Regiani (57) showed that concentrations of α_1 -antitrypsin less than 250 $\mu\text{g}/\text{ml}$ can be inhibited by a PMN-mediated H_2O_2 -dependent mechanism. Additionally, Matheson et al. (58) demonstrated that oxidation of α_1 -antitrypsin by myeloperoxidase is optimized at acidic pH (6.2) such as might be found in an area of thrombosis or inflammation. Conditions present during coagulation might thus facilitate the elastase-mediated proteolysis of platelets as detailed in this study.

The interaction of physiologic concentrations of elastase with platelets is rapid. At an elastase concentration of 50 nM (3,000 elastase molecules/platelet), major quantities of glycoproteins Ib were degraded within 60 s. This correlates with functional studies detailing loss of thrombin responsiveness within this time interval when similar elastase concentrations are used. The finding that glycoprotein Ib is affected by elastase treatment remains compatible with the hypothesis that thrombin might interact with glycoprotein Ib (59–62).

Elastase treatment of glycoprotein immunopurified with anti-glycoprotein Ib antibody produced a peptide with the same

electrophoretic mobility as the $M_r \sim 102,000$ glycopeptide on the surface of elastase-treated platelets, suggesting that it is a glycoprotein Ib fragment. However, the origin of the $M_r = 88,000$ protein in the supernatant of elastase-treated platelets remains unknown. The difference in the molecular weights of these two peptides is similar to that between the α -chain of glycoprotein Ib ($M_r = 143,000$) (51, 52) and glycolaligin ($M_r = 135,000$) (47, 53). This raises the possibility that the $M_r = 88,000$ peptide might result from elastase-mediated cleavage of glycolaligin. Proof of this hypothesis requires further experimental studies.

As previously reported (30, 63–65), analysis of thrombin-platelet binding kinetics demonstrates both high and low affinity thrombin binding sites. The number of low affinity binding sites and the K_d reported in this study ($4,600 \pm 5,100$ sites/platelet, $K_d = 98 \pm 90$ nM) is compatible with the findings of the above authors. However, using computer analysis we have found both the number of high affinity sites per platelet (31 ± 10) and the K_d (0.10 ± 0.06) to be less than the values previously published (290 to 1,500 sites/platelet; $K_d = 0.2$ to 3.2 nM). We attribute these differences strictly to a more accurate resolution of kinetic constants by the Ligand program for fitting multiple binding site data (34), which was not available to the above investigators. (In studies not shown, analysis using the technique of Chamness and McGuire (66) revealed two classes of binding sites, 340 sites with a K_d of ~ 1.6 nM and 14,000 sites with a K_d of ~ 180 nM, values similar to those reported for the high and low affinity sites.) It appears then that accurate resolution of kinetic constants for the high affinity thrombin receptor by linear regression and extrapolation of the Scatchard plot results in a 10-fold higher value compared to mathematical resolution of the same data using the Ligand program.

A criterion for defining "specific" binding of thrombin to platelets is that it is correlated with physiologic effects (30). Both platelet secretion and platelet aggregation occur at thrombin concentration of 0.1 to 1 nM (Figs. 1 and 4). From thrombin binding studies (Fig. 5) it can be calculated that this range of concentrations encompasses the steep portion of the thrombin binding curve. (An added thrombin concentration of about 1 nM corresponds approximately to a bound/free ratio of 0.10 and to a bound thrombin concentration of 0.1 nM. At a platelet concentration of 2×10^9 /ml, this is equivalent to 30 molecules thrombin/platelet). Approximately 30 thrombin molecules binding per platelet will thus maximize platelet secretion. This suggests that the high affinity sites also are the specific receptors for both the aggregation and secretion reactions. As noted by others (30) it remains possible that the binding of additional thrombin molecules per platelet may produce other as yet unidentified effects on the platelet.

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