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Research Article

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Estimated glucose production at 5-6 wk of starvation is reduced to approximately 86 g/24 hr. Of this amount the liver contributes about one-half and the kidney the remainder. Approximately all of the lactate, pyruvate, glycerol, and amino acid carbons which are removed by liver and kidney are converted into glucose, as evidenced by substrate balances across these organs.

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Liver and Kidney Metabolism during Prolonged Starvation

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ABSTRACT This study quantifies the concentrations of circulating insulin, growth hormone, glucose, free fatty acids, glycerol, β -hydroxybutyrate, acetoacetate, and alpha amino nitrogen in 11 obese subjects during prolonged starvation. The sites and estimated rates of gluconeogenesis and ketogenesis after 5–6 wk of fasting were investigated in five of the subjects.

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Estimated glucose production at 5–6 wk of starvation is reduced to approximately 86 g/24 hr. Of this amount the liver contributes about one-half and the kidney the remainder. Approximately all of the lactate, pyruvate, glycerol, and amino acid carbons which are removed by liver and kidney are converted into glucose, as evidenced by substrate balances across these organs.

INTRODUCTION

Both liver and kidney have the enzymatic machinery to synthesize glucose (1, 2) and the liver is generally recognized as the major source of blood glucose after a brief postabsorptive period. Estimations of net splanchnic glucose production in man after an overnight fast vary from 122 to 420 g/day (3–7), an amount which corresponds to approximately one-half of the basal caloric turnover. Were glucose production from amino acids to continue at this rate, all body protein would be consumed within several weeks. That a diminution of hepatic gluconeogenesis during prolonged starvation does

occur is evidenced, however, by a reduction in urinary nitrogen to 3–5 g/day (8, 9). In addition, ammonia replaces urea as the principal excreted nitrogenous product (8), and in view of the recent observations coupling renal ammoniogenesis with gluconeogenesis (10, 11), an evaluation of the renal and splanchnic roles in overall substrate balance is warranted. Preliminary reports of some of the data to be described have appeared (12, 13).

METHODS

Subjects. 11 obese subjects were admitted to the Clinical Center of the Peter Bent Brigham Hospital for therapeutic starvation (Table I). Each had previously been on various dietary regimes without success and volunteered for the studies described. They were informed of the nature, purpose, and risks involved in both the starvation and catheterization procedures.

On admission all had normal hemograms, urinalyses, chest and abdominal roentgenograms, electrocardiograms (except F.N. who had left ventricular hypertrophy), thyroid hormone levels, serum enzymes (alkaline phosphatase, lactic dehydrogenase, glutamic-oxalacetic transaminase), and normal concentrations of serum protein, cholesterol, phospholipids, triglycerides, electrolytes, urea nitrogen, and creatinine. Daily intake during starvation consisted of one multivitamin capsule (Unicap, Upjohn Co., Kalamazoo, Mich.), 17 mEq of NaCl, 1500 ml of water, and, intermittently, 13 mEq of KCl. Subjects were encouraged to participate in occupational therapy and daily walks. Of more importance, a strong rapport between physician and patient was established, leading, in most cases, to maintenance of weight reduction after termination of the studies.

Blood and urine collections. During the prestarvation period, an intravenous glucose tolerance test was performed in each subject by rapid intravenous injection of glucose (0.5 g/kg body weight) with blood drawn for analysis at 10 min intervals for 1 hr after the infusion. For the remainder of the studies blood specimens were obtained at 7 a.m. before the subjects arose from bed, at 3 days prestarvation, on the morning fasting was initiated, 3 days after initiation of starvation, and at weekly intervals there-

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TABLE I
Clinical Data

Subject	Age	Sex	Height	Weight			Loss	Final body surface	i.v. GTT*	Duration of starvation
				Initial	Final	Difference				
	<i>yr</i>		<i>cm</i>	<i>kg</i>	<i>kg</i>	<i>kg</i>	<i>%</i>	<i>m²</i>	<i>kg</i>	<i>day</i>
A. B.	19	M	180.3	125.2	101.8	23.4	20.5	2.25	1.2	35
R. S.	21	M	180.0	160.6	138.2	22.4	14.0	2.53	1.4	35
R. R.	28	M	177.8	178.6	159.6	19.0	10.6	2.70	0.9	35
F. F.	32	M	180.3	127.2	105.6	21.6	16.9	2.25	1.0	39
P. C.	33	M	170.8	169.0	148.7	20.3	12.0	2.49	1.4	35
F. N.	49	M	177.5	132.6	109.8	22.8	17.2	2.26	0.6	38
C. R.	16	F	167.6	104.1	88.4	15.7	15.0	1.97	2.1	35
D. D.	20	F	159.0	108.9	93.0	15.9	14.6	1.94	1.7	35
M. L.	26	F	176.0	147.3	124.2	23.1	15.7	2.37	1.2	40
S. C.	33	F	163.0	117.0	103.0	14.0	12.0	2.07	0.7	35
M. B.	43	F	173.0	123.5	98.8	24.7	20.0	2.14	1.0	41

* Intravenous glucose tolerance test (0.5 g/kg) performed before weight reduction. Values expressed as absolute per cent disappearance per min.

after. These were usually taken from either antecubital vein using a tourniquet only when necessary. Whole blood, plasma, or serum were analyzed for acetoacetate, β -hydroxybutyrate, glycerol, glucose, free fatty acids, α -amino nitrogen, insulin, and growth hormone. Urine was collected in refrigerated plastic containers, and at the end of 24 hr (7:00 a.m.) the volume was measured and aliquots taken for analysis of total nitrogen, urea, ammonia, creatinine, uric acid, acetoacetate, and β -hydroxybutyrate. The various techniques have been described (8, 14) including that of growth hormone (15). During the prestarvation period, subjects were maintained on a 2500 kcal per day diet consisting of 300 g carbohydrate, 100 g protein, and 85 g fat. All had previously been on a random diet with neither weight gain nor loss immediately before the study.

Blood flow. Effective renal blood flow (ERBF) was estimated in five subjects by clearances of sodium *para*-aminohippurate (16). Subject F.N. had a single determination after 28 days of starvation. They were encouraged to drink water freely on the night before the clearance study, and after rising and emptying the bladder at 7:00 a.m., they received 1000 ml of water by mouth during a 15–20 min period, followed by supplemental amounts to maintain urine flow greater than 5 ml/min. One antecubital vein was used for blood sampling and the indwelling needle kept patent with 0.9% sodium chloride. In the contralateral antecubital vein an indwelling needle was used for a priming dose of 0.04 ml/kg body weight of a 20% solution of sodium *para*-aminohippurate (PAH) and for a continuous infusion calculated to maintain a plasma concentration of PAH of 0.01–0.02 mg/ml. After 45 min for equilibration, three urine specimens were collected at successive 30-min intervals, and blood specimens were obtained midway between voidings. Effective renal blood flow was calculated by the formula: $ERBF = C_{PAH} / (1 - \text{hematocrit})$.

Catheterization. After five subjects had fasted 35–41 days they were given 100 mg of sodium pentobarbital and 50 mg of meperidine sulfate intramuscularly and then brought to the catheterization laboratory at 8 a.m. A polyethylene catheter was advanced percutaneously into the left brachial artery. A left median antecubital vein was exposed and a

Goodale-Lubin catheter (No. 7–8) was inserted and advanced to a loosely wedged position in the right hepatic vein. The femoral vein was catheterized percutaneously and a Goodale-Lubin catheter (No. 7–8) advanced to the right renal vein. The catheters were kept patent with 0.9% sodium chloride, and their positions were repeatedly ascertained by fluoroscopy with image intensification.

Blood samples were obtained simultaneously from the arterial, renal, and hepatic veins every 15 min for four to six periods in most cases. The samples were immediately prepared for analysis of oxygen, carbon dioxide, and metabolic substrates. Blood pressure and heart rate were monitored continuously during the procedure and did not change significantly. The patients received an isovolumetric replacement of the withdrawn blood by 5% human albumin in 0.9% sodium chloride.

Hepatic blood flow was estimated at the time of catheterization in two subjects (M.B. and F.F.) using Indocyanine green (17). After injection of 0.5 mg/kg body weight of dye, arterial and hepatic plasma concentrations were determined at 3-min intervals for 21 min at 805 $m\mu$ using a Beckman DU spectrophotometer. Plasma volume was calculated from the zero-time intercept of the arterial clearance curve. Hematocrit was measured in an arterial blood sample and corrected for trapped plasma and body hematocrit according to Chaplin, Mollison, and Vetter (18). Estimated hepatic blood flow (EHBf) was calculated from the clearance constant of dye for arterial plasma (k), blood volume (V), and the fractional extraction of dye in the liver (E) according to the equation: $EHBf = (kV)/E$.

Cardiac output was determined from arterial dye dilution curves after single intravenous injections of 5 mg of Indocyanine green. The area under the curve was measured by planimetry. Calibration was made by analyses of two blood samples with known dye concentrations. Statistical calculations were made according to Snedecor (19).

RESULTS

In Table II are presented the circulating levels of substrates and the concentrations of insulin and growth hor-

TABLE II
Circulating Substrate and Hormone Concentrations (Venous) in 11 Subjects

Days of Fasting...	-3	0	3	10	17	24	31	35-38
Plasma free fatty acids*	0.66 ±0.06	0.71 ±0.05	1.25 ±0.09	1.360 ±0.11	1.464 ±0.12	1.55 ±0.13	1.48 ±0.09	1.60 ±0.14
Blood β-hydroxybutyrate*	0.06 ±0.01	0.07 ±0.01	1.21 ±0.19	4.30 ±0.40	5.10 ±0.26	5.78 ±0.27	5.84 ±0.39	5.85 ±0.38
Blood acetoacetate*	0.03 ±0.01	0.03 ±0.01	0.41 ±0.06	1.00 ±0.11	1.12 ±0.11	1.27 ±0.11	1.37 ±0.13	1.34 ±0.14
Blood glycerol*	0.090 ±0.009	0.092 ±0.008	0.108 ±0.017	0.110 ±0.014	0.132 ±0.027	0.154 ±0.028	0.114 ±0.013	0.124 ±0.013
Serum α-amino nitrogen*	4.38 ±0.11	4.39 ±0.12	4.32 ±0.16	4.41 ±0.14	4.15 ±0.11	3.85 ±0.09	4.01 ±0.09	3.85 ±0.11
Blood glucose*	4.95 ±0.30	4.79 ±0.20	3.63 ±0.26	3.79 ±0.27	3.76 ±0.19	3.76 ±0.19	3.78 ±0.18	3.70 ±0.18
Serum insulin‡	45 ±6	37 ±7	20 ±5	20 ±3	17 ±4	13 ±3	17 ±3	14 ±2
Serum§ growth hormone	0.6 ±0.2	1.0 ±0.5	1.4 ±0.6	1.2 ±0.3	0.7 ±0.2	0.5 ±0.1	0.7 ±0.2	0.4 ±0.1

* Values in millimoles per liter ±SEM.
‡ Values in microunits per milliliter.
§ Values in millimicrogram per ml.

mone in the 11 subjects. There was a slight but insignificant fall in glucose and insulin concentrations between the start and the end of the dietary pretreatment (day 3 to day 0). Of note, however, is the higher level of both glucose [86.2 ±1.6 (SEM) mg/100 ml, n = 11] and insulin (37 ±7 μU/ml, n = 11) compared to an age and

sex-matched, nonobese population studied in this laboratory (glucose = 76.8 ±0.8 mg/100 ml, n = 119 and insulin = 15.8 ±1.0 μU/ml, n = 69, unpublished data, courtesy of Dr. J. S. Soeldner). As shown in Fig. 1, glucose concentration reached a nadir after 3 days of fasting and stayed unchanged thereafter. The initial fall

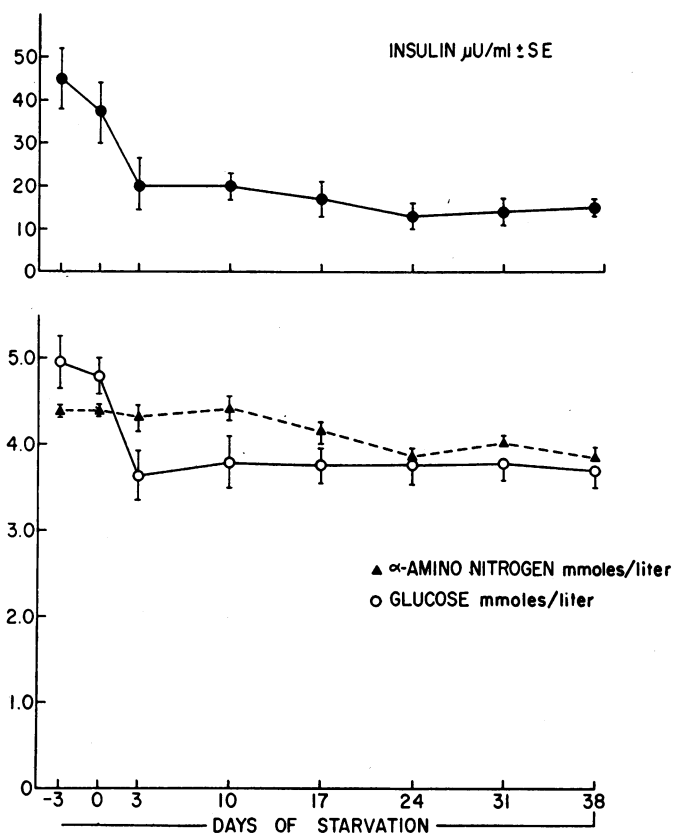


FIGURE 1 Concentration of blood glucose, serum insulin, and serum α-aminonitrogen in 11 obese subjects at various times throughout a prolonged fast.

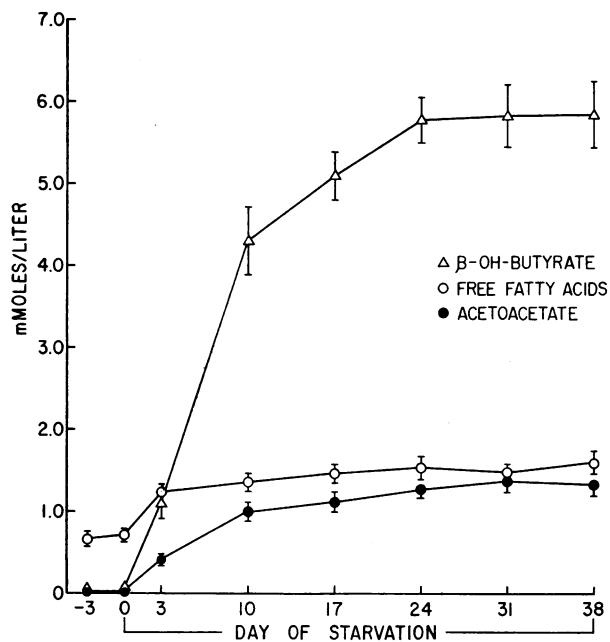


FIGURE 2 Concentration of plasma free fatty acids and blood acetoacetate and β -hydroxybutyrate in 11 obese subjects at various times throughout a prolonged fast.

in serum immunoreactive insulin paralleled that of glucose and then decreased gradually as starvation and weight loss continued. Using the paired *t* test the decrease in insulin concentration between day 3 or 10 vs. day 24 is significant ($P < 0.05$ and $P < 0.01$ respectively).

Serum levels of α -amino nitrogen also fell and the decrease is significant after 17 days of fasting ($P < 0.05$). It should be stressed that this determination is a gross representation of all the amino acids and does not reflect the individual fluctuations which may be marked, a topic currently under study (20). The concentrations of β -hydroxybutyrate and acetoacetate rose slowly for the 1st 3 days (Fig. 2), and between days 3 and 24 a greater rise occurred, especially when con-

sidered in relation to the simultaneous free fatty acid concentrations. It is interesting to note that the rise in β -hydroxybutyrate and acetoacetate was associated with an increase in the ratio between the two metabolites, changing from approximately 2 at the beginning to 4.5 on the 17th day. The changes in venous glycerol levels (Table II) were not statistically significant.

In Fig. 3 is presented the urinary nitrogen excretion of a typical subject (F.N.). During the control period there were fluctuations, primarily in urea, which comprised the major component. Daily nitrogen excretion decreased progressively for the 1st 4 wk of fasting after which it remained fairly constant at about 3–6 g/day. In the five subjects who underwent catheterization studies, the mean nitrogen excretion fell to 4.7 g/day of which 42% was ammonia (Table III). Urea excretion decreased to an average of 1.55 g/day, 33% of the total.

In Table IV the ammonia excretion after 5–6 wk of starvation has been converted into mmoles/24 hr and compared to the corresponding values for excreted milliequivalents of β -hydroxybutyrate and acetoacetate. The urinary β -hydroxybutyrate:acetoacetate ratio is 8:1 which is greater than that of arterial blood (4.5:1). Mean total β -hydroxybutyrate and acetoacetate excretion was 114 mmoles and that of ammonia 139 mmoles, the difference probably owing to the presence of other anions as well as a possible small degradatory loss of β -hydroxybutyrate and acetoacetate in the urine in spite of storage in a refrigerator at 4°C during the 24 hr collection period.

The hemodynamic data for the five subjects who underwent catheterization are presented in Table V. For the three subjects who did not have hepatic flow determined directly and for the subject in whom no renal flow determination was made, it was assumed that 20% of the cardiac output for each represented an approximate estimation for liver and kidney blood flow (21), and further extrapolations were based on this assumption.

Successive estimates of renal blood flow were made during starvation. It is interesting to note that they failed to demonstrate any significant change (Fig. 4).

TABLE III
Grams of Urinary Nitrogen/24 hr after 5–6 wk of Starvation*

Patient	Total	Ammonia	Urea	Uric acid	Creatinine	Unidentified
A. B.	6.63	1.47	3.84	0.11	0.70	0.51
F. F.	4.53	2.70	0.88	0.09	0.67	0.19
F. F.	4.30	1.91	1.43	0.13	0.59	0.25
M. L.	2.77	1.56	0.39	0.06	0.39	0.37
M. B.	5.06	2.11	1.24	0.09	0.91	0.71
Mean \pm SEM	4.66 \pm 0.62	1.95 \pm 0.22	1.55 \pm 0.60	0.10 \pm 0.01	0.65 \pm 0.08	0.41 \pm 0.09

* Values are the mean of the last 3 days of starvation.

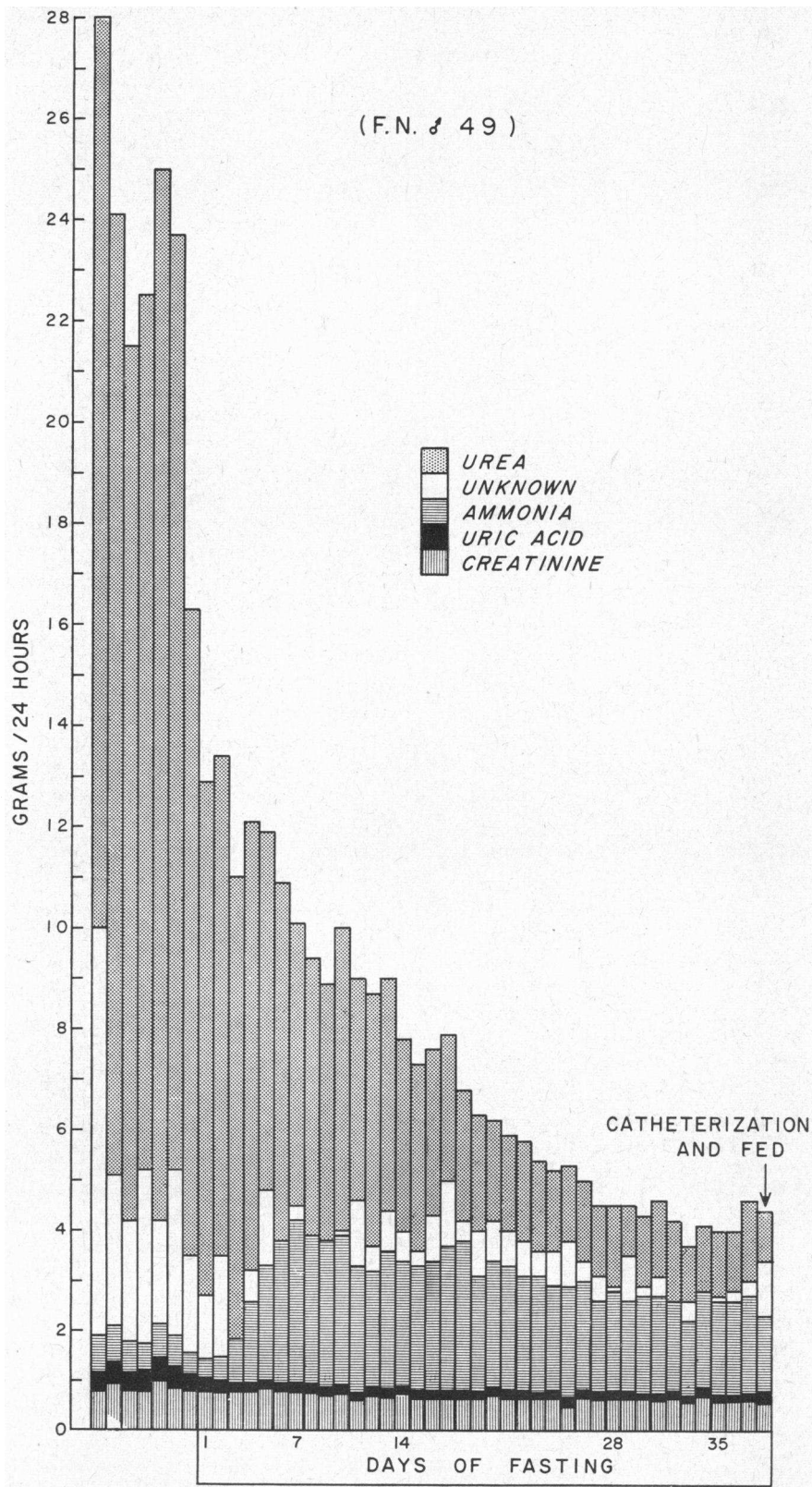


FIGURE 3 Daily urinary nitrogen excretion in a male subject fasted for 5-6 wk showing baseline fluctuations during the control period and the dramatic decrease with progressive starvation with ammonia becoming the primary excreted product.

TABLE IV
Urinary β -Hydroxybutyrate, Acetoacetate, and Ammonia Excretion (mmoles/24 hr) after 5-6 wk of Starvation

Patient	β -hydroxybutyrate	Acetoacetate	β -hydroxybutyrate and acetoacetate	Ammonia
	mmoles/24 hr			
A. B.	85	8	93	105
F. F.	138	9	147	193
F. N.	85	16	101	136
M. L.	85	—	—	111
M. B.	100	15	115	151
Mean \pm SEM	99 \pm 10	12 \pm 2	114 \pm 12	139 \pm 16

* Values are the mean for the last 3 days of starvation.

The arterial levels of the various substrates after 35-41 days of starvation are presented in Table VI, as are the balances across the splanchnic and renal beds. The oxygen content and lactate and pyruvate concentrations are in good agreement with standard values obtained in nonfasting subjects, and the decreased CO_2 content is in keeping with the ketoacidosis. Of note is the low splanchnic respiratory quotient. Significant extractions of lactate, pyruvate, α -amino nitrogen, glycerol, and free fatty acids and production of β -hydroxybutyrate, acetoacetate, and glucose are observed. Although the lactate:pyruvate ratio is equal in the hepatic vein and arterial blood, the ratio of β -hydroxybutyrate:acetoacetate is lower in hepatic vein than in arterial blood, reflecting a splanchnic production of these two compounds in a ratio of 1.8:1. Lactate, pyruvate, glycerol, and α -amino nitrogen are extracted both by the splanchnic bed and by the kidney. Free fatty acids and β -hydroxybutyrate are also extracted by the kidney, but there is a surprising and unexpected production of acetoacetate.

TABLE V
Blood Flow Rates after 5-6 wk of Starvation*

Patient	Cardiac output	Hepatic blood flow	Renal blood flow
	liters/min		
A. B.	6.1	1.22 \ddagger	1.53
F. F.	4.7	1.25	1.21
F. N.	4.2	0.84 \ddagger	1.47
M. L.	4.8	0.96 \ddagger	0.96 \ddagger
M. B.	4.5	1.13	0.88
Mean \pm SEM	4.8 \pm 0.3	1.08 \pm 0.08	1.21 \pm 0.13

* Values in liters/min.

\ddagger Regional flow estimated at 20% of cardiac output (21).

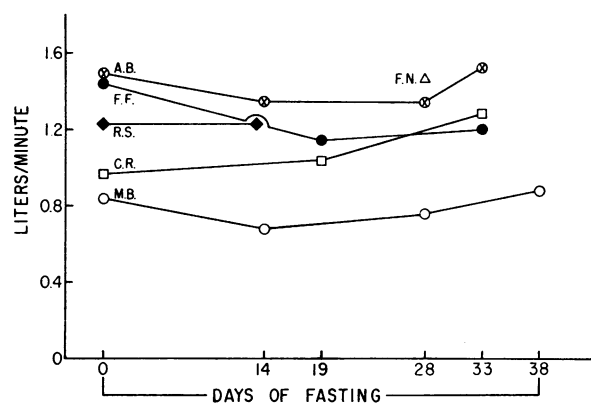


FIGURE 4 Successive determinations of effective renal plasma flow (C_{PAH}) in five subjects during starvation.

DISCUSSION

Blood insulin and substrate concentrations. Many substances including pituitary hormones, glucagon, corticosteroids, catecholamines, and prostaglandins have been shown to influence the release of free fatty acids from adipose tissue, but the possible role of insulin as the primary hormonal regulator of tissue catabolism, and thus peripheral fuel mobilization, has again been emphasized (14).

Since methods for determination of insulin in serum have become available, it has been demonstrated that obese subjects, with or without decreased glucose tolerance, have elevated insulin concentrations (22-25). Hirsch and Gallian have shown that adipose tissue cells from obese subjects exhibit hypertrophy as well as hyperplasia (26), and that insulin insensitivity is correlated with an increased lipid content in each cell (27). After the subjects lost weight and reduced their adipose cell size, insulin sensitivity and serum immunoreactive insulin levels returned to normal.

When the present obese subjects are compared to previously studied normal subjects in our laboratory (14), the obese patients exhibit the expected increased basal levels of insulin, glucose, and free fatty acids after an overnight fast. However, after 3 and 7 days of starvation substrate levels were not different between normals and obese subjects. On the other hand, insulin concentrations were 8.0 ± 0.7 and $7.7 \pm 0.5 \mu\text{U/ml}$ in the normals compared with 20.0 ± 5.0 and $20.0 \pm 3.0 \mu\text{U/ml}$ in the obese. Thus the persistent hyperinsulinemia observed in the starved obese subjects suggests a "relative" insulin resistance of the lipolytic mechanism of their adipose tissue during fasting. Similar results have been observed by Solomon, Ensinck, and Williams (28).

Substrate balances. Lactate, pyruvate, α -amino nitrogen, and glycerol are essentially three carbon substrates, and their sum divided by two gives an esti-

TABLE VI
Blood, Serum, and Plasma Levels

	O ₂	CO ₂	RQ†	Lactate	Pyruvate
Arterial concentrations	7.45 ±0.26	14.62 ±0.76	—	0.47 ±0.04	0.055 ±0.006
Splanchnic arterial-venous differences	2.25 ±0.07	-0.81 ±0.13	0.37 ±0.06	0.19 ±0.05	0.019 ±0.007
Renal arterial-venous differences	0.92 ±0.12	-0.61 ±0.12	0.77 ±0.21	0.043 ±0.007	0.004 ±0.001

* Values in mmoles per liter whole blood (plasma for free fatty acids and serum for alpha amino nitrogen). Each value is the mean with SEM for five subjects (A. B., F. F., F. N., M. L., and M. B.). For each subject, the value is the mean of four to six observation periods.

† Respiratory quotient.

mate of the amount of glucose that could be formed from these precursors if all were converted into hexose. In the splanchnic bed, 0.193 mmoles of glucose could be added to each liter of blood by these substrates, a value close to the observed 0.172 mmoles/liter. Across the kidney, the observed glucose production was 0.122 mmoles/liter and the extrapolated value from substrate differences, 0.087 mmoles/liter. These data support an optimal conversion of glucogenic substrate into glucose, presumably facilitated by the elevated levels of free fatty acids present during starvation (See Fig. 2 and Table II). Free fatty acid oxidation inhibits rate-limiting glycolytic enzymes (29), promotes increased activity of key gluconeogenic enzymes (30), and furnishes reduced pyridine nucleotides essential for glucose synthesis (31, 32).

Table VII presents balance data of free fatty acid,

TABLE VII
Balances of Free Fatty Acids, β -hydroxybutyrate, Acetoacetate, and Respiratory gases

	Splanchnic*	Renal*
	mmoles/liter	
Free fatty acids	0.208	0.085
β -hydroxybutyrate	-0.335	0.279
Urine	—	-0.079
Metabolic balance	-0.335	0.191
Acetoacetate	-0.180	-0.047
Urine	—	-0.008
Metabolic balance	-0.180	-0.055
Oxygen equivalents‡	2.56	2.49
Observed oxygen balance	2.25	0.92
Carbon dioxide equivalents§	-1.26	-1.90
Observed CO ₂ balance	-0.81	-0.61

* Values as mmoles per liter in whole blood.

‡ Oxygen equivalent: 23 for free fatty acid, 4.5 for β -hydroxybutyrate, and 4.0 for acetoacetate.

§ Carbon dioxide equivalent: 16 for free fatty acid, 4.0 for β -hydroxybutyrate, and 4.0 for acetoacetate.

β -hydroxybutyrate, acetoacetate, and respiratory gases. In the present study the liver extracted 19% and the kidney 8% of the arterial free fatty acid content. The latter figure, however, is probably high, the mean being elevated primarily by data from subject MB whose kidney inexplicably extracted 0.332 mmoles free fatty acids/liter of plasma. These findings and the quantities of oxygen consumed and carbon dioxide produced by the splanchnic and renal tissues are in good agreement with the data of others (4, 5, 7, 32, 33). Yet, there are stoichiometric imbalances among free fatty acids, β -hydroxybutyrate, acetoacetate, and respiratory gases. The data show greater arterio-venous differences for free fatty acids across the splanchnic and renal beds than can be accounted for by the production of β -hydroxybutyrate, acetoacetate, and carbon dioxide, and the consumption of oxygen. Possible explanations for these imbalances are repetitive errors in analysis (particularly free fatty acids such as in subject M.B.) or the lack of information pertaining to other reactions such as triglyceride synthesis (34). Of interest is the renal uptake of β -hydroxybutyrate and the production of acetoacetate, accentuated all the more when corrected for urinary loss. The data show that renal handling of these substrates is not a process of simple filtration with reabsorption, but is complicated by metabolism and interconversion.

Total balance. Using the flow data from Table V, and appreciating that these are approximations in several cases, the net exchanges for carbohydrate, β -hydroxybutyrate, and acetoacetate, and other substrates can be extrapolated to a 24 hr period of time. It must be emphasized that these calculations do not take into account diurnal changes, if they exist, changes due to activity or other inaccuracies inherent in using arterio-venous differences, and a single flow determination, particularly in a sedated subject undergoing an experimental procedure.

Certain tissues, especially the cellular blood elements, derive their energy from glycolysis without terminal

after 5-6 wk of Starvation*

α -amino nitrogen	Glycerol	Free fatty acids	β -hydroxy butyrate	Acetoacetate	Glucose
3.61 \pm 0.19	0.148 \pm 0.018	1.86 \pm 0.22	6.48 \pm 0.65	1.42 \pm 0.23	4.03 \pm 0.36
0.14 \pm 0.045	0.092 \pm 0.016	0.346 \pm 0.039	-0.335 \pm 0.063	-0.180 \pm 0.059	-0.172 \pm 0.033
0.16 \pm 0.04	0.040 \pm 0.007	0.142 \pm 0.052	0.270 \pm 0.057	-0.047 \pm 0.009	-0.122 \pm 0.023

oxidation of glucose to carbon dioxide and water. The cyclic process of glucose breakdown in peripheral tissue with the formation of lactate and pyruvate which are transported to the liver and resynthesized into glucose has been quantitatively estimated in man from glucose-1-¹⁴C turnover studies (14, 35). These values for recycling metabolites vary between 27 and 58 g/day and do not appear to be related to the nutritional status of the subjects since one study was done in fasting and the other in postabsorptive subjects. A more direct estimation of anaerobic glycolysis was obtained in our study. The combined removal of lactate and pyruvate from arterial blood by the liver and kidney was estimated to be 430 mmoles/day. These precursors could theoretically form 215 mmoles or 39 g of glucose/day, in rough agreement with the ¹⁴C turnover studies previously reported (14, 35).

As discussed in a previous publication, a reduction in protein catabolism during prolonged starvation is mandatory for survival (36). Data in Fig. 3 and Table III corroborate this fact, showing the total mean urinary nitrogen excretion falling to 4.66 g/day. Urea excretion decreased the most, suggesting diminished hepatic gluconeogenesis from amino acids. In contrast, the need to titrate the urinary loss of β -hydroxybutyrate and acetoacetate with ammonium cations resulted in an increased ammonia excretion amounting to 42% of the total nitrogen. In vitro experiments using rat renal cortical tissue have coupled ammoniogenesis with gluconeogenesis (10, 11), and the substrate balance data across the kidney are further support for this relationship.

The amount of glucose derived from amino acids can be estimated from urinary nitrogen excretion or from α -amino nitrogen differences across the liver and kidney. 1 g of nitrogen is equal to 6.25 g of protein, and theoretically 4.66 g of nitrogen from catabolized protein should yield about 16 g of glucose (37). Approximately 26 g of glucose can be synthesized from the amino acids

taken up by these organs as approximated by the arterio-venous differences in our studies. These estimates of glucose production are in reasonable agreement, since some of the amino acids taken up by the liver are ketogenic rather than glucogenic, and complete conversion of amino acids to glucose appears improbable.

These obese subjects, fasted 5-6 wk, mobilize approximately 190 g of triglyceride/day, as calculated by indirect calorimetry¹ yielding 19 g of glycerol from adipose tissue which can serve as a glucose precursor. In the present study the liver and kidneys together removed approximately 213 mmoles or 19 g of glycerol from the blood per day, capable of producing 19 g of glucose, a surprising agreement. This is about one and one-half times the amount Borchgrevink and Havel reported for splanchnic and renal uptake of glycerol after an overnight fast (38).

The present study has demonstrated that there is a reduction in the total amount of glucose produced during prolonged starvation. The estimated hepatic-renal glucose production calculated from the limited flow data was 86 g/24 hr. The liver contributes approximately 55% and the kidney 45% of the total. The estimated glucose equivalent is 83 g/24 hr calculated from the measured hepatic-renal uptake of lactate, pyruvate, glycerol, and α -amino nitrogen from arterial blood. About one-half of the glucose formed, as discussed above, is derived from recycled lactate and pyruvate. The remaining glucose, derived from glycerol released from triglycerides and amino acids mobilized from proteins, can be terminally combusted to CO₂. The central nervous system, deriving the major proportion of its energy from β -hydroxybutyrate and acetoacetate (32 and 6 g/day, respectively) (8), continues to utilize the major part of this glucose, about 42 g/day (8). Recent studies by Ide, Steinke, and Cahill² have shown that rat brain has a

¹ A. P. Morgan. Manuscript in preparation.

² Ide, T., J. Steinke, and G. F. Cahill, Jr. 1968. The metabolic interactions of glucose, lactate, and β -hydroxybutyrate in rat brain. *Amer. J. Physiol.* Submitted for publication.

small but significant obligatory requirement for glucose for optimal metabolism of β -hydroxybutyrate; the human brain may behave accordingly.

The β -hydroxybutyrate and acetoacetate balances cannot be explained in light of present knowledge. Splanchnic production of β -hydroxybutyrate (54 g/day) is approximately equalled by renal extraction of this substrate (49 g/day). Yet, as shown previously (8), brain requires this fuel at the rate of approximately 32 g/day. Acetoacetate production by the splanchnic bed approximates 29 g and kidney adds to this another 8 g for a total of 37 g. β -hydroxybutyrate and acetoacetate are readily interconvertible and, if treated as a single entity, production of ketoacids is approximately equal to brain and kidney consumption plus urinary loss. But to make such a balance one must first consider that all the other tissues, particularly muscle, have virtually ceased to extract these two substrates with prolonged starvation, and preliminary studies on myocardial metabolism have suggested this to be true.³ Secondly, a conversion of acetoacetate to β -hydroxybutyrate by one or more tissues not directly studied in these experiments must be postulated, if the approximations are indeed valid. That this may be possible is suggested by the studies of Gammeltoft (39) who showed that *in situ* perfusion of starving cat liver and hind limb for 220 min gave β -hydroxybutyrate: acetoacetate ratios of 1.7 and 10.8, respectively. Furthermore, Hagenfeldt and Wahren (40) have reported net production of β -hydroxybutyrate and/or acetoacetate by exercising skeletal muscle.

General comments. We have specifically limited the discussion to the physiological observations and extrapolations derived therefrom without expanding upon mechanisms. All tissues metabolize fat or fat-derived products with but few exceptions. The glycolytic tissues which consume glucose and produce lactate derive their energy from fat metabolism in liver and kidney, the glucose-to-lactate-to-glucose cycle serving as an energy shuttle. Protein is catabolized in the minimum amount needed to provide ammonia for excretion with urinary organic acids. The deaminated and deamidated residues are converted into glucose, and this glucose plus that amount derived from glycerol is consumed by brain as its minimal obligatory amount, the other and major proportion of brain's fuel being β -hydroxybutyrate and acetoacetate, which are, again, fat-derived products.

These quantitative observations and approximations provide little direct insight, however, into the control mechanisms, but may serve, instead, as baselines for further studies into obesity, diabetes, hormonal effects, and protein catabolic states (such as trauma or sepsis), to name a few. The fact that gluconeogenesis by liver can be so markedly attenuated, and the fact that brain

³ Owen, O. E., and J. Wahren. Manuscript in preparation.

can adapt to ketoacid utilization both require a reevaluation of substrate requirements and enzyme patterns of specific cell types, particularly in man. Lastly, man and his hypertrophied central nervous system may have necessitated these unique metabolic adaptations, since to date, similar adaptations in experimental animals have not been found.

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REFERENCES

1. Krebs, H. A. 1964. Gluconeogenesis. *Proc. Roy. Soc. Ser. B.* **159**: 545.
2. Krebs, H. A. 1963. Renal gluconeogenesis. In *Advances in Enzyme Regulation*. G. Weber, editor. Pergamon Press Inc., New York and Oxford. **1**: 385.
3. Bondy, P. K., D. F. James, and B. W. Farrar. 1949. Studies of the role of the liver in human carbohydrate metabolism by the venous catheter technic. I. Normal subjects under fasting conditions and following the injection of glucose. *J. Clin. Invest.* **28**: 238.
4. Myers, J. D. 1950. Net splanchnic glucose production in normal man and in various disease states. *J. Clin. Invest.* **29**: 1421.
5. Bearn, A. G., B. H. Billing, and S. Sherlock. 1951. Hepatic glucose output and hepatic insulin sensitivity in diabetes mellitus. *Lancet.* **2**: 698.
6. Werk, E. E., Jr., H. T. McPherson, L. W. Hamrick, Jr., J. D. Myers, and F. L. Engel. 1955. Studies on ketone metabolism in man. I. A method for the quantitative estimation of splanchnic ketone production. *J. Clin. Invest.* **34**: 1256.
7. Kibler, R. F., W. J. Taylor, and J. D. Myers. 1964. The effect of glucagon on net splanchnic balances of glucose, amino acid nitrogen, urea, ketones, and oxygen in man. *J. Clin. Invest.* **43**: 904.
8. Owen, O. E., A. P. Morgan, H. G. Kemp, J. M. Sullivan, M. G. Herrera, and G. F. Cahill, Jr. 1967. Brain metabolism during fasting. *J. Clin. Invest.* **46**: 1589.
9. Drenick, E. J., M. E. Swendseid, W. H. Blahd, and S. G. Tuttle. 1964. Prolonged starvation as treatment of severe obesity. *J. Amer. Med. Assoc.* **187**: 100.
10. Goodman, A. D., R. E. Fuisz, and G. F. Cahill, Jr. 1966. Renal gluconeogenesis in acidosis, alkalosis, and potassium deficiency: its possible role in the regulation of renal ammonia production. *J. Clin. Invest.* **45**: 612.
11. Goorno, W. E., F. C. Rector, Jr., and D. W. Seldin. 1967. Relation of renal gluconeogenesis to ammonia production in the dog and rat. *Amer. J. Physiol.* **213**: 969.

12. Owen, O. E., J. Wahren, P. Felig, and G. F. Cahill, Jr. 1968. Changing patterns of gluconeogenesis during prolonged starvation. *Clin. Res.* **16**: 348.
13. Cahill, G. F., Jr., O. E. Owen, and A. P. Morgan. 1968. The consumption of fuels during prolonged starvation in *Advances in Enzyme Regulation*, G. Weber, editor, Pergamon Press, Inc., New York. **6**: 143-150.
14. Cahill, G. F., Jr., M. G. Herrera, A. P. Morgan, J. S. Soeldner, J. Steinke, P. L. Levy, G. A. Reichard, Jr., and D. M. Kipnis. 1966. Hormone-fuel interrelationships during fasting. *J. Clin. Invest.* **45**: 1751.
15. Boden, G., and J. S. Soeldner. 1967. A sensitive double antibody radioimmunoassay for human growth hormone (HGH): levels of serum HGH following rapid tolbutamide infusion. *Diabetologia.* **3**: 413.
16. Smith, H. W., N. Finkelstein, L. Aliminosa, B. Crawford, and M. Graber. 1945. The renal clearances of substituted hippuric acid derivatives and other aromatic acids in dog and man. *J. Clin. Invest.* **24**: 388.
17. Caesar, J., S. Shaldon, L. Chianducci, L. Guevara, and S. Sherlock. 1961. The use of indocyanine green in the measurement of hepatic blood flow and as a test of hepatic function. *Clin. Sci.* **21**: 43.
18. Chaplin, H., Jr., P. L. Mollison, and H. Vetter. 1953. The body/venous hematocrit ratio: its constancy over a wide hematocrit range. *J. Clin. Invest.* **32**: 1309.
19. Snedecor, G. W. 1956. *Statistical Methods*. Iowa State College Press, Ames. 5th edition. 35.
20. Felig, P., O. E. Owen, J. Wahren, and G. F. Cahill, Jr. 1969. Amino acid metabolism during prolonged starvation. *J. Clin. Invest.* **48**: 584.
21. Wade, O. L., and J. M. Bishop. 1962. *Cardiac Output and Regional Blood Flow*. F. A. Davis Company, Philadelphia. 86.
22. Karam, J. H., G. M. Grodsky, and P. H. Forsham. 1963. Excessive insulin response to glucose in obese subjects as measured by immunochemical assay. *Diabetes.* **12**: 197.
23. Sussman, K. E. 1966. Effect of prolonged fasting glucose and insulin metabolism in exogenous obesity. *Arch. Intern. Med.* **117**: 343.
24. Bagdade, J. D., E. L. Bierman, and D. Porte, Jr. 1967. The significance of basal insulin levels in the evaluation of the insulin response to glucose in diabetic and non-diabetic subjects. *J. Clin. Invest.* **46**: 1549.
25. Rabinowitz, D., and K. L. Zierler. 1962. Forearm metabolism in obesity and its response to intra-arterial insulin: characterization of insulin resistance and evidence for adaptive hyperinsulinism. *J. Clin. Invest.* **41**: 2173.
26. Hirsch, J., and E. Gallian. 1968. Methods for the determination of adipose cell size and cell number in man and animals. *J. Lipid Res.* **9**: 110.
27. Salans, L. B., J. L. Knittle, and J. Hirsch. 1968. The role of adipose cell enlargement in the carbohydrate intolerance of human obesity. *J. Clin. Invest.* **47**: 153.
28. Solomon, S. S., J. W. Ensink, and R. H. Williams. 1968. Effect of starvation on plasma immunoreactive insulin and non-suppressible insulin-like activity in normal and obese humans. *Metabolism.* **17**: 528.
29. Lea, M. A. and G. Weber. 1968. Role of enzymes in homeostasis. VIII. Inhibition of the activity of glycolytic enzymes by free fatty acids. *J. Biol. Chem.* **243**: 1096.
30. Weber, G., M. A. Lea, H. J. H. Convery, and N. B. Stamm. 1967. Regulation of gluconeogenesis and glycolysis: studies of mechanisms controlling enzyme activity. *In Advances in Enzyme Regulation*, Vol. 5, G. Weber, editor, Academic Press Inc., London and New York. 257.
31. Williamson, J. R., R. A. Kreisberg, and P. W. Felts. 1966. Mechanism for the stimulation of gluconeogenesis by fatty acids in perfused rat liver. *Proc. Nat. Acad. Sci. U.S.A.* **56**: 247.
32. Neith, H., and P. Schollmeyer. 1966. Substrate utilization of the human kidney. *Nature (London)*. **209**: 1244.
33. Leevy, C. M., W. George, W. Lesko, M. Deysine, C. C. Abbott, and E. J. Halligan. 1961. Observations on hepatic oxygen metabolism in man. *J. Amer. Med. Assoc.* **178**: 565.
34. Baker, N., A. S. Garfinkel, and M. G. Schotz. 1968. Hepatic triglyceride secretion in relation to lipogenesis and free fatty acid mobilization in fasted and glucose-refed rats. *J. Lipid Res.* **9**: 1.
35. Reichard, G. A., Jr., N. F. Moury, N. J. Hochella, A. L. Patterson, and S. Weinhouse. 1963. Quantitative estimation of the Cori cycle in the human. *J. Biol. Chem.* **238**: 495.
36. Cahill, G. F., Jr., and O. E. Owen. 1967. Starvation and survival. *Trans. Amer. Clin. Climat. Ass.* **79**: 13.
37. Krebs, H. A. 1964. The metabolic fate of amino acids in Mammalian Protein Metabolism. H. N. Munro and J. B. Allison, editors. Academic Press Inc., New York and London. **1**: 125.
38. Borchgrevink, C. F., and R. J. Havel. 1963. Transport of glycerol in human blood. *Proc. Soc. Exp. Biol. Med.* **113**: 946.
39. Gammeltoft, A. 1951. The ratio of β -hydroxybutyric acid to acetoacetic acid in blood under various experimental conditions. *Acta Physiol. Scand.* **24**: 35.
40. Hagenfeldt, L., and J. Wahren. 1968. Human forearm muscle metabolism during exercise III. Uptake, release and oxidation of β -hydroxybutyrate and observations on the β -hydroxybutyrate/acetoacetate ratio. *Scand. J. Clin. Lab. Invest.* **21**: 314.